Phthalic Acid Esters

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Phthalic acid esters (PAEs) are a class of lipophilic chemicals widely used as plasticizers and additives to improve various products' mechanical extensibility and flexibility. At present, synthesized PAEs, which are considered to cause potential hazards to ecosystem functioning and public health, have been easily detected in the atmosphere, water, soil, and sediments; PAEs are also frequently discovered in plant and microorganism sources, suggesting the possibility that they might be biosynthesized in nature.

Keywords: phthalic acid esters ; biological activity

1. Physicochemical Properties and Applications of PAEs

Phthalic acid esters (dialkyl or alkyl aryl esters of 1,2-benzenedicarboxylic acid), usually called PAEs, phthalate esters, or just phthalates, are a group of important derivatives of phthalic acids which are synthesized from phthalic anhydride and specific alcohols by Fischer esterification ^{[1][2]}. PAEs based on hydrogen bond and van der Waals force interconnection are hydrophobic compounds with log K_{ow} ranging from 1.6 to 12 ^[3]. Most of the phthalate esters are colorless liquids with a low volatility, high boiling point, and poor solubility in water, but they are soluble in organic solvents and oils ^[4]. These esters' general chemical structure consists of a rigid planar aromatic ring and two malleable nonlinear fatty side chains. The two side-chain groups can be the same or not, and there are approximately 30 types of different side chains, ranging from dimethyl phthalate to tridecyl ester ^[5]. Due to phthalate esters being widespread in the biosphere and potential hazards in relation to ecosystem functioning and public health, six PAEs have been listed as priority pollutants by the United States Environmental Protection Agency and the European Union ^{[3][6]}, including dimethyl phthalate, diethyl phthalate, di-*n*-octyl phthalate, butyl benzyl phthalate, di(2-ethylhexyl) phthalate, and di-*n*-octyl phthalate. These phthalate esters' physicochemical properties and common applications are summarized in **Table 1** and **Figure 1**.

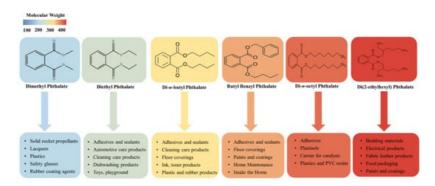


Figure 1. The application of six PAEs listed as priority pollutants.

PAEs are a class of lipophilic chemicals widely used in the plastics manufacturing industries as plasticizers and additives to improve the mechanical extensibility and flexibility of various products, such as plastics, paints, and synthetic fibers ^[Z]. Phthalates of lower molecular weight, such as dimethyl phthalate, diethyl phthalate, and di-*n*-butyl phthalate, are widely used in cosmetics and personal care products; dimethyl phthalate and diethyl phthalate allow perfume fragrances to evaporate more slowly, making the scent linger longer, and a small amount of di-*n*-butyl phthalate can make nail polish chip-resistant. Di-*n*-butyl phthalate is also used in cellulose esters, printing inks, latex adhesives, and insect repellents ^[3].

Higher phthalate molecules, such as di(2-ethylhexyl) phthalate, diisononyl phthalate, and butyl benzyl phthalate, have a wide range of applications as plasticizers in the polymer industry to improve flexibility, workability, and general handling properties, and about 80% of PAEs are used for this purpose ^{[3][10]}. The stability, fluidity and low volatility of these compounds make them very suitable for manufacturing PVC and other resins, such as polyvinyl acetates and polyurethanes ^[11]. One of the most widespread phthalate plasticizers, di(2-ethylhexyl) phthalate, has several useful

applications in numerous consumer products, commodities, and building materials ^[12]. Diisononyl phthalate is commonly used in garden hoses, pool liners, flooring tiles, tarps, and toys. Additionally, butyl benzyl phthalate, as a component of materials, is extensively used in vinyl flooring, synthetic leather, inks, and adhesives ^[2]. Phthalates are not covalently bound to the polymer matrix, rather they usually remain present as a freely mobile and leachable phase; therefore, they can be lost from soft plastic over time and released to the environment during production and manufacture. Not surprisingly, phthalates can often be found in freshwater lakes and oceans ^{[13][14]}, urban and suburban soil ^{[15][16]}, the atmosphere ^{[12][18]}, and sediments ^{[19][20]}. Bu et al. (2020) ^[21] summarized the concentrations of six representative phthalates from published papers in the last twenty years (2000–2019) to analyze the pollution characteristics of phthalates worldwide and found that their mean concentration in settled dust was 500.02 µg/g in North America, 580.12 µg/g in Europe, and 945.45 µg/g in Asia, with DEHP being the most predominant phthalate, with mean and median values of 615.78 µg/g and 394.03 µg/g, respectively; the mean concentration of six representative phthalates in indoor air was 598.14 ng/m³ in North America, 823.98 ng/m³ in Europe, and 1710.26 ng/m³ in Asia. In another study, Hu et al. (2020) ^[22] detected 8 PAEs in 67 sediment samples collected from Hangzhou Bay, Taizhou Bay, and Wenzhou Bay in China; the total concentrations of detected PAEs were in the range of 654–2603 ng/g, with di(2-ethylhexyl) phthalate being the predominant PAE (mean 663 ng/g, accounting for a mean of 52% of total PAEs).

PAEs	Molecular Formula	Molecular Weight	CAS Registration Number	Specific Gravity (20 °C)	Water Solubility (mg/L)	log K _{ow}	Melting Point (°C)	Application	References
Dimethyl phthalate	C ₁₀ H ₁₀ O ₄	194.18	131-11-3	1.19	4000	1.47	5.5	Insect repellent, personal care products, etc.	[23]
Diethyl phthalate	C ₁₂ H ₁₄ O ₄	222.24	84-66-2	1.12	1000	2.38	-40	Personal care products, plasticizers, cosmetics, etc.	[24]
Di- <i>n</i> -butyl phthalate	C ₁₄ H ₃₈ O ₄	278.35	84-74-2	1.05	11.2	3.74	-35	PVC plastics, explosive materials, nail paints, etc.	[25]
Butyl benzyl phthalate	C ₁₉ H ₂₀ O ₄	302.39	85-68-7	1.11	2.7	4.59	-35	Rapping materials, food conveyor belts, artificial letter, traffic cones, etc.	[<u>26]</u>
Di(2- ethylhexyl) phthalate	C ₂₄ H ₃₈ O ₄	390.62	117-81-7	0.99	0.003	7.5	-40	Medical devices, food packaging, building products, children's products, etc.	[27]
Di- <i>n</i> -octyl phthalate	C ₂₄ H ₃₈ O ₄	390.62	117-84-0	0.99	0.0005	8.06	-25	Conveyor belts, pool liners, garden hoses, etc.	[6]

Table 1. Physicochemical properties and application of six PAEs listed as priority pollutants.

2. Biological Activities of PAEs

2.1. Allelopathic/Phytotoxic Activity

Allelopathy refers to any direct or indirect harmful or beneficial effect exerted by one plant on another through the production of chemical compounds that are released into the environment. In some cases, allelopathy is suspected to contribute to the establishment of dominance of certain plant species, including some invasive alien species. Due to the phytotoxic property of allelochemicals, they are often considered valuable candidates for environmentally friendly bioherbicies [28][29]. Di-n-octyl phthalate isolated from Fimbristylis miliacea can remarkably inhibit the seed germination of tested weed species Ludwigia hysopifolia, Echinochloa colonum, Cyperus iria, and Paspalam digitatum ^[30]. Zhu et al. (2014) [31] isolated two allelochemicals, di(2-ethylhexyl) phthalate and di-n-butyl phthalate, from the root exudates of the invasive plant, Ageratina adenophora. In a bioassay, di-n-butyl phthalate was found to possess a significant inhibitory effect on seed germination and seedling growth of A. adenophora. Meanwhile, these two compounds significantly increased the superoxide dismutase (SOD) activity of A. adenophora's leaves and caused lipid peroxidation and cell membrane damage. Xuan et al. (2006) [32] identified the derivatives of phthalic acids from root exudates of Echinochloa crusgalli and found that diethyl phthalate strongly affects the seedling growth of alfalfa, Indian jointvetch, lettuce, monochorea, and sesame. Huang et al. (2017) [33] analyzed the extracts of aerial parts plants, root exudates, and plant rhizosphere soil of Chrysanthemum indicum to determine the effect of the allelochemical diethyl phthalate, and the results show that it has a noticeable impact on promoting the fresh weight of lettuce, as well as the root growth of lettuce and rape. Shanab et al. (2010) [34] extracted four phthalate derivatives from Eichhornia crassipes, including di-n-octyl phthalate, mono (2-ethylhexyl) phthalate, methyl dioctyl phthalate, and diisooctyl phthalateis, which possess strong inhibitory effects on Chlorella vulgaris.

Physiological studies have indicated that PAEs can influence enzyme activity, which might be at least one of their phytotoxicity mechanisms. Deng et al. (2017) ^[35] revealed that as the concentration of PAEs secreted by tobacco roots increased, the rate of production of superoxide anion radicals, the concentration of malondialdehyde, and the activity of peroxidase and SOD in tobacco root increased significantly. A series of changes could reduce the root system's antioxidant properties and cause oxidative damage to the apical cell membrane system, thereby affecting root absorption and ultimately showing autotoxicity. Dong et al. (2016) ^[36] extracted diisobutyl phthalate and di-*n*-butyl phthalate from the ethyl acetate extract of *C. fracta*, both of which show a strong inhibitory effect on the growth of *Heterosigma akashiwo* and *Gymnodinium breve*, which may be related to the production of reactive oxygen species (ROS) induced by diisobutyl phthalate and di-*n*-butyl phthalate in algal cells. Excessive ROS inhibits the activities of catalase and SOD, leading to lipid oxidation and the destruction of algae cell membranes.

2.2. Antimicrobial Activity

Natural products, including secondary metabolites produced by plants and microorganisms, have long been studied for their antimicrobial activity in the search for eco-friendly substitutes for synthesized chemicals [37]. Di(2-ethylhexyl) phthalate and di-n-butyl phthalate isolated from *B. mcbrellneri* show broad-spectrum antibacterial activity [38]. Di(2ethylhexyl) phthalate can inhibit the growth of gram-positive (S. epidermidis, MIC of 9.37 µg/mL; S. aureus, MIC of 18.75 µg/mL) and gram-negative bacteria (E. coli, MIC of 37.5 µg/mL; P. aeruginosa and Klebsiella pneumoniae, MIC at 75 µg/mL for both). Di-n-butyl phthalate also inhibits the growth of gram-positive (Bacillus subtilis and S. epidermidis, MIC at 18.75 µg/mL for both) as well as gram-negative bacteria (E. coli and P. aeroginosa, MIC at 37.5 µg/mL for both). Di(2ethylhexyl) phthalate isolated from the flowers of Calotropis gigantean exerts antimicrobial activity against B. subtilis with a MIC of 32 µg/mL ^[39]. There are also reports on the antimicrobial activity of di-n-butyl phthalate isolated from Streptomyces albidoflavus showing a MIC for E. coli of 53 μ g/mL, with B. subtilis at 84 μ g/mL ^[40]. Four phthalate derivatives isolated from E. crassipes also exert significant antibacterial activity against gram-positive bacteria (B. subtilis and Streptococcus faecalis) and gram-negative bacteria E. coli, and antifungal activity against Candida albicans [34]. In another study, El-Mehalawy et al. (2008) [41] found that di(2-ethylhexyl) phthalate could be produced by certain bacteria, including Tsukamurella inchonensis, Corynebacterium nitrilophilus, and Cellulosimicrbium cellulans, and di(2-ethylhexyl) phthalate has the function to inhibit fungal spore germination, cell membrane growth, and the production of total lipids and total protein. Li et al. (2021) [42] isolated di-n-butyl phthalate from a new marine Streptomyces sp. and found this compound significantly inhibited spore germination and mycelial growth of Colletotrichum fragariae. In addition to this, an obvious decrease was detected in sugar and protein contents of C. fragariae mycelia. Other studies have shown similar results. For instance, di-n-butyl phthalate was reported to inhibit spore germination and mycelium growth of Colletotrichum gloeosporioides, Colletotrichum musae, and Gaeumannomyces graminis [43][44][45].

Janu and Jayanthy (2014) found that diethyl phthalate derived from the fungus *Aspergillus* sp. increased the superoxide production and exerted ROS generated oxidative stress in the cytoplasm of bacterial cells, which eventually led to cell death ^[46]. In addition, diethyl phthalate with antimicrobial properties was reported for its ability to interfere with quorum sensing mediated virulence factors and biofilm formation in *Pseudomonas aeruginosa* ^{[47][48]}. Another study demonstrated that dimethyl phthalate (concentration ranged from 20 to 40 mg/L) greatly inhibited the growth and glucose utilization of *Pseudomonas fluorescens*, meanwhile the surface hydrophobicity and membrane permeability of *P. fluorescens* were also increased. Dimethyl phthalate could lead to deformation of the cell membrane and misopening of membrane channels. Additionally, RNA-Seq and RT-qPCR results revealed that the expression of some genes in *P. fluorescens* were altered, including the genes involved in energy metabolism, ATP-binding cassette transporting, and two-component systems by dimethyl phthalate ^[49].

2.3. Insecticidal Activity

In addition to their phytotoxic and antimicrobial activity, PAEs were also found to be insecticidal; attributed to inhibition of acetylcholinesterase enzyme activity, they possess significant mosquito larvicidal activity. Therefore, some phthalates, such as synthetic diethyl phthalate and dimethyl phthalate, have been used as active ingredients in insect repellents ^[50]. Previously, Adsul et al. (2012) ^[52] isolated di-*n*-butyl phthalate from the leaf extract of *Ipomoea carnea* via column chromatography, and this compound ws found to be lethal to the fourth instar larvae of *Aedes aegypti* and *Culex quinquefasciatus*, with the lethal concentrations of LC₅₀ being 81.43 and 109.64 ppm, respectively. Di-*n*-butyl phthalate and di(2-ethylhexyl) phthalate were isolated from the bacterium *B. mcbrellneri*; because of their significant acetylcholinesterase inhibitory activity, they are also active against the fourth instar of *A. aegypti* after 24 h of exposure ^[38]. On the other hand, various PAEs were also constantly reported as possessing other biological activities, such as anti-inflammatory, antiviral, anti-tumor, for a sub-activity, etc., indicating their valuable potential to be explored further in capacities other than plasticizers ^[53] (**Figure 2**).

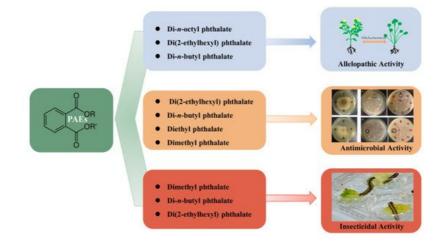


Figure 2. Biological activities of PAEs in living organisms.

References

- 1. Mathur, S.P. Phthalate esters in the environment: Pollutants or natural products? J. Environ. Qual. 1974, 3, 189–197.
- Gomez-Hens, A.; Aguilar-Caballos, M.P. Social and economic interest in the control of phthalic acid esters. TrAC Trends Anal. Chem. 2003, 22, 847–857.
- 3. Net, S.; Sempere, R.; Delmont, A.; Paluselli, A.; Ouddane, B. Occurrence, Fate, Behavior and Ecotoxicological State of Phthalates in Different Environmental Matrices. Environ. Sci. Technol. 2015, 49, 4019–4035.
- Gani, K.M.; Tyagi, V.K.; Kazmi, A.A. Occurrence of phthalates in aquatic environment and their removal during wastewater treatment processes: A review. Environ. Sci. Pollut. Res. 2017, 24, 17267–17284.
- 5. Autian, J. Toxicity and health threats of phthalate esters: Review of the literature. Environ. Health Perspect. 1973, 4, 3– 26.
- Das, M.T.; Kumar, S.S.; Ghosh, P.; Shah, G.; Malyan, S.K.; Bajar, S.; Thakur, I.S.; Singh, L. Remediation strategies for mitigation of phthalate pollution: Challenges and future perspectives. J. Hazard. Mater. 2020, 124496.
- 7. Hu, A.P.; Liu, Y.L.; Shi, L.K. Widespread occurrence of phthalic acid esters in raw oilseeds in China used for edible vegetable oil production. Food Addit. Contam. Part A 2016, 33, 1421–1427.

- 8. Heudorf, U.; Mersch-Sundermann, V.; Angerer, J. Phthalates: Toxicology and exposure. Int. J. Hyg. Environ. Health 2007, 210, 623–634.
- 9. Giuliani, A.; Zuccarini, M.; Cichelli, A.; Khan, H.; Reale, M. Critical review on the presence of phthalates in food and evidence of their biological impact. Int. J. Environ. Res. Public Health. 2020, 17, 5655.
- Vieira, M.G.A.; da Silva, M.A.; dos Santos, L.O.; Beppu, M.M. Natural-based plasticizers and biopolymer films: A review. Eur. Polym. J. 2011, 47, 254–263.
- 11. Alnaimat, A.S.; Barciela-Alonso, M.C.; Bermejo-Barrera, P. Development of a sensitive method for the analysis of four phthalates in tea samples: Tea bag contribution to the total amount in tea infusion. Food Addit. Contam. Part A 2020, 37, 1719–1729.
- Martínez, M.; Rovira, J.; Sharma, R.P.; Nadal, M.; Schuhmacher, M.; Kumar, V. Comparing dietary and non-dietary source contribution of BPA and DEHP to prenatal exposure: A Catalonia (Spain) case study. Environ. Res. 2018, 166, 25–34.
- Chen, H.; Mao, W.; Shen, Y.; Feng, W.; Mao, G.; Zhao, T.; Yang, L.; Yang, L.; Meng, C.; Li, Y.; et al. Distribution, source, and environmental risk assessment of phthalate esters (PAEs) in water, suspended particulate matter, and sediment of a typical Yangtze River Delta City, China. Environ. Sci. Pollut. Res. 2019, 26, 24609–24619.
- 14. Sha, Y.; Xia, X.; Yang, Z.; Huang, G.H. Distribution of PAEs in the middle and lower reaches of the Yellow River, China. Environ. Monit. Assess. 2007, 124, 277–287.
- 15. Liu, Y.; Ji, C.; Yu, Y.; Liu, H.; Shen, Y. Distribution characteristics and health risk assessment of PAEs in urban soils of Changji City, Xinjiang, China. Chin. J. Nat. 2020, 22, 67–74.
- 16. Wang, H.; Liang, H.; Gao, D.W. Occurrence and risk assessment of phthalate esters (PAEs) in agricultural soils of the Sanjiang Plain, northeast China. Environ. Sci. Pollut. Res. 2017, 24, 19723–19732.
- 17. He, Y.; Wang, Q.; He, W.; Xu, F. Phthalate esters (PAEs) in atmospheric particles around a large shallow natural lake (Lake Chaohu, China). Sci. Total Environ. 2019, 687, 297–308.
- Wang, P.; Wang, S.L.; Fan, C.Q. Atmospheric distribution of particulate- and gas-phase phthalic esters (PAEs) in a Metropolitan City, Nanjing, East China. Chemosphere 2008, 72, 1567–1572.
- 19. Zhao, X.; Jin, H.; Ji, Z.; Li, D.; Kaw, H.Y.; Chen, J.; Xie, Z.; Zhang, T. PAES and PAHs in the surface sediments of the East China Sea: Occurrence, distribution and influence factors. Sci. Total Environ. 2020, 703, 134763.
- 20. Arfaeinia, H.; Fazlzadeh, M.; Taghizadeh, F.; Saeedi, R.; Spitz, J.; Dobaradaran, S. Phthalate acid esters (PAEs) accumulation in coastal sediments from regions with different land use configuration along the Persian Gulf. Ecotoxicol. Environ. Saf. 2019, 169, 496–506.
- 21. Bu, S.B.; Wang, Y.L.; Wang, H.Y.; Wang, F.; Tan, Y.F. Analysis of global commonly-used phthalates and non-dietary exposure assessment in indoor environment. Build. Environ. 2020, 177, 106853.
- 22. Hu, H.; Fang, S.; Zhao, M.; Jin, H. Occurrence of phthalic acid esters in sediment samples from East China Sea. Sci. Total Environ. 2020, 722, 137997.
- 23. Wang, Y.; Zhang, G.; Wang, L. Potential Toxicity of Phthalic Acid Esters Plasticizer: Interaction of Dimethyl Phthalate with Trypsin in Vitro. J. Agric. Food Chem. 2015, 63, 75–84.
- 24. Keire, D.A.; Anton, P.; Faull, K.F.; Ruth, E.; Walsh, J.H.; Chew, P.; Quisimoro, D.; Territo, M.; Reeve, J.R. Diethyl phthalate, a chemotactic factor secreted by Helicobacter pylori. J. Biol. Chem. 2001, 276, 48847–48853.
- 25. Li, F.M.; Wu, M.; Yao, Y.; Zheng, X.; Zhao, J.; Wang, Z.Y.; Xing, B.S. Inhibitory effects and oxidative target site of dibutyl phthalate on Karenia brevis. Chemosphere 2015, 132, 32–39.
- Daiem, M.M.A.; Rivera-Utrilla, J.; Ocampo-Perez, R.; Mendez-Diaz, J.D.; Sanchez-Polo, M. Environmental impact of phthalic acid esters and their removal from water and sediments by different technologies—A review. J. Environ. Manag. 2012, 109, 164–178.
- 27. Martinez-Razo, L.D.; Martinez-Ibarra, A.; Vazquez-Martinez, E.R.; Cerbon, M. The impact of Di-(2-ethylhexyl) Phthalate and Mono(2-ethylhexyl) Phthalate in placental development, function, and pathophysiology. Environ. Int. 2021, 146, 106228.
- Callaway, R.M.; Aschehoug, E.T. Invasive plants versus their new and old neighbors: A mechanism for exotic invasion. Science 2000, 290, 521–523.
- 29. Shao, H.; Huang, X.; Wei, X.; Zhang, C. Phytotoxic effects and a phytotoxin from the invasive plant Xanthium italicum Moretti. Molecules 2012, 17, 4037–4046.
- Ismail, B.S.; Siddique, A.B. Identification of allelochemicals from Fimbristylis miliacea and their allelopathic potential against weed species. Allelopath. J. 2012, 30, 311–318.

- 31. Zhu, X.Z.; Guo, J.; Shao, H.; Yang, G.Q. Effects of allelochemicals from Ageratina adenophora (Spreng.) on its own autotoxicity. Allelopath. J. 2014, 34, 253–264.
- 32. Xuan, T.D.; Chung, I.M.; Khanh, T.D.; Tawata, S. Identification of phytotoxic substances from early growth of barnyard grass (Echinochloa crusgalli) root exudates. J. Chem. Ecol. 2006, 32, 895–906.
- 33. Huang, X.; Li, Y.; Jiang, P.; Zhang, X.; Zhang, X.; Tan, P.; Tian, W. Identification of chrysanthemum root exudates and allelopathic effects of the three plants. Hubei Agric. Sci 2017, 56, 1061–1071.
- Shanab, S.M.; Shalaby, E.A.; Lightfoot, D.A.; El-Shemy, H.A. Allelopathic effects of water hyacinth [Eichhornia crassipes]. PLoS ONE 2010, 5, e13200.
- 35. Deng, J.; Zhang, Y.; Hu, J.; Jiao, J.; Hu, F.; Li, H.; Zhang, S. Autotoxicity of Phthalate Esters in Tobacco Root Exudates: Effects on Seed Germination and Seedling Growth. Pedosphere 2017, 27, 1073–1082.
- 36. Dong, S.J.; Bi, X.D.; Wang, N.; Song, L.; Dai, W.; Zhang, S.L. Algicidal activities of Cladophora fracta on red tideforming microalgae Heterosigma akashiwo and Gymnodinium breve. Allelopath. J. 2016, 37, 231–240.
- Atanasov, A.G.; Waltenberger, B.; Pferschy-Wenzig, E.M.; Linder, T.; Wawrosch, C.; Uhrin, P.; Temml, V.; Wang, L.; Schwaiger, S.; Heiss, E.H.; et al. Discovery and resupply of pharmacologically active plant-derived natural products: A review. Biotechnol. Adv. 2015, 33, 1582–1614.
- Rajamanikyam, M.; Vadlapudi, V.; Parvathaneni, S.P.; Koude, D.; Sripadi, P.; Misra, S.; Amanchy, R.; Upadhyayula, S.M. Isolation and characterization of phthalates from Brevibacterium mcbrellneri that cause cytotoxicity and cell cycle arrest. EXCLI J. 2017, 16, 375–387.
- Habib, M.R.; Karim, M.R. Antimicrobial and Cytotoxic Activity of Di-(2-ethylhexyl) Phthalate and Anhydrosophoradiol-3acetate Isolated from Calotropis gigantea (Linn.) Flower. Mycobiology 2009, 37, 31–36.
- Roy, R.N.; Laskar, S.; Sen, S.K. Dibutyl phthalate, the bioactive compound produced by Streptomyces albidoflavus 321.2. Microbiol. Res. 2006, 161, 121–126.
- 41. El-Mehalawy, A.; Gebreel, H.; Rifaat, H.; El-Kholy, I.; Humid, A. Effect of antifungal compounds produced by certain bacteria on physiological activities of human and plant pathogenic fungi. J. Appl. Sci. Res. 2008, 4, 425–432.
- 42. Li, X.; Jing, T.; Zhou, D.; Zhang, M.; Qi, D.; Zang, X.; Zhao, Y.; Li, K.; Tang, W.; Chen, Y.; et al. Biocontrol efficacy and possible mechanism of Streptomyces sp. H4 against postharvest anthracnose caused by Colletotrichum fragariae on strawberry fruit. Postharvest Biol. Technol. 2021, 175, 111401.
- 43. Qi, G.; Liu, H.; Li, Z.; Long, L.; Wang, M. Active constituents of garlic bulb extract and its inhibition on Colletotrichum gloeosporioides. J. Henan Agri. Uni. 2020, 54, 59–63.
- 44. Zhang, Z.; Ao, W.; Xiong, Y.; Yan, R.; Wang, Y.; Zhu, D. Identification of antagonistic endophytic actinomycete FRo2 and isolation of its antimicrobial composition. Microbiology China. 2014, 41, 1574–1581.
- 45. Liang, C.; Chen, Y.; Li, C.; Song, X.; Xie, C.; Zhu, C.; Sun, R. Antifungal activity of extracts of Helicteres ngustifolia against ten plant pathogenic fungi. Guihaia 2020, 40, 715–726.
- 46. Janu, N.P.; Jaynthy, C. Antimicrobial activity of diethyl phthalate: An insilico approach. Asian J. Pharm. Clin. Res. 2014, 7, 141–142.
- 47. Gayatri, K.V.; Soundhari, C.; Pavithra, B.P. Biofilm inhibitory effect of Chlorella extracts on Pseudomonas aeruginosa. Int. J. Pharm. Sci. Res. 2019, 10, 1966–1971.
- Rashiya, N.; Padmini, N.; Ajilda, A.A.K.; Prabakaran, P.; Durgadevi, R.; Ravi, A.V.; Ghosh, S.; Sivakumar, N.; Selvakumar, G. Inhibition of biofilm formation and quorum sensing mediated virulence in Pseudomonas aeruginosa by marine sponge symbiont Brevibacterium casei strain Alu 1. Microb. Pathog. 2021, 150, 104693.
- 49. Wang, Z.; Wang, C.; You, Y.; Xu, W.; Lv, Z.; Liu, Z.; Chen, W.; Shi, Y.; Wang, J. Response of Pseudomonas fluorescens to dimethyl phthalate. Ecotoxicol. Environ. Saf. 2019, 167, 36–43.
- 50. Xu, H.; He, X.Q. Natural products-based insecticidal agents 6. Design, semisynthesis, and insecticidal activity of novel monomethyl phthalate derivatives of podophyllotoxin against Mythimna separata Walker in vivo. Bioorg. Med. Chem. Lett. 2010, 20, 4503–4506.
- 51. Brown, M.; Hebert, A.A. Insect repellents: An overview. J. Am. Acad. Dermatol. 1997, 36, 243–249.
- 52. Adsul, V.B.; Khatiwora, E.; Torane, R.C.; Deshpande, N.R. Isolation and characterization of dibutyl phthalate from leaves of Ipomoea carnea. Chem. Nat. Compd. 2012, 48, 712–713.
- 53. Roy, R.N. Bioactive natural derivatives of phthalate ester. Crit. Rev. Biotechnol. 2020, 40, 913–929.