

Glomerular Autophagy in Diabew

Subjects: **Pathology**

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Glomerular injury is a central feature of diabetic nephropathy. A growing body of evidence indicates a critical role of autophagy in maintaining podocyte integrity and renal function. This process is vital for highly differentiated post-mitotic cells, such as neurons and podocytes. Sodium-glucose cotransporter-2 (SGLT2) inhibitors and dipeptidylpeptidase-4 (DPP4) inhibitors are promising antidiabetic agents introduced into clinical practice in the last decade. The effect of SGLT2 and DPP4 inhibition on glomerular autophagy has not been studied yet. This entry summarizes the study, which have demonstrated, that the SGLT2 inhibitor empagliflozin, the DPP4 inhibitor linagliptin, and a combination of these agents, could reactivate glomerular autophagy in db/db mice, a model of type 2 diabetes. The effect is associated with mitigation of renal hypertrophy, an improvement in glomerular morphology, including a decrease in the severity of podocytopathy, and a slowdown in the growth of albuminuria.

autophagy

podocyte

type 2 diabetes

diabetic nephropathy

empagliflozin

linagliptin

1. Introduction

Diabetic nephropathy is the most common cause for end-stage renal disease worldwide^[1]. The pathogenesis of diabetic nephropathy is being studied extensively. A number of studies have identified podocyte injury as a cornerstone in pathogenesis of diabetic kidney disease^{[2][3]}. Podocytes are terminally differentiated cells that wrap the glomerular capillary loops from outside and form slit diaphragms by podocyte foot processes (FPs). Diabetes-induced podocyte injury results in several phenotypic changes, including effacement of FPs, hypertrophy, detachment, death, and/or dedifferentiation. In its turn, the loss or damage of podocytes causes dysfunction of the filtration barrier and increases albuminuria^[2].

Some recent studies have indicated an emerging role of autophagy downregulation in diabetic podocytopathy^{[4][5]}
^[6]. Autophagy is a cellular recycling process involving self-degradation and reconstruction of damaged organelles and proteins^[6]. The process is vital for highly differentiated post-mitotic cells, such as neurons and podocytes^[7]. A growing body of evidence indicates a critical role of autophagy in maintaining podocyte integrity and renal function^[6]. Mice with podocyte-specific deletion of autophagic regulators, such as class III PI3K vacuolar protein sorting 34 (Vps34) and the Atg5 gene, develop early proteinuria, progressive glomerulosclerosis, and renal failure^[6]. Accordingly, autophagy is considered a potential therapeutic target for renal protection^{[8][9]}.

Sodium-glucose cotransporter-2 (SGLT2) inhibitors and dipeptidylpeptidase-4 (DPP4) inhibitors are promising antidiabetic agents introduced into clinical practice in the last decade. The antihyperglycemic effect of SGLT2 inhibitors is mediated by increment of glucosuria, while DPP4 inhibitors realize their activity through an increase in the half-life of incretin hormones. Both SGLT2 and DPP4 inhibitors demonstrated renal protective activity in large-scaled randomized clinical trials. Specifically, in the Empagliflozin Cardiovascular Outcome Event Trial in Type 2 Diabetes Mellitus Patients (EMPA-REG OUTCOME trial) SGLT2 inhibitor empagliflozin reduced the risk of progression to macroalbuminuria, a doubling of the serum creatinine, initiation of renal replacement therapy or death from renal disease in patients with type 2 diabetes [10]. In the Cardiovascular and Renal Microvascular Outcome Study with Linagliptin in Patients with Type 2 Diabetes Mellitus (CARMELINA study), the DPP4 inhibitor linagliptin retarded the albuminuria progression in patients with type 2 diabetes and high cardiovascular risk [11]. It is assumed that double inhibition of SGLT2 and DPP4 may have advantages in terms of renoprotection [12][13].

The mechanisms of protective action of SGLT2 inhibitors and DPP4 inhibitors on diabetic kidney are still a matter for debate. SGLT2 inhibitors have been shown to enhance sodium excretion and suppress glomerulosclerosis and interstitial fibrosis, inflammation and oxidative stress in diabetic kidney [14][15][16]. In addition, DPP4 inhibitors have been shown to reduce glomerular hypertension and hyperfiltration [17][18][19]. Moreover, empagliflozin has been shown to activate autophagy in tubular cells [20][21]. However, the effect of SGLT2 and DPP4 inhibition on glomerular autophagy has not been studied yet [22].

Therefore, we aimed to assess the effect of SGLT2 inhibitor empagliflozin, DPP4 inhibitor linagliptin, and a combination of both agents on glomerular autophagy in a model of type 2 diabetic kidney disease.

2. Data, Model, Applications and Influences

2.1. Methods of the study

The study was carried out at the Center for Genetic Resources of Laboratory Animals at the IC&G SB RAS (Novosibirsk, Russia) with *db/db*-specific pathogen-free mice (BKS.Cg- Dock7^{m/+}/+Lepr^{db}/J, stock #000642) obtained from Jackson Laboratory (Bar Harbor, Maine, USA). The design of the study is presented in Figure 1. After randomization, diabetic *db/db* mice received empagliflozin (Boehringer Ingelheim, Ingelheim, Germany), linagliptin (Boehringer Ingelheim, Ingelheim, Germany), or the combination of both agents, at a dose of 10 mg/kg of body weight diluted in 200 μ l of saline intragastrically. Vehicle-treated mice received only 200 μ l intragastrically. All animals were treated once per day for 56 days from 8 to 16 weeks. The control group included the littermate non-diabetic *db/+* male mice.

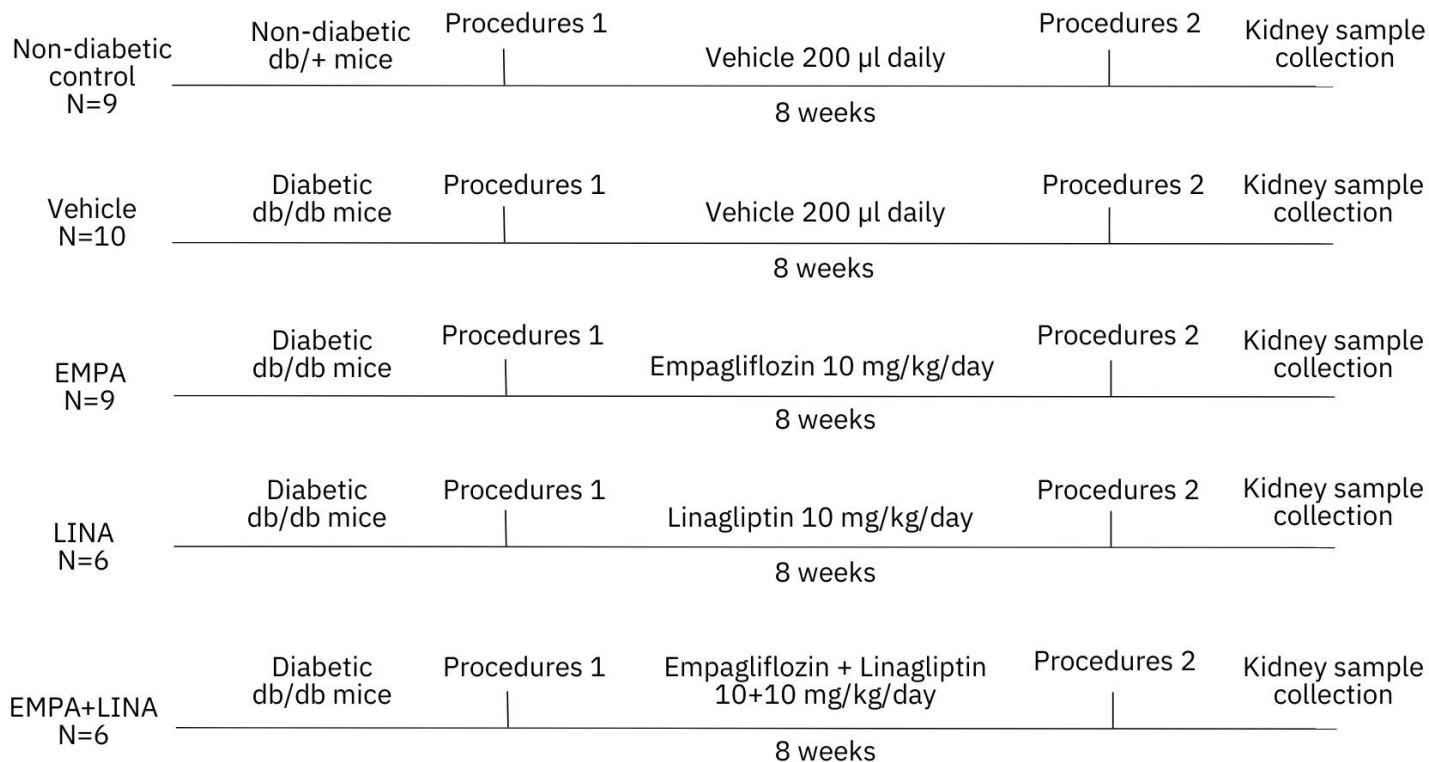


Figure 1. Flowchart of the study. The experiment was started with 8-week-old non-diabetic *db*+ and diabetic *db/db* mice. Diabetic mice were randomly assigned for treatment with a vehicle, empagliflozin, linagliptin, or combination of empagliflozin and linagliptin. The total duration of the experiment was 8 weeks. Procedures 1 and 2 included body weight measurement, body composition assessment, blood plasma and urine sampling. EMPA, empagliflozin-treated *db/db* mice; LINA, linagliptin-treated *db/db* mice; EMPA + LINA, empagliflozin-linagliptin-treated *db/db* mice.

Animals were weighed weekly. Body composition was assessed at week 8 and 16 of age by MRI body composition analyzer (Echo Medical Systems, Houston, TX, USA). On the same days, prior to the weighting and body composition study, random blood and urine samples. At week 16, all mice were sacrificed by decapitation under anesthesia. Following laboratory investigations includes plasma levels of glucose, fructosamine, glycated albumin, creatinine, leptin, insulin, ghrelin and PAI-1, urinary albumin-creatinine ratio (UACR).

Kidney samples were obtained for histological assessment, ultrastructural examination, western blot and IHC. To assess renal hypertrophy, kidneys were weighted and adjusted to body weight and lean mass. Mesangial hypertrophy was assessed as volume density (V_v) of mesangii in glomeruly. The width of glomerular basement membrane (GBM), mean width and numerical density (N_A) of podocyte FPs was assessed in transmission electron microscopy (TEM) images.

Autophagy and apoptosis regulators in renal cortex were assessed by western blotting using anti-LAMP1, anti-Caspase-3, anti-Bcl-2, anti-LC3B antibody (Abcam, Cambridge, UK, ab25245, ab13847, ab692 and ab48394).

Glomerular autophagy was assessed by with staining for beclin-1 and LAMP-1 (ab62557 and ab25245, respectively, Abcam, Cambridge, UK). The V_V of mesangium, glomerular beclin-1, and LAMP-1 positive areas was calculated.

Autophagic compartments (autophagosomes, autolysosomes and lysosomes) in podocytes were defined according to the guidelines for the use and interpretation of assay for monitoring autophagy (3rd edition)^[23]. Autophagosomes were identified as compartments with a double membrane, which is separated by an electron-translucent cleft and encircling cytosol and/or organelles appearing morphologically intact^{[23][24]}. Single-membrane, relatively electron-dense organelles, homogenously filled with very tiny granules, were counted as lysosomes (Figure 2a)^[25]. Single-membrane compartments with electron dense cytoplasmic material and/or organelles at various stages of degradation were recognized as autolysosomes (Figure 2b)^{[23][24]}. The V_V of autophagosomes, autolysosomes and lysosomes in the podocyte cytoplasm was assessed.

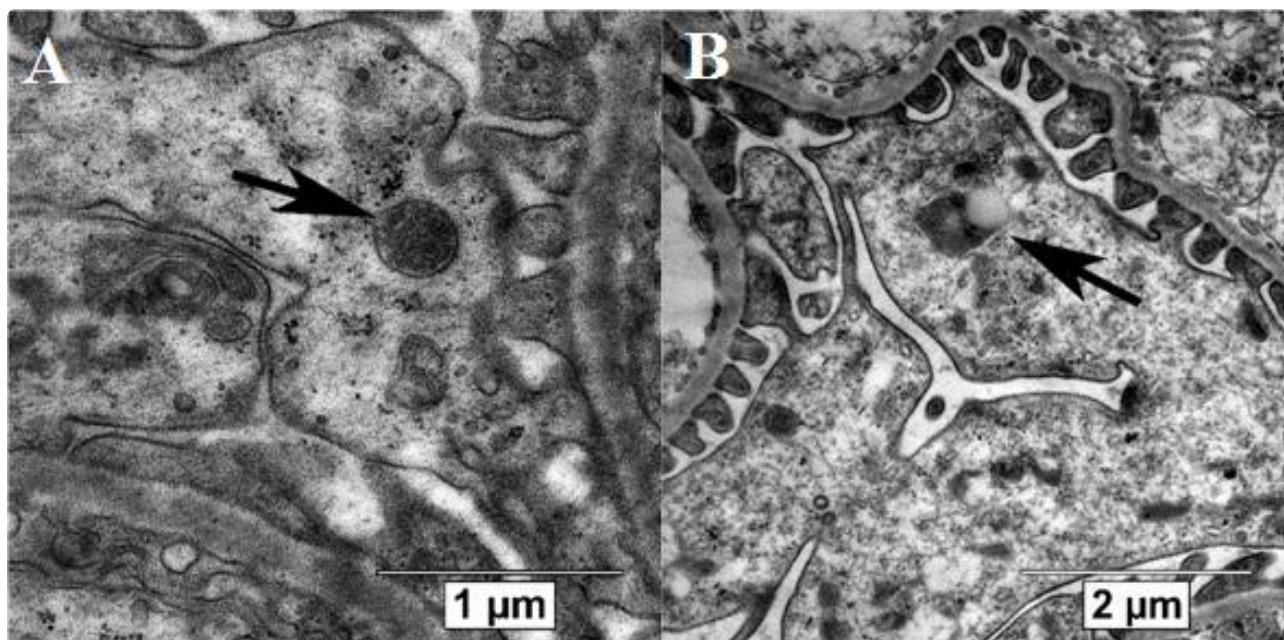


Figure 2. Autophagic compartments in mouse podocytes: lysosomes (black arrow, **a**) and autolysosome (black arrow, **b**). TEM, scale bars: 1 μ m (a) and 2 μ m (b).

2.2. Body Composition and Biochemical Parameters

Diabetic *db/db* mice had become obese and hyperglycemic prior to the start of experiment. When compared to *db/+* controls, 8-week-old *db/db* mice demonstrated increase in their fat mass and reduction of lean mass and water content (Table 1). In vehicle-treated mice, these changes remained stable throughout the experiment. In actively treated animals, especially in empagliflozin groups, a further increase in the body weight and fat mass was observed.

Table 1. Body weight and body composition in *db/+* and *db/db* mice.

Parameter	Week of age	<i>db/+</i> mice		<i>db/db</i> mice		
		(non- diabetic control)	Vehicle	EMPA	LINA	EMPA+LINA
		n	9	10	9	6
Body weight, g	8	26.8 (25.0– 31.1)	37.1 (32.1– 41.6)***	37.4 (28.5– 41.5)***	36.5 (29.6– 40.3)**	37.3 (35.9– 39.5)***
	16	24.8 (21.6– 27.7) ⁺	37.8 (34.3– 40.4)***	52.6 (42.5– 62.2)***##++	45.3 (39.5– 47.5)***\$§ ⁺	50.4 (46.9– 52.3)***#ØØ ⁺
Fat mass, g	8	3.1 (1.9– 4.1)	18.3 (16.2– 22.5)***	18.4 (13.5– 20.6)***	18.0 (15.1– 19.5)***	18.1 (16.7– 19.5)***
	16	2.7 (2.1– 4.5)	19.6 (17.4– 22.7)***	28.0 (22.2– 35.3)***##++	23.4 (20.3– 26.4)***## ⁺	26.7 (23.2– 28.3)***## ⁺
Lean mass, g	8	22.2 (21.0– 24.8)	17.1 (16.3– 18.7)***	18.2 (13.4– 19.8)***	16.0 (13.8– 17.2)***	18.5 (15.8– 19.1)***
	16	21.2 (19.1– 22.5) ⁺	16.1 (14.3– 22.4)**	21.4 (19.1–24.4) ⁺⁺	17.6 (14.9– 20.7)**\$§ ⁺	20.6 (19.5–21.5) ^Ø ⁺

		8	16	Total water, g		
		19.5 (18.4– 21.5)	14.8 (14.5– 16.4)***	15.8 (11.6– 17.6)***	12.5 (9.6– 14.1)***	16.0 (13.7– 16.5)***
				18.4 (16.5– 19.4) ⁺	14.1 (12.3– 19.2) [*]	18.8 (16.7–21.4) ⁺⁺

* $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$ vs. non-diabetic control ($db/+$); # $p < 0.05$, ## $p < 0.01$, ### $p < 0.001$ vs. vehicle-treated db/db mice; \S $p < 0.05$, $\S\S$ $p < 0.01$ vs. empagliflozin group (Mann–Whitney U-test); \emptyset $p < 0.05$, $\emptyset\emptyset$ $p < 0.01$ vs. linagliptin group (Mann–Whitney U-test); $^+$ $p < 0.05$, $^{++}$ $p < 0.01$ vs. with week 8 (Wilcoxon test). EMPA, empagliflozin-treated db/db mice; LINA, linagliptin-treated db/db mice; EMPA+LINA, empagliflozin-linagliptin-treated db/db mice. Data are presented as medians (min – max values).

Diabetic mice demonstrated increased plasma glucose, fructosamine, glycated albumin, leptin, insulin and PAI-1 levels; meanwhile, ghrelin concentrations were decreased significantly (Table 2). Groups of animals were heterogeneous for plasma glucagon levels at week 8 and week 16. Severe hyperglycemia persisted throughout the experiment in vehicle-treated and linagliptin-treated db/db mice.

Table 2. Plasma biochemical parameters and hormones in db/db and $db/+$ mice.

Parameter	Week of age	<i>db/+</i> mice (non- diabetic control)	<i>db/db</i> mice			
			Vehicle	EMPA	LINA	EMPA+LINA
<i>n</i>		9	10	9	6	6

Glucose, mmol/l	8	9.5 (5.1–11.4)	28.5 (16.8– 40.1)***	23.1 (15.3– 35.4)***	27.5 (20.6– 48.8)***
	16	9.5 (8.5–12.2)	32.7 (22.5– 53.1) ***	16.1 (9.9– 23.6)***## ⁺	29.9 (25.8– 34.6)*** ^{SS}
Fructosamine, μmol/l	8	237 (217–249)	456 (424– 511)***	480 (425– 579)***	436 (383– 487)***
	16	239 (222–296)	622 (524– 672)***++	468 (341– 491)***	380 (342– 691)*** ^{SS} ⁺
Glycated albumin, μmol/l	8	107 (103–127)	227 (206– 239)***	235 (217– 261)***	209 (156– 256)***
	16	117 (109–133)	283 (252– 349)***++	210 (166– 245)***## ⁺	178 (157– 376)*** ^{SS}
Leptin, ng/ml	8	3.30 (1.40– 6.54)	97.1 (53.2– 114.4)***	93.4 (80.8– 133.1)***	92.9 (76.0– 118.4)***
	16	3.80 (1.50– 6.30)	90.0 (21.2– 151.4)***	136.4 (53.0– 171.2)***## ⁺	96.5 (51.6– 172.6)***
Insulin, ng/ml	8	5.59 (3.43– 18.0)	25.6 (11.5– 45.2)***	21.2 (10.8– 40.8)**	22.7 (15.3– 54.0)**
	16	10.0 (2.10– 22.2)	22.1 (9.8– 33.2)**	20.8 (5.95– 56.7)*	13.9
					33.3 (13.6– 72.9)** ^Ø

					(7.2–23.7)
		370	660	450	375
	8	(250–2670)	(190–2760)	(290–2180)	(260–2090)
Glucagon, ng/ml					(170–780)
		370	605	390	325
	16	(150–2120)	(260–2960)	(240–860)	(190–810)
					(200–460) [#]
	8	0.99 (0.24– 4.27)	0.42 (0.10– 1.57)**	0.50 (0.25– 1.15)**	0.37 (0.25– 0.82)**
Ghrelin, ng/ml					0.39 (0.32–1.13)**
	16	1.47 (0.76– 6.31)	1.27 (0.57– 2.26)*	0.81 (0.28–2.58)	0.45 (0.20– 1.14)*** [§]
					0.77 (0.08–1.47)*
	8	1.52 (0.70– 3.24)	2.76 (1.41– 4.23)*	2.07 (1.00–6.01)	1.84 (0.48–4.20)
PAI-1, ng/ml					2.25 (1.72–3.91)*
	16	3.22 (0.40– 4.36)	1.64 (0.09– 3.76)	2.22 (0.79–5.12)	1.85 (0.63–4.17)
					2.82 (0.98–6.05)

* $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$ vs. non-diabetic control ($db/+$); [#] $p < 0.05$, ^{##} $p < 0.01$, ^{###} $p < 0.001$ vs. vehicle-treated db/db mice; [§] $p < 0.05$, ^{§§} $p < 0.01$ vs. empagliflozin group (Mann–Whitney U-test) \emptyset $p < 0.05$, $\emptyset\emptyset$ $p < 0.01$ vs. linagliptin group (Mann–Whitney U-test); ⁺ $p < 0.05$, ⁺⁺ $p < 0.01$, ⁺⁺⁺ $p < 0.001$ vs. week 8 (Wilcoxon test). EMPA, empagliflozin-treated db/db mice; LINA, linagliptin-treated db/db mice; EMPA+LINA, empagliflozin–linagliptin-treated db/db mice. PAI-1, plasminogen activator inhibitor-1. Data are presented as medians (min – max values).

Under the treatment with empagliflozin or empagliflozin–linagliptin combination, db/db mice demonstrated improvement in their glycemic status, which was documented by the reduction in their plasma glucose, fructosamine and glycated albumin (empagliflozin: $p = 0.02$, $p = 0.02$, and $p = 0.02$, respectively; combination: $p =$

0.03, $p = 0.046$, and $p = 0.046$, respectively, vs. week 8). Meanwhile, no significant changes in glycemic control were observed in the linagliptin group. The levels of insulin, glucagon, ghrelin and PAI-1 did not change significantly throughout the experiment in all groups ($p > 0.05$ week 16 vs. week 8). However, plasma leptin levels were increased significantly in the empagliflozin group and tended to increase in the empagliflozin–linagliptin group ($p = 0.03$ and $p = 0.08$, respectively, week 16 vs. week 8). Meanwhile, empagliflozin–linagliptin-treated *db/db* mice demonstrated lower plasma levels of glucagon as compared with vehicle-treated *db/db* mice ($p = 0.04$).

2.3. Albuminuria and Renal Function

A markedly increased albuminuria was detected in all diabetic mice groups at the start of experiment (all $p < 0.001$, Table 3). Administration of empagliflozin, linagliptin, or both agents decreased albumin excretion ($p = 0.007$, $p = 0.03$, and $p = 0.04$, respectively, vs. week 8). No significant changes in the levels of plasma creatinine were detected throughout the study.

Table 3. Albuminuria and plasma creatinine in *db/+* and *db/db* mice.

Parameter	Week of age	<i>db/+</i> mice		<i>db/db</i> mice		
		(non-diabetic control)	Vehicle	EMPA	LINA	EMPA + LINA
<i>n</i>		9	10	9	6	6
UACR, mg/mmol	8	1.50 (0.60–4.00)	14.8 (10.0–26.9)***	16.3 (12.8–30.0)***	26.8 (19.4–54.7)***	23.5 (14.3–41.4)***
	16	1.80 (0.10–3.10)	21.4 (14.4–29.4)***	11.4 (4.00–22.7)****#+	6.9 (4.40–22.4)****#+	10.6 (4.50–17.6)****#+
Plasma creatinine, μmol/l	8	67.0 (47.0–78.0)	76.4 (70.2–84.2)	70.5 (54.0–82.2)	70.3 (52.8–103.5)	70.4 (66.6–82.2)
	16	61.8 (54.0–71.7)	75.6 (63.9–89.4)**	81.3 (55.8–91.2)*	71.7 (67.8–99.0)**	80.4 (65.7–91.2)**

* $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$ vs. non-diabetic control ($db/+$ mice), Mann–Whitney U -test; ## $p < 0.01$, ### $p < 0.001$ vs. vehicle-treated db/db mice; + $p < 0.05$, ++ $p < 0.01$ vs. with week 8, Wilcoxon test. EMPA, empagliflozin-treated db/db mice; LINA, linagliptin-treated db/db mice; EMPA+LINA, empagliflozin–linagliptin-treated db/db mice. UACR, urinary albumin-to-creatinine ratio. Data are presented as medians (min - max values).

2.4. Renal Hypertrophy, Glomerular and Podocyte Morphology

Vehicle-treated db/db mice demonstrated increase in their kidney weight and kidney weight/lean mass ratio, whereas kidney weight/body weight ratio was decreased ($p = 0.048$, $p = 0.0007$ and $p = 0.00002$ vs. non-diabetic $db/+$ mice, respectively, Figure 3). The kidney weight/body weight and kidney weight/lean mass ratio was lower in empagliflozin and linagliptin-treated mice as compared to vehicle-treated animals (all $p < 0.05$).

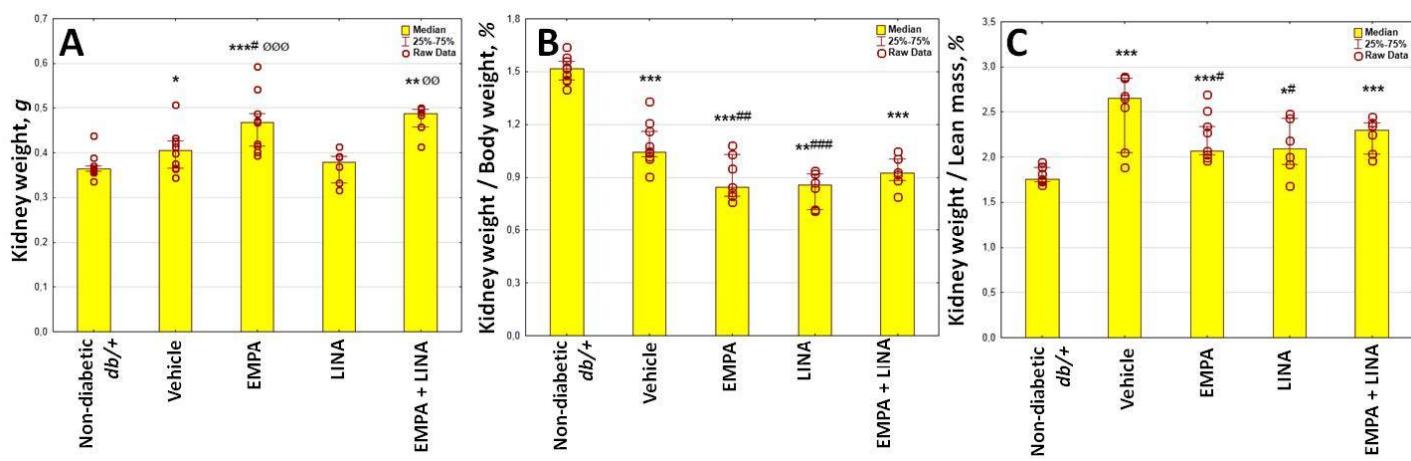


Figure 3. Parameters of kidney weight in non-diabetic $db/+$ mice and diabetic db/db mice: (a) kidney weight, g; (b) kidney weight / body weight, %; (c) kidney weight / lean mass ratio, %. EMPA, empagliflozin-treated db/db mice; LINA, linagliptin-treated db/db mice; EMPA+LINA, empagliflozin–linagliptin-treated db/db mice. Data are presented as a bar graph (median, lower and upper quartile) with individual data set (dots); * $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$ vs. non-diabetic control ($db/+$ mice), # $p < 0.05$, ## $p < 0.01$, ### $p < 0.001$ vs. vehicle-treated db/db mice, ØØ $p < 0.01$, ØØØ $p < 0.001$ vs. linagliptin-treated db/db mice.

Vehicle-treated db/db mice demonstrated typical morphological signs of diabetic nephropathy, including mesangial expansion, thickening of glomerular basement membrane (GBM), and effacement of podocyte foot processes (FPs). These changes were quantified by morphometric analysis (Table 4, Figure 4).

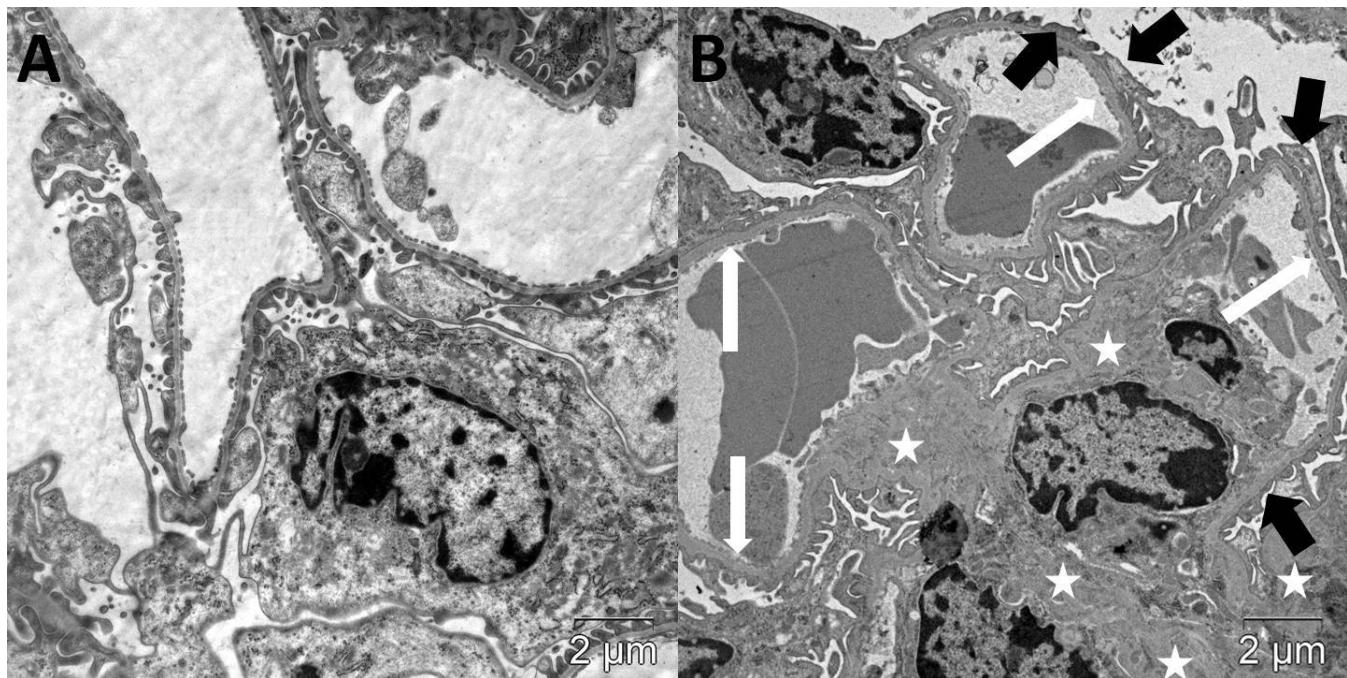


Figure 4. Glomerular structure in non-diabetic *db/+* mice (a) and vehicle-treated diabetic *db/db* mice (b). Diabetic mice demonstrated thickening of GBM (white arrows), effacement of podocyte FPs (black arrows) and mesangial expansion (white stars). Scale bar 2 μ m. TEM.

Table 4. Glomerular structural parameters in *db/+* and *db/db* mice.

Parameter	<i>db/+</i> mice (non-diabetic control)	<i>db/db</i> mice			
		Vehicle	EMPA	LINA	EMPA+LINA
<i>n</i>	9	10	9	6	6
Mesangial fractional volume, %	14.4 (9.8–18.5)	38.6 (34.5–42.7)***	25.5 (20.4–35.8)***##	31.1 (27.3–34.6)##	28.5 (26.1–30.0)##
GBM width, nm	135 (116–157)	163 (135–203)*	139 (116–225)##	150 (126–165)##	146 (108–152)##
Width of podocyte FPs, nm	220 (191–242)	372 (299–426)**	203 (162–317)##	189 (172–243)##	202 (165–252)##

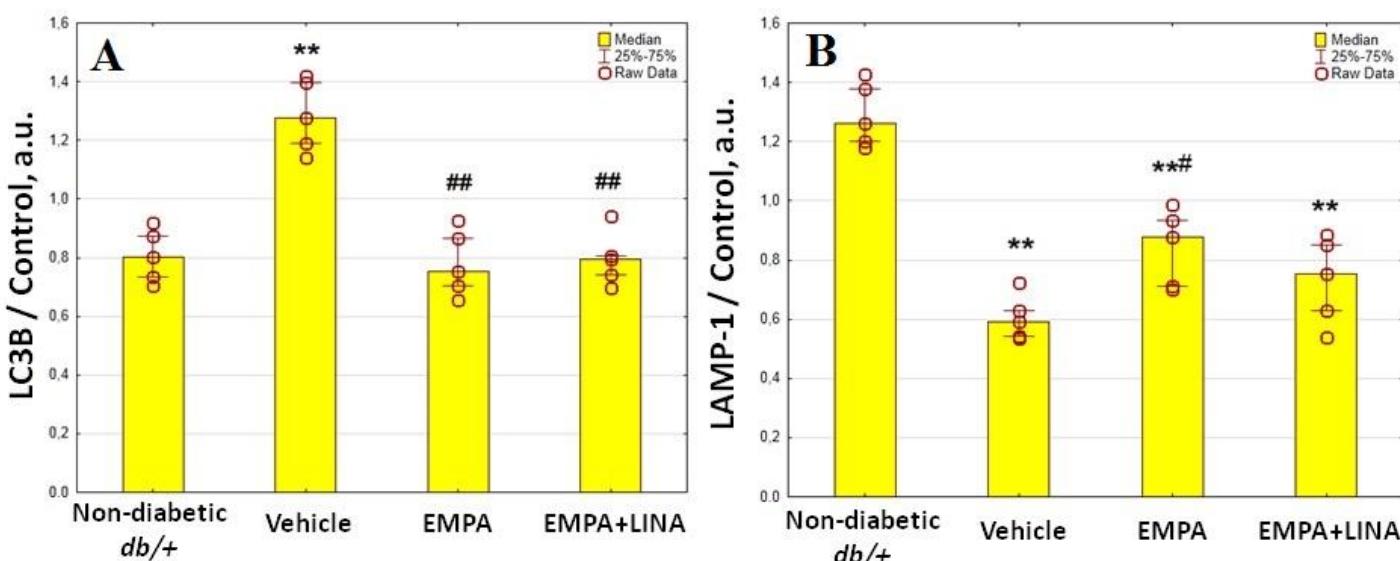
N _A of podocyte	3.39	2.73	3.31	3.33	3.25
FPs, nm ⁻¹	(3.00–3.79)	(1.92–3.51)*	(2.32–4.18) [#]	(2.85–4.07) [#]	(2.58–3.56) [#]

* $p < 0.05$, ** $p < 0.01$, *** $p < 0.001$ vs. non-diabetic control ($db/+$); [#] $p < 0.05$, ^{##} $p < 0.01$, ^{###} $p < 0.001$ vs. vehicle-treated db/db mice; EMPA, empagliflozin-treated db/db mice; LINA, linagliptin-treated db/db mice; EMPA+LINA, empagliflozin–linagliptin-treated db/db mice; GBM, glomerular basement membrane; FPs, foot processes; N_A, numerical density. Data are presented as medians (min–max values).

The structural changes in the kidneys were mitigated by empagliflozin, linagliptin, and combined treatment. Specifically, fractional mesangial volume and mean GBM width were reduced ($p = 0.0008$ and $p = 0.02$ for empagliflozin; $p = 0.002$ and $p = 0.04$ for linagliptin; $p = 0.03$ and $p = 0.04$ for combination, respectively). The mean width of podocyte FPs decreased in empagliflozin-treated, linagliptin-treated and empagliflozin–linagliptin-treated mice ($p = 0.005$, $p = 0.02$, and $p = 0.004$, respectively); accordingly, N_A of FPs increased in these groups ($p = 0.03$, $p = 0.04$, and $p = 0.03$, respectively).

2.5. Expression of Autophagy and Apoptosis Regulators in the Renal Cortex

Western blotting analysis showed protein expression of LC3B, an autophagosome marker, to be significantly elevated in renal cortex in vehicle-treated db/db mice compared to controls ($p=0.008$, Figure 5). We found decrease in the protein expression of LC3B in db/db mice treated with empagliflozin and the combination of empagliflozin and linagliptin (both $p = 0.008$ vs. vehicle group). Protein expression of LAMP-1, a lysosomal marker, was reduced significantly in vehicle-treated db/db mice when compared to non-diabetic $db/+$ mice ($p = 0.008$). In the empagliflozin group, the amount of LAMP-1 was restored ($p = 0.03$ vs. vehicle-treated db/db mice). Although empagliflozin–linagliptin-treated db/db mice demonstrated similar changes in LAMP-1 protein expression, statistical significance was not achieved ($p = 0.15$ vs. vehicle-treated db/db mice).



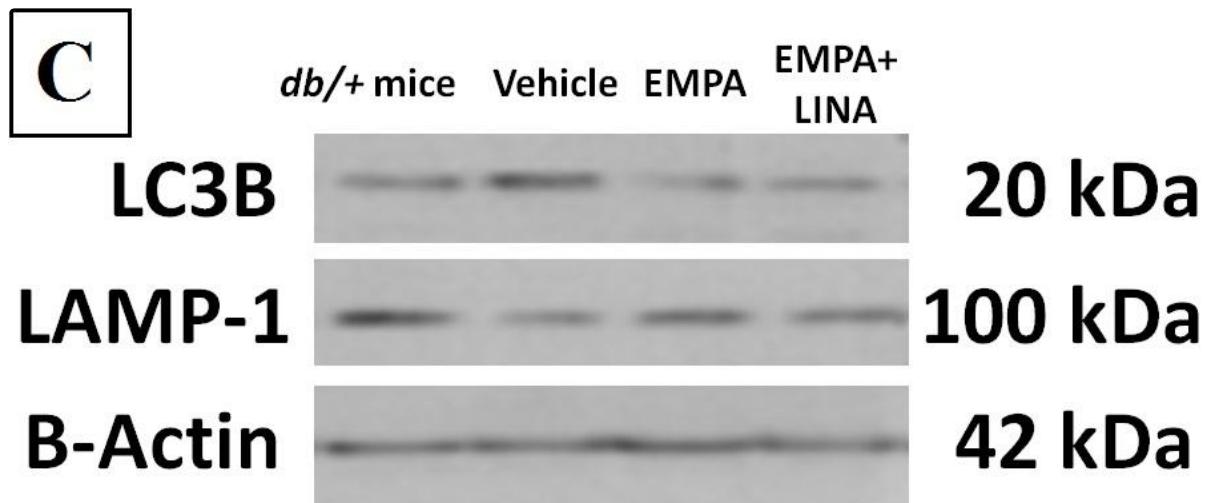
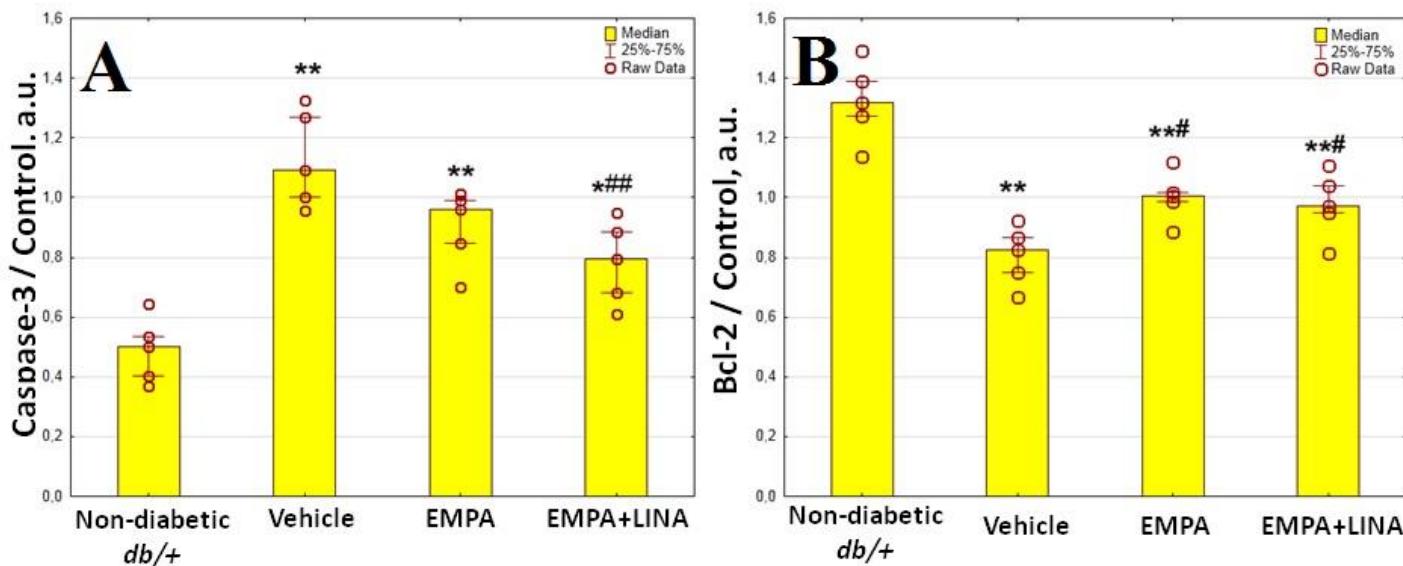


Figure 5. Protein expression of LC3B, an autophagosome marker (a), and LAMP-1, a lysosome marker (b), in the renal cortex from non-diabetic *db/+* mice and diabetic *db/db* mice treated with vehicle, empagliflozin, or empagliflozin–linagliptin; (c) representative immunoblots of LC3B, LAMP-1 and β -actin as the control. Data are presented as a bar graph (median, lower and upper quartile) with individual data set (dots); a.u., arbitrary unit; ** $p < 0.01$ vs. non-diabetic *db/+* mice; # $p < 0.05$, ## $p < 0.01$ vs. vehicle-treated *db/db* mice (Mann–Whitney U-test).

The levels of caspase-3 expression, an apoptosis marker, were elevated markedly in vehicle-treated *db/db* mice ($p = 0.008$, Figure 6). The expression of caspase-3 tended to be lower in empagliflozin-treated *db/db* mice ($p = 0.095$ vs. vehicle-treated *db/db* mice); it was significantly lower in *db/db* mice treated with empagliflozin and linagliptin ($p = 0.008$ vs. vehicle-treated *db/db* mice).



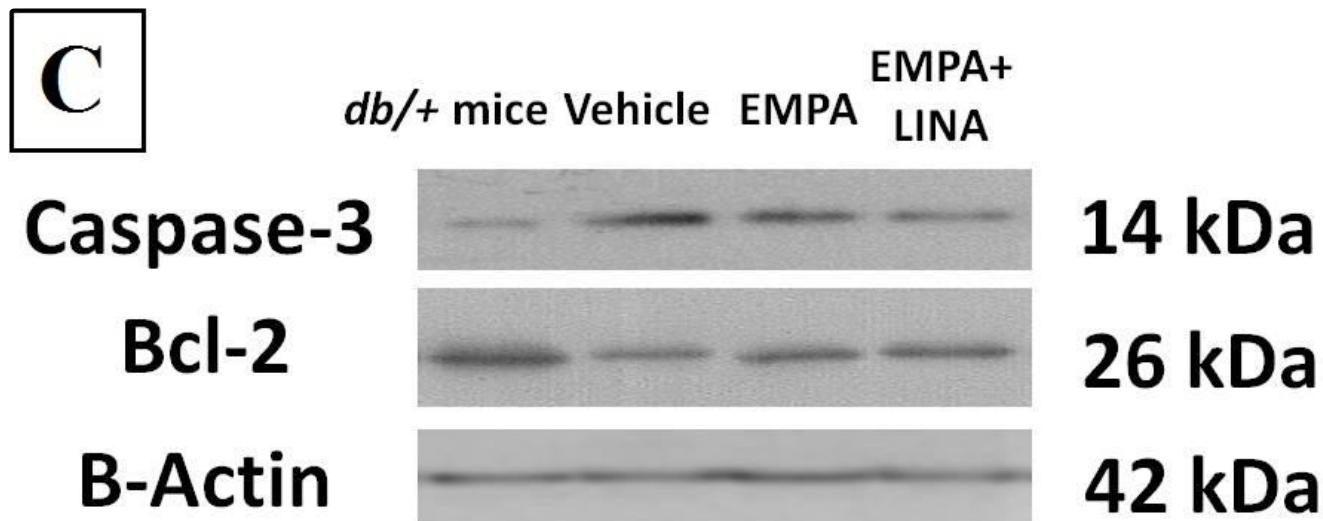


Figure 6. Protein expression of caspase-3, a regulator of apoptosis (a), and Bcl-2, an anti-apoptotic protein (b) in the renal cortex from non-diabetic *db/+* mice and diabetic *db/db* mice treated with vehicle, empagliflozin, and empagliflozin–linagliptin; (c) representative immunoblots of caspase-3, Bcl-2 and β -actin as the control. Data are presented as a bar graph (median, lower and upper quartile) with individual data set (dots); a.u., arbitrary unit; * $p < 0.05$, ** $p < 0.01$ vs. non-diabetic *db/+* mice; # $p < 0.05$, ## $p < 0.01$ vs. vehicle-treated *db/db* mice (Mann–Whitney U -test).

The expression of Bcl-2, an anti-apoptotic protein, was lower in the kidneys of vehicle-treated *db/db* mice as compared to *db/+* controls ($p = 0.008$, Figure 6). The *db/db* mice treated with empagliflozin and empagliflozin–linagliptin demonstrated increase in Bcl-2 expression when compared to vehicle-treated animals ($p = 0.02$ and $p = 0.046$, respectively).

2.6. Glomerular Expression of the Autophagy Markers

The glomerular expression of beclin-1 and LAMP-1, two markers attributed to autophagy, was assessed by immunohistochemistry (IHC). Vehicle-treated *db/db* mice demonstrated a reduction of beclin-1-positive areas in glomeruli ($p = 0.00002$ vs. non-diabetic *db/+* mice). This reduction was mitigated by empagliflozin, linagliptin, or empagliflozin–linagliptin treatment ($p = 0.0003$, $p = 0.001$ and $p = 0.001$, respectively vs. vehicle-treated *db/db* mice, Figure 7).

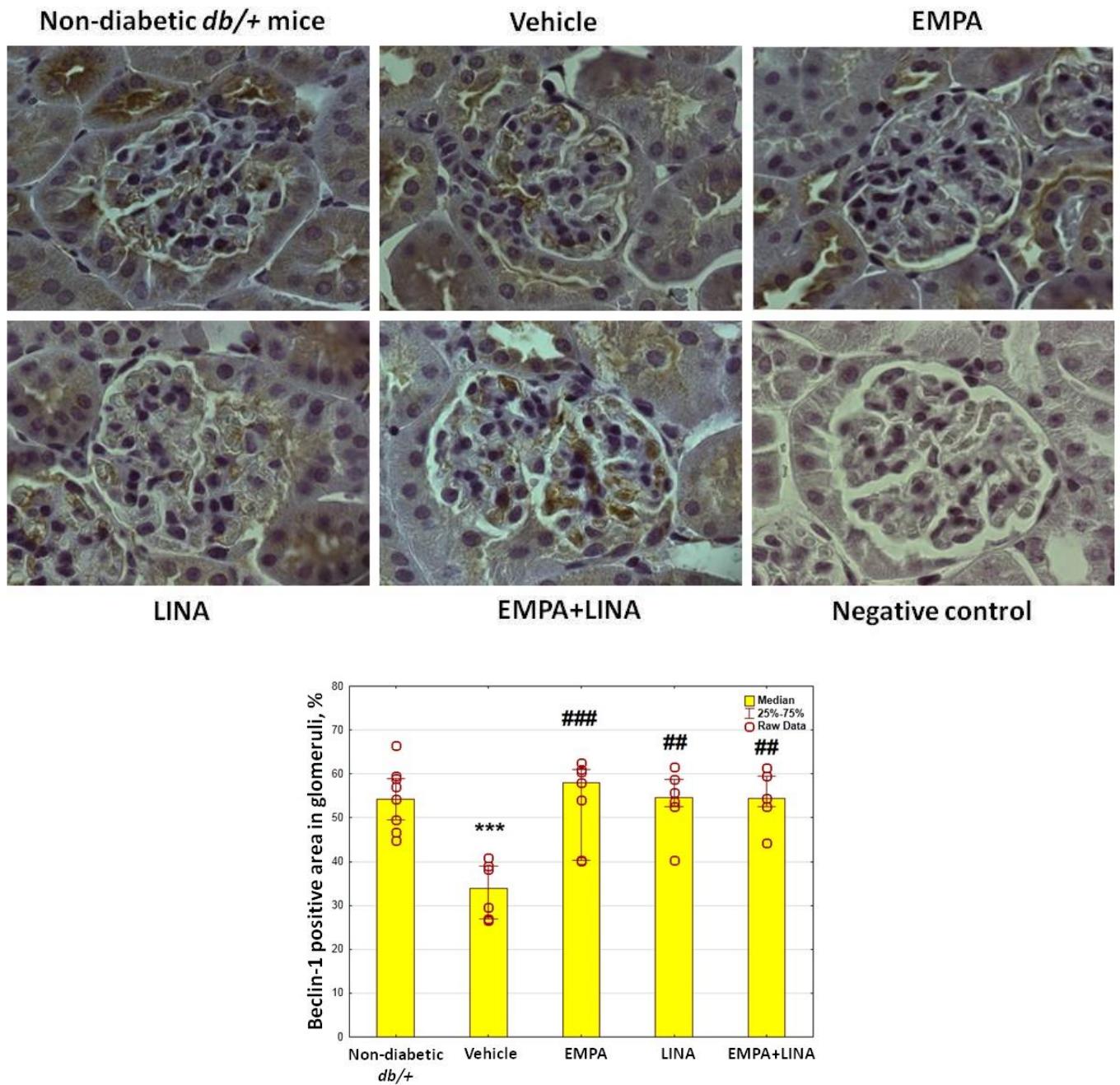


Figure 7. Glomerular beclin-1 expression in non-diabetic *db*/+ mice and diabetic *db*/*db* mice. **Top:** IHC-pictures of glomeruli of non-diabetic *db*/+ mice and *db*/*db* mice treated by vehicle, empagliflozin, linagliptin, or empagliflozin–linagliptin; magnification $\times 400$; **Bottom:** Quantitative analysis of beclin-1-positive area in glomeruli. Data are presented as a bar graph (median, lower and upper quartile) with individual data set (dots); *** $p < 0.001$ vs. non-diabetic *db*/+ mice; ## $p < 0.01$, ### $p < 0.001$ vs. vehicle-treated *db*/*db* mice (Mann-Whitney *U*-test).

The area of the staining for LAMP-1 was reduced in vehicle-treated *db*/*db* mice as compared to non-diabetic *db*/+ controls ($p = 0.01$). Administration of empagliflozin, linagliptin or both agents preserved this reduction ($p = 0.002$, $p = 0.04$ and $p = 0.01$ vs. vehicle-treated *db*/*db* mice respectively, Figure 8).

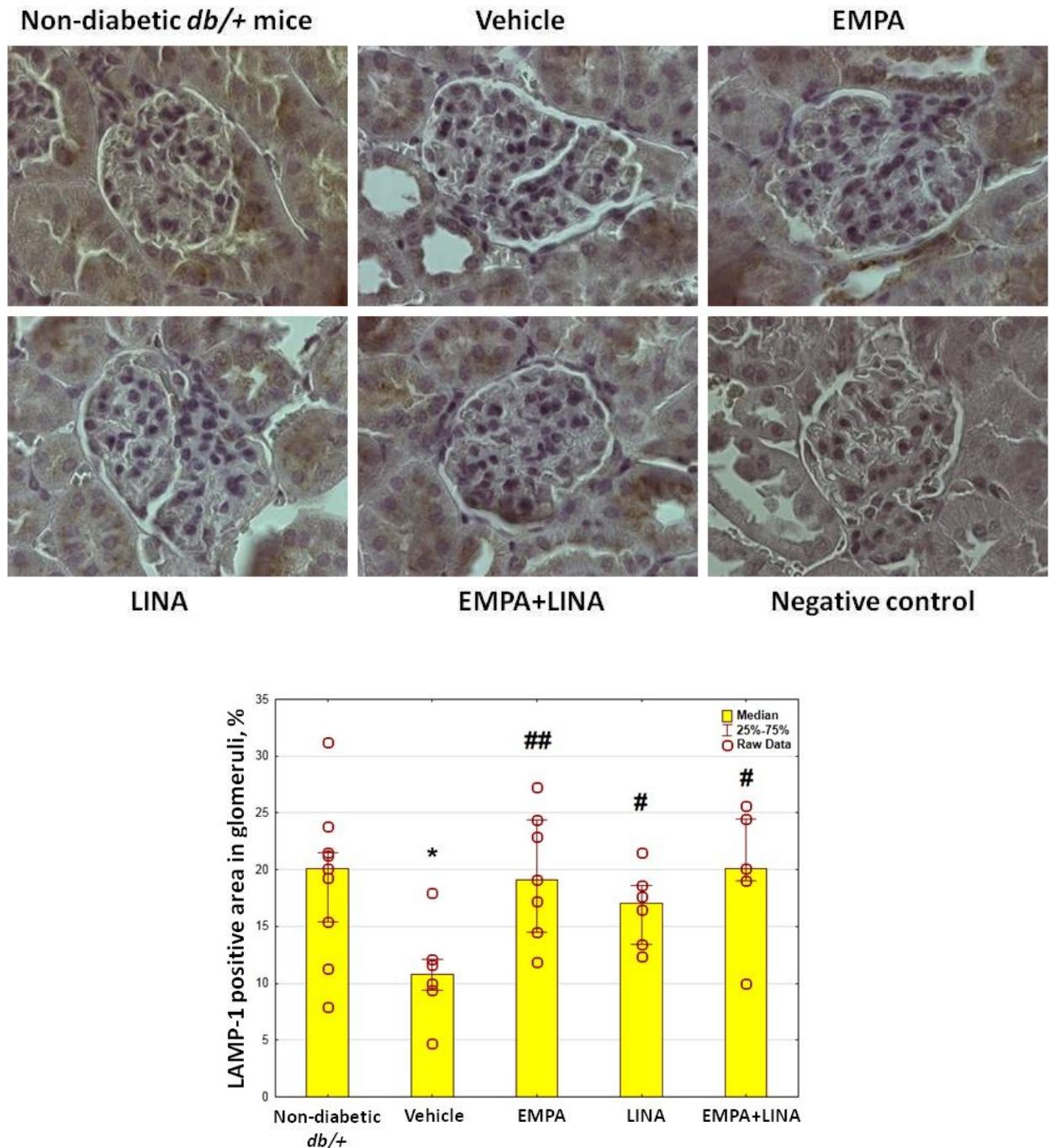


Figure 8. Glomerular LAMP-1 expression in non-diabetic *db/+* mice and diabetic *db/db* mice. **Top:** IHC-pictures of glomeruli of non-diabetic *db/+* mice and *db/db* mice treated by vehicle, empagliflozin, linagliptin, or empagliflozin–linagliptin; magnification $\times 400$; **Bottom:** Quantitative analysis of LAMP-1-positive areas in glomeruli. Data are presented as a bar graph (median, lower and upper quartile) with individual data set (dots); * $p < 0.05$ vs. non-diabetic *db/+* mice; # $p < 0.05$, ## $p < 0.01$ vs. vehicle-treated *db/db* mice (Mann–Whitney *U*-test).

2.7. Autophagy in Podocytes

The V_V of autophagosomes, autolysosomes and lysosomes in podocytes was assessed from TEM images. Vehicle-treated diabetic *db/db* mice had reduced V_V of autophagosomes in podocytes ($p = 0.00002$); administration

of empagliflozin, linagliptin, or both agents increased V_V of autophagosomes significantly ($p = 0.00004$, $p = 0.0003$, and $p = 0.0005$, respectively, Table 5).

Table 5. The volume density (V_V) of autophagosomes, autolysosomes and lysosomes in podocytes in *db/+* and *db/db* mice.

Parameter	<i>db/+</i> mice (non-diabetic control)	<i>db/db</i> mice groups			
		Vehicle	EMPA	LINA	EMPA+LINA
		<i>n</i>	5	6	5
Autophagosomes, V_V , %	2.10 (1.80–2.86)	0.80 (0.61–1.68)***	2.09 (1.65–2.5)###	2.23 (1.73–2.35)###	2.14 (1.57–2.31)###
Autolysosomes, V_V , %	2.37 (1.85–3.21)	1.09 (0.84–1.69)***	2.25 (1.37–3.09)###	2.37 (1.58–2.96)###	2.22 (1.58–3.04)###
Lysosomes, V_V , %	2.94 (2.42–4.00)	2.22 (1.22–2.92)***	3.16 (2.06–3.83)##	2.64 (2.08–3.52)##	2.89 (2.44–3.58)###

*** $p < 0.001$ vs. non-diabetic control (*db/+*); # $p < 0.05$, ## $p < 0.01$, ### $p < 0.001$ vs. vehicle-treated *db/db* mice; EMPA, empagliflozin-treated *db/db* mice; LINA, linagliptin-treated *db/db* mice; EMPA + LINA, empagliflozin–linagliptin-treated *db/db* mice; V_V , volume density. Data are presented as medians (min–max values).

There was a reduction in the count of autolysosomes in vehicle-treated *db/db* mice ($p = 0.00002$, Table 5). Nevertheless, diabetic *db/db* mice treated with empagliflozin, linagliptin or the combination of both demonstrated an increase in V_V of autolysosomes ($p = 0.0003$, $p = 0.0005$, and $p = 0.0005$, respectively). The changes in the count of lysosomes under these treatments were similar ($p = 0.002$, $p = 0.04$, and $p = 0.0005$, respectively).

2.8. Relationships between Autophagy Intensity, Glomerular Morphology and Biochemical Parameters

In *db/+* and *db/db* mice, both beclin-1 and LAMP-1 correlated positively with V_V of autophagosomes and autolysosomes in podocytes (Table 6). Additionally, the area of the staining for LAMP-1 and a protein amount of LAMP-1 demonstrated a correlation with V_V of the lysosomes. The V_V of autophagosomes and autolysosomes

correlated positively with renal LAMP-1 and Bcl-2. However, V_V of autophagosomes, autolysosomes and lysosomes correlated negatively with renal LC3B and caspase-3.

Table 6. Relationships between renal autophagy and apoptosis markers and V_V of autophagic compartments in podocytes.

Parameter	Autophagosomes, V_V	Autolysosomes, V_V	Lysosomes, V_V
Glomerular beclin-1 (IHC)	$r = 0.58$ $p = 0.0001, n = 26$	$r = 0.48$ $p = 0.002, n = 26$	$r = 0.25$ $p > 0.05, n = 26$
Glomerular LAMP-1 (IHC)	$r = 0.46$ $p = 0.003, n = 26$	$r = 0.50$ $p = 0.001, n = 26$	$r = 0.42$ $p = 0.008, n = 26$
Renal LC3B (WB)	$r = -0.57$ $p = 0.009, n = 20$	$r = -0.69$ $p = 0.0008, n = 20$	$r = -0.55$ $p = 0.01, n = 20$
Renal LAMP-1 (WB)	$r = 0.50$ $p = 0.02, n = 20$	$r = 0.55$ $p = 0.01, n = 20$	$r = 0.63$ $p = 0.003, n = 20$
Renal caspase-3 (WB)	$r = -0.35$ $p > 0.05, n = 20$	$r = -0.56$ $p = 0.01, n = 20$	$r = -0.44$ $p > 0.05, n = 20$
Renal Bcl-2 (WB)	$r = 0.54$ $p = 0.01, n = 20$	$r = 0.57$ $p = 0.009, n = 20$	$r = 0.63$ $p = 0.003, n = 20$

Data are presented as Spearman's rank correlation coefficients, p -value and number of observations; IHC, immunohistochemistry; V_V , volume density; WB, western blot.

Kidney weight/lean mass ratio, considered as an indicator of renal hypertrophy, was found to correlate negatively with V_V of autolysosomes and lysosomes in podocytes (Table 7). Mesangial fractional volume correlated negatively with all studied autophagy parameters. A similar pattern of relationships was demonstrated for GBM width. The correlations between metrics of podocyte FPs and autophagy indicators were also revealed.

Table 7. Relationships between glomerular and podocyte autophagy characteristics and other parameters of renal morphology.

Parameter	Glomerular beclin-1, %	Glomerular LAMP-1, %	Autophagosomes in podocytes, V_V	Autolysosomes in podocytes, V_V	Lysosomes in podocytes, V_V
Kidneys weight/lean mass ratio	$r = -0.13$ $p > 0.05, n = 33$	$r = -0.19$ $p > 0.05, n = 33$	$r = -0.31$ $p > 0.05, n = 26$	$r = -0.43$ $p = 0.007, n = 26$	$r = -0.45$ $p = 0.007, n = 26$
Mesangial fractional volume	$r = -0.39$ $p = 0.02, n = 40$	$r = -0.37$ $p = 0.03, n = 40$	$r = -0.36$ $p = 0.03, n = 40$	$r = -0.50$ $p = 0.002, n = 40$	$r = -0.32$ $p = 0.009, n = 40$
Mean width of GBM	$r = -0.41$ $p = 0.03, n = 26$	$r = -0.40$ $p = 0.03, n = 26$	$r = -0.54$ $p = 0.003, n = 26$	$r = -0.45$ $p = 0.01, n = 26$	$r = -0.26$ $p > 0.05, n = 26$
Mean width of podocyte FPs	$r = -0.62$ $p = 0.0004, n = 26$	$r = -0.29$ $p > 0.05, n = 26$	$r = -0.58$ $p = 0.0009, n = 26$	$r = -0.49$ $p = 0.007, n = 26$	$r = -0.41$ $p = 0.03, n = 26$
N_A of podocyte FPs	$r = 0.27$ $p > 0.05, n = 26$	$r = 0.32$ $p > 0.05, n = 26$	$r = 0.54$ $p = 0.003, n = 26$	$r = 0.47$ $p = 0.01, n = 26$	$r = 0.35$ $p > 0.05, n = 26$

Data are presented as Spearman's rank correlation coefficients; GBM, glomerular basement membrane; FPs, foot processes; N_A , numerical density; V_V , volume density

Renal cortex expression of caspase-3, LAMP-2 and Bcl-2 was associated with kidney weight/lean mass ratio (Table 8). The mesangial fractional volume and GBM width correlated positively with renal LC3B and caspase-3. Contrary, renal LAMP-1 and Bcl-2 demonstrated negative correlations with mesangial fractional volume and GBM width. Renal LC3B and caspase-3 were associated with the changes in podocyte FPs.

Table 8. Relationships between expression of autophagy and apoptosis regulators in the renal cortex and parameters of renal and glomerular morphology.

Parameter	Renal LC3B	Renal LAMP-1	Renal caspase-3	Renal Bcl-2
Kidney weight/lean mass ratio	$r = 0.40$ $p > 0.05$	$r = -0.60$ $p = 0.005$	$r = 0.64$ $p = 0.002$	$r = -0.64$ $p = 0.002$
Mesangial fractional volume	$r = 0.45$ $p = 0.045$	$r = -0.71$ $p = 0.0004$	$r = 0.67$ $p = 0.001$	$r = -0.71$ $p = 0.0004$
Mean width of GBM	$r = 0.67$ $p = 0.002$	$r = -0.45$ $p = 0.045$	$r = 0.69$ $p = 0.002$	$r = -0.53$ $p = 0.02$
Mean width of podocyte FPs	$r = 0.56$ $p = 0.02$	$r = -0.38$ $p > 0.05$	$r = 0.52$ $p = 0.02$	$r = -0.44$ $p = 0.048$
N_A of podocyte FPs	$r = -0.46$ $p = 0.04$	$r = 0.38$ $p > 0.05$	$r = -0.62$ $p = 0.004$	$r = 0.46$ $p = 0.04$

Data are presented as Spearman's rank correlation coefficients. $n = 20$. GBM, glomerular basement membrane; FPs, foot processes; N_A , numerical density; V_V , volume density

There were negative correlations between indicators of autophagy and glycemic control parameters. Specifically, plasma fructosamine and glycated albumin correlated with renal LAMP-1 ($r = -0.65, p = 0.002$ and $r = -0.42, p = 0.007$ respectively), glomerular beclin-1 ($r = -0.34, p = 0.04$ for both) and LAMP-1 ($r = -0.41, p = 0.008$; $r = -0.42, p = 0.007$), podocyte V_V of autophagosomes ($r = -0.34, p = 0.03$; $r = -0.32, p = 0.046$), autolysosomes ($r = -0.34, p = 0.03$; $r = -0.31, p > 0.05$) and lysosomes ($r = -0.36, p = 0.02$; $r = -0.33, p = 0.04$). No correlations were found with plasma leptin, insulin, ghrelin and PAI-1. However, plasma glucagon concentrations showed negative correlations with glomerular LAMP-1 ($r = -0.39, p = 0.03$), V_V of autophagosomes ($r = -0.44, p = 0.005$) and lysosomes ($r = -0.38, p = 0.02$).

Renal caspase-3 demonstrated strong positive correlations with fructosamine and glycated albumin levels ($r = 0.91, p < 0.0001$ and $r = 0.89, p < 0.0001$, respectively); renal Bcl-2 correlated with these parameters negatively ($r = -0.66, p = 0.001$; $r = -0.67, p = 0.001$). We found no correlations between autophagy and apoptosis parameters and plasma creatinine levels. Meanwhile, UACR correlated negatively with renal LAMP-1 ($r = -0.76, p = 0.001$), Bcl-2 ($r = -0.81, p < 0.0001$), glomerular beclin-1 ($r = -0.44, p = 0.006$) and V_V of autophagosomes ($r = -0.47, p = 0.002$), autolysosomes ($r = -0.48, p = 0.002$) and lysosomes ($r = -0.53, p = 0.0004$) in podocytes. There were positive correlations between UACR and renal LC3B ($r = 0.50, p = 0.03$) and caspase-3 expression ($r = 0.66, p = 0.002$).

2.9. Suppression of Renal Autophagy in Diabetes: Markers and Mechanisms

The suppression of macroautophagy in *db/db* diabetic mice was documented by the reduction of glomerular IHC-staining for beclin-1 and LAMP-1, two autophagy markers, and decrease in the volume density of the autophagic compartments (autophagosomes, autolysosomes, and lysosomes) in the podocytes. We have also revealed the reduced expression of LAMP-1 in the renal cortex by western blotting. The results are in agreement with the data from other researches recorded downregulation of autophagy in diabetic kidneys [4][26][27][28][29].

The data on the enhanced expression of LC3B in the renal cortex, obtained by western blot, at first glance are out of the picture. The LC3 molecule is commonly considered a marker of autophagosomes, although it could also be detected in phagophores [23]. LC3-II is generated by the conjugation of cytosolic LC3-I to phosphatidylethanolamine on the surface of nascent autophagosomes. Accordingly, LC3-II is relatively specifically associated with autophagosomes and autolysosomes [30]. In previous studies, both a decrease in LC3-II/LC3-I ratio [31][32] and an increase in LC3-I/II expression [33] in the kidneys of *db/db* mice was reported. In podocyte cell culture, the expression of LC3-II was increased under high glucose condition [34]. Similarly, an increased LC3B/LC3A ratio was found in high-fat-diet-induced, obesity-related glomerulopathy in mice [35]. Increased LC3-II expression was previously described in tubular epithelial cells in diabetes [36]. Accumulation of LC3-II could be observed when the fusion of autophagosomes with lysosomes was interrupted by chemical agents [23]. It was demonstrated that endogenous LC3-positive puncta become larger in cells where autophagosome formation is abrogated and are prominent even when LC3-II is not formed. This occurs even with transient and incomplete inhibition of autophagosome biogenesis. This phenomenon is due to LC3-I sequestration to p62 aggregates, which accumulate when autophagy is impaired [30]. In this study, we found negative correlations between LC3B levels in

the renal cortex and the volume density of lysosomes and autolysosomes in the podocytes. Taking into account a decrease in V_V of autophagosomes and lysosomes, as well as a decrease in the renal expression of other markers of autophagy, we can assume that an increase in LC3B protein could be attributed to compromised autophagy in diabetic kidney.

The mechanisms of autophagy suppression in diabetic kidneys remain a matter of debate. Hyperglycemia was considered an autophagy suppressor in the studies with cultured podocytes [27][37], glomerular endothelial cells [38] and mesangial cells [39]. Kidney macroautophagy is regulated by mammalian target of rapamycin (mTOR) [8][40][41], AMP-activated protein kinase (AMPK)[8], SIRT1[8], Wnt/β-catenin [42], and TGF-β [43] signaling pathways. The downregulation of glomerular autophagy under hyperglycemic condition is considered a result of activation of mTOR pathway [8][44] or suppression of AMPK activity and SIRT1 signaling [39][45][46]. In our study, glomerular autophagy was related negatively to parameters of glycemic status, fructosamine and glycated albumin. As it has been demonstrated previously, advanced glycation end-products impair autophagic flux in the podocytes via mTOR activation and inhibition of the nuclear translocation and activity of the pro-autophagic transcription factor EB (TFEB) [47]. In cultured mesangial cells, advanced glycation end-products inhibited autophagy via the RAGE/STAT5 axis [48]. Therefore, hyperglycemia and related activation of non-enzymatic glycation could be a cornerstone in glomerular autophagy suppression.

In this study, we used leptin receptor-defective *db/db* mice as a model of type 2 diabetes. In these mice, we found extremely high levels of leptin, which can be explained by obesity and defective leptin signaling. The recent data indicate that leptin plays an important role in the neuroendocrine control of autophagy. It was demonstrated that intravenous or intraperitoneal administration of recombinant leptin induced autophagy in mouse peripheral tissues, including skeletal muscle, heart and liver. Moreover, leptin stimulated canonical autophagy in cultured human or mouse cell lines, a phenomenon that was coupled to the activation of AMPK, as well as the inhibition of mTOR, and this was confirmed by autophagic flux measurements [49]. Thereby, it can be assumed that the defect of leptin receptor is another factor contributing to the suppression of autophagy in *db/db* mice.

The recorded high insulin levels in plasma and severe hyperglycemia in *db/db* mice undoubtedly indicate insulin resistance, which is consistent with the literature [50]. Intrinsic interactions between insulin signaling and podocyte autophagy have been proposed. The recent findings suggested that downregulation of podocyte autophagy, observed after long-term exposure to high glucose, results from suppressed sensitivity to insulin [51]. Podocytes express all the elements of the insulin-signaling cascade, such as functional insulin receptor substrate-1 and insulin receptor, and are capable of increasing their glucose uptake when they are stimulated with insulin through glucose transporters, primarily glucose transporter type 4 (GLUT4) [52]. Glomeruli from mice with podocyte-specific knockout of GLUT4 are protected from diabetes-induced hypertrophy, mesangial expansion and albuminuria, and fail to activate the mTOR pathway [53]. The latter is activated in podocytes in diabetic conditions that decrease autophagy activity [44]. Interestingly, rapamycin-induced autophagy increased the glucose uptake in the podocytes and phosphorylated insulin receptor, which caused an increase in insulin sensitivity [27].

In addition to insulin, the activity of autophagy in diabetes can be modified by other hormonal regulators. We recorded a significant decrease in concentrations of ghrelin in the plasma of *db/db* mice. Ghrelin is a proven enhancer of autophagy, mediating its effect through the activation of AMPK [54]. Additionally, PAI-1 is considered an autophagy activator in immune cells [55]. In a model of vinyl-chloride-induced renal fibrosis, activation of autophagy corresponded to increased PAI-1 expression, while expression of PAI-1 gene in HK-2 cell culture was increased under beclin-1 siRNA exposure [56]. Nevertheless, in our study we failed to find any relationship between plasma PAI-1 levels and autophagy parameters.

Glucagon is shown to regulate autophagy by increasing the number and fragility of lysosomes and apoptotic vacuoles. These effects were primary described in liver and suggested to be organ-specific [57]. In our study, plasma glucagon levels correlated negatively with glomerular LAMP-1 and volume density of autophagosomes and lysosomes in the podocytes. These results do not prove causality; the role of glucagon in regulation of renal autophagy in diabetes requires further research.

2.10. Renal Autophagy Suppression in Pathophysiology of Diabetic Kidney Disease

In our study, the suppression of autophagy was related to severity of the classical signs of diabetic nephropathy, including mesangial expansion, GBM thickening and podocytopathy. It was suggested that the presence or activation of autophagy could play a protective role in human podocytes against high glucose-induced insulin resistance and cell injury [27]. In its turn, downregulation of autophagy is critical for podocyte survival in hyperglycemic conditions [51]. In the study with autophagy-deficient models of diabetes, blocking of autophagy in podocytes or glomerular endothelial cells caused podocytopathy and leaky glomerular filtration barriers [38]. The deficiency of autophagy activation in KkAy diabetic mice resulted in more severe proteinuria and impaired renal function [29]. In agreement with these data, we found an inverse association between morphometric parameters of glomerular autophagy, podocytopathy and albuminuria in the model of type 2 diabetes.

Apoptosis is considered another contributor to podocyte depletion in diabetes [44]. In this study, we investigated the expression of caspase-3, an apoptosis regulator, and Bcl-2, an anti-apoptotic protein, in the renal cortex of *db/db* mice. We found increased protein expression of caspase-3; meanwhile, Bcl-2 expression was diminished. Thus, we recorded reciprocal changes in the levels of autophagy and apoptosis markers. It was demonstrated recently that autophagy can regulate apoptosis following molecular interactions between the key proteins of these processes, including members of Bcl-2 family, autophagy proteins, and caspases [58]. Firstly, beclin-1 can bind to Bcl-2, which results in inhibition of autophagy [58]. Contrary, release of beclin-1 from the complex with Bcl-2 and phosphorylation of beclin-1 or Bcl-2 promote autophagy [23]. In addition, cleavage of beclin-1 by caspase-3 inactivates autophagy and promotes apoptosis [59]. This could be an explanation for the concomitant increase in caspase-3 and decrease in beclin-1 expression recorded in our study.

2.11. Reactivation of Autophagy: a Missing Link in the Mechanism of Renoprotection under SGLT2 and DPP4 Inhibition?

In this work, we demonstrate that empagliflozin and linagliptin, both as monotherapy and in combination, reactivate glomerular autophagy in *db/db* diabetic mice. In the previous studies, empagliflozin restored autophagy in the tubular cells in a model of streptozotocin-induced diabetes [20], and in the heart in a model of myocardial infarction and type 2 diabetes [60]. Canagliflozin, another SGLT2 inhibitor, activated autophagy in immune cells [31]. A negative correlation between beclin-1 and DPP4 activity was observed in patients with coronary heart disease [61]. The DPP4 inhibitors vildagliptin and sitagliptin improved cardiac function after myocardial infarction through activation of autophagy in the rodent models of diabetes [62][63]. Sitagliptin activated autophagy in cardiac muscle in Zucker diabetic fatty rats, in high-glucose cultured embryonic heart myoblastic (H9c2) cells [61] and in the liver of *ob/ob* mice, a mouse model of genetic obesity and diabetes [64].

According to our data, empagliflozin, both alone and in combination with linagliptin, contributed to preventing changes in the renal expression of apoptosis regulators, caspase-3 and Bcl-2. In the previous studies, empagliflozin have demonstrated decreased expression of pro-apoptotic proteins in cell cultures of human renal proximal tubular cells (hRPTCs) [20] and in the kidney in streptozotocin-induced diabetic rats [22]. Taking into account the close relationships in the regulation of autophagy and apoptosis, normalization of the expression of apoptosis regulators could also contribute to the reactivation of autophagy under empagliflozin treatment.

It is well known that both SGLT2 and DPP4 inhibitors generate glucose-dependent and glucose-independent effects in the targeted organs [14][15][16][17][18][19]. In our study, autophagy was reactivated by both empagliflozin and linagliptin, while glucose levels were reduced, but not to the normal range, only under empagliflozin administration. We did not observe antihyperglycemic effects of linagliptin in *db/db* mice in our study, although we proved renal benefits of the agent. Similarly, in previous studies, linagliptin ameliorated glomerular changes in *db/db* mice, despite of the lack of antihyperglycemic activity [65][66]. This suggests a glucose-independent mechanism of autophagy reactivation. It was demonstrated that vildagliptin suppressed Beclin-1–Bcl-2 interaction and increased both LC3-II protein level and autophagosomes in the diabetic heart [62]. In the liver of *ob/ob* mice, sitagliptin increased the levels of phosphorylation of AMPK and decreased those of mTOR [64]. The activation of the AMPK signaling pathway in immune cells was proven for canagliflozin [31]. Further identification of the molecular mechanisms for autophagy activation under SGLT2 and DPP4 inhibition is a challenge for future research.

3. Conclusions

SGLT2 inhibitors and DPP4 inhibitors are relatively new classes of antihyperglycemic agents with distinct renoprotective actions. Identifying the renal effects of these drugs is a challenge to modern medicine. In our study, we demonstrated that both empagliflozin, a SGLT2 inhibitor, and linagliptin, a DPP4 inhibitor, attenuate diabetic kidney disease by promoting glomerular autophagy in *db/db* mice, a model of type 2 diabetes. The acceleration of autophagy flux under empagliflozin and linagliptin treatment is associated with mitigation of glomerular sclerosis, preservation of podocyte morphology, as well as with a slowdown in the growth of albuminuria. Upregulation of beclin-1 and downregulation of caspase-3 could be suggested as possible pathways in autophagy promotion. The data provide a new explanation for the mechanism of the renoprotective action of SGLT2 inhibitors and DPP4 inhibitors in diabetic kidney disease.

4. Abbreviations

AMPK	AMP-activated protein kinase
DPP4	Dipeptidylpeptidase-4
GLUT4	Glucose transporter type 4
IHC	Immunohistochemistry
mTOR	mammalian Target of rapamycin
N _A	Numerical density
LAMP-1	Lysosomal-associated membrane protein-1
PAI-1	Plasminogen activator inhibitor-1
PFs	Foot processes
SGLT2	Sodium-glucose cotransporter-2
TEM	Transmission electron microscopy
UACR	Urinary albumin-to-creatinine ratio

V _v	Volume density
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