

# Biodiversity Contribution in Beer Inputs

Subjects: Food Science & Technology | Agronomy | Biotechnology & Applied Microbiology  
Contributor: Vittorio Capozzi, Nicola De Simone, Giuseppe Spano

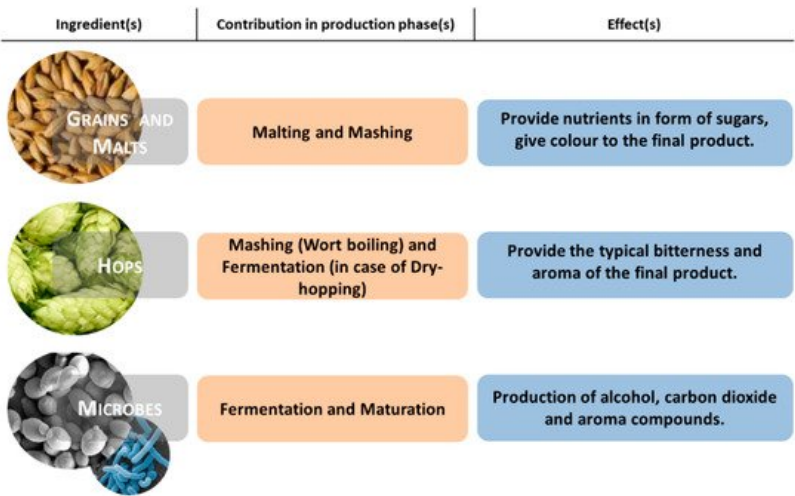
Selected biological resources used as raw materials in beer production are important drivers of innovation and segmentation in the dynamic market of craft beers. Among these resources, local/regional ingredients have several benefits, such as strengthening the connection with territories, enhancing the added value of the final products, and reducing supply costs and environmental impacts. It is assumed that specific ingredients provide differences in flavours, aromas, and, more generally, sensory attributes of the final products. In particular, of interest are ingredients with features attributable and/or linked to a specific geographical origin.

Keywords: craft beer ; ingredient ; autochthonous ; regionalisation ; brewing ; cereal ; hop ; yeasts ; bacteria ; biodiversity

## 1. Introduction

Beer is among the most appreciated and consumed beverages worldwide. The global beer market is constantly growing, supported by emerging segments (such as non-alcoholic, gluten-free, organic, and craft beers) <sup>[1]</sup>. In particular, craft beers are produced by small and independent microbreweries, which have rapidly risen worldwide due to increased interest in artisanal products <sup>[2]</sup>. Nevertheless, several factors, such as the competitive market, energy costs, scale, and taxation, appear to undermine the economic sustainability of microbreweries. For these reasons, the diversification and extension of the range of products are becoming increasingly important, leading to interest in finding new beer styles and improving the added value of marketed beer.

Considering the above, there is increased interest in the ingredients and raw materials involved in the beer production process, which could be simplified into four steps: malting, mashing, fermentation, and maturation. The principal ingredients are mainly water, barley malt, hops, and yeast. **Figure 1** shows the participation of the production phases of the main raw materials and their effects on the final product. In addition, the list of traditional ingredients can be extended according to the beer style and the breweries. In fact, it is assumed that specific raw materials could provide different flavours, aromas, and sensory attributes <sup>[3]</sup>. In this regard, the use of autochthonous biological inputs could provide several benefits, e.g., reducing the production and supply costs and creating a strong connection with territories that give added value to the final products.



**Figure 1.** Contribution of the main ingredients in the beer production process and their effects on the final product.

Water (including mineral water) is also used in the mashing stage, and is often crucial in connecting with a given site. Water chemical parameters influence several parameters, such as wort pH, yeast flocculation,  $\alpha$ -amylase activity, hop utilisation, colour, mouthfeel, and palatability of the final product <sup>[3][4][5][6]</sup>. Therefore, the water used at famous brewing

cities, e.g., in Dublin, Dortmund, Vienna, Munich, London, and Edinburgh, are utilized as references for the respective beer styles [7]. However, mineral composition, and the concentration of certain ions, such as carbonate, sulphate, iron, and manganese, are often adjusted to avoid unfavourable sensory attributes, colours, and tastes [8], and to comply with regional guidelines (such as in Germany) [9], or to suit the parameters given by regions/cities in order to obtain the same beer styles [4].

The brewing features of autochthonous biological resources, such as cereals, malts, hops, microbes, and adjuncts of biological origin, are differently influenced by specific soil features and environmental factors [9][10], demonstrating that regional attributes may also be promoted differently in the beer production chain. This association—known as *terroir*—was already described for other products related to specific regions, having particular interest in the wine industry, and could take advantage of the production of regional craft beers [11].

## 2. Ingredients

### 2.1. Grains and Malted Cereals

Cereals are the main ingredients for beer production; they provide nutritional sources for the fermentation progress, principally in the form of sugars. The main carbohydrate in the grain is starch, which is unfermentable and unusable by brewing yeasts. For this reason, prior to being used in the brewing process, cereals must be malted to promote the synthesis of the enzymes required for starch hydrolysis in fermentable sugars. After the germination, malts are dried and kilned. At this stage, different kilning procedures (time and temperature) allow obtaining different malt types, such as base malts (pale), speciality (caramel), and roasted (chocolate, coffee) [12]. Moreover, a wide range of malts, covering a plethora of beer styles, is available on the market.

Barley (*Hordeum vulgare* L.) and wheat (*Triticum durum* Desf. and *Triticum aestivum* L.) are the main cereals used in the malt industries. Among these, barley malt accounts for about 90% of worldwide beer production [12]. Whereas wheat malt is used in proportion with barley for the production of wheat (or sometimes ‘weiss’ or ‘weizen’) beers, characterised by distinctive cloudiness and persistent foam due to the major protein content of wheat malt [13]. Belgium and Northern Germany are historically known for the production of traditional beers brewed with raw (Belgian Witbier) and malted (German Weißbier) wheat, which are included in the list of classical beer styles, and are now produced and exported worldwide [13].

The use of autochthonous cultivars of cereals is one of the most explored strategies for the regionalisation of beer production. In this regard, cereals used in brewing vary among countries and as a function of the desired features of the final products (Table 1). Moreover, some of these cereals could be useful in producing beer with particular added value (e.g., gluten-free beer [14]), providing different nutritional and sensory properties [15], and reducing production costs [16]. Nevertheless, integrating the malting stage is considered a ‘bottleneck’ in the regionalisation of the beer industry [17], and autochthonous cereals are often used raw and unmalted as partial substitutes for barley malt.

**Table 1.** Cultivars, origins, and impact of grains that were recently investigated for their brewing potential.

Grains	Regions	Impact on Beer Quality	Ref.
Durum wheat (cv. Senatore Cappelli)	Sardinia (IT)	High polyphenol content, balanced taste, low sweetness.	[18]
Einkorn, emmer and spelt	Italy, Hungary	Higher antioxidant activity and polyphenol content, more fibre, lower gluten content, and low extract yields.	[19][20] [21]
Tritordeum (cv. Bulel)	Spain	Addition of slight acidity and higher free amino nitrogen.	[22]
Triticale (cv. Remiko)	Poland	Higher acidity, lower esters and isoamyl alcohol content.	[23]
Rice (cv. Loto)	Italy	Acceptable alcoholic content (3.5–4.5% vol.), good foam stability, rather poor in body and mouthfeel.	[24]
Rice (cv. Centauro)	Italy	Pale colour, not persistent foam, flat sensory characteristics. Optimisation of malt production improves colour and flavour.	[25][26] [27][28]
Oat (cv. Koneser)	Finland	Increased protein content in the wort and prolonged filtration time.	[15]
Oat (cv. Raisio)	Finland	High pH and low alcohol content, strong berry flavour and low amount of staling compounds during ageing.	[29]

Grains	Regions	Impact on Beer Quality	Ref.
Corn (cv. Nzaka-nzaka)	Congo	Poor foam stability, saccharification needs of exogenous $\alpha$ -amylase.	[30]
Pigmented Corn (cv. Chalqueño)	Mexico	Low-alcohol beer with polyphenols and anthocyanins with antioxidant properties.	[31][32]
Sorghum	African countries	Slight alcoholic (3.6%), brown colour, and acid pH (4.15 on average) due to the alcoholic and lactic co-fermentation. Incomplete saccharification, residue of insoluble materials, increased viscosity.	[33][34]
Red Sorghum (cv. DKS-74)	Mexico	Exogenous enzymes treatment yields glucose and alcohol content similar to barley-malt beer.	[34][35] [36][37]
Quinoa	Bolivia	Higher foam stability, lower level of soluble nitrogen, and more than twice the amount of fat; positive effect on the overall sensorial quality.	[38]
Rye (cv. Dukato)	Belgium	Increased beer viscosity, higher palate-fullness.	[39]
Teff (cv. Witkop)	South Africa	Higher content of glucose and a lower content of maltose, higher sweetness, fruity aroma, with little body.	[40]

The Mediterranean area is characterised by the presence of several cereal cultivars, in some cases only locally diffused, which could be exploited to improve segmentation in brewing regionalisation. Ancient wheat varieties have recently gained popularity due to their nutraceutical properties, from their higher concentrations of flavonoids, fibre, and minerals [41][42][43]. In Sardinia, Italy, a craft beer brewed with the old Italian wheat cultivar 'Senatore Cappelli' as an unmalted supplement (40%) was compared with two industrial wheat beers, resulting in higher polyphenol content and more balanced taste [18]. Other *Triticum* spp., such as einkorn (*Triticum monococcum* L.), emmer (*Triticum dicoccum* L.) and spelt (*Triticum spelta* L.), are used to obtain malts with higher antioxidant activities and total polyphenol content [20], characterised by the presence of more fibre, lower gluten content [19], and low extract yields [21]. New hybrids were recently assessed for malt and beer production. Among those, Tritordeum (*x Tritordeum martinii*), obtained by crossing wild barley with wheat, was comparable to barley in saccharification time, lautering, as well as colour and turbidity, but a slightly acidifying effect and higher free amino nitrogen were observed [22]. Whereas, Triticale (*x Triticosecale*), a hybrid of durum wheat and rye, is suitable for malt production because of its high extraction capacity, high diastatic power, and short saccharification time, but with higher acidity and lower esters and isoamyl alcohol content [23].

Oat (*Avena sativa* L.) is considered a functional cereal due to its pronounced level of antioxidants, fibre, and  $\beta$ -glucan. Compared to barley, oat is characterised by a higher husk content, which improves lautering performance, but leads to lower extract content, malted or unmalted [15][29]. Beer produced entirely from oat malts also has higher pH, lower alcohol content, and an intense berry flavour [29], while any significant difference can be found in the protein profile and in the fermentation trend [29]. The enhancement of extract and alcohol content by using exogenous enzymes, such as  $\beta$ -glucanases, amylases, neutral proteases, and hemicellulases, was reported on in the literature [15].

Rice (*Oryza sativa* L.), due to its high starch content, is sometimes used in brewing as a supplement to increase the sugar content of the mash [24][25]. Different trials were carried out in the production of rice beers [24][25][26][27][28]. Nevertheless, its protein and enzymatic profiles are lower than barley, and, furthermore, its starch does not entirely break up during the malting and mashing stage [44]. Despite the flat sensory profile and soft pale colour being reported in the first trials [24][25][26], optimisation of rice malt production resulted in an enhancement of flavour, taste, colour, and body of all rice beers, with encouraging results recently obtained [27][28].

Other grains, such as corn and sorghum, although they are considered as supplements for beer production, are commonly used to ferment traditional beers in South America and Africa, respectively, where they are the most cultivated cereals. Corn (*Zea mays*) is used to produce several indigenous South- and Mesoamerican beers, commonly known as Chicha [45]. Among these, *Chicha de Guinapo*, traditional of Arequipa (Peru), is based on malted-pigmented corn [46]. Recently, pigmented corn malt was proposed as the main ingredient in different beer-style productions [31][32]. Whereas sorghum-based African beers are characterised by a slight alcohol content (3.6% ethanol), brown colour, and acid pH (4.15 on average) due to the alcoholic and lactic co-fermentation [33]. However, some problems, such as incomplete saccharification, residue of insoluble materials, increased viscosity, and low free amino nitrogen content, occur in brewing with sorghum malt [34]. For these reasons, it is necessary to develop an appropriate process to improve malting and mashing conditions and the final sensory aspects. In this regard, the use of different formulations of exogenous enzymes has been evaluated. For example,  $\beta$ -amylase increases the amount of fermentable sugars [35], whereas amyloglucosidase

treatment improves wort yield, resulting in higher alcohol content <sup>[35][36]</sup>. Alternatively, the  $\alpha$ -amylase,  $\beta$ -amylase, and amyloglucosidase activities of sorghum malts could be enhanced by the addition of koji (*Aspergillus oryzae*) <sup>[37]</sup>.

## 2.2. Hops

Hops are the unfertilised female inflorescence of the perennial climbing vine *Humulus lupulus* L., belonging to the family Cannabaceae. The importance of hop is related to its bitterness, which contributes to the characteristic aroma and flavour of beer. Hops also enhance foam formation and stability and have antibacterial properties protecting against spoilage by certain microorganisms <sup>[47]</sup>. Their key compounds for brewing are resins and essential oils. Among the resin fraction, iso- $\alpha$ -acids, which originate when hops are added during wort boiling, are the most significant bittering compounds. Essential oils, mainly terpenoids, are extracted during late- and dry-hopping and are responsible for the beer aroma <sup>[48]</sup>. The perceived sensorial attributes depend on the hop varieties <sup>[49]</sup> and different commercial preparations (whole leaf, pellets, extracts) <sup>[3]</sup>. Brewers formerly divide hops into two groups based on their bittering or aroma contributions <sup>[50]</sup>. Among the most known cultivars, *Target*, *Admiral*, *Nugget*, *Pride of Ringwood*, and *Super Pride* release a high quantity of bittering compounds, while *Fuggles*, *Goldings*, *Saaz*, *Willamette*, *Cascade*, and *Cluster* varieties provide a pleasant hoppy aroma <sup>[3]</sup>. Hop cultivars have a strong connection with the geographical contest. In fact, they are also divided into groups, such as American, German, British, European, and others, according to their origin. For example, aroma hop varieties, such as *Hallertau* and *Hersbrucker*, are denominated according to a German region and city, respectively, and are among the most cultivated hops in Germany. *Saaz* is termed with the German name Žatec (*Saaz*), a city in the Czech Republic, and it is a traditional ingredient of Pilsner beer. Nevertheless, the United States and Germany are the most important producers, with about 50,000 tons for each, and Germany as the larger exporter, with 25,000 tons exported worldwide <sup>[51]</sup>. In this regard, there is growing interest in the definition of hop *terroir* and in presenting the differences among cultivars grown in different geographical contests. **Table 2** shows the difference in terms of quality among hops from different *terroir*. The dual-purpose American hop *Amarillo* grown in Idaho has lower citrusy and floral notes but a more fruity, spicy, and resinous odour than Washington <sup>[52]</sup>. In the same regions, the effect of *terroir* was later confirmed for hexyl glucoside content, a green leaf volatile with a grassy aroma, in twenty-three hop cultivars <sup>[53]</sup>. Similarly, *Cascade* from the Hallertau has more polyphenols and esters, but lower terpene content than those grown in Yakima (Washington, DC, USA) <sup>[54]</sup>. On the contrary, the same variety cultivated in Sardinia (Italy) had essential oils and acid content comparable to those farmed in the US <sup>[55]</sup>. In this way, recent research compared the volatile fingerprint and the acid profile of 15 commercial international hop cultivars, grown in an experimental field of Central Italy, with their standard characteristics, discovering desirable acids and terpene content in the cultivars *Chinook*, *Yeoman*, and *Hallertau* <sup>[56]</sup>.

**Table 2.** Cultivars, origins, and impact of hops recently investigated for their brewing potential.

Hops' Varieties	Region	Impact on Beer Quality	Ref.
Kazbek	Czech Republic	Low content of alpha acids, citrus-like aroma due to geranyl esters content of essential oil fraction.	[57]
Aramis	France	Terpenoid profile similar to the parental variety Strisselspalt, gives spicy and herbal notes to the beer.	[58]
Triskel	France	High concentration of monoterpenoids, especially linalool, which bring a floral note to beer.	[58]
Amarillo	Idaho (US)	Lower citrusy and floral notes, but higher fruity, spicy, and resinous odour descriptions.	[52]
Cascade	Hallertau (DE)	Higher content of polyphenols and esters, such as isobutyl-isobutyrate and 2-methylbutyl-2-methylpropanoate.	[54]
Cascade	Washington (US)	Higher linalool contents with respect to those grown in Hallertau (DE).	[54]
Cascade	Sardinia (IT)	Essential oil and the $\alpha$ / $\beta$ -acids in the same range of those cultivated in the US.	[55]
Cascade	Brazil	Higher content of farnesene and selinene, but lower levels of humulene and myrcene respect to the US grown crops.	[59]
Sorachi Ace	Hokkaido (JP)	It contains a unique volatile compound, geranic acid, which enhances the aroma contribution of terpenoids at sub-threshold levels.	[60]
Wild hops	Italy	Selinenes, $\alpha$ -acids, trans- $\beta$ -farnesene, and $\alpha$ -caryophyllene/ $\beta$ -humulene ratio are the main contributors and have a higher content of xanthohumol and $\alpha$ -acids among European wild hops.	[61]
Wild hops	Canada	High content of myrcene and low contents of humulene, farnesene, and selinenes.	[62]

Hops' Varieties	Region	Impact on Beer Quality	Ref.
Wild hops	Caucasus	Significantly lower cohumulone content.	[62]

The breeding of new varieties is another way to expand the plethora of hop cultivars, and to select those suitable for specific regions. In France, the variety *Strisselspalt* was used as a parental strain to obtain the new ones, *Bouclier* and *Triskel*, which showed significantly different terpenoid profiles [58]. While the variety *Kazbek* was bred in Czech Republic as the first aroma hops variety with a specific citrus-like aroma for Pilsner beer [57]. *Sorachi Ace*, the most known Japanese hop, has a significant amount of geranic acid, enhancing the varietal aroma of the hop-derived terpenoids [60].

Evaluating the brewing potential of wild hop varieties is one of the new tools used in the regionalisation of beer production. Mongelli et al. [61] characterised the aromatic profiles of 22 Italian wild hop genotypes. The low essential oils and bitter acidic content of some of these ecotypes suggest a potential exploitation, for a dual-purpose or for dry hopping hops. In addition, a comparison between wild hops collected from different regions of North America and the Caucasus showed significant differences in the contents of  $\alpha$ - and  $\beta$ -acids, cohumulone, and colupulone amount [62].

### 2.3. Microbes

Among microbes, yeast plays the most important role in beer production, as it ferments the wort, metabolises the sugar, and produces the compounds that define the peculiar characteristics of the beverage. It is also crucial for defining the flavour and aroma of the final products, including the synthesis of higher alcohols, esters, aldehydes, and organic acids [63], but also the bioconversion of hop-derived compounds [64][65][66]. For this reason, yeast starter cultures, in some cases, are protected and included in the list of autochthonous and/or traditional ingredients of regional beers (e.g., Münchener Bier) [67]. In brewing, yeasts are formerly divided into two major groups, based on beer-style, working temperatures, and flocculation ability. In the first group, there are those belonging to the species *Saccharomyces cerevisiae*, which are suitable for ale beers, capable of fermenting at warmer temperatures (16–22 °C) and able to flocculate or aggregate at the top of the vessel once fermentation is complete. In the second group, there are natural hybrids between *S. cerevisiae* and *S. bayanus* species, known as *S. pastorianus* (syn. *S. carlsbergensis*), suitable for lager-style; they ferment at lower temperatures (6–16 °C) and settle to the bottom of the vessel at the end of fermentation [68]. The two styles also need different periods of maturation: lager beers undergo a long, low-temperature period of ageing (known as lagering), while ale beers are usually mature in a short period [68]. The world beer production is represented by 90% for lager, and 5% for ale, while the other 5% is produced by spontaneous fermentation [68]. The latter has a particularly sour taste resulting from the sequential or contemporary fermentation by different microbes, among which, *Saccharomyces*, *Dekkera/Brettanomyces*, and lactic acid bacteria (LAB), such as *Lactobacillus* and *Pediococcus*, are considered the most important for the final beer character [69]. In the traditional Belgian lambic style, and its analogous American coolship ale, the wort is exposed to air during cooling and then transferred to wood barrels used in previous fermentations, resulting in a spontaneous inoculation by a consortium of microorganisms [69][70][71][72][73]. A microbial population of more than 2000 strains has been documented in lambic fermentation [69]. Nevertheless, the complete maturation of these kinds of beers can take many years, making it difficult to obtain a suitable quantity of products for commercial purposes [74].

The demand for novel starter cultures for brewing is increasing, and brewers and scientists are converging on the selection of those that could bring added value to the final products. Nowadays, the definition of microbial *terroir* has assumed relevant significance in wine production [75][76][77][78]; however, this approach has only recently gained popularity among breweries. In brewing, microbial *terroir* could be associated with the use of native microbes, isolated from traditional beer ingredients, but also with those strains isolated from other autochthonous biological resources. In fact, different research trends are focusing on improving the microbial biodiversity useful for beer production, including the exploration of the brewing potential of different groups of microorganisms, such as *Saccharomyces* strains isolated from other fermented food and beverage hybrids of the *Saccharomyces* genus and non-*Saccharomyces* species [79]. Thus, the features of relevant interest comprise sugar utilisation (mainly maltose and maltotriose), hops and ethanol tolerance, and ethanol and flavouring compound production (e.g., esters and higher alcohols) [80]. **Table 3** reassumes the main microbial strains recently investigated for their impact on beer quality.

**Table 3.** Species, strains, origin, and impact of microbes recently investigated for their brewing potential.

	Species/Strains	Source	Region	Impact on the Beer Quality	Ref.
Saccharomyces spp.	<i>S. cerevisiae</i>	Wine	Italy	Higher fruity and flowery aroma compounds in bottle re-fermentation.	[81]
	<i>S. cerevisiae</i> S-42	Sourdough	Sardinia (IT)	Similar sensorial profile, higher acidity, higher ethanol and esters content.	[82] [83] [84]
	<i>S. bayanus</i> × <i>S. cerevisiae</i>	De novo hybridisation	Italy	Efficient consumption of maltotriose, appreciable level of aroma compounds.	[85]
Non-Saccharomyces	<i>Hanseniaspora guilliermondii</i> IST315	Grape	Portugal	Increasing eight times the content of phenylethyl acetate, associated with rose and honey aroma.	[86]
	<i>Hanseniaspora vineae</i> T02/05	Grape	Uruguay	High ester production, fruity aroma suitable for low-alcohol beer production.	[63]
	<i>Kazachstania servazzii</i>	Rye malt Sourdough	Finland	Clean flavour profile and tolerance to low-temperature conditions.	[87]
	<i>Lachancea fermentati</i> KBI 12.1	Kombucha	Ireland	Lactic acid production, lower alcohol level, fruity aroma.	[88]
	<i>Lachancea thermotolerans</i> MN477031	Grape must	Slovakia	Low lactic acid production with a minor impact on pH of the beer.	[89]
	<i>Mrakia gelida</i> DBVPG 5952	Glacial melting water	Italy	Low alcohol production and low diacetyl, and appreciable organoleptic characteristics.	[90]
	<i>Pichia fermentans</i>	Sourdough	Finland	Production of the spice/clove aroma 4-vinylguaicol, suitable for low-alcohol wheat beers.	[87]
	<i>Saccharomycodes ludwigii</i> DBVPG 3010	Grape must	Italy	Production of low-alcohol beer, higher content of esters, and lower amount of diacetyl.	[91]
Lactic Acid Bacteria	<i>Torulaspora delbrueckii</i> DiSVA 254	Papaya leaves	Cameron	Increase of aromatic compounds, emphasised fruity/citric and fruity/esters notes.	[92] [93] [94]
	<i>L. brevis</i> BSO 464	Collection strain	Not reported	High flavour intensity, acidic taste, and astringency in co-fermentation.	[95] [96]
	<i>Lactobacillus amylovorus</i> FST2.11	Brewing environment	Ireland	High sensitive to hops, acidification of unhopped wort until 5–6 g/L of lactic acid.	[97]
	<i>Pediococcus acidilactici</i> K10	Kimchi	Korea	Starter for malt acidification; provides bioprotection against spoilage bacteria.	[98]
	<i>P. acidilactici</i> HW01	Malt	Korea	In malt acidification, improves microbiological stability, viscosity, and filtration time.	[99] [100]

The *Saccharomyces* species are responsible for the primary fermentation of a large variety of fermented food and beverages. However, the ability to metabolise maltose and maltotriose is not widespread, thus restricting the brewing potential only to a few species [80]. Rossi and co-workers [101] compared the fermentative ability in laboratory-scale fermentation and volatile profiles of different *S. cerevisiae* strains isolated from grape must, bakery, and wine and apple stillage. The authors selected a baking yeast as the most promising strain, leading to features in line with ale profiles and with a pronounced contribution in esters (above threshold). In this contest, sourdough could represent an important source of biodiversity when selecting autochthonous strains suitable for craft beer production. This idea is supported by the fact that some regional beers, such as Finnish Sahti beers, recognised in the European Union as ‘Traditional Speciality Guaranteed’, are traditionally fermented using baking yeast strains [102]. In this light, Ripari et al. [103][104] reported for the first time the utilisation of the whole microbiota of artisanal sourdough, comprised of yeasts and LAB, to obtain sour beers inspired by lambic fermentation. Other studies confirmed the potential of sourdough yeasts. For example, in Sardinia (Italy), two *S. cerevisiae* strains from artisanal sourdoughs were selected to produce wheat beers

from the Italian cultivars 'Senatore Cappelli' [82][83][84]. Among those, *S. cerevisiae* S-38 has shown physical–chemical, volatile, and sensory characteristics comparable to the commercial strain Safale S-33 (Fermentis, Lesaffre, Marcq-en-Baroeul, France) [83], whereas *S. cerevisiae* S-42 had higher ethanol content, lower pH, and a higher content of esters [82]. While Catallo et al. [85] used a sourdough strain as a parental strain in a de novo hybridisation to obtain a hybrid for lager beer fermentation. The authors reported that the hybrid inherited the ability to produce positive aroma compounds, such as 3-methylbutylacetate, ethyl acetate, and ethyl hexanoate by the sourdough strain, together with the efficient utilisation of maltotriose from the other parental strain [85].

Non-*Saccharomyces* yeasts have limited fermentation performance and are less tolerant to ethanol, but they produce volatile compounds, contributing to the sensory characteristics of several fermented beverages [105]. For this reason, different genera have recently been investigated for their contributions to brewing [106]. Among these, *Dekkera/Brettanomyces* strains are the most known as they are considered the main spoilers in beer and wine production, but if applied correctly, they contribute to the production of exotic flavours and the aroma complexity of speciality beers [107]. The enhancement of aromatic flavour in beer by different *Torulaspora delbrueckii* strains of oenological origins, to the production of isoamyl acetate, has been well documented, in both pure and mixed fermentations with commercial starters [92][93][94][108][109]. Meanwhile, the strains unable to ferment maltose *Saccharomycodes ludwigii* and *Zygosaccharomyces rouxii* have been studied to produce special beer styles, such as low-alcohol and alcohol-free beer [91]. Differently, the use of species able in producing lactic acid, such as *Lachancea thermotolerans* [89][110], *L. fermentati*, *Hanseniaspora vineae*, *Schizosaccharomyces japonicus*, and *Wickerhamomyces anomalus* [111] has been investigated for the production of sour beers in a single step of fermentation. Different methods for producing sour beers, such as kettle souring, sequenced, and mixed controlled fermentation, were recently explored [73][112]. Sour beers, such as the German Berliner Weisse and Gose, and the spontaneously fermented Belgian lambic and American Coolship Ale, have an intentional acidic taste given by the fermentation of acid-producing bacteria, mainly LAB, aside from brewing yeasts [112]. In addition, since LAB could be isolated from several food matrices, including brewing ingredients [97], they could contribute essentially to the regionalisation. In this regard, *L. amylovorus* FST2.11, isolated from the brewing environment, was tested with different acidification methods, suggesting kettle souring as the best practice to obtain sour beer with minimal organoleptic failures [97]. Contrarily, Dysvik et al. [95] showed that beer co-fermented with LAB strains, such as *L. plantarum* and *L. brevis* BSO464, had an increased intensity in fruity odour and higher total flavour intensity, respectively. These suggest that organoleptic attributes could be species- and/or strain-dependent, rather than method-dependent, since pre-acidification of the wort did not surpass that obtained by chemical acidification [113].

## 2.4. Adjuncts

The use of unusual ingredients is widespread among breweries, and although they are not essential for beer production—they are often used for the production of speciality beers. However, concerning the high number of commercially available products belonging to this category, only a few data are available in the literature. Among adjuncts, fruits and spices are often used to enrich the flavour complexity of certain beer styles. For example, orange peel and coriander are traditional ingredients of Belgian Witbier [114]. Fruits (as whole or as juices) are among the most studied supplements, and they are already present in many commercial products [115]. In Belgian lambic-style, for example, whole raspberries (*Rubus idaeus* L.) and tart cherries (*Prunus cerasus* L.) are traditionally used in the maturation of Framboise and Kriek beers, respectively [74]. Recently, the Beer Judge Certification Program recognised the Italian Grape Ale (IGA) as the first Italian style, which could be an expression of the regional biodiversity, promoted by the wide availability of grape cultivars [116]. In IGA, grapes can be added up to 40% during mashing, fermentation, and maturation. In addition, the use of several fruits, such as bananas or persimmon, has been reported in the literature. In both forms, fruit supplementation increases fermentable sugars [117][118], adds flavour and fruity aroma [117], modifies the colour due to the solubilisation of pigments, such as carotenoids [117] and anthocyanins [119][120], enhances antioxidant activity and total phenolics [115], increases bioactive compounds [115][121], adds a different degree of acidity [117][119], and increases alcohol content [118]. In addition, these supplementary ingredients could represent relevant links to specific regions (Table 4).

**Table 4.** Cultivars, origins, and impact of adjuncts recently investigated for their brewing potential.

	Adjuncts	Regions	Impact on Beer Quality	Ref.
Fruits	Banana (cv. Prata)	Brazil	Increasing of fermentable sugars and ethanol production.	[118]
	Persimmon (cv. Rojo Brillante)	Spain	Addition of malic and citric acid, light orange colour by solubilisation of carotenoids, increase of fermentable sugars.	[117]
	Hawthorn fruit (cv. Aurea)	Not reported	Increase of antioxidant activity, polyphenols, and volatile aroma compounds.	[122]
	Cornelian cherry (cv. Podolski)	Poland	Increase of polyphenols and antioxidant activity, addition of anthocyanins and sour taste.	[119] [120]
	Chestnut	Croatia	Slightly higher alcohol content, higher colour index.	[123]
	Cocoa (VP 1151)	Brazil	Increasing of wort viscosity, higher mineral, glucose and fructose content, higher ethanol production.	[124] [125]
	Soursop	Brazil	Lesser variation in beer standard parameters, good acceptance in sensorial attribute.	[126]
Vegetables	Carqueja ( <i>Baccharis trimera</i> )	Brazil	Addition of bittering compounds, total substitution of hop has shown no negative effects, sensorial acceptance similar to commercial beers.	[127]
	Artichoke	Brazil	Bittering effect, suitable for total hop substitution, good sensorial acceptance.	[128]
	Purple sweet potato (ST-13)	India	High content of anthocyanins and antioxidant compounds, peculiar pink colour.	[129]
	Birch-derived Xylooligosaccharides	Norway	Promoting secondary fermentation by LAB in sour beer.	[96]
	Olive leaves	Italy	Increase polyphenol content but not antioxidant activity, sour/astringent taste and herbal aroma at 10 g/L, pleasant sensory profile at 5 g/L.	[130]
	Eggplant (cv. Classic) peel extract	Romania	Increase of antioxidant activity, phenolics, and flavonoids content, reddish colour due to the release of anthocyanins.	[131]

Other ingredients can also be added to contribute to bitterness. In fact, before hops introduction, beers were initially flavoured with a mix of herbs and spices called gruit, which contained several herbs and spices [50]. Therefore, other bittering plants were investigated as hops substitutes, including where hop cultivation was not suitable. For instance, artichoke and carqueja were positively evaluated in Brazil as total substitutes because of their good sensorial acceptance and the absence of negative effects on physicochemical characteristics [127][128].

The regionality of the adjuncts is only poorly evaluated; however, intriguingly, attention is placed on the use of by-products from the food industry [96][132], with the incremental aim of reducing food waste and its economic weight.

## References

1. Allied Market Research. Available online: <https://www.alliedmarketresearch.com/beer-market> (accessed on 21 May 2021).
2. Aquilani, B.; Laureti, T.; Poponi, S.; Secondi, L. Beer Choice and Consumption Determinants When Craft Beers Are Tasted: An Exploratory Study of Consumer Preferences. *Food Qual. Prefer.* 2015, 41, 214–224.
3. Parker, D.K. 6-Beer: Production, sensory characteristics and sensory analysis. In *Alcoholic Beverages*; Piggott, J., Ed.; Woodhead Publishing Series in Food Science, Technology and Nutrition; Woodhead Publishing: Sawston, UK, 2012; pp. 133–158. ISBN 978-0-85709-051-5.
4. Briggs, D.E.; Brookes, P.A.; Stevens, R.; Boulton, C.A. *Brewing: Science and Practice*; Elsevier: Amsterdam, The Netherlands, 2004.
5. Omari, I.O.; Charnock, H.M.; Fugina, A.L.; Thomson, E.L.; McIndoe, J.S. Magnesium-Accelerated Maillard Reactions Drive Differences in Adjunct and All-Malt Brewing. *J. Am. Soc. Brew. Chem.* 2020, 79, 145–155.
6. Bastgen, N.; Becher, T.; Titze, J. Influencing Factors on Hop Isomerization Beyond the Conventional Range. *J. Am. Soc. Brew. Chem.* 2019, 77, 126–133.

7. Palmer, J.; Kaminski, C. *Water: A Comprehensive Guide for Brewers*; Brewers Publications: Boulder, CO, USA, 2013.
8. Wunderlich, S.; Back, W. 1-Overview of Manufacturing Beer: Ingredients, Processes, and Quality Criteria. In *Beer in Health and Disease Prevention*; Preedy, V.R., Ed.; Academic Press: San Diego, CA, USA, 2009; pp. 3–16. ISBN 978-0-12-373891-2.
9. Herb, D.; Filichkin, T.; Fisk, S.; Helgersen, L.; Hayes, P.; Benson, A.; Vega, V.; Carey, D.; Thiel, R.; Cistue, L. Malt Modification and Its Effects on the Contributions of Barley Genotype to Beer Flavor. *J. Am. Soc. Brew. Chem.* 2017, 75, 354–362.
10. Herb, D.; Filichkin, T.; Fisk, S.; Helgersen, L.; Hayes, P.; Meints, B.; Jennings, R.; Monsour, R.; Tynan, S.; Vinkemeier, K. Effects of Barley (*Hordeum vulgare* L.) Variety and Growing Environment on Beer Flavor. *J. Am. Soc. Brew. Chem.* 2017, 75, 345–353.
11. Berbegal, C.; Spano, G.; Tristezza, M.; Grieco, F.; Capozzi, V. Microbial Resources and Innovation in the Wine Production Sector. *South Afr. J. Enol. Vitic.* 2017, 38, 156–166.
12. Mallett, J. *Malt: A Practical Guide from Field to Brewhouse*; Brewers Publications: Boulder, CO, USA, 2014; Volume 4.
13. Hieronymus, S. *Brewing with Wheat: The 'wit' and 'weizen' of World Wheat Beer Styles*; Brewers Publications: Boulder, CO, USA, 2010.
14. Cela, N.; Condelli, N.; Caruso, M.C.; Perretti, G.; Di Cairano, M.; Tolve, R.; Galgano, F. Gluten-Free Brewing: Issues and Perspectives. *Fermentation* 2020, 6, 53.
15. Kordialik-Bogacka, E.; Bogdan, P.; Diowski, A. Malted and Unmalted Oats in Brewing. *J. Inst. Brew.* 2014, 120, 390–398.
16. Goode, D.L.; Arendt, E.K. 3-Developments in the supply of adjunct materials for brewing. In *Brewing*; Bamforth, C.W., Ed.; Woodhead Publishing Series in Food Science, Technology and Nutrition; Woodhead Publishing: Sawston, UK, 2006; pp. 30–67. ISBN 978-1-84569-003-8.
17. Maier, P.; Klein, O.; Schumacher, K.P. Ecological Benefits through Alternative Food Networks? Prospects of Regional Barley-Malt-Beer Value Chains in Bavaria, Germany. *J. Clean. Prod.* 2020, 265, 121848.
18. Mascia, I.; Fadda, C.; Dostálek, P.; Olšovská, J.; Caro, A.D. Preliminary Characterization of an Italian Craft Durum Wheat Beer. *J. Inst. Brew.* 2014, 120, 495–499.
19. Cooper, R. Re-Discovering Ancient Wheat Varieties as Functional Foods. *J. Tradit. Complement. Med.* 2015, 5, 138–143.
20. Fogarasi, A.-L.; Kun, S.; Tankó, G.; Stefanovits-Bányai, É.; Hegyesné-Vecseri, B. A Comparative Assessment of Antioxidant Properties, Total Phenolic Content of Einkorn, Wheat, Barley and Their Malts. *Food Chem.* 2015, 167, 1–6.
21. Mayer, H.; Marconi, O.; Perretti, G.; Sensidoni, M.; Fantozzi, P. Investigation of the Suitability of Hulled Wheats for Malting and Brewing. *J. Am. Soc. Brew. Chem.* 2011, 69, 116–120.
22. Zdaniewicz, M.; Pater, A.; Szczepanik, O.; Duliński, R.; Cioch-Skoneczny, M. Tritordeum Malt: An Innovative Raw Material for Beer Production. *J. Cereal Sci.* 2020, 96, 103095.
23. Cioch-Skoneczny, M.; Zdaniewicz, M.; Pater, A.; Skoneczny, S. Impact of Triticale Malt Application on Physiochemical Composition and Profile of Volatile Compounds in Beer. *Eur. Food Res. Technol.* 2019, 245, 1431–1437.
24. Ceppi, E.L.M.; Brenna, O.V. Brewing with Rice Malt—A Gluten-Free Alternative. *J. Inst. Brew.* 2010, 116, 275–279.
25. Mayer, H.; Marconi, O.; Regnicoli, G.F.; Perretti, G.; Fantozzi, P. Production of a Saccharifying Rice Malt for Brewing Using Different Rice Varieties and Malting Parameters. *J. Agric. Food Chem.* 2014, 62, 5369–5377.
26. Mayer, H.; Ceccaroni, D.; Marconi, O.; Sileoni, V.; Perretti, G.; Fantozzi, P. Development of an All Rice Malt Beer: A Gluten Free Alternative. *LWT-Food Sci. Technol.* 2016, 67, 67–73.
27. Ceccaroni, D.; Marconi, O.; Sileoni, V.; Wray, E.; Perretti, G. Rice Malting Optimization for the Production of Top-Fermented Gluten-Free Beer. *J. Sci. Food Agric.* 2019, 99, 2726–2734.
28. Ceccaroni, D.; Sileoni, V.; Marconi, O.; De Francesco, G.; Lee, E.G.; Perretti, G. Specialty Rice Malt Optimization and Improvement of Rice Malt Beer Aspect and Aroma. *LWT* 2019, 99, 299–305.
29. Klose, C.; Mauch, A.; Wunderlich, S.; Thiele, F.; Zarnkow, M.; Jacob, F.; Arendt, E.K. Brewing with 100% Oat Malt. *J. Inst. Brew.* 2011, 117, 411–421.
30. Diakabana, P.; Mvoulatsieri, M.; Dhellot, J.; Kobawila, S.; Louembé, D. Physico-Chemical Characterization of Brew during the Brewing Corn Malt in the Production of Maize Beer in Congo. *Adv. J. Food Sci. Technol.* 2013, 5, 671–677.
31. Flores-Calderón, A.M.D.; Luna, H.; Escalona-Buendía, H.B.; Verde-Calvo, J.R. Chemical Characterization and Antioxidant Capacity in Blue Corn (*Zea mays* L.) Malt Beers. *J. Inst. Brew.* 2017, 123, 506–518.

32. Romero-Medina, A.; Estarrón-Espinosa, M.; Verde-Calvo, J.R.; Lelièvre-Desmas, M.; Escalona-Buendía, H.B. Renewing Traditions: A Sensory and Chemical Characterisation of Mexican Pigmented Corn Beers. *Foods* 2020, 9, 886.
33. Lyumugabe, F.; Gros, J.; Nzungize, J.; Bajyana, E.; Thonart, P. Characteristics of African Traditional Beers Brewed with Sorghum Malt: A Review. *Biotechnol. Agron. Société Et Environ.* 2012, 16, 509–530.
34. Taylor, J.R.N.; Dlamini, B.C.; Kruger, J. 125th Anniversary Review: The Science of the Tropical Cereals Sorghum, Maize and Rice in Relation to Lager Beer Brewing. *J. Inst. Brew.* 2013, 119, 1–14.
35. Espinosa-Ramírez, J.; Pérez-Carrillo, E.; Serna-Saldívar, S.O. Production of Brewing Worts from Different Types of Sorghum Malts and Adjuncts Supplemented with  $\beta$ -Amylase or Amyloglucosidase. *J. Am. Soc. Brew. Chem.* 2013, 71, 49–56.
36. Espinosa-Ramírez, J.; Pérez-Carrillo, E.; Serna-Saldívar, S.O. Maltose and Glucose Utilization during Fermentation of Barley and Sorghum Lager Beers as Affected by  $\beta$ -Amylase or Amyloglucosidase Addition. *J. Cereal Sci.* 2014, 60, 602–609.
37. Heredia-Olea, E.; Cortés-Ceballos, E.; Serna-Saldívar, S.O. Malting Sorghum with *Aspergillus oryzae* Enhances Gluten-Free Wort Yield and Extract. *J. Am. Soc. Brew. Chem.* 2017, 75, 116–121.
38. Kordialik-Bogacka, E.; Bogdan, P.; Pielech-Przybylska, K.; Michałowska, D. Suitability of Unmalted Quinoa for Beer Production. *J. Sci. Food Agric.* 2018, 98, 5027–5036.
39. Langenaeken, N.A.; De Schutter, D.P.; Courtin, C.M. Arabinoxylan from Non-Malted Cereals Can Act as Mouthfeel Contributor in Beer. *Carbohydr. Polym.* 2020, 239, 116257.
40. Di Ghionno, L.; Sileoni, V.; Marconi, O.; De Francesco, G.; Perretti, G. Comparative Study on Quality Attributes of Gluten-Free Beer from Malted and Unmalted Teff [*Eragrostis Tef* (Zucc.) Trotter]. *LWT* 2017, 84, 746–752.
41. Leoncini, E.; Prata, C.; Malaguti, M.; Marotti, I.; Segura-Carretero, A.; Catizone, P.; Dinelli, G.; Hrelia, S. Phytochemical Profile and Nutraceutical Value of Old and Modern Common Wheat Cultivars. *PLoS ONE* 2012, 7, e45997.
42. Dinelli, G.; Segura-Carretero, A.; Di Silvestro, R.; Marotti, I.; Arráez-Román, D.; Benedettelli, S.; Ghiselli, L.; Fernandez-Gutierrez, A. Profiles of Phenolic Compounds in Modern and Old Common Wheat Varieties Determined by Liquid Chromatography Coupled with Time-of-Flight Mass Spectrometry. *J. Chromatogr. A* 2011, 1218, 7670–7681.
43. Migliorini, P.; Spagnolo, S.; Torri, L.; Arnoulet, M.; Lazzerini, G.; Ceccarelli, S. Agronomic and Quality Characteristics of Old, Modern and Mixture Wheat Varieties and Landraces for Organic Bread Chain in Diverse Environments of Northern Italy. *Eur. J. Agron.* 2016, 79, 131–141.
44. Yu, W.; Quek, W.P.; Li, C.; Gilbert, R.G.; Fox, G.P. Effects of the Starch Molecular Structures in Barley Malts and Rice Adjuncts on Brewing Performance. *Fermentation* 2018, 4, 103.
45. Hayashida, F.M. Ancient Beer and Modern Brewers: Ethnoarchaeological Observations of Chicha Production in Two Regions of the North Coast of Peru. *J. Anthropol. Archaeol.* 2008, 27, 161–174.
46. Vargas-Yana, D.; Aguilar-Morón, B.; Pezo-Torres, N.; Shetty, K.; Ranilla, L.G. Ancestral Peruvian Ethnic Fermented Beverage “Chicha” Based on Purple Corn (*Zea mays* L.): Unraveling the Health-Relevant Functional Benefits. *J. Ethn. Foods* 2020, 7, 1–12.
47. Sakamoto, K.; Konings, W.N. Beer Spoilage Bacteria and Hop Resistance. *Int. J. Food Microbiol.* 2003, 89, 105–124.
48. Van Opstaele, F.; De Rouck, G.; De Clippeleer, J.; Aerts, G.; De Cooman, L. Analytical and Sensory Assessment of Hoppy Aroma and Bitterness of Conventionally Hopped and Advanced Hopped Pilsner Beers. *J. Inst. Brew.* 2010, 116, 445–458.
49. Česlová, L.; Holčápek, M.; Fidler, M.; Dršticzková, J.; Lísa, M. Characterization of Prenylflavonoids and Hop Bitter Acids in Various Classes of Czech Beers and Hop Extracts Using High-Performance Liquid Chromatography–Mass Spectrometry. *J. Chromatogr. A* 2009, 1216, 7249–7257.
50. Hornsey, I.S. *A History of Beer and Brewing*; Royal Society of Chemistry: London, UK, 2003; Volume 34.
51. FAO. FAOSTAT Statistical Database. Available online: <http://www.fao.org/faostat/en/#compare> (accessed on 1 June 2021).
52. Van Holle, A.; Van Landschoot, A.; Roldán-Ruiz, I.; Naudts, D.; De Keukeleire, D. The Brewing Value of Amarillo Hops (*Humulus lupulus* L.) Grown in Northwestern USA: A Preliminary Study of Terroir Significance. *J. Inst. Brew.* 2017, 123, 312–318.
53. Morcol, T.B.; Negrin, A.; Matthews, P.D.; Kennelly, E.J. Hop (*Humulus lupulus* L.) Terroir Has Large Effect on a Glycosylated Green Leaf Volatile but Not on Other Aroma Glycosides. *Food Chem.* 2020, 321, 126644.

54. Forster, A.; Gahr, A. A Comparison of the Analytical and Brewing Characteristics of Cascade and Comet Hop Varieties as Grown in Yakima (USA) and Hallertau (Germany). *Brew. Sci.* 2014, 67, 137–148.
55. Forteschi, M.; Porcu, M.C.; Fanari, M.; Zinellu, M.; Secchi, N.; Buiatti, S.; Passaghe, P.; Bertoli, S.; Pretti, L. Quality Assessment of Cascade Hop (*Humulus lupulus* L.) Grown in Sardinia. *Eur. Food Res. Technol.* 2019, 245, 863–871.
56. Mozzon, M.; Foligni, R.; Mannozi, C. Brewing Quality of Hop Varieties Cultivated in Central Italy Based on Multivolatile Fingerprinting and Bitter Acid Content. *Foods* 2020, 9, 541.
57. Krofta, K.; Patzak, J.; Sedlák, T.; Mikyška, A.; Štěřba, K.; Jurková, M. Kazbek—The First Czech Aroma “Flavor Hops” Variety: Characteristics and Utilization. *KVASNY PRUMYSL* 2019, 65, 72–83.
58. Steyer, D.; Clayeux, C.; Laugel, B. Characterization of the Terpenoids Composition of Beers Made with the French Hop Varieties: Strisselspalt, Aramis, Triskel and Bouclier. *Brew. Sci.* 2013, 66, 192–197.
59. da Rosa Almeida, A.; Maciel, M.V.D.O.B.; Cardoso Gasparini Gandolpho, B.; Machado, M.H.; Teixeira, G.L.; Bertoldi, F.C.; Noronha, C.M.; Vitali, L.; Block, J.M.; Barreto, P.L.M. Brazilian Grown Cascade Hop (*Humulus lupulus* L.): LC-ESI-MS-MS and GC-MS Analysis of Chemical Composition and Antioxidant Activity of Extracts and Essential Oils. *J. Am. Soc. Brew. Chem.* 2020, 79, 156–166.
60. Sanekata, A.; Tanigawa, A.; Takoi, K.; Nakayama, Y.; Tsuchiya, Y. Identification and Characterization of Geranic Acid as a Unique Flavor Compound of Hops (*Humulus lupulus* L.) Variety Sorachi Ace. *J. Agric. Food Chem.* 2018, 66, 12285–12295.
61. Mongelli, A.; Rodolfi, M.; Ganino, T.; Marieschi, M.; Dall’Asta, C.; Bruni, R. Italian Hop Germplasm: Characterization of Wild *Humulus lupulus* L. Genotypes from Northern Italy by Means of Phytochemical, Morphological Traits and Multivariate Data Analysis. *Ind. Crop. Prod.* 2015, 70, 16–27.
62. Patzak, J.P.; Nesvadba, V.N.; Krofta, K.K.; Henychova, A.H.; Marzoev, A.I.M.I.; Richards, K.R. Evaluation of Genetic Variability of Wild Hops (*Humulus lupulus* L.) in Canada and the Caucasus Region by Chemical and Molecular Methods. *Genome* 2010, 53, 545–557.
63. Larroque, M.N.; Carrau, F.; Fariña, L.; Boido, E.; Dellacassa, E.; Medina, K. Effect of *Saccharomyces* and Non-*Saccharomyces* Native Yeasts on Beer Aroma Compounds. *Int. J. Food Microbiol.* 2021, 337, 108953.
64. Takoi, K.; Koie, K.; Itoga, Y.; Katayama, Y.; Shimase, M.; Nakayama, Y.; Watari, J. Biotransformation of Hop-Derived Monoterpene Alcohols by Lager Yeast and Their Contribution to the Flavor of Hopped Beer. *J. Agric. Food Chem.* 2010, 58, 5050–5058.
65. Takoi, K.; Itoga, Y.; Koie, K.; Takayanagi, J.; Kaneko, T.; Watanabe, T.; Matsumoto, I.; Nomura, M. Systematic Analysis of Behaviour of Hop-Derived Monoterpene Alcohols during Fermentation and New Classification of Geraniol-Rich Flavour Hops. *BrewingScience* 2017, 70, 177–186.
66. Ohashi, Y.; Huang, S.; Maeda, I. Biosyntheses of Geranic Acid and Citronellic Acid from Monoterpene Alcohols by *Saccharomyces Cerevisiae*. *Biosci. Biotechnol. Biochem.* 2021, 85, 1530–1535.
67. Capozzi, V.; Russo, P.; Spano, G. Microbial Information Regimen in EU Geographical Indications. *World Pat. Inf.* 2012, 34, 229–231.
68. Petruzzi, L.; Corbo, M.R.; Sinigaglia, M.; Bevilacqua, A. Brewer’s Yeast in Controlled and Uncontrolled Fermentations, with a Focus on Novel, Nonconventional, and Superior Strains. *Food Rev. Int.* 2016, 32, 341–363.
69. Spitaels, F.; Wieme, A.D.; Janssens, M.; Aerts, M.; Daniel, H.-M.; Van Landschoot, A.; De Vuyst, L.; Vandamme, P. The Microbial Diversity of Traditional Spontaneously Fermented Lambic Beer. *PLoS ONE* 2014, 9, e95384.
70. Shayevitz, A.; Harrison, K.; Curtin, C.D. Barrel-Induced Variation in the Microbiome and Mycobiome of Aged Sour Ale and Imperial Porter Beer. *J. Am. Soc. Brew. Chem.* 2021, 79, 33–40.
71. Bokulich, N.A.; Bamforth, C.W.; Mills, D.A. Brewhouse-Resident Microbiota Are Responsible for Multi-Stage Fermentation of American Coolship Ale. *PLoS ONE* 2012, 7, e35507.
72. Roos, J.D.; Vuyst, L.D. Microbial Acidification, Alcoholization, and Aroma Production during Spontaneous Lambic Beer Production. *J. Sci. Food Agric.* 2019, 99, 25–38.
73. Dysvik, A.; Rosa, S.L.L.; Rouck, G.D.; Rukke, E.-O.; Westereng, B.; Wicklund, T. Microbial Dynamics in Traditional and Modern Sour Beer Production. *Appl. Environ. Microbiol.* 2020, 86.
74. De Keersmaecker, J. The Mystery of Lambic Beer. *Sci. Am.* 1996, 275, 74–80.
75. Grieco, F.; Tristezza, M.; Vetrano, C.; Bleve, G.; Panico, E.; Mita, G.; Logrieco, A. Exploitation of Autochthonous Micro-Organism Potential to Enhance the Quality of Apulian Wines. *Ann. Microbiol.* 2011, 61, 67–73.
76. Tristezza, M.; Fantastico, L.; Vetrano, C.; Bleve, G.; Corallo, D.; Grieco, F.; Mita, G.; Grieco, F. Molecular and Technological Characterization of *Saccharomyces cerevisiae* Strains Isolated from Natural Fermentation of

77. Tufariello, M.; Chiriatti, M.A.; Grieco, F.; Perrotta, C.; Capone, S.; Rampino, P.; Tristezza, M.; Mita, G. Influence of Autochthonous *Saccharomyces cerevisiae* Strains on Volatile Profile of Negroamaro Wines. LWT-Food Sci. Technol. 2014, 58, 35–48.
78. Capozzi, V.; Garofalo, C.; Chiriatti, M.A.; Grieco, F.; Spano, G. Microbial Terroir and Food Innovation: The Case of Yeast Biodiversity in Wine. Microbiol. Res. 2015, 181, 75–83.
79. Iattici, F.; Catallo, M.; Solieri, L. Designing New Yeasts for Craft Brewing: When Natural Biodiversity Meets Biotechnology. Beverages 2020, 6, 3.
80. Cubillos, F.A.; Gibson, B.; Grijalva-Vallejos, N.; Krogerus, K.; Nikulin, J. Bioprospecting for Brewers: Exploiting Natural Diversity for Naturally Diverse Beers. Yeast 2019, 36, 383–398.
81. Canonico, L.; Comitini, F.; Ciani, M. Dominance and Influence of Selected *Saccharomyces cerevisiae* Strains on the Analytical Profile of Craft Beer Refermentation. J. Inst. Brew. 2014, 120, 262–267.
82. Mascia, I.; Fadda, C.; Dostálek, P.; Karabín, M.; Zara, G.; Budroni, M.; Caro, A.D. Is It Possible to Create an Innovative Craft Durum Wheat Beer with Sourdough Yeasts? A Case Study. J. Inst. Brew. 2015, 121, 283–286.
83. Marongiu, A.; Zara, G.; Legras, J.-L.; Del Caro, A.; Mascia, I.; Fadda, C.; Budroni, M. Novel Starters for Old Processes: Use of *Saccharomyces cerevisiae* Strains Isolated from Artisanal Sourdough for Craft Beer Production at a Brewery Scale. J. Ind. Microbiol. Biotechnol. 2015, 42, 85–92.
84. Mascia, I.; Fadda, C.; Karabín, M.; Dostálek, P.; Del Caro, A. Aging of Craft Durum Wheat Beer Fermented with Sourdough Yeasts. LWT-Food Sci. Technol. 2016, 65, 487–494.
85. Catallo, M.; Iattici, F.; Randazzo, C.; Caggia, C.; Krogerus, K.; Magalhães, F.; Gibson, B.; Solieri, L. Hybridization of *Saccharomyces Cerevisiae* Sourdough Strains with Cryotolerant *Saccharomyces bayanus* NBRC1948 as a Strategy to Increase Diversity of Strains Available for Lager Beer Fermentation. Microorganisms 2020, 9, 514.
86. Bourbon-Melo, N.; Palma, M.; Rocha, M.P.; Ferreira, A.; Bronze, M.R.; Elias, H.; Sá-Correia, I. Use of *Hanseniaspora Guilliermondii* and *Hanseniaspora opuntiae* to Enhance the Aromatic Profile of Beer in Mixed-Culture Fermentation with *Saccharomyces Cerevisiae*. Food Microbiol. 2021, 95, 103678.
87. Johansson, L.; Nikulin, J.; Juvonen, R.; Krogerus, K.; Magalhães, F.; Mikkelsen, A.; Nuppenen-Puputti, M.; Sohlberg, E.; de Francesco, G.; Perretti, G.; et al. Sourdough Cultures as Reservoirs of Maltose-Negative Yeasts for Low-Alcohol Beer Brewing. Food Microbiol. 2021, 94, 103629.
88. Bellut, K.; Michel, M.; Hutzler, M.; Zarnkow, M.; Jacob, F.; Schutter, D.P.D.; Daenen, L.; Lynch, K.M.; Zannini, E.; Arendt, E.K. Investigation into the Potential of *Lachancea fermentati* Strain KBI 12.1 for Low Alcohol Beer Brewing. J. Am. Soc. Brew. Chem. 2019, 77, 157–169.
89. Zdaniewicz, M.; Satora, P.; Pater, A.; Bogacz, S. Low Lactic Acid-Producing Strain of *Lachancea thermotolerans* as a New Starter for Beer Production. Biomolecules 2020, 10, 256.
90. De Francesco, G.; Sannino, C.; Sileoni, V.; Marconi, O.; Filippucci, S.; Tasselli, G.; Turchetti, B. *Mrakia gelida* in Brewing Process: An Innovative Production of Low Alcohol Beer Using a Psychrophilic Yeast Strain. Food Microbiol. 2018, 76, 354–362.
91. De Francesco, G.D.; Turchetti, B.; Sileoni, V.; Marconi, O.; Perretti, G. Screening of New Strains of *Saccharomycodes judwigii* and *Zygosaccharomyces Rouxii* to Produce Low-alcohol Beer. J. Inst. Brew. 2015, 121, 113–121.
92. Canonico, L.; Comitini, F.; Ciani, M. *Torulaspora delbrueckii* Contribution in Mixed Brewing Fermentations with Different *Saccharomyces Cerevisiae* Strains. Int. J. Food Microbiol. 2017, 259, 7–13.
93. Canonico, L.; Agarbati, A.; Comitini, F.; Ciani, M. *Torulaspora Delbrueckii* in the Brewing Process: A New Approach to Enhance Bioflavour and to Reduce Ethanol Content. Food Microbiol. 2016, 56, 45–51.
94. Canonico, L.; Ciani, E.; Galli, E.; Comitini, F.; Ciani, M. Evolution of Aromatic Profile of *Torulaspora delbrueckii* Mixed Fermentation at Microbrewery Plant. Fermentation 2020, 6, 7.
95. Dysvik, A.; La Rosa, S.L.; Liland, K.H.; Myhrer, K.S.; Østlie, H.M.; De Rouck, G.; Rukke, E.-O.; Westereng, B.; Wicklund, T. Co-Fermentation Involving *Saccharomyces cerevisiae* and *Lactobacillus* Species Tolerant to Brewing-Related Stress Factors for Controlled and Rapid Production of Sour Beer. Front. Microbiol. 2020, 11, 279.
96. Dysvik, A.; La Rosa, S.L.; Buffetto, F.; Liland, K.H.; Myhrer, K.S.; Rukke, E.-O.; Wicklund, T.; Westereng, B. Secondary Lactic Acid Bacteria Fermentation with Wood-Derived Xylooligosaccharides as a Tool To Expedite Sour Beer Production. J. Agric. Food Chem. 2020, 68, 301–314.
97. Peyer, L.C.; Zarnkow, M.; Jacob, F.; Schutter, D.P.D.; Arendt, E.K. Sour Brewing: Impact of *Lactobacillus amylovorus* FST2.11 on Technological and Quality Attributes of Acid Beers. J. Am. Soc. Brew. Chem. 2017, 75, 207–216.

98. Kim, M.; Choi, E.; Kim, J.; Ahn, H.; Han, H.; Kim, W.J. Effect of Bacteriocin-Producing *Pediococcus acidilactici* K10 on Beer Fermentation. *J. Inst. Brew.* 2016, 122, 422–429.
99. Kim, D.-Y.; Kim, J.; Kim, J.H.; Kim, W.J. Malt and Wort Bio-Acidification by *Pediococcus acidilactici* HW01 as Starter Culture. *Food Control* 2021, 120, 107560.
100. Ahn, H.; Kim, J.; Kim, W.J. Isolation and Characterization of Bacteriocin-Producing *Pediococcus acidilactici* HW01 from Malt and Its Potential to Control Beer Spoilage Lactic Acid Bacteria. *Food Control* 2017, 80, 59–66.
101. Rossi, S.; Turchetti, B.; Sileoni, V.; Marconi, O.; Perretti, G. Evaluation of *Saccharomyces cerevisiae* Strains Isolated from Non-Brewing Environments in Beer Production. *J. Inst. Brew.* 2018, 124, 381–388.
102. Catallo, M.; Nikulin, J.; Johansson, L.; Krogerus, K.; Laitinen, M.; Magalhães, F.; Piironen, M.; Mikkelsen, A.; Randazzo, C.L.; Solieri, L. Sourdough Derived Strains of *Saccharomyces cerevisiae* and Their Potential for Farmhouse Ale Brewing. *J. Inst. Brew.* 2020, 126, 168–175.
103. Ripari, V.; Tomassetti, M.; Cecchi, T.; Enrico, B. Recipe, Volatiles Profile, Sensory Analysis, Physico-Chemical and Microbial Characterization of Acidic Beers from Both Sourdough Yeasts and Lactic Acid Bacteria. *Eur. Food Res. Technol.* 2018, 244, 2027–2040.
104. Ripari, V.; Tomassetti, M.; Cecchi, T.; Berardi, E. First Study of Sourdough Beer Aging Via the Chemical Fingerprint of Volatile Markers. *Food Anal. Methods* 2019, 12, 2459–2468.
105. Ciani, M.; Comitini, F.; Mannazzu, I.; Domizio, P. Controlled Mixed Culture Fermentation: A New Perspective on the Use of Non-*Saccharomyces* Yeasts in Winemaking. *FEMS Yeast Res.* 2010, 10, 123–133.
106. Capece, A.; Romaniello, R.; Siesto, G.; Romano, P. Conventional and Non-Conventional Yeasts in Beer Production. *Fermentation* 2018, 4, 38.
107. Serra Colomer, M.; Funch, B.; Forster, J. The Raise of *Brettanomyces* Yeast Species for Beer Production. *Curr. Opin. Biotechnol.* 2019, 56, 30–35.
108. Holt, S.; Mukherjee, V.; Lievens, B.; Verstrepen, K.J.; Thevelein, J.M. Bioflavoring by Non-Conventional Yeasts in Sequential Beer Fermentations. *Food Microbiol.* 2018, 72, 55–66.
109. Tataridis, P.; Kanelis, A.; Logotetis, S.; Nerancis, E. Use of Non-*Saccharomyces Torulaspora delbrueckii* Yeast Strains in Winemaking and Brewing. *Zb. Matice Srp. Za Prir. Nauk.* 2013, 124, 415–426.
110. Domizio, P.; House, J.F.; Joseph, C.M.L.; Bisson, L.F.; Bamforth, C.W. *Lachancea thermotolerans* as an Alternative Yeast for the Production of Beer. *J. Inst. Brew.* 2016, 122, 599–604.
111. Osburn, K.; Amaral, J.; Metcalf, S.R.; Nickens, D.M.; Rogers, C.M.; Sausen, C.; Caputo, R.; Miller, J.; Li, H.; Tennesen, J.M.; et al. Primary Souring: A Novel Bacteria-Free Method for Sour Beer Production. *Food Microbiol.* 2018, 70, 76–84.
112. Bossaert, S.; Crauwels, S.; De Rouck, G.; Lievens, B. The Power of Sour—a Review: Old Traditions, New Opportunities. *BrewingScience* 2019, 72, 78–88.
113. Dysvik, A.; Liland, K.H.; Myhrer, K.S.; Westereng, B.; Rukke, E.-O.; De Rouck, G.; Wicklund, T. Pre-Fermentation with Lactic Acid Bacteria in Sour Beer Production. *J. Inst. Brew.* 2019, 125, 342–356.
114. Strong, G.; England, K. Beer Judge Certification Program: 2015 Style Guidelines. *Brew. Assoc.* 2015, 47. Available online: [https://www.bjcp.org/docs/2015\\_Guidelines\\_Beer.pdf](https://www.bjcp.org/docs/2015_Guidelines_Beer.pdf) (accessed on 1 June 2021).
115. Nardini, M.; Garaguso, I. Characterization of Bioactive Compounds and Antioxidant Activity of Fruit Beers. *Food Chem.* 2020, 305, 125437.
116. Garavaglia, C. The Emergence of Italian Craft Breweries and the Development of Their Local Identity. In *The Geography of Beer: Culture and Economics*; Hoalst-Pullen, N., Patterson, M.W., Eds.; Springer International Publishing: Cham, Switzerland, 2020; pp. 135–147. ISBN 978-3-030-41654-6.
117. Martínez, A.; Vegara, S.; Martí, N.; Valero, M.; Saura, D. Physicochemical Characterization of Special Persimmon Fruit Beers Using Bohemian Pilsner Malt as a Base. *J. Inst. Brew.* 2017, 123, 319–327.
118. Carvalho, G.B.; Silva, D.P.; Bento, C.V.; Vicente, A.A.; Teixeira, J.A.; Maria das Graças, A.F.; e Silva, J.B.A. Banana as Adjunct in Beer Production: Applicability and Performance of Fermentative Parameters. *Appl. Biochem. Biotechnol.* 2009, 155, 53–62.
119. Kawa-Rygielska, J.; Adamenko, K.; Kucharska, A.Z.; Prorok, P.; Piórecki, N. Physicochemical and Antioxidative Properties of Cornelian Cherry Beer. *Food Chem.* 2019, 281, 147–153.
120. Adamenko, K.; Kawa-Rygielska, J.; Kucharska, A.Z. Characteristics of Cornelian Cherry Sour Non-Alcoholic Beers Brewed with the Special Yeast *Saccharomyces ludwigii*. *Food Chem.* 2020, 312, 125968.

121. Adadi, P.; Kovaleva, E.G.; Glukhareva, T.V.; Shatunova, S.A.; Petrov, A.S. Production and Analysis of Non-Traditional Beer Supplemented with Sea Buckthorn. *Agron. Res.* 2017, 15, 1831–1845.
122. Gasiński, A.; Kawa-Rygielska, J.; Szumny, A.; Gąsior, J.; Głowacki, A. Assessment of Volatiles and Polyphenol Content, Physicochemical Parameters and Antioxidant Activity in Beers with Dotted Hawthorn (*Crataegus punctata*). *Foods* 2020, 9, 775.
123. Velić, N.; Mujčić, I.; Krstanović, V.; Velić, D.; Franić, M.; Zec Sombol, S.; Mastanjević, K. Chestnut in Beer Production: Applicability and Effect on Beer Quality Parameters. *Acta Horti* 2018, 1220, 209–214.
124. Nunes, C.S.O.; da Silva, M.L.C.; Camilloto, G.P.; Machado, B.A.S.; Hodel, K.V.S.; Koblit, M.G.B.; Carvalho, G.B.M.; Uetanabaro, A.P.T. Potential Applicability of Cocoa Pulp (*Theobroma cacao* L.) as an Adjunct for Beer Production. *Sci. World J.* 2020, 2020, e3192585.
125. Nunes, C.D.S.O.; de Carvalho, G.B.M.; da Silva, M.L.C.; da Silva, G.P.; Machado, B.A.S.; Uetanabaro, A.P.T. Cocoa Pulp in Beer Production: Applicability and Fermentative Process Performance. *PLoS ONE* 2017, 12, e0175677.
126. Alves, M.D.M.; Rosa, M.D.S.; SANTOS, P.P.A.D.; PAZ, M.F.D.; Morato, P.N.; Fuzinato, M.M. Artisanal Beer Production and Evaluation Adding Rice Flakes and Soursop Pulp (*Annona muricata* L.). *Food Sci. Technol.* 2020, 40, 545–549.
127. Schuina, G.L.; Quelhas, J.O.F.; de Carvalho, G.B.M.; Bianchi, V.L.D. Use of Carqueja (*Baccharis Trimeria* (Less.) DC. Asteraceae) as a Total Substitute for Hops in the Production of Lager Beer. *J. Food Process. Preserv.* 2020, 44, e14730.
128. Schuina, G.L.; Quelhas, J.O.F.; CASTILHOS, M.B.M.D.; CARVALHO, G.B.M.D.; Del Bianchi, V.L. Alternative Production of Craft Lager Beers Using Artichoke (*Cynara scolymus* L.) as a Hops Substitute. *Food Sci. Technol.* 2020, 40, 157–161.
129. Panda, S.K.; Panda, S.H.; Swain, M.R.; Ray, R.C.; Kayitesi, E. Anthocyanin-Rich Sweet Potato (*Ipomoea batatas* L.) Beer: Technology, Biochemical and Sensory Evaluation. *J. Food Process. Preserv.* 2015, 39, 3040–3049.
130. Guglielmotti, M.; Passaghe, P.; Buiatti, S. Use of Olive (*Olea europaea* L.) Leaves as Beer Ingredient, and Their Influence on Beer Chemical Composition and Antioxidant Activity. *J. Food Sci.* 2020, 85, 2278–2285.
131. Horincar, G.; Enachi, E.; Bolea, C.; Râpeanu, G.; Aprodu, I. Value-Added Lager Beer Enriched with Eggplant (*Solanum melongena* L.) Peel Extract. *Molecules* 2020, 25, 731.
132. Pereira, I.M.C.; Matos, J.D.; Figueiredo, R.W.; Carvalho, J.D.G.; Figueiredo, E.A.T.D.; Menezes, N.V.S.D.; Gaban, S.V.F. Physicochemical Characterization, Antioxidant Activity, and Sensory Analysis of Beers Brewed with Cashew Peduncle (*Anacardium occidentale*) and Orange Peel (*Citrus Sinensis*). *Food Sci. Technol.* 2020, 40, 749–755.