

Dysregulation of miRNA in Leukemia

Subjects: Cell Biology

Contributor: Luisa Anelli

Micro RNAs (miRNAs) are a class of small non-coding RNAs that have a crucial role in cellular processes such as differentiation, proliferation, migration, and apoptosis. miRNAs may act as oncogenes or tumor suppressors; therefore, they prevent or promote tumorigenesis, and abnormal expression has been reported in many malignancies. The role of miRNA in leukemia pathogenesis is still emerging, but several studies have suggested using miRNA expression profiles as biomarkers for diagnosis, prognosis, and response to therapy in leukemia.

Keywords: circulating miRNAs ; epigenetic miRNAs ; biomarkers ; dysregulated expression

1. Introduction

Micro RNAs (miRNAs) are a subset of human non-coding RNA (ncRNA) that plays an essential role in regulating gene expression, RNA maturation, and protein synthesis ^{[1][2]}. ncRNAs have long been considered as “junk” elements; they account for about 75–90% of the human genome and are classified in two main groups according to their length: small (<200 nucleotides; miRNAs) and long (>200 nucleotides; lncRNAs) ^{[2][3]}. About 19–22 nt that play a crucial role in cell growth, development, and differentiation by regulating gene expression ^{[4][5]}.

Most human miRNAs map in the introns of coding genes; some may overlap with the exons, less frequently in non-coding regions or next to the 3'-UTR sites, and they are often located in the co-transcribed clusters ^{[6][7][8]}. miRNA distribution in the human genome is not random, and some chromosomes such as 1, 2, 19, and X have higher numbers of miRNAs than others ^[9]. Several miRNAs are located next to fragile chromosomal sites or breakpoints frequently involved in leukemia rearrangements ^{[10][11]}. Most of the mature miRNAs bind to the “seed” region (5–8 nt long) in the 3'UTR of target mRNA molecules, induce silencing complex (miRISC) and act as post-transcriptional regulators causing mRNA degradation or translational repression via deadenylation, decapping, and exonucleolytic processes ^{[12][13][14][15]}.

Moreover, other regions, such as gene promoters, 5' UTR, or coding regions, may also be linked by miRNAs ^{[16][17]}. One miRNA can inhibit many different mRNA transcripts, often with similar functions, and control multiple signaling pathways ^[18]; conversely, one mRNA transcript can be targeted by several miRNAs ^[19]. miRNA expression is highly tissue-specific, some miRNAs being expressed in a specific cell or tissue type; deregulation of miRNA expression has been associated with several diseases and cancers. Almost 50% of miRNAs are located near or within genes translocated in cancer ^[20].

By down-regulating the expression of oncogenes or tumor suppressors, miRNAs can prevent or promote tumorigenesis; therefore, they may act as oncogenes (onco-miRs) or tumor suppressors ^[21]. Abnormal expression of miRNA has been reported in many malignancies, in which tumor suppressors are downregulated and oncogenic. The roles of miRNA in leukemia pathogenesis are still emerging, and several studies have suggested miRNA expression profiles using as biomarkers for diagnosis, prognosis, and response to therapy in leukemia. These aspects will be discussed in the present review considering their mechanisms of action and the miRNAs most frequently deregulated in myeloid or lymphoid leukemias.

2. Epigenetic miRNAs (epimiRNAs)

However, some other miRNAs, defined as epigenetic-miRNAs (epi-miRNAs or epi-miRs), can directly or indirectly influence the expression of known epigenetic regulators such as DNA methyltransferases (DNMTs), HDACs, and components of PRC ^{[22][23]}. The first identified example of epi-miRNAs was the miR-29 family members (29a, 29b, and 29c); they regulate the expression of DNMT3A and DNMT3B in lung cancer and acute myeloid leukemia (AML). In vitro experiments showed that the exogenous introduction of these miRNAs in lung cancer or AML cell lines led to the reversion of the neoplastic phenotype by inhibiting different DNA methyltransferases and the consequent hypomethylation and reactivation of other target genes ^{[24][25][26]}. Increasing the expression of this epi-miRNA, apoptosis and differentiation are stimulated in leukemic cells due to inhibition of the AML1-ETO and other epigenetic regulators ^[27].

3. miRNAs and lncRNAs Interaction

Several data showed that many lncRNAs have multiple miRNA response elements (MRE), that are regions mediating reciprocal interaction; lncRNAs could therefore influence miRNA expression as they can act as sponges in both normal and cancer cells, determining several interactive networks. [28] HOTAIRM1 also showed an essential role in the pathogenesis of AML cases with the t(15;17) translocation, acting as a microRNA sponge sequestering several miRNAs [29]. Several recent papers reported different miRNA and lncRNA interactions in AML, such as LINC01018 and miR-499a-5p, with LINC01018 acting as a sponge of miR-499a-5p, which in turn targets PDCD4 gene [30]. Both LINC01018 overexpression and miR-499a-5p knockdown suppressed AML cell proliferation and induced apoptosis, whereas miR-499a-5p transfection and silencing of PDCD4 reversed these effects [30].

Moreover, lncRNA and miRNA interaction seem to be frequently involved in chemoresistance mechanisms in AML, as for lncRNA-UCA1/miR-125a/hexokinase 2 or HOXA-AS2/miR-520c-3p [31][32]. UCA1 expression was found upregulated following adriamycin (ADR)-based chemotherapy, and UCA1 knockdown. On the other hand, lncRNA HOXA-AS2 was significantly upregulated in BM cells from AML cases after ADR-based chemotherapy, and its knockdown inhibited cell proliferation and induced apoptosis. These data showed that both lncRNAs UCA1 and HOXA-AS2 may represent useful therapeutic targets for overcoming ADR-chemoresistance in AML [32].

Another interesting example is represented by the network between lncRNAs and MYC expression in both myeloid and lymphoid malignancies, which contributes to inhibiting apoptosis, stimulates cell proliferation, induces genomic instability and resistance to therapy [33]. In AML, lncRNA CCAT1 interacts and inhibits miR-155, whose targets are MYC, AP-1, FOS, and c-JUN, which regulate myeloid cell differentiation [34][35]. CCAT1 sponges miR-155 and stimulates MYC expression; interestingly, in previous studies, CCAT1 has been reported to be activated by MYC, suggesting the existence of a MYC/CCAT1/miR-155 feedback loop [35]. Other examples of lncRNA and miRNA interaction include MEG3/miR-147 and UCA1/miR-16 that have been detected in CML and are considered possible therapeutic targets in blast crisis or imatinib (IM) resistance, respectively [36][37]. Moreover, in lymphoma, the interaction between HOTAIR and miR-148b regulates apoptosis and the cell cycle progression of B cells.

4. miRNAs Expression Profile in Leukemia

as different miRNAs are expressed in a specific cell or tissue type; miRNAs play a crucial role in regulating gene expression during normal hematopoiesis, acting on the self-renewal capacity of hematopoietic stem cell (HSC) and the differentiation of lineage-restricted progenitors [38][39]. Thirty-three miRNAs were identified as specifically expressed in CD34+ hematopoietic stem-progenitor cells (HSPCs) [40]. Some miRNAs such as miRNA-17, -24, 146, -155, -128, and -181, were found to be expressed in early hematopoietic cells whereas other miRNAs, such as miRNA155, were able to control specific processes, as myelopoiesis and erythropoiesis [40], or as miR-34a and miR-17-92 clusters that have an essential role in the pro- to pre-B-cell differentiation [41]. miR-181 cluster also plays a critical role in the differentiation of hematopoietic cells as T, B, and natural killer cells or megakaryocytes [42].

Deregulation of miRNA expression has been associated with several diseases and cancers, and specific miRNA expression profiles have been reported in several hematologic neoplasms [43][44]. Microarray technologies, next-generation sequencing (NGS), and quantitative real-time PCR (qRT-PCR) are the most valuable methodologies for identifying reproducible dysregulated expression profiles in specific leukemia types (**Figure 1**). Unlike microarray analysis, NGS by miRNA-Seq shows high sensitivity in discovering new miRNAs and detecting whole-genome miRNA transcripts (miRNoma) with no need for previous selection. Several of these miRNAs may potentially be used as prognostic biomarkers, either as single miRNAs or as miRNA expression profiles.

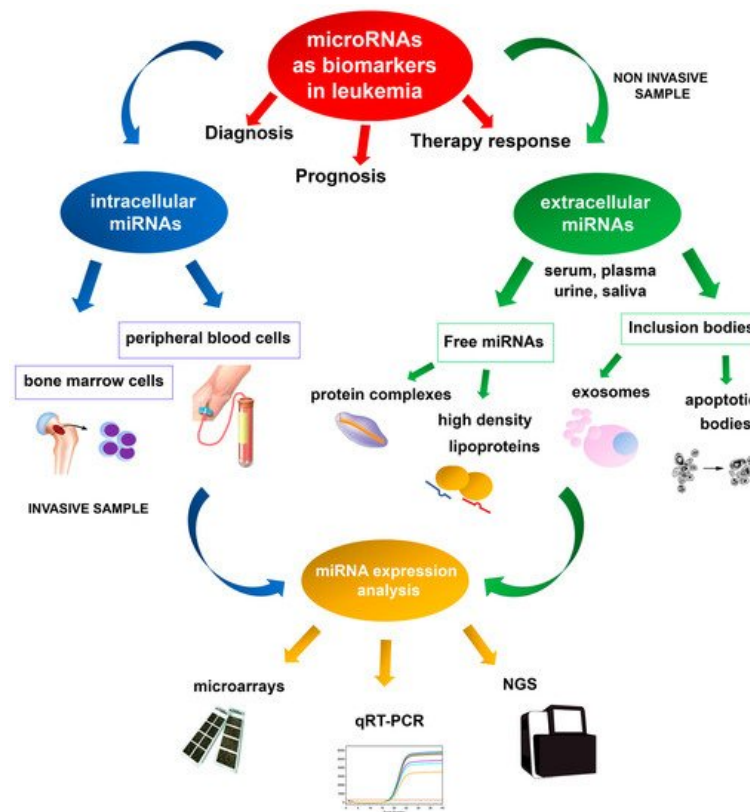


Figure 1. Schematic representation of miRNA employment as biomarkers in leukemia, based on invasive and non-invasive approaches for expression analysis.

The first evidence of the involvement of miRNAs in leukemogenesis was reported in a study of CLL aimed at characterizing a 30 Kb deletion on 13q14; the study did not identify any protein-coding genes, but a cluster of two miRNAs, miR-15a and miR-16-1, was found to be deleted or down-regulated in most of the CLL cases examined [45]. Moreover, these miRNAs have also been shown to be frequently deregulated in other solid and hematological cancer types [46][47][48], and can modulate cell cycle progression and induce apoptosis by targeting pivotal genes such as BCL2, MCL1, CCND1, or WNT3A [49][50][51].

Different studies reported specific miRNA expression profiles that distinguish between AML and acute lymphoblastic leukemia (ALL), some miRNAs being reported in different studies, such as miR-23a, miR-27a/b, miR-128a, miR-128b, miR-221, miR-222, miR-223, and let-7b [52][53]. In a study by Mi et al., four miRNAs were sufficient to distinguish between AML and ALL with an accuracy of greater than 95%, as let-7b/miR-223 being significantly upregulated and miR-128a/miR-128b downregulated in AML comparing to ALL [52]. Together, the above work showed that these identified miRNAs could be new potential markers for ALL and AML classification and diagnosis [54].

One of the miRNAs most frequently involved in AML pathogenesis is miR-155, which is also commonly overexpressed in B-cell neoplasms, where it is considered an oncogenic driver of B-cell lymphoma [55]. This miRNA shows a contrasting dose-dependent function as onco-miRNA or tumor suppressor according to the expression levels. In short, a high level of expression is correlated with the antitumor effect and inhibition of AML cell proliferation. In contrast, an intermediate-low level of expression induces oncogenesis [56] and has been associated with poor prognosis in AML irrespective of specific cytogenetic or molecular aberrations [56].

Several studies have identified specific miRNA signatures in different AML subgroups defined by the classification of myeloid neoplasms [57] (Table 1), and a correlation was detected with cytogenetics alterations [58][59][60], prognosis, and clinical characteristics [61].

Table 1. miRNAs most frequently involved in acute leukemia pathogenesis.

AML	miRNAs	Expression Data	References
t(15;17)(q24;q21) <i>PML-RARA</i>	miR-127, miR-154, miR-154*, miR-299, miR-323, miR-368, miR-370, miR-382, miR-134, miR-376a, miR-127, miR-299-5p, miR-323, miR-224	Upregulation of miRNAs mapping in in the 14q32 imprinted region	[58][59][60]

AML	miRNAs	Expression Data	References
t(8;21) (q22;q22.1) <i>RUNX1-RUNX1T1</i>	miR-126, miR-146a, miR-133a, let-7b, let-7c	Overexpression of miR-126 and miR-146a; downregulation of miR-133a let-7b and let-7c	[59][60][62]
inv(16) (p13;q22) <i>CBFB-MYH11</i>	miR-99a, miR-100, miR-224, miR-126	miRNA signature sometimes overlapping with t(8;21) AML	[58][59][60]
<i>FLT3-ITD</i>	miR-155, miR-10a, miR-10b	Upregulation	[59][63]
Mutated <i>NPM1</i>	miR-10a and b, let-7, miR-29, miR-15a/16-1, miR-17-18a-19a-20a, miR-204 and miR-128a	Upregulation of miR-10a and b, let-7, miR-29, miR-15a/16-1, and miR-17-18a-19a-20a; downregulation of miR-204 and miR-128a	[63][64][65]
<i>MLL</i> rearranged	miR-34b, miR-15a, let-7 family, miR-196, miR-10a, miR-331, and miR-340, miR-17-92, miR-126, -126*, -224, -368, -382, 17-5p, and -20a	Downregulation of miR-34b, miR-15a, let-7, and miR-196; upregulation of miR-17-92, miR-126, -126*, -224, -368, -382, 17-5p, and -20a	[58][60][66]
Mutated <i>IDH2</i>	miR-125b, miR-1, miR-133, miR-194-1, miR-526, miR-520a-3p, miR-548b	Upregulation of miR-125b, miR-1 and miR-133; downregulation of miR-194-1, miR-526, miR-520a-3p, and miR-548b	[67]
Mutated <i>RUNX1</i>	miR-223, let-7, miR-211, miR-220, miR-595	Downregulation of miR-223 and let-7; upregulation of miR-211, miR-220, and miR-595	[68]
Normal karyotype	miR-181a/b, miR-124, miR-128-1, miR-194, miR-219-5p, miR-220a, miR-320	Upregulation	[69]
ALL	miRNAs	Expression Data	References
<i>MLL</i> rearranged	miR-128b, miR-708, let-7b	Downregulation	[52][70]
t(12;21) (p13;q22) <i>ETV6-RUNX1</i>	miR-100, miR-125b, miR-99a, miR-126, let-7c, miR-181a	Upregulation of miR-100, miR-125b, miR-99a, miR-126, let-7c; downregulation of miR-181a	[71][72][73][74]
t(9;22)(q34;q11) <i>BCR-ABL1</i>	miR-125b-2, miR-203	Overexpression of miR-125b-2; downregulation of miR-203	[75]
hyperdiploid karyotype	miR-222, miR-223, miR-374, miR-660, miR-98 and miR-511	Upregulation	[71]
t(1;19)(q23;p13) <i>TCF3-PBX1</i>	miR-126, miR-146a, miR-511, miR-545, miR-365, miR-24, miR-30d, miR193, miR-181, miR-708	Downregulation of miR-126, miR-146a, miR-511, miR-545, miR-365, miR-24, miR-30d, miR193; upregulation of miR-181, miR-708	[72][76]
T-ALL	miR-17-92, miR-708, miR-196b, miR-128, miR-181, miR-29, miR-150, miR-99a and miR-708	Overexpression of miR-17-92, miR-708, miR-196b, miR-128, miR-181; Downregulation of miR-29	[77][78][79]

In AML with the t(15;17) translocation, the upregulation of miRNAs located in the 14q32 imprinted domain (miR-127, miR-154, miR-154*, miR-299, miR-323, miR-368, and miR-370) was reported in a first study [58], whereas a set of partially overlapping strongly upregulated microRNAs (miR-382, miR-134, miR-376a, miR-127, miR-299–5p, and miR-323) was described by Jongen-Lavrencic et al. [59]. In another study, the overexpression of miR-224, miR-368, and miR-382 was detected [60].

AML with t(8;21) showed high miR-126/126* [60] and miR-146a expression with decreased miR-133a [58]; other evidence showed a set of down-regulated miRNAs, including two members of a known tumor suppressor microRNA family, let-7b and let-7c [59], that was previously found to be involved in other cancers [62].

In AML with inv(16), a high level of miR-99a, miR-100, and miR-224 expression or of miR-126/126* was observed by different investigators [58][60]. Overall, AML with inv(16) showed a miRNA signature that sometimes overlapped with t(8;21) AML; this is not surprising as both these AML subtypes belong to the CBF group [59].

In AML with FLT3-internal tandem duplication (FLT3-ITD), miR-155, miR-10a, and miR-10b were found to be upregulated [59][63].

In AML with NPM1 mutations, a specific miRNA-based expression signature was revealed with upregulation of miR-10a and b, members of the let-7 and miR-29 families, miR-15a/16-1 and miR-17-18a-19a-20a clusters, and downregulation of miR-204 and miR-128a, predicted to target HOX genes known to be upregulated in NPM1-mutated AML [63][64]. A further

study was based on an integrative approach based on both microRNA and gene expression profiles. Several interesting microRNA-target mRNA interactions, such as IRF2-miR-20a, KIT-miR-20a, and MN1-miR-15a, were identified. This study also showed a deregulated expression of tumor suppressor microRNAs, such as miR-29a and miR-30c, that seem to be involved in sensitivity to therapy [65].

In AML with balanced 11q23 translocations and KMT2A (MLL1) rearrangements, the downregulation of several tumor suppressor miRNAs such as miR-34b, miR-15a, the let-7 family, and miR-196, targeting several known target genes such as CDK4 and CCNE2, BCL2, RAS, and HOX genes was reported; other evidence showed that AML with the MLL rearrangement were characterized by the loss of miR-10a, miR-331, and miR-340 expression [58]. Other authors revealed the overexpression of miRNAs from polycistronic cluster miR-17-92, and a minimal class predictor with only seven miRNAs (miR-126, -126*, -224, -368, -382, 17-5p, and -20a) was identified [60]. Leukemic cells with higher expression levels of miR-17-92 showed arrested differentiation and increased proliferation in concomitance with reduced expression of p21, a downstream target of polycistronic miR-17-92 [66].

In AML with IDH2 mutations, a specific signature for R172 IDH2-AML was identified with the upregulation of miR-125b, which targets the TP53 gene and inhibits myeloid differentiation, miR-1 and miR-133, involved in embryonic stem-cell differentiation, and downregulation of miR-194-1, miR-526, miR-520a-3p, and miR-548b, not previously associated with normal hematopoiesis or AML [67].

In AML with RUNX1 mutations, miR-223 and two members of the let-7 tumor suppressor family were downregulated, whereas three miRNAs with an unknown role in leukemogenesis, miR-211, miR-220, and miR-595, were found to be upregulated [68].

MiRNA expression analysis performed on normal karyotype AML (CN-AML) revealed a prognostic relevant miRNA signature, as the upregulation of miR-181a/b and miR-124, miR-128-1, miR-194, miR-219-5p, miR-220a, and miR-320 was associated with a low or increased risk of failure to achieve complete remission (CR), of relapse or death, respectively [69]. Some miR-181 putative targets were genes involved in innate immunity, encoding interleukins, caspases, and Toll-like receptors [69].

miRNAs deregulation is a common event in B- and T-cell malignancies in which they act as either oncomiRs or tumor suppressors [41][71][80]. ALL [71]; miR-155, that induces pre-B cells clonal expansion and is overexpressed in different pediatric ALL subtypes [55]; miR-128b, that allows differentiation with AML cases and is downregulated in ALL with the MLL-AF4 translocation [52][70]. Moreover, miR-203 is silenced through epigenetic mechanisms [75]; it has been shown that by enhancing miR-203 expression, BCR-ABL1 transcript level is reduced, cell proliferation is inhibited, and resistance to TKI can be overcome [81]. Regarding ALL with hyperdiploid karyotype, miR-222, miR-223, miR-374, miR-660, miR-98, and miR-511 were found upregulated, probably as a consequence of their mapping location in chromosomes X and 10 that are frequently present as extra copies [71].

miR-708 was found upregulated when comparing T-ALL with healthy individuals and downregulated when comparing T-ALL with different leukemia subtypes [71]; miR-708 downregulation is a poor prognostic factor of T-ALL, as it induces an increased expression of CD47 and promotes the evasion of leukemic cells from macrophage-mediated phagocytosis [82]. Low levels of miR-150 expression were associated with poor prognosis as correlating with relapse, high-risk and high WBCs at diagnosis; the association of miR-150 downregulation and poor prognosis was also revealed in other hematologic malignancies as AML, CLL, and different lymphoma subtypes [41]. Regarding miR-99, different studies revealed that upregulation and downregulation were correlated with favorable and poor prognosis, respectively [83][84]. Finally, miR-708 upregulation was associated with a good prognosis, as lower relapse risk, low WBC count, and better overall survival were detected in ALL cases; on the contrary, miR-708 downregulation was revealed in poor prognosis subtypes as T-ALL and cases with MLL gene rearrangement [71][84].

Moreover, recent data revealed that miR-300 is a tumor suppressor miRNA inducing quiescence in CML leukemic stem cells (LSCs) [85], and that miR-126-3p influences both quiescence and self-renewal of CML LSCs [86] (Table 2). Another recent study showed a global decrease in microRNA levels in LSC-enriched CD34+CD38-CD26+ and HSC from CML-CP patients compared to those from healthy donors HSC [87]. Ederne San José-Enériz et al. identified a group of 19 miRNAs that may predict clinical resistance to IM in patients with newly diagnosed CML [88]. Another study revealed that miR-30 induces the degradation of BCR/ABL1 mRNAs by binding directly to their 3'UTR, which was downregulated in CML patients less responsive to IM [89].

Table 2. miRNAs most frequently involved in chronic leukemia pathogenesis.

CML	miRNAs	Expression Data	References
pathogenesis	miR-155, miR-300, miR-126-3p	Upregulation of miR-155, miR-300; Downregulation of miR-126-3p	[90][85][86]
TKI resistance	miR-30, miR-26a, miR-29c, miR-130b, miR-146a, miR-142-5, miR-365a-3p, miR-153-3p, miR-185	Downregulation	[89][91][92][93]
TKI discontinuation	miR-148b and miR-215	Downregulation	[89][91][92][93] [94]
CLL	miRNAs	Expression Data	References
13q deletion	miR-15a/miR-16-1 cluster, miR-34a/miR-34b/miR-34c cluster	Downregulation of miR-15 and miR-16; upregulation of miR-34 cluster	[45][95][96][97]
Trisomy 12	miR-181a	Upregulation	[96]
17p deletion	miR-15a, miR-21, miR-34a, miR-155, and miR-181b	Upregulation of miR-21, miR-34a, miR-155, miR-181b	[98]
11q deletion	miR-34b/miR-34c cluster	Downregulation	[97]

In contrast, another recent investigation did not find any significant differences in miRNA expression patterns between TKI responder and non-responder patients [99]. Different studies analyzing miRNA expression profiles at CML diagnosis and in cases showing resistance considered only a small series of CML cases and produced contrasting results. More extensive analyses are therefore needed to verify whether the aforementioned miRNAs may be used to discriminate between the responders and non-responders among CML patients as well as the predictive biomarkers for TKI resistance. The miRNA involvement has also been investigated in CML to identify possible biomarkers for TKI discontinuation; two miRNAs, miR-148b and miR-215, showed downregulated expression in CML cases with successful IM discontinuation, suggesting that these miRNAs may contribute to immune surveillance in CML patients showing safe TKI discontinuation [94][100].

miRNAs are involved in the regulation of B lymphocyte development and can be altered in different B-cell malignancies. Other miRNAs frequently involved in CLL and other B-cell malignancies are miR-150, miR-155, and the miR-17-92 cluster; they regulate the expression of crucial transcription factors involved in normal or malignant B-cells development [101][102][103]. miR-150 is considered a lymphopoietic-specific miRNA, as its overexpression inhibits the pro-B to pre-B transition, probably by targeting forkhead box P1 (FOXP1) and GRB2-associated binding protein 1 (GAB1), an important transcription factor involved in B-cell differentiation [104][105][106]. It has been shown that the treatment with “BCR inhibitor” ibrutinib induces the downregulation of several miRNAs involved in B cell activation, as miR-22, miR-34a, miR-146b, and miR-181b, whereas the expression of several target genes including ARID1BATM, HDAC1, CYLD, FOXP1, IBTK, ARID2, PTEN, and SMAD4 is activated [107].

5. Conclusions

Increasing evidence shows that miRNAs have a role as potential biomarkers in leukemia, allowing a better subtype classification, prognostic stratification, and predicting the response to treatment. An ideal biomarker should have a specific expression in patients compared to normal controls, should allow early diagnosis and minimal residual disease monitoring during the follow-up, with a possible non-invasive, simple and accurate detection method. About this, miRNAs can be extracted and analyzed from peripheral blood or bone marrow cells of leukemia patients, while circulating miRNAs can be examined by non-invasive methods based on liquid biopsy analysis and usually show expression profiles overlapping with neoplastic cells.

References

- Morris, K.V.; Mattick, J.S. The rise of regulatory RNA. *Nat. Rev. Genet.* 2014, 15, 423–437.
- Palazzo, A.F.; Lee, E.S. Non-coding RNA: What is functional and what is junk? *Front. Genet.* 2015, 5.
- Collins, L.J.; Penny, D. The RNA infrastructure: Dark matter of the eukaryotic cell? *Trends Genet.* 2009, 25, 120–128.
- Bartel, D.P. MicroRNAs: Genomics, Biogenesis, Mechanism, and Function. *Cell* 2004, 116, 281–297.
- Liu, M.; Zhou, K.; Cao, Y. MicroRNA-944 affects cell growth by targeting EPHA7 in non-small cell lung cancer. *Int. J. Mol. Sci.* 2016, 17, 1493.

6. Rodriguez, A.; Griffiths-Jones, S.; Ashurst, J.L.; Bradley, A. Identification of mammalian microRNA host genes and transcription units. *Genome Res.* 2004, 14, 1902–1910.
7. Altuvia, Y.; Landgraf, P.; Lithwick, G.; Elefant, N.; Pfeffer, S.; Aravin, A.; Brownstein, M.J.; Tuschl, T.; Margalit, H. Clustering and conservation patterns of human microRNAs. *Nucleic Acids Res.* 2005, 33, 2697–2706.
8. Albano, F.; Anelli, L.; Zagaria, A.; Coccaro, N.; Casieri, P.; Minervini, A.; Specchia, G. SETBP1 and miR-4319 dysregulation in primary myelofibrosis progression to acute myeloid leukemia. *J. Hematol. Oncol.* 2012, 5, 1–4.
9. Grimwood, J.; Gordon, L.A.; Olsen, A.; Terry, A.; Schmutz, J.; Lamerdin, J.; Hellsten, U.; Goodstein, D.; Couronne, O.; Tran-Gyam, M.; et al. The DNA sequence and biology of human chromosome 19. *Nature* 2004, 428, 529–535.
10. Calin, G.A.; Sevignani, C.; Dumitru, C.D.; Hyslop, T.; Noch, E.; Yendamuri, S.; Shimizu, M.; Rattan, S.; Bullrich, F.; Negrini, M.; et al. Human microRNA genes are frequently located at fragile sites and genomic regions involved in cancers. *Proc. Natl. Acad. Sci. USA* 2004, 101, 2999–3004.
11. Starczynowski, D.T.; Morin, R.; McPherson, A.; Lam, J.; Chari, R.; Wegrzyn, J.; Kuchenbauer, F.; Hirst, M.; Tohyama, K.; Humphries, R.K.; et al. Genome-wide identification of human microRNAs located in leukemia-associated genomic alterations. *Blood* 2011, 117, 595–607.
12. Tan, L.; Yu, J.T.; Tan, L. Causes and Consequences of MicroRNA Dysregulation in Neurodegenerative Diseases. *Mol. Neurobiol.* 2015, 51, 1249–1262.
13. Pfaff, J.; Meister, G. Argonaute and GW182 proteins: An effective alliance in gene silencing. *Biochem. Soc. Trans.* 2013, 41, 855–860.
14. Fabian, M.R.; Sonenberg, N.; Filipowicz, W. Regulation of mRNA translation and stability by microRNAs. *Annu. Rev. Biochem.* 2010, 79, 351–379.
15. Chen, C.Y.A.; Shyu, A. Bin Mechanisms of deadenylation-dependent decay. *Wiley Interdiscip. Rev. RNA* 2011, 2, 167–183.
16. Lytle, J.R.; Yario, T.A.; Steitz, J.A. Target mRNAs are repressed as efficiently by microRNA-binding sites in the 5' UTR as in the 3' UTR. *Proc. Natl. Acad. Sci. USA* 2007, 104, 9667–9672.
17. Forman, J.J.; Legesse-Miller, A.; Collier, H.A. A search for conserved sequences in coding regions reveals that the let-7 microRNA targets Dicer within its coding sequence. *Proc. Natl. Acad. Sci. USA* 2008, 105, 14879–14884.
18. El-Daly, S.M.; Abba, M.L.; Patil, N.; Allgayer, H. MiRs-134 and-370 function as tumor suppressors in colorectal cancer by independently suppressing EGFR and PI3K signalling. *Sci. Rep.* 2016, 6.
19. Bartel, D.P. Metazoan MicroRNAs. *Cell* 2018, 173, 20–51.
20. Laganà, A.; Russo, F.; Sismeiro, C.; Giugno, R.; Pulvirenti, A.; Ferro, A. Variability in the incidence of miRNAs and genes in fragile sites and the role of repeats and CpG islands in the distribution of genetic material. *PLoS ONE* 2010, 5.
21. Lu, J.; Getz, G.; Miska, E.A.; Alvarez-Saavedra, E.; Lamb, J.; Peck, D.; Sweet-Cordero, A.; Ebert, B.L.; Mak, R.H.; Ferrando, A.A.; et al. MicroRNA expression profiles classify human cancers. *Nature* 2005, 435, 834–838.
22. Iorio, M.V.; Piovani, C.; Croce, C.M. Interplay between microRNAs and the epigenetic machinery: An intricate network. *Biochim. Biophys. Acta Gene Regul. Mech.* 2010, 1799, 694–701.
23. Sandoval, J.; Esteller, M. Cancer epigenomics: Beyond genomics. *Curr. Opin. Genet. Dev.* 2012, 22, 50–55.
24. Amodio, N.; Rossi, M.; Raimondi, L.; Pitari, M.R.; Botta, C.; Tagliaferri, P.; Tassone, P. miR-29s: A family of epi-miRNAs with therapeutic implications in hematologic malignancies. *Oncotarget* 2015, 6, 12837–12861.
25. Fabbri, M.; Garzon, R.; Cimmino, A.; Liu, Z.; Zanesi, N.; Callegari, E.; Liu, S.; Alder, H.; Costinean, S.; Fernandez-Cymering, C.; et al. MicroRNA-29 family reverts aberrant methylation in lung cancer by targeting DNA methyltransferases 3A and 3B. *Proc. Natl. Acad. Sci. USA* 2007, 104, 15805–15810.
26. Garzon, R.; Liu, S.; Fabbri, M.; Liu, Z.; Heaphy, C.E.A.; Callegari, E.; Schwind, S.; Pang, J.; Yu, J.; Muthusamy, N.; et al. MicroRNA-29b induces global DNA hypomethylation and tumor suppressor gene reexpression in acute myeloid leukemia by targeting directly DNMT3A and 3B and indirectly DNMT1. *Blood* 2009, 113, 6411–6418.
27. Li, Y.; Gao, L.; Luo, X.; Wang, L.; Gao, X.; Wang, W.; Sun, J.; Dou, L.; Li, J.; Xu, C.; et al. Epigenetic silencing of microRNA-193a contributes to leukemogenesis in t(8;21) acute myeloid leukemia by activating the PTEN/PI3K signaling pathway. *Blood* 2013, 121, 499–509.
28. Xing, C.-Y.; Hu, X.-Q.; Xie, F.-Y.; Yu, Z.-J.; Li, H.-Y.; Zhou, B.; Wu, J.-B.; Tang, L.-Y.; Gao, S.-M. Long non-coding RNA HOTAIR modulates c-KIT expression through sponging miR-193a in acute myeloid leukemia. *FEBS Lett.* 2015, 589, 1981–1987.

29. Chen, Z.H.; Wang, W.T.; Huang, W.; Fang, K.; Sun, Y.M.; Liu, S.R.; Luo, X.Q.; Chen, Y.Q. The lncRNA HOTAIRM1 regulates the degradation of PML-RARA oncoprotein and myeloid cell differentiation by enhancing the autophagy pathway. *Cell Death Differ.* 2017, 24, 212–224.
30. Zhou, H.; Shi, P.; Jia, X.; Xue, Q. Long non-coding RNA LINC01018 inhibits the progression of acute myeloid leukemia by targeting miR-499a-5p to regulate PDCD4. *Oncol. Lett.* 2021, 22, 541.
31. Zhang, Y.; Liu, Y.; Xu, X. Knockdown of lncRNA-UCA1 suppresses chemoresistance of pediatric AML by inhibiting glycolysis through the microRNA-125a/hexokinase 2 pathway. *J. Cell. Biochem.* 2018, 119, 6296–6308.
32. Dong, X.; Fang, Z.; Yu, M.; Zhang, L.; Xiao, R.; Li, X.; Pan, G.; Liu, J. Knockdown of long noncoding RNA HOXA-AS2 suppresses chemoresistance of acute myeloid leukemia via the miR-520c-3p/S100A4 Axis. *Cell. Physiol. Biochem.* 2018, 51, 886–896.
33. Benetatos, L.; Benetatos, A.; Vartholomatos, G. Long non-coding RNAs and MYC association in hematological malignancies. *Ann. Hematol.* 2020, 99, 2231–2242.
34. Palma, C.A.; Al Sheikh, D.; Lim, T.K.; Bryant, A.; Vu, T.T.; Jayaswal, V.; Ma, D.D.F. MicroRNA-155 as an inducer of apoptosis and cell differentiation in Acute Myeloid Leukaemia. *Mol. Cancer* 2014, 13.
35. Chen, L.X.; Wang, W.; Cao, L.; Li, Z.; Wang, X. Long non-coding RNA CCAT1 acts as a competing endogenous RNA to regulate cell growth and differentiation in acute myeloid leukemia. *Mol. Cells* 2016, 39, 330–336.
36. Li, Z.-Y.; Yang, L.; Liu, X.-J.; Wang, X.-Z.; Pan, Y.-X.; Luo, J.-M. The Long Noncoding RNA MEG3 and its Target miR-147 Regulate JAK/STAT Pathway in Advanced Chronic Myeloid Leukemia. *EBioMedicine* 2018, 34, 61–75.
37. Xiao, Y.; Jiao, C.; Lin, Y.; Chen, M.; Zhang, J.; Wang, J.; Zhang, Z. lncRNA UCA1 contributes to imatinib resistance by acting as a ceRNA against miR-16 in chronic myeloid leukemia cells. *DNA Cell Biol.* 2017, 36, 18–25.
38. O'Connell, R.M.; Zhao, J.L.; Rao, D.S. MicroRNA function in myeloid biology. *Blood* 2011, 118, 2960–2969.
39. Zhang, L.; Sankaran, V.G.; Lodish, H.F. MicroRNAs in erythroid and megakaryocytic differentiation and megakaryocyte-erythroid progenitor lineage commitment. *Leukemia* 2012, 26, 2310–2316.
40. Georgantas, R.W.; Hildreth, R.; Morisot, S.; Alder, J.; Liu, C.G.; Heimfeld, S.; Calin, G.A.; Croce, C.M.; Civin, C.I. CD34 + hematopoietic stem-progenitor cell microRNA expression and function: A circuit diagram of differentiation control. *Proc. Natl. Acad. Sci. USA* 2007, 104, 2750–2755.
41. Musilova, K.; Mraz, M. MicroRNAs in B-cell lymphomas: How a complex biology gets more complex. *Leukemia* 2015, 29, 1004–1017.
42. Chen, C.Z.; Li, L.; Lodish, H.F.; Bartel, D.P. MicroRNAs Modulate Hematopoietic Lineage Differentiation. *Science* 2004, 303, 83–86.
43. Benetatos, L.; Vartholomatos, G. Deregulated microRNAs in multiple myeloma. *Cancer* 2012, 118, 878–887.
44. Ward, B.P.; Tsongalis, G.J.; Kaur, P. MicroRNAs in chronic lymphocytic leukemia. *Exp. Mol. Pathol.* 2011, 90, 173–178.
45. Calin, G.A.; Dumitru, C.D.; Shimizu, M.; Bichi, R.; Zupo, S.; Noch, E.; Aldler, H.; Rattan, S.; Keating, M.; Rai, K.; et al. Frequent deletions and down-regulation of micro-RNA genes miR15 and miR16 at 13q14 in chronic lymphocytic leukemia. *Proc. Natl. Acad. Sci. USA* 2002, 99, 15524–15529.
46. Zhang, L.; Zhou, L.; Shi, M.; Kuang, Y.; Fang, L. Downregulation of miRNA-15a and miRNA-16 promote tumor proliferation in multiple myeloma by increasing CABIN1 expression. *Oncol. Lett.* 2018, 15, 1287–1296.
47. Bhattacharya, R.; Nicoloso, M.; Arvizo, R.; Wang, E.; Cortez, A.; Rossi, S.; Calin, G.A.; Mukherjee, P. MiR-15a and MiR-16 control Bmi-1 expression in ovarian cancer. *Cancer Res.* 2009, 69, 9090–9095.
48. Xu, F.; Zhang, X.; Lei, Y.; Liu, X.; Liu, Z.; Tong, T.; Wang, W. Loss of repression of HuR translation by miR-16 may be responsible for the elevation of HuR in human breast carcinoma. *J. Cell. Biochem.* 2010, 111, 727–734.
49. Linsley, P.S.; Schelter, J.; Burchard, J.; Kibukawa, M.; Martin, M.M.; Bartz, S.R.; Johnson, J.M.; Cummins, J.M.; Raymond, C.K.; Dai, H.; et al. Transcripts Targeted by the MicroRNA-16 Family Cooperatively Regulate Cell Cycle Progression. *Mol. Cell. Biol.* 2007, 27, 2240–2252.
50. Liu, Q.; Fu, H.; Sun, F.; Zhang, H.; Tie, Y.; Zhu, J.; Xing, R.; Sun, Z.; Zheng, X. miR-16 family induces cell cycle arrest by regulating multiple cell cycle genes. *Nucleic Acids Res.* 2008, 36, 5391–5404.
51. Cimmino, A.; Calin, G.A.; Fabbri, M.; Iorio, M.V.; Ferracin, M.; Shimizu, M.; Wojcik, S.E.; Aqeilan, R.I.; Zupo, S.; Dono, M.; et al. miR-15 and miR-16 induce apoptosis by targeting BCL2. *Proc. Natl. Acad. Sci. USA* 2005, 102, 13944–13949.
52. Mi, S.; Lu, J.; Sun, M.; Li, Z.; Zhang, H.; Neilly, M.B.; Wang, Y.; Qian, Z.; Jin, J.; Zhang, Y.; et al. MicroRNA expression signatures accurately discriminate acute lymphoblastic leukemia from acute myeloid leukemia. *Proc. Natl. Acad. Sci. US*

53. Wang, Y.; Li, Z.; He, C.; Wang, D.; Yuan, X.; Chen, J.; Jin, J. MicroRNAs expression signatures are associated with line age and survival in acute leukemias. *Blood Cells Mol. Dis.* 2010, 44, 191–197.
54. Zhi, F.; Cao, X.; Xie, X.; Wang, B.; Dong, W.; Gu, W.; Ling, Y.; Wang, R.; Yang, Y.; Liu, Y. Identification of Circulating MicroRNAs as Potential Biomarkers for Detecting Acute Myeloid Leukemia. *PLoS ONE* 2013, 8, e56718.
55. Eis, P.S.; Tam, W.; Sun, L.; Chadburn, A.; Li, Z.; Gomez, M.F.; Lund, E.; Dahlberg, J.E. Accumulation of miR-155 and BCL2 RNA in human B cell lymphomas. *Proc. Natl. Acad. Sci. USA* 2005, 102, 3627–3632.
56. Narayan, N.; Morenos, L.; Phipson, B.; Willis, S.N.; Brumatti, G.; Eggers, S.; Lalaoui, N.; Brown, L.M.; Kosasih, H.J.; Bartolo, R.C.; et al. Functionally distinct roles for different miR-155 expression levels through contrasting effects on gene expression, in acute myeloid leukaemia. *Leukemia* 2017, 31, 808–820.
57. Arber, D.A.; Orazi, A.; Hasserjian, R.; Thiele, J.; Borowitz, M.J.; Le Beau, M.M.; Bloomfield, C.D.; Cazzola, M.; Vardiman, J.W. The 2016 revision to the World Health Organization classification of myeloid neoplasms and acute leukemia. *Blood* 2016, 127, 2391–2405.
58. Dixon-McIver, A.; East, P.; Mein, C.A.; Cazier, J.B.; Molloy, G.; Chaplin, T.; Lister, T.A.; Young, B.D.; Debernardi, S. Distinctive patterns of microRNA expression associated with karyotype in acute myeloid leukaemia. *PLoS ONE* 2008, 3.
59. Jongen-Lavrencic, M.; Sun, S.M.; Dijkstra, M.K.; Valk, P.J.M.; Löwenberg, B. MicroRNA expression profiling in relation to the genetic heterogeneity of acute myeloid leukemia. *Blood* 2008, 111, 5078–5085.
60. Li, Z.; Lu, J.; Sun, M.; Mi, S.; Zhang, H.; Luo, R.T.; Chen, P.; Wang, Y.; Yan, M.; Qian, Z.; et al. Distinct microRNA expression profiles in acute myeloid leukemia with common translocations. *Proc. Natl. Acad. Sci. USA* 2008, 105, 15535–15540.
61. Trino, S.; Lamorte, D.; Caivano, A.; Laurenzana, I.; Tagliaferri, D.; Falco, G.; Del Vecchio, L.; Musto, P.; De Luca, L. MicroRNAs as new biomarkers for diagnosis and prognosis, and as potential therapeutic targets in acute myeloid leukemia. *Int. J. Mol. Sci.* 2018, 19, 460.
62. Shell, S.; Park, S.M.; Radjabi, A.R.; Schickel, R.; Kistner, E.O.; Jewell, D.A.; Feig, C.; Lengyel, E.; Peter, M.E. Let-7 expression defines two differentiation stages of cancer. *Proc. Natl. Acad. Sci. USA* 2007, 104, 11400–11405.
63. Garzon, R.; Garofalo, M.; Martelli, M.P.; Briesewitz, R.; Wang, L.; Fernandez-Cymering, C.; Volinia, S.; Liu, C.G.; Schnittger, S.; Haferlach, T.; et al. Distinctive microRNA signature of acute myeloid leukemia bearing cytoplasmic mutated nucleophosmin. *Proc. Natl. Acad. Sci. USA* 2008, 105, 3945–3950.
64. Alcalay, M.; Tiacci, E.; Bergomas, R.; Bigerna, B.; Venturini, E.; Minardi, S.P.; Meani, N.; Diverio, D.; Bernard, L.; Tizzoni, L.; et al. Acute myeloid leukemia bearing cytoplasmic nucleophosmin (NPMc+ AML) shows a distinct gene expression profile characterized by up-regulation of genes involved in stem-cell maintenance. *Blood* 2005, 106, 899–902.
65. Russ, A.C.; Sander, S.; Lück, S.C.; Lang, K.M.; Bauer, M.; Rücker, F.G.; Kestler, H.A.; Schlenk, R.F.; Döhner, H.; Holzmann, K.; et al. Integrative nucleophosmin mutation-associated microRNA and gene expression pattern analysis identifies novel microRNA—Target gene interactions in acute myeloid leukemia. *Haematologica* 2011, 96, 1783–1791.
66. Wong, P.; Iwasaki, M.; Somervaille, T.C.P.; Ficara, F.; Carico, C.; Arnold, C.; Chen, C.Z.; Cleary, M.L. The miR-17-92 microRNA polycistron regulates MLL leukemia stem cell potential by modulating p21 expression. *Cancer Res.* 2010, 70, 3833–3842.
67. Handschuh, L.; Lonetti, A. Not only Mutations Matter: Molecular Picture of Acute Myeloid Leukemia Emerging from Transcriptome Studies. *J. Oncol.* 2019, 2019, 7239206.
68. Mendl, J.H.; Maharry, K.; Radmacher, M.D.; Mrózek, K.; Becker, H.; Metzeler, K.H.; Schwind, S.; Whitman, S.P.; Khalfell, J.; Kohlschmidt, J.; et al. RUNX1 mutations are associated with poor outcome in younger and older patients with cytogenetically normal acute myeloid leukemia and with distinct gene and microRNA expression signatures. *J. Clin. Oncol.* 2012, 30, 3109–3118.
69. Marcucci, G.; Radmacher, M.D.; Maharry, K.; Mrózek, K.; Ruppert, A.S.; Paschka, P.; Vukosavljevic, T.; Whitman, S.P.; Baldus, C.D.; Langer, C.; et al. MicroRNA expression in cytogenetically normal acute myeloid leukemia. *N. Engl. J. Med.* 2008, 358, 1919–1928.
70. Kotani, A.; Ha, D.; Schotte, D.; Den Boer, M.L.; Armstrong, S.A.; Lodish, H.F. A novel mutation in the miR-128b gene reduces miRNA processing and leads to glucocorticoid resistance of MLL-AF4 acute lymphocytic leukemia cells. *Cell Cycle* 2010, 9, 1037–1042.
71. Gutierrez-Camino, A.; Garcia-Obregon, S.; Lopez-Lopez, E.; Astigarraga, I.; Garcia-Orad, A. MiRNA deregulation in childhood acute lymphoblastic leukemia: A systematic review. *Epigenomics* 2019, 12, 69–80.

72. Szczepanek, J. Role of microRNA dysregulation in childhood acute leukemias: Diagnostics, monitoring and therapeutic s: A comprehensive review. *World J. Clin. Oncol.* 2020, 11, 348–369.
73. Yang, Y.L.; Yen, C.T.; Pai, C.H.; Chen, H.Y.; Yu, S.L.; Lin, C.Y.; Hu, C.Y.; Jou, S.T.; Lin, D.T.; Lin, S.R.; et al. A double negative loop comprising ETV6/RUNX1 and MIR181A1 contributes to differentiation block in t(12;21)-positive acute lymphoblastic leukemia. *PLoS ONE* 2015, 10.
74. Gefen, N.; Binder, V.; Zaliouva, M.; Linka, Y.; Morrow, M.; Novosel, A.; Edry, L.; Hertzberg, L.; Shomron, N.; Williams, O.; et al. Hsa-mir-125b-2 is highly expressed in childhood ETV6/RUNX1 (TEL/AML1) leukemias and confers survival advantage to growth inhibitory signals independent of p53. *Leukemia* 2010, 24, 89–96.
75. Grobbelaar, C.; Ford, A.M. The Role of MicroRNA in Paediatric Acute Lymphoblastic Leukaemia: Challenges for Diagnosis and Therapy. *J. Oncol.* 2019, 2019, 8941471.
76. Organista-Nava, J.; Gómez-Gómez, Y.; Illades-Aguilar, B.; Leyva-Vázquez, M.A. Regulation of the miRNA expression by TEL/AML1, BCR/ABL, MLL/AF4 and TCF3/PBX1 oncoproteins in acute lymphoblastic leukemia (Review). *Oncol. Rep.* 2016, 36, 1226–1232.
77. Yamada, N.; Noguchi, S.; Kumazaki, M.; Shinohara, H.; Miki, K.; Naoe, T.; Akao, Y. Epigenetic regulation of microRNA-128a expression contributes to the apoptosis-resistance of human T-cell leukaemia Jurkat cells by modulating expression of Fas-associated protein with death domain (FADD). *Biochim. Biophys. Acta Mol. Cell Res.* 2014, 1843, 590–602.
78. Yan, Z.X.; Zheng, Z.; Xue, W.; Zhao, M.Z.; Fei, X.C.; Wu, L.L.; Huang, L.M.; Leboeuf, C.; Janin, A.; Wang, L.; et al. MicroRNA181a is overexpressed in t-cell leukemia/lymphoma and related to chemoresistance. *Biomed Res. Int.* 2015, 2015, 197241.
79. Oliveira, L.H.; Schiavinato, J.L.; Fráguas, M.S.; Lucena-Araujo, A.R.; Haddad, R.; Araújo, A.G.; Dalmazzo, L.F.; Rego, E.M.; Covas, D.T.; Zago, M.A.; et al. Potential roles of microRNA-29a in the molecular pathophysiology of T-cell acute lymphoblastic leukemia. *Cancer Sci.* 2015, 106, 1264–1277.
80. Zaliouva, M.; Stuchly, J.; Winkowska, L.; Musilova, A.; Fiser, K.; Slamova, M.; Starkova, J.; Vaskova, M.; Hrusak, O.; Sramkova, L.; et al. Genomic landscape of pediatric B-other acute lymphoblastic leukemia in a consecutive European cohort. *Haematologica* 2019.
81. Li, Y.; Yuan, Y.; Tao, K.; Wang, X.; Xiao, Q.; Huang, Z.; Zhong, L.; Cao, W.; Wen, J.; Feng, W. Inhibition of BCR/ABL Protein Expression by miR-203 Sensitizes for Imatinib Mesylate. *PLoS ONE* 2013, 8, e61858.
82. Huang, W.; Wang, W.T.; Fang, K.; Chen, Z.H.; Sun, Y.M.; Han, C.; Sun, L.Y.; Luo, X.Q.; Chen, Y.Q. MIR-708 promotes phagocytosis to eradicate T-ALL cells by targeting CD47. *Mol. Cancer* 2018, 17, 12.
83. Li, X.J.; Luo, X.Q.; Han, B.W.; Duan, F.T.; Wei, P.P.; Chen, Y.Q. MicroRNA-100/99a, deregulated in acute lymphoblastic leukaemia, suppress proliferation and promote apoptosis by regulating the FKBP51 and IGF1R/mTOR signalling pathways. *Br. J. Cancer* 2013, 109, 2189–2198.
84. Avigad, S.; Verly, I.R.; Lebel, A.; Kordi, O.; Shichrur, K.; Ohali, A.; Hameiri-Grossman, M.; Kaspers, G.J.; Cloos, J.; Fronkova, E.; et al. miR expression profiling at diagnosis predicts relapse in pediatric precursor B-cell acute lymphoblastic leukemia. *Genes Chromosom. Cancer* 2016, 55, 328–339.
85. Silvestri, G.; Trotta, R.; Stramucci, L.; Ellis, J.J.; Harb, J.G.; Neviani, P.; Wang, S.; Eisfeld, A.-K.; Walker, C.J.; Zhang, B.; et al. Persistence of Drug-Resistant Leukemic Stem Cells and Impaired NK Cell Immunity in CML Patients Depend on MIR300 Antiproliferative and PP2A-Activating Functions. *Blood Cancer Discov.* 2020, 1, 48–67.
86. Zhang, B.; Nguyen, L.X.T.; Li, L.; Zhao, D.; Kumar, B.; Wu, H.; Lin, A.; Pellicano, F.; Hopcroft, L.; Su, Y.L.; et al. Bone marrow niche trafficking of miR-126 controls the self-renewal of leukemia stem cells in chronic myelogenous leukemia. *Nat. Med.* 2018, 24, 450–462.
87. Ruiz, M.S.; Sánchez, M.B.; Bonecker, S.; Furtado, C.; Koile, D.; Yankilevich, P.; Cranco, S.; del Custidiano, M.R.; Freitas, J.; Moiraghi, B.; et al. miRNome profiling of LSC-enriched CD34+CD38–CD26+ fraction in Ph+ CML-CP samples from Argentinean patients: A potential new pharmacogenomic tool. *Front. Pharmacol.* 2021, 11, 2231.
88. San José-Enériz, E.; Román-Gómez, J.; Jiménez-Velasco, A.; Garate, L.; Martin, V.; Cordeu, L.; Vilas-Zornoza, A.; Rodríguez-Otero, P.; José Calasanz, M.; Prósper, F.; et al. MicroRNA expression profiling in Imatinib-resistant Chronic Myeloid Leukemia patients without clinically significant ABL1-mutations. *Mol. Cancer* 2009, 8, 69.
89. Hershkovitz-Rokah, O.; Modai, S.; Pasmanik-Chor, M.; Toren, A.; Shomron, N.; Raanani, P.; Shpilberg, O.; Granot, G. MiR-30e induces apoptosis and sensitizes K562 cells to imatinib treatment via regulation of the BCR-ABL protein. *Cancer Lett.* 2015, 356, 597–605.
90. Mahdloo, T.; Sahami, P.; Ramezani, R.; Jafarinia, M.; Goudarzi, H.; Babashah, S. Up-regulation of miR-155 potentiates CD34+ CML stem/progenitor cells to escape from the growth-inhibitory effects of TGF-β1 and BMP signaling. *EXCLI J.* 2021, 20, 748–763.

91. Ferreira, A.F.; Moura, L.G.; Tojal, I.; Ambrósio, L.; Pinto-Simões, B.; Hamerschlag, N.; Calin, G.A.; Ivan, C.; Covas, D.T.; Kashima, S.; et al. ApoptomiRs expression modulated by BCR-ABL is linked to CML progression and imatinib resistance. *Blood Cells Mol. Dis.* 2014, 53, 47–55.
92. Klümper, T.; Bruckmueller, H.; Diewock, T.; Kaehler, M.; Haenisch, S.; Pott, C.; Bruhn, O.; Cascorbi, I. Expression differences of miR-142-5p between treatment-naïve chronic myeloid leukemia patients responding and non-responding to imatinib therapy suggest a link to oncogenic ABL2, SRI, cKIT and MCL1 signaling pathways critical for development of therapy resistance. *Exp. Hematol. Oncol.* 2020, 9, 1–15.
93. Li, Y.L.; Tang, J.M.; Chen, X.Y.; Luo, B.; Liang, G.H.; Qu, Q.; Lu, Z.Y. MicroRNA-153-3p enhances the sensitivity of chronic myeloid leukemia cells to imatinib by inhibiting B-cell lymphoma-2-mediated autophagy. *Hum. Cell* 2020, 33, 610–618.
94. Ohyashiki, J.H.; Ohtsuki, K.; Mizoguchi, I.; Yoshimoto, T.; Katagiri, S.; Umez, T.; Ohyashiki, K. Downregulated microRNA-148b in circulating PBMCs in chronic myeloid leukemia patients with undetectable minimal residual disease: A possible biomarker to discontinue imatinib safely. *Drug Des. Dev. Ther.* 2014, 8, 1151–1159.
95. Calin, G.A.; Ferracin, M.; Cimmino, A.; Di Leva, G.; Shimizu, M.; Wojcik, S.E.; Iorio, M.V.; Visone, R.; Sever, N.I.; Fabbri, M.; et al. A MicroRNA Signature Associated with Prognosis and Progression in Chronic Lymphocytic Leukemia. *N. Engl. J. Med.* 2005, 353, 1793–1801.
96. Visone, R.; Rassenti, L.Z.; Veronese, A.; Taccioli, C.; Costinean, S.; Aguda, B.D.; Volinia, S.; Ferracin, M.; Palatini, J.; Balatti, V.; et al. Karyotype-specific microRNA signature in chronic lymphocytic leukemia. *Blood* 2009, 114, 3872–3879.
97. Fabbri, M.; Bottoni, A.; Shimizu, M.; Spizzo, R.; Nicoloso, M.S.; Rossi, S.; Barbarotto, E.; Cimmino, A.; Adair, B.; Wojcik, S.E.; et al. Association of a microRNA/TP53 feedback circuitry with pathogenesis and outcome of B-cell chronic lymphocytic leukemia. *JAMA J. Am. Med. Assoc.* 2011, 305, 59–67.
98. Balatti, V.; Acunzo, M.; Pekarky, Y.; Croce, C.M. Novel mechanisms of regulation of miRNAs in CLL. *Trends Cancer* 2016, 2, 134–143.
99. Lavrov, A.V.; Chelysheva, E.Y.; Adilgereeva, E.P.; Shukhov, O.A.; Smirnikhina, S.A.; Kochergin-Nikitsky, K.S.; Yakushina, V.D.; Tsaurov, G.A.; Mordanov, S.V.; Turkina, A.G.; et al. Exome, transcriptome and miRNA analysis don't reveal any molecular markers of TKI efficacy in primary CML patients. *BMC Med. Genom.* 2019, 12.
100. Ohyashiki, K.; Umez, T.; Katagiri, S.; Kobayashi, C.; Azuma, K.; Tauchi, T.; Okabe, S.; Fukuoka, Y.; Ohyashiki, J.H. Downregulation of plasma miR-215 in chronic myeloid leukemia patients with successful discontinuation of imatinib. *Int. J. Mol. Sci.* 2016, 17, 570.
101. Ferrajoli, A.; Shanafelt, T.D.; Ivan, C.; Shimizu, M.; Rabe, K.G.; Nouraei, N.; Ikuo, M.; Ghosh, A.K.; Lerner, S.; Rassenti, L.Z.; et al. Prognostic value of miR-155 in individuals with monoclonal B-cell lymphocytosis and patients with B chronic lymphocytic leukemia. *Blood* 2013, 122, 1891–1899.
102. Psathas, J.N.; Doonan, P.J.; Raman, P.; Freedman, B.D.; Minn, A.J.; Thomas-Tikhonenko, A. Lymphoid neoplasia: The Myc-miR-17-92 axis amplifies B-cell receptor signaling via inhibition of ITIM proteins: A novel lymphomagenic feed-forward loop. *Blood* 2013, 122, 4220–4229.
103. Xiao, C.; Calado, D.P.; Galler, G.; Thai, T.H.; Patterson, H.C.; Wang, J.; Rajewsky, N.; Bender, T.P.; Rajewsky, K. MiR-150 Controls B Cell Differentiation by Targeting the Transcription Factor c-Myb. *Cell* 2007, 131, 146–159.
104. Zhou, B.; Wang, S.; Mayr, C.; Bartel, D.P.; Lodish, H.F. miR-150, a microRNA expressed in mature B and T cells, blocks early B cell development when expressed prematurely. *Proc. Natl. Acad. Sci. USA* 2007, 104, 7080–7085.
105. Hu, H.; Wang, B.; Borde, M.; Nardone, J.; Maika, S.; Allred, L.; Tucker, P.W.; Rao, A. Foxp1 is an essential transcriptional regulator of B cell development. *Nat. Immunol.* 2006, 7, 819–826.
106. Mraz, M.; Chen, L.; Rassenti, L.Z.; Ghia, E.M.; Li, H.; Jepsen, K.; Smith, E.N.; Messer, K.; Frazer, K.A.; Kipps, T.J. MiR-150 influences B-cell receptor signaling in chronic lymphocytic leukemia by regulating expression of GAB1 and FOXP1. *Blood* 2014, 124, 84–94.
107. Saleh, L.M.; Wang, W.; Herman, S.E.M.; Saba, N.S.; Anastas, V.; Barber, E.; Corrigan-Cummins, M.; Farooqui, M.; Sun, C.; Sarasua, S.M.; et al. Ibrutinib downregulates a subset of miRNA leading to upregulation of tumor suppressors and inhibition of cell proliferation in chronic lymphocytic leukemia. *Leukemia* 2017, 31, 340–349.