

Food–Drug Interactions with Fruit Juices

Subjects: Pharmacology & Pharmacy | Biochemistry & Molecular Biology

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Fruit juices contain a large number of phytochemicals that, in combination with certain drugs, can cause food–drug interactions that can be clinically significant and lead to adverse events. The mechanisms behind such interactions are in most cases related to phytochemical interference with the activity of cytochrome P450 metabolizing enzymes (CYPs) or drug transporters. Moreover, alterations in their activity can have a clinical relevance if systemic exposure to the drug is decreased or increased, meaning that the pharmacological drug effects are suboptimal, or the drug will cause toxicity. In general, the common pharmacokinetic parameters found to be altered in food–drug interactions regarding fruit juices are the area under the concentration–time curve, bioavailability, and maximum plasma concentration. In most cases, the results from the drug interaction studies with fruit juices provide only limited information due to the small number of subjects, which are also healthy volunteers. Moreover, drug interactions with fruit juices are challenging to predict due to the unknown amounts of the specific phytochemicals responsible for the interaction, as well as due to the inter-individual variability of drug metabolism, among others. Therefore, this work aims to raise awareness about possible pharmacological interactions with fruit juices.

Keywords: fruit juices ; drugs ; pharmacological interactions ; phytochemicals ; pharmacokinetics ; pharmacodynamics

1. Introduction

By definition, drug interactions (DIs) occur when the pharmacological effect of one drug is altered by the presence of another drug or xenobiotic, which includes herbal medicine, food or drink bioactive components, or any other chemical agents. DIs, if considered to be clinically significant, pose a risk for human health as they can have a direct effect on the therapeutic outcome and even cause life-threatening adverse drug reactions. Traditionally, mechanisms of DIs are classified as pharmacokinetic or pharmacodynamic, depending on the nature of the interactions. Pharmacokinetic DIs involve processes related to drug absorption, distribution, metabolism, and elimination, whereas pharmacodynamic DIs are those in which the drug effects are changed due to the presence of another drug or xenobiotic at its site of action^[1]. In other words, depending on the nature of the DIs, different outcomes can be expected, such as decreased drug effectiveness with(out) increased drug toxicity or increased drug effectiveness with(out) increased drug toxicity. Hence, in the context of food–drug interactions, the clinical consequences can be the same, or expected, as in drug–drug interactions ^{[2][3]}.

As a healthy lifestyle is nowadays becoming an imperative for many people, fruit juices recently came in the spotlight as a novel class of functional beverages, as they are promising carriers of biologically active compounds from many other food sources ^[4]. Namely, fruit juices, due to the recent advances in food (bio)technology, are now able to provide even more nutritional and health benefits to those seeking a well-balanced diet ^[5]. Given the fact that many bioactive compounds can be added to fruit juices, it is important to emphasize their clinical, pharmacological, and toxicological aspects regarding the potential of their involvement in DIs ^{[2][3][6]}.

2. Absorption, Distribution, Metabolism and Elimination

Pharmacokinetics (PK) is a subdiscipline of pharmacology that quantitatively studies how a drug behaves in the body, i.e., what the body does to the drug. More specifically, the acronym ADME (absorption, distribution, metabolism and elimination) is often used to describe the PK processes of many drugs. PK principles can also be applied to patients (clinical pharmacokinetics) in order to provide safe and effective pharmacotherapy. To summarize, PK provides a quantitative relationship between a given dose and observed concentrations of a drug as a function of time, to provide optimization of dosage regimens. Pharmacodynamics (PD), on the other hand, is a subdiscipline of pharmacology, which studies the relationship between the drug concentration (at the site of the action, i.e., receptors) and the drug effects (response); i.e., what the drug does to the body. The pharmacological drug effect (or response) can be therapeutic and/or toxic, depending on the drug exposure. Hence, the PK/PD relationship can be viewed as an exposure–response

relationship. Additionally, the relationship between the therapeutic and toxic dose of a drug is expressed by the therapeutic index, which tells about the relative safety of a drug or the narrowness of the therapeutic index, where a relatively small increase in the plasma drug concentration can lead to adverse effects and cause toxicity^{[7][8][9][10][11]}.

When the drug is orally taken, it must be absorbed, and this fraction is then carried into the hepatic portal system and liver before reaching the site of action in the unchanged state (systemic circulation). However, there is a possibility of drug metabolism (loss of a drug) along that path in gastrointestinal tissues and the liver. This loss of a drug is called the (presystemic) first-pass effect (metabolism). First-pass metabolism is an enzyme-catalyzed process, where the most common enzymes are of the cytochrome P450 (CYP) type. Besides, various isozymes, such as CYP3A4 (55% of all drugs) and CYP2D6 (30% of all drugs), among many others, are involved in first-pass drug metabolism and show inter-individual variability; i.e., differing enzymatic activity. This activity is of particular interest in determining many DIs, as drugs, bioactive compounds from food (fruit juices), and other xenobiotics, when co-administered, can inhibit or induce CYP activity (CYP inhibitors and CYP inducers), consequently affecting the pharmacodynamic outcome^{[7][8][9][10][11]}. One of the PK parameters often mentioned in this paper is the area under the drug concentration–time curve (AUC), which is used to calculate the bioavailability (F) of a drug. The AUC shows how much of a drug is in the body or describes the total systemic exposure to the drug. Furthermore, F is the fraction (or percentage) of the absorbed drug that will be available at the site of action. In other words, F tells us about the rate and extent of drug absorption. F is a PK parameter that also indicates the extent of the systemic exposure to the drug. Additionally, for the intravenous route of drug administration (i.v.), F is by definition 100% ($F = 1$). The AUC of the orally administered drug relative to the AUC of the i.v. route gives us the possibility to calculate the absolute oral bioavailability of the drug (F_{oral}). F_{oral} can also be viewed as the product of the fractions of a drug dose that escapes metabolism by the gut (FG) and liver (FH); i.e., $F_{oral} = F_{abs} \times FG \times FH$, where F_{abs} is the fraction of a dose that is absorbed intact across the enterocytes^{[7][8][9][10][11]}.

Many factors can influence the rate of drug absorption, such as P-glycoprotein (drug transporter), and many factors can influence the extent of the drug absorption, such as the first-pass metabolism. Another PK parameter that is mentioned in the paper is the elimination half-life ($t_{1/2}$), which can be obtained from the concentration–time profile of a drug, which gives us information of the time needed to eliminate half of the drug in the body (after drug distribution is over); simply put, it describes the decay of the drug. Moreover, the half-life determines the duration of drug-action for many medications. Half-life is dependent on drug clearance (CL), as CL relates to the rate of drug elimination and drug systemic exposure. The higher the CL, the lower the systemic exposure (AUC). In addition, C_{max} is the peak concentration, or maximum plasma concentration of the drug, which can be easily obtained from the AUC^{[7][8][9][10][11]}. Finally, the number of possible DIs is without a limit, but as a general statement, it can be said that the most pronounced DIs will be those that are affecting oral bioavailability and clearance of drugs, particularly by affecting drug metabolism. DIs are one of the important sources of drug toxicity, as well as responsible for the variable therapeutic responses among individuals^{[7][8][9][10][11]}.

3. Drug Interactions with Common Fruit Juices

The most extensively described DIs with fruit juices are those with grapefruit juice (and grapefruit pulp), for which it is reported to have more than 40 DIs in humans ^[12]. Therefore, grapefruit juice that was consumed with the medicine lovastatin was used as a representative example for illustration of a DI, shown in [Figure 1](#). The figure shows the CYP3A4 inhibition by the grapefruit juice, leading to development of adverse effects of lovastatin; i.e., myopathy and rhabdomyolysis. Additionally, a human PK study confirmed that C_{max} and the AUC of lovastatin, when taken with grapefruit juice, were increased about 12-fold and 15-fold, respectively^{[13][14][15]}.

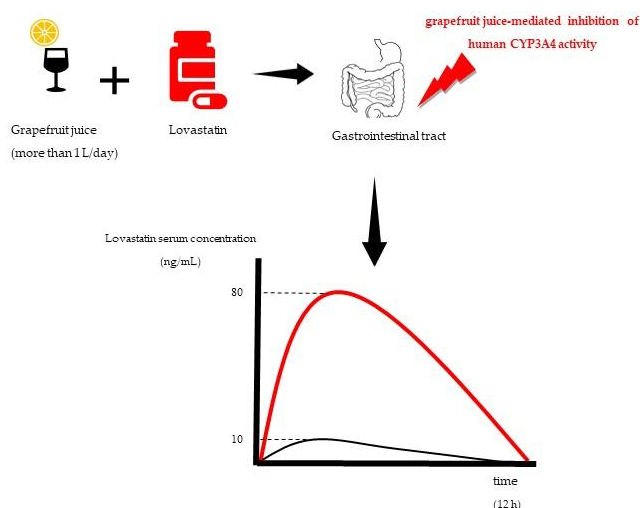


Figure 1. Hypothetical PK profile of lovastatin taken with water (black curve) and with grapefruit juice (red curve).

Notice the increase of AUC (red curve) which implicates the increase of systemic drug exposure that leads to development of adverse effects.

The discovery of the grapefruit interaction with drugs was an unexpected result of an interaction study with a focus on ethanol and felodipine (a calcium-channel antagonist) that targeted masking the taste of ethanol with grapefruit juice. The results showed an increase in felodipine bioavailability and C_{max} , as a consequence of irreversible degradation of an intestinal CYP3A by the grapefruit compounds^{[14][15]}. Moreover, grapefruit juice was found to have an impact on the efflux transport via P-glycoprotein in vitro, but also the influx transport via organic anion-transporting polypeptides—OATPs, esterases and sulfotransferases—although the clinical relevance of grapefruit juice on these drug transporters and enzymes was never determined^[16]. Other drug classes that are known to have a clinically relevant interaction with grapefruit juice include 3-hydroxy-3-methylglutaryl coenzyme A (HMG-CoA) reductase inhibitors (simvastatin), immunosuppressives (cyclosporine), antiarrhythmics (amiodarone) and anticonvulsants (carbamazepine). However, in some other cases with other drugs, even if the pharmacokinetic parameters were altered by the interaction with grapefruit, the clinical significance was still considered insignificant^{[16][17]}. In addition, it should be kept in mind that, due to the variable amounts of phytochemicals in juices, the results between the different pharmacokinetic studies of the same fruit juices cannot be compared, but the outcome can still be intuitively predicted.

The literature suggests that the specific components in grapefruit juice related to DIs could involve furanocoumarins, namely, bergamottin and 6'7'-dihydroxybergamottin^[18], but also naringin, which can inhibit OATP1A2 activity^[19]. It is speculated that even micromolar concentrations of naringin were responsible for a DI with fexofenadine, resulting in a decrease of its bioavailability^[20], which in this case could be clinically significant but was not confirmed by other authors.

There are some indications that grapefruit juice can reduce the levels of CYP3A4 activity by 47% only four hours after consumption, and these effects are persistent in the intestinal and liver cells at least 24 h after ingestion, meaning that grapefruit juice, even without co-administration with the drug, can lead to a DI or alter the metabolism of any CYP3A4 substrate for longer periods of time after consumption^[21]. Proof for that can be seen in the example of a DI with tacrolimus (immunosuppressant), where a delayed increase in systemic exposure to tacrolimus, e.g., from 4.7 ng/mL to 47.4 ng/mL, happened one week after the last grapefruit juice intake (250 mL, 4 times a day for 3 days). The patient developed severe headache and nausea, luckily without nephrotoxicity^{[12][22]}. Additionally, there is a case report of statin-associated rhabdomyolysis triggered by grapefruit consumption. Rhabdomyolysis is a rare but very serious adverse effect associated with statin therapy, which could cause kidney failure and death. In this case, the metabolism of simvastatin (CYP3A4 substrate) was altered due to CYP3A4 inactivation by the grapefruit, resulting in toxic systemic exposure to simvastatin^[23]. What could be expected from this data is that other statins, such as atorvastatin, which are CYP3A4 substrates, will have the same unwanted clinical outcomes. However, statins that are not substrates for the same CYP, such as rosuvastatin, are not expected to have this DI.

Altogether, it is clear that grapefruit juice interactions with orally given drugs cannot be generalized, and clinical relevance cannot be precisely determined. Hence, it is better to avoid grapefruit consumption with drugs that are substrates for an intestinal CYP3A4 and/or P-gp, especially in the cases when a drug has a narrow therapeutic index and a poor oral bioavailability due to the high pre-systemic (first-pass) metabolism mediated via CYP3A4, because even a single consumption of grapefruit juice can lead, in some cases, to drug toxicity. Additionally, the concentration/amount of the grapefruit extracts and as well interindividual variability of intestinal CYP3A4 activity among humans also contribute to the severity of the unwanted outcome [17][24].

Besides grapefruit, other citrus juices, such as orange, lemon, pomelo, and lime, were also reported to cause DIs in some clinical studies. As previous results were contradictory, or the clinical relevance could not be determined, a meta-analysis from Sridharan et al. provides a good summary of the DIs with selected citrus juices and cyclosporine (substrate for CYP3A4). Cyclosporine is an immunosuppressive drug with a narrow therapeutic index, with variable pharmacokinetics from person to person. Pooled results showed that no significant changes in the AUC and C_{max} of cyclosporine were observed with orange juice as compared to the controls, while pomelo juice, on the other hand, increased the AUC and C_{max} and decreased the elimination $t_{1/2}$ of cyclosporine[25]. Hence, orange juice did not have any significant interaction with cyclosporine. Interestingly, an earlier review from different authors[26], regarding orange juice, reported quite the opposite trend. Furthermore, orange juice does not contain bergamottin derivatives that are present in grapefruit juice, which may answer why orange juice did not have any significant interaction with cyclosporine. Pharmacologically, it would be expected that DIs with citrus juices (except orange juice) would have a more significant meaning in the case of liver disease and the concomitant use of other drugs (often seen in elderly patients) that share the same enzymatic pathway.

Orange juice was in one study found to decrease the oral absorption of alendronate (bisphosphonate agent for the treatment of osteoporosis) by approx. 60% compared to water [27]. This is very important, as alendronate, if taken on an empty stomach (after an overnight fast), 2 h before any meal (breakfast), has by default a very poor absolute oral bioavailability—approx. only 0.75% of the total dose[28]. Hence, taking orange juice with alendronate should be avoided.

Seville (sour) orange juice was found to have the same mechanism of CYP3A4 inactivation as grapefruit juice, and thus interacts with felodipine[29]. However, Seville orange juice did not alter the bioavailability of cyclosporine despite the CYP3A4 activity being significantly reduced, in contrast to grapefruit juice[30]. Therefore, it seems that grapefruit juice, besides the inactivation of intestinal CYP3A4, alters some other enzymes or transporters.

Seville orange and lime juices were in one clinical study inspected for a DI with sildenafil, a drug mostly known as Viagra®, which is an agent that improves penile erectile function[31]. Sildenafil has an extensive first-pass metabolism, which results in a relatively low absolute oral bioavailability ($F = 40\%$). Namely, for healthy subjects that received sildenafil (single dose of 50 mg) for three consecutive days and drank 250 mL of juice (or water) just before the drug was taken, the results showed that Seville orange juice increased the AUC and C_{max} of sildenafil by 44%. Although Seville orange juice is considered a moderate CYP3A4 inhibitor, this interaction did not have any adverse effects. The increase in systemic exposure is believed to be due to the intestinal inhibition of CYP3A4 and P-gp. On the other hand, lemon juice did not show any effects on the PK parameters of sildenafil.

Pomelo fruit juice (250 mL) was also inspected for a DI with sildenafil in another clinical study [32]. Surprisingly, pomelo juice decreased the systemic exposure to sildenafil. Namely, the bioavailability of sildenafil with pomelo juice was 60% lower compared with water. This is explained by a possibility that there was some sort of physicochemical interaction with one of the phytochemicals from pomelo juice, or an interaction with some drug transporter. However, the clinical significance of this interaction was not established, but the authors suggested avoiding taking pomelo juice with sildenafil.

Based on the previous information, it can be predicted that sildenafil, as a substrate for CYP3A4, will also have a DI with grapefruit juice, resulting in an increase in the systemic exposure of sildenafil. This was also confirmed in humans, but the adverse effects were not noticed[33]. However, it seems reasonable to avoid this combination. Namely, there are three independent case reports of DI-induced priapism (a persistent and painful penile erection, which is an emergency due to the risk of impotency) due to the concomitant use of pomegranate fruit juice (a high content of flavonoids) and sildenafil[34]. Although evidence of this interaction is based on circumstances, it is still a good example of how DIs with fruit juices are clinically significant, and adverse effects that are generally rare, such as priapism, are believed to be precipitated by a fruit juice. The proposed mechanism behind this interaction is due to CYP3A4 inhibition by phytochemicals from pomegranate juice.

Dresser et al. examined the effect of apple and orange juice on OATP uptake transporters[35]. Namely, in a human volunteer study, subjects were given two fexofenadine tablets (60 mg) (antihistaminic, sold by the trade name Allegra®) with 300 mL of fruit juice, up to a total volume of 1.2 L. The results of the PK study showed that both juices significantly decreased the fexofenadine concentrations in plasma compared with water (AUC (fexofenadine) apple juice = 434 ± 53 ;

AUC (fexofenadine) orange juice = 494 ± 16 ; and AUC (fexofenadine) water = 1616 ± 120 , $p < 0.001$). Among the tested individuals, the extent of the decreased fexofenadine concentrations was variable, or individuals with the highest fexofenadine AUC with water had the greatest decrease with the juices. Fexofenadine is a substrate of P-gp, but as well as of OATP uptake transporters, so Dresser et al. suggest the necessity to determine the individual contribution of each transporter in this case.

Apple juice was also found to have a DI with atenolol, which is antihypertensive drug [36]. In a human volunteer PK study, atenolol (50 mg) was taken with apple juice (600 mL and 1200 mL) and it was shown that atenolol systemic exposure was inversely proportional to the amount of consumed apple juice as compared to water (AUC (atenolol) 600 mL apple juice = 885.3; AUC (atenolol) 1200 mL apple juice = 389.7; and AUC (atenolol) water = 2110). In other words, the AUC of atenolol was decreased by 82% after the ingestion of 1200 mL apple juice, but there were no observed changes in the pharmacodynamic outcome even though there is an evident dose–response relationship. This DI is believed to be due to inhibition of intestinal drug transporter OATP2B1, but it is also believed that the higher acidity in the gastrointestinal tract (a large amount of apple juice) could also have an impact on atenolol absorption.

Other drugs reported as having a decreased systemic exposure with apple juice were montelukast (an anti-asthma agent) and aliskiren (an antihypertensive agent), which are also substrates for uptake transporters [37] [54]. In addition, decreasing the systemic exposure of atenolol and montelukast was also noticed with orange juice [38][39]. However, to our knowledge, there are no documented cases of a DI (from clinical practice) with apple and orange juice to date. One exception could be in the case of calcium-fortified orange juice, where, due to chelation of the drugs with calcium, alterations in the C_{max} and AUC of the fluoroquinolones (antibiotics) could decrease the antibiotic effects, leading to antibiotic resistance [40][41]. The latter statement could be applied to all calcium-fortified fruit juices.

It is worth mentioning that orange juice was found to increase the systemic exposure to aluminum [42] and iron [43] by enhancing their absorption. In the case of aluminum, there is, in theory, a risk of aluminum toxicity in a patient with renal disease, as it was found that urinary excretion of aluminum increased 10 times when it was taken with orange juice.

Fruits, such as oranges, bananas and prunes, along with their fruit juices, or in common combinations with vegetable juices such as carrot juice and tomato juice, contain very high amounts of potassium, which in combination with potassium-sparing drugs, such as angiotensin-converting enzyme inhibitors (ramipril), diuretics (spironolactone, triamterene) and especially in patients with kidney disease (hemodialysis patients) and with hypoaldosteronism, could lead to life-threatening hyperkalemia, so it is advisable to always seek advice from a pharmacist prior to consumption [44] [45][46]. Prune juice is best known for its laxative effect, so it is often sold as an over-the-counter (OTC) dietary supplement in many pharmacies and specialized stores, where consumers/patients might be unaware of the possible side effects [47] [48].

Plum and avocado juices were found to contain higher amounts of biogenic amine, tyramine, as compared to other fruit sources [49]. In theory, monoamine oxidase inhibitors (MAOIs) can inhibit tyramine degradation, which leads to its increased systemic exposure with a consequence of developing a hypertensive crisis [50]. However, over the last decades, the use of MAOIs has decreased and documented cases of their toxicity (due to the tyramine overexposure) are very rare, but the caution is still advised, as even 8 mg of tyramine was historically known to cause hypertension with some MAOIs [51][52].

Cranberry fruit juice is nowadays a very popular prophylactic treatment for the infections of the urinary tract. There have been some safety concerns regarding the cranberry fruit juice intake and using warfarin—an anticoagulant—which is known for its narrow therapeutic index and life-threatening side effects in case of increased systemic exposure [53][54]. The main reason for concern was a clinical case report that reported a change in the INR (International Normalized Ratio), a biochemical marker of warfarin's antithrombotic effects, related to cranberry juice intake. INR was in some patients decreased, meaning that the blood coagulates too easily, so the risk for developing blood clots rises [55], or it was increased, meaning that there is increased risk of bleeding [56]. However, in a randomized, double-blind clinical study, this interaction was not confirmed [57]. Ansell et al. commented that cranberry juice was not the one to blame for the results, as the reported cases were not convincing beyond doubt. Namely, patients had a variety of illnesses, the amount of ingested cranberry juice was not known in some cases, as well as the other dietary components, nor was the patients' compliance known, but most importantly, the pharmacogenetic profiling was not done either [58]. Although, it is well-established that polymorphisms of CYP2C9 and vitamin K epoxide reductase (VKORC) affect the INR values and clinical outcome of pharmacotherapy with warfarin. Detailed discussion of this interaction was provided by Zikria et al. [59]. In the literature, there are other described cases of other fruit juices that have in vitro effects on drug-metabolizing enzymes or transporters; however, their clinical significance was never firmly confirmed.

Tangerine fruit juice was reported to upregulate CYP3A4 activity and inhibit P-glycoprotein due to the high content of flavonoid tangeretin^{[60][61]}. Interestingly, some older data reported the opposite effect—tangerine (tangeretin) inhibits CYP3A4 and CYP1A2 in human liver microsomes ^[62].

Black mulberry was reported to inhibit CYP3A (and OATP-B), but also was wild grape ^[63]. Black mulberry is traditionally promoted as a fruit with a high content of iron, so it has a beneficial effect on the treatment of anemia. In addition, some studies showed that black mulberry has a beneficial effect on cholesterol levels, liver tissue (hepatoprotective effect) and even anti-obesity potential ^[64]. In theory, large amounts of black mulberry fruit juice could interact with CYP3A4 substrates, leading to an increase in systemic exposure of such drugs, but clinical relevance was never determined in humans.

Mango stem-bark was found to inhibit CYP1A1, CYP1A2, CYP3A1, CYP2C6, CYP2E1 and P-glycoprotein ^[65], while grape, due to the resveratrol, was implied in inhibition of CYP1A1/1A2 isoforms and CYP2E1, which are needed for the activation of procarcinogens (polycyclic arylamines, polyaromatic hydrocarbons, aflatoxin B1 and *N*-nitrosamines, respectively). Hence, besides having protective role as antioxidants, it is suggested that their fruit juices could alter the CYP3A4 activity, which is an important biotransformation pathway for many drugs ^[66]. The same was reported for papaya^[67] and black raspberry; however, a recent study with taxane agents (anticancer agents) did not confirm altered CYP3A4 activity^[68].

Tropical fruit juices, such as pineapple, papaya, litchi, kiwi, starfruit and passion fruit, were implicated in DIs in vitro due to their inhibitory effects on CYP2C9 or CYP3A4. Pineapple fruit juice, due to a high bromelain content was found to have the most pronounced inhibitory properties on CYP2C9, compared to other fruit juices. The effect of the inhibition was proportionally dependent on the increase in the amount of pineapple juice. Moreover, starfruit juice was found to be a very potent inhibitor of CYP3A4 compared to grapefruit juice. Namely, an assay of midazolam 1-hydroxylase activity of human CYP3A showed that residual activity of midazolam 1-hydroxylase (%) with starfruit juice was only 0.1 ± 0.0 , as compared to the 14.7 ± 0.5 for grapefruit juice. It would be interesting to study those inhibitory activities in vivo to determine the clinical relevance of tropical fruit juices on DI.

The next interesting question to address is, could fruit juices be exploited for enhancing the positive pharmacotherapy outcome in some cases? The answer is yes^[69].

Grapefruit juice was shown to be a drug-sparing agent (an agent that decreases the therapeutic dose of another drug) in a case of concomitant use with cyclosporine. In other words, patients could avoid the dose-related side effects of cyclosporine ^{[70][71]}. In another study, grapefruit enhanced oral bioavailability of artemether, which is antimalarial agent with generally high presystemic metabolism via CYP3A4, indicating more effective treatments of malaria ^[72]. Therefore, grapefruit juice could be useful in maintaining the effectiveness and efficacy of some drugs.

Lime juice and artemisinin combination therapy (antimalarial agents) were given in one study to children (61 males and 50 females) with acute uncomplicated malaria^[73]. It was observed that the artemisinin with lime juice caused more rapid clearance of parasites; also, the lime juice is believed to prevent resistance. The proposed explanation for this phenomenon is high amounts of vitamin C and flavonoids present in lime juice, which, besides its low pH, contributed to antioxidant activity. Similar results were shown recently on murine models, but with a lemon decoction resulting in a suppression of parasites by 39% and rapid early parasite clearance, as compared to the controls. However, there is a need to determine the exact effects of lime and lemon juice as supplement treatments in malaria by doing further investigations.

Blueberry juice, prepared from fresh blueberries, was given to 201 children that were receiving etanercept as a treatment for their juvenile idiopathic arthritis ^[74]. The study showed that the blueberry juice treatment combined with etanercept improved the symptoms of the disease, but as well decreased the side-effects of etanercept.

Regarding the previously mentioned increase of iron absorption with orange juice, it seems that it has a beneficial effect and such a combination could contribute to a better response to iron-deficiency anemia, which is a common problem, especially in children, but also in adults^[75]. As DIs precipitated by fruit juices can sometimes be predicted, [Table 1](#) summarizes previously described examples of potentially relevant drug interactions that should be considered in everyday clinical practice.

Table 1. Examples of potentially relevant drug interactions precipitated by fruit juices in humans.

References

1. Thanacoody, R. Drug interactions. In *Clinical Pharmacy and Therapeutics*; Whittlesea, C., Hodson, K., Eds.; Elsevier: Amsterdam, The Netherlands, 2019; pp. 53–65.
2. Amadi, C.N.; Mgbahurike, A.A. Selected food/herb–drug interactions. *Am. J. Ther.* 2018, 25, 423–433.
3. Mouly, S.; Lloret-Linares, C.; Sellier, P.-O.; Sene, D.; Bergmann, J.F. Is the clinical relevance of drug-food and drug-herb interactions limited to grapefruit juice and saint-john's wort? *Pharmacol. Res.* 2017, 118, 82–92.
4. Bursać Kovačević, D.; Brdar, D.; Fabečić, P.; Barba, F.J.; Lorenzo, J.M.; Putnik, P. Strategies to achieve a healthy and balanced diet: Fruits and vegetables as a natural source of bioactive compounds. In *Agri-Food Industry Strategies for Healthy Diets and Sustainability*; Barba, F.J., Putnik, P., Bursać Kovačević, D., Eds.; Elsevier: Amsterdam, The Netherlands, 2020; pp. 51–88.
5. Putnik, P.; Pavlić, B.; Šojić, B.; Zavadlav, S.; Žuntar, I.; Kao, L.; Kitonić, D.; Kovačević, D.B. Innovative hurdle technologies for the preservation of functional fruit juices. *Foods* 2020, 9, 699.
6. Zawiah, M.; Yousef, A.-M.; Khan, A.H.; Al-Ashwal, F.Y.; Matar, A.; Alkhawaldeh, B.; Nassar, R.; Abduljabbar, R.; Abdo Ahmed, A.A. Food-drug interactions: Knowledge among pharmacists in Jordan. *PLoS ONE* 2020, 15, e0234779.
7. Tozer, T.N.; Rowland, M. *Essentials of Pharmacokinetics and Pharmacodynamics*; Lippincott Williams & Wilkins: Philadelphia, PA, USA, 2016.
8. Tozer, W.K.; Shargel, L.; Yu, A.B.C. *Applied Biopharmaceutics & Pharmacokinetics*; Lippincott Williams & Wilkins Publishers: Philadelphia, PA, USA, 2016.
9. Beringer, P. *Winter's Basic Clinical Pharmacokinetics*; Lippincott Williams & Wilkins: Philadelphia, PA, USA, 2018.
10. Persky, A.M.; Pollack, G.M. *Foundations in Pharmacokinetics*; UNC Press: Chapel Hill, NC, USA, 2017.
11. Derendorf, H.; Schmidt, S. Rowland and Tozer's *Clinical Pharmacokinetics and Pharmacodynamics: Concepts and Applications*, 5th ed.; LWW Lippincott Williams and Wilkins: Philadelphia, PA, USA, 2019.
12. Rodríguez-Fragoso, L.; Martínez-Arismendi, J.L.; Orozco-Bustos, D.; Reyes-Esparza, J.; Torres, E.; Burchiel, S.W. Potential risks resulting from fruit/vegetable-drug interactions: Effects on drug-metabolizing enzymes and drug transporters. *J. Food Sci.* 2011, 76, R112–R124.
13. Kantola, T.; Kivistö, K.T.; Neuvonen, P.J. Grapefruit juice greatly increases serum concentrations of lovastatin and lovastatin acid. *Clin. Pharmacol. Ther.* 1998, 63, 397–402.

14. Bailey, D.G.; Malcolm, J.; Arnold, O.; David Spence, J. Grapefruit juice–drug interactions. *Br. J. Clin. Pharmacol.* 2002, 46, 101–110.
15. Bailey, D.G. Predicting clinical relevance of grapefruit–drug interactions: A complicated process. *J. Clin. Pharm. Ther.* 2017, 42, 125–127.
16. Hanley, M.J.; Cancalon, P.; Widmer, W.W.; Greenblatt, D.J. The effect of grapefruit juice on drug disposition. *Expert Opin. Drug Metab. Toxicol.* 2011, 7, 267–286.
17. Mertens-Talcott, S.U.; Zadezensky, I.; De Castro, W.V.; Derendorf, H.; Butterweck, V. Grapefruit–drug interactions: Can interactions with drugs be avoided? *J. Clin. Pharmacol.* 2006, 46, 1390–1416.
18. Paine, M.F.; Widmer, W.W.; Hart, H.L.; Pusek, S.N.; Beavers, K.L.; Criss, A.B.; Brown, S.S.; Thomas, B.F.; Watkins, P.B. A furanocoumarin-free grapefruit juice establishes furanocoumarins as the mediators of the grapefruit juice–felodipine interaction. *Am. J. Clin. Nutr.* 2006, 83, 1097–1105.
19. Farkas, D.; Greenblatt, D.J. Influence of fruit juices on drug disposition: Discrepancies between *in vitro* and clinical studies. *Expert Opin. Drug Metab. Toxicol.* 2008, 4, 381–393.
20. Bailey, D.G.; Dresser, G.K.; Leake, B.F.; Kim, R.B. Naringin is a major and selective clinical inhibitor of organic anion-transporting polypeptide 1a2 (oatp1a2) in grapefruit juice. *Clin. Pharmacol. Ther.* 2007, 81, 495–502.
21. Fukuda, K.; Guo, L.; Ohashi, N.; Yoshikawa, M.; Yamazoe, Y. Amounts and variation in grapefruit juice of the main components causing grapefruit–drug interaction. *J. Chromatogr. B* 2000, 741, 195–203.
22. Fukatsu, S.; Fukudo, M.; Masuda, S.; Yano, I.; Katsura, T.; Ogura, Y.; Oike, F.; Takada, Y.; Inui, K.-i. Delayed effect of grapefruit juice on pharmacokinetics and pharmacodynamics of tacrolimus in a living-donor liver transplant recipient. *Drug Metab. Pharmacokinet.* 2006, 21, 122–125.
23. Dreier, J.P.; Endres, M. Statin-associated rhabdomyolysis triggered by grapefruit consumption. *Neurology* 2004, 62, 670.
24. Koziolok, M.; Alcaro, S.; Augustijns, P.; Basit, A.W.; Grimm, M.; Hens, B.; Hoad, C.L.; Jedamzik, P.; Madla, C.M.; Maliepaard, M.; et al. The mechanisms of pharmacokinetic food–drug interactions—A perspective from the ungap group. *Eur. J. Pharm. Sci.* 2019, 134, 31–59.
25. Grenier, J.; Fradette, C.; Morelli, G.; Merritt, G.; Vrandeć, M.; Ducharme, M. Pomelo juice, but not cranberry juice, affects the pharmacokinetics of cyclosporine in humans. *Clin. Pharmacol. Ther.* 2006, 79, 255–262.
26. Sermsappasuk, P.; Chaiyakunapruk, N.; Lohitnavy, M.; Kitiyakara, K. Assessing clinical evidence of drug interactions between citrus juices and cyclosporine. *Asian Biomed.* 2013, 7, 477–489.
27. Gertz, B.J.; Holland, S.D.; Kline, W.F.; Matuszewski, B.K.; Freeman, A.; Quan, H.; Lassetter, K.C.; Mucklow, J.C.; Porras, A.G. Studies of the oral bioavailability of alendronate*. *Clin. Pharmacol. Ther.* 1995, 58, 288–298.
28. Gertz, B.J.; Holland, S.D.; Kline, W.F.; Matuszewski, B.K.; Porras, A.G. Clinical pharmacology of alendronate sodium. *Osteoporos. Int.* 1993, 3, 13–16.
29. Malhotra, S. Seville orange juice–felodipine interaction: Comparison with dilute grapefruit juice and involvement of furocoumarins. *Clin. Pharmacol. Ther.* 2001, 69, 14–23.
30. Edwards, D.; Fitzsimmons, M.; Schuetz, E.; Yasuda, K.; Ducharme, M.; Warbasse, L.; Woster, P.; Schuetz, J.; Watkins, P. 6',7'-dihydroxybergamottin in grapefruit juice and seville orange juice: Effects on cyclosporine disposition, enterocyte cyp3a4, and p-glycoprotein. *Clin. Pharmacol. Ther.* 1999, 65, 237–244.
31. Abdelkawy, K.S.; Donia, A.M.; Turner, R.B.; Elbarbry, F. Effects of lemon and seville orange juices on the pharmacokinetic properties of sildenafil in healthy subjects. *Drugs R D* 2016, 16, 271–278.
32. Al-Ghazawi, M.A.; Tutunji, M.S.; AbuRuz, S.M. The effects of pummelo juice on pharmacokinetics of sildenafil in healthy adult male Jordanian volunteers. *Eur. J. Clin. Pharmacol.* 2009, 66, 159–163.
33. Žuntar, I.; Krivohlavek, A.; Kosić-Vukšić, J.; Granato, D.; Bursać Kovačević, D.; Putnik, P. Pharmacological and toxicological health risk of food (herbal) supplements adulterated with erectile dysfunction medications. *Curr. Opin. Food Sci.* 2018, 24, 9–15.
34. Senthilkumaran, S.; Suresh, P.; Thirumalaikolundusubramanian, P.; Balamurugan, N. Priapism, pomegranate juice, and sildenafil: Is there a connection? *Urol. Ann.* 2012, 4, 108.
35. Dresser, G. Fruit juices inhibit organic anion transporting polypeptide-mediated drug uptake to decrease the oral availability of fexofenadine. *Clin. Pharmacol. Ther.* 2002, 71, 11–20.
36. Jeon, H.; Jang, I.-J.; Lee, S.; Ohashi, K.; Kotegawa, T.; Ieiri, I.; Cho, J.-Y.; Yoon, S.H.; Shin, S.-G.; Yu, K.-S.; et al. Apple juice greatly reduces systemic exposure to atenolol. *Br. J. Clin. Pharmacol.* 2013, 75, 172–179.

37. Tapaninen, T.; Neuvonen, P.J.; Niemi, M. Orange and apple juice greatly reduce the plasma concentrations of the oap2b1 substrate aliskiren. *Br. J. Clin. Pharmacol.* 2011, 71, 718–726.
38. Lilja, J. Orange juice substantially reduces the bioavailability of the β -adrenergic-blocking agent celiprolol. *Clin. Pharmacol. Ther.* 2004, 75, 184–190.
39. Mougey, E.B.; Lang, J.E.; Wen, X.; Lima, J.J. Effect of citrus juice and slco2b1 genotype on the pharmacokinetics of montelukast. *J. Clin. Pharmacol.* 2011, 51, 751–760.
40. Wallace, A.W.; Victory, J.M.; Amsden, G.W. Lack of bioequivalence when levofloxacin and calcium-fortified orange juice are coadministered to healthy volunteers. *J. Clin. Pharmacol.* 2003, 43, 539–544.
41. Neuhofer, A.L.; Wilton, J.H.; Victory, J.M.; Hejmanowski, L.G.; Amsden, G.W. Lack of bioequivalence of ciprofloxacin when administered with calcium-fortified orange juice: A new twist on an old interaction. *J. Clin. Pharmacol.* 2002, 42, 461–466.
42. Fairweather-Tait, S.; Hickson, K.; McGaw, B.; Reid, M. Orange juice enhances aluminium absorption from antacid preparation. *Eur. J. Clin. Nutr.* 1994, 48, 71–73.
43. Balay, K.S.; Hawthorne, K.M.; Hicks, P.D.; Griffin, I.J.; Chen, Z.; Westerman, M.; Abrams, S.A. Orange but not apple juice enhances ferrous fumarate absorption in small children. *J. Pediatr. Gastroenterol. Nutr.* 2010, 50, 545–550.
44. Aronson, J.K. *Meyler's Side Effects of Drugs: The International Encyclopedia of Adverse Drug Reactions and Interactions*; Aronson, J.K., Ed.; Elsevier Science: Amsterdam, The Netherlands, 2016; pp. 1030–1053.
45. Weaver, C.M. Potassium and health. *Adv. Nutr.* 2013, 4, 368S–377S.
46. St-Jules, D.E.; Goldfarb, D.S.; Sevcik, M.A. Nutrient non-equivalence: Does restricting high-potassium plant foods help to prevent hyperkalemia in hemodialysis patients? *J. Renal Nutr.* 2016, 26, 282–287.
47. Stacewicz-Sapuntzakis, M.; Bowen, P.E.; Hussain, E.A.; Damayanti-Wood, B.I.; Farnsworth, N.R. Chemical composition and potential health effects of prunes: A functional food? *Crit. Rev. Food Sci. Nutr.* 2001, 41, 251–286.
48. Wallace, T. Dried plums, prunes and bone health: A comprehensive review. *Nutrients* 2017, 9, 401.
49. Sánchez-Pérez, S.; Comas-Basté, O.; Rabell-González, J.; Veciana-Nogués, M.; Latorre-Moratalla, M.; Vidal-Carou, M. Biogenic amines in plant-origin foods: Are they frequently underestimated in low-histamine diets? *Foods* 2018, 7, 205.
50. Bushra, R.; Aslam, N.; Khan, A. Food drug interactions. *Oman Med. J.* 2011, 26, 77–83.
51. Garcia, E.; Santos, C. Monoamine Oxidase Inhibitor Toxicity. Available online: <https://www.ncbi.nlm.nih.gov/books/nbk459386/> (accessed on 24 October 2020).
52. Chen, J.J. Pharmacologic safety concerns in parkinson's disease: Facts and insights. *Int. J. Neurosci.* 2011, 121, 45–52.
53. Aston, J.L.; Lodolce, A.E.; Shapiro, N.L. Interaction between warfarin and cranberry juice. *Pharmacotherapy* 2006, 26, 1314–1319.
54. Mergenhagen, K.A.; Sherman, O. Elevated international normalized ratio after concurrent ingestion of cranberry sauce and warfarin. *Am. J. Health-Syst. Pharm.* 2008, 65, 2113–2116.
55. Medicines and Healthcare Products Regulatory Agency/Committee on Safety of Medicines. Interaction between warfarin and cranberry juice: New advice. *Curr. Probl. Pharmacovigil.* 2004, 30:10.
56. Suvarna, R. Possible interaction between warfarin and cranberry juice. *BMJ* 2003, 327, 1454–1454.
57. Ansell, J.; McDonough, M.; Zhao, Y.; Harmatz, J.S.; Greenblatt, D.J. The absence of an interaction between warfarin and cranberry juice: A randomized, double-blind trial. *J. Clin. Pharmacol.* 2009, 49, 824–830.
58. Johnson, J.A.; Cavallari, L.H. Warfarin pharmacogenetics. *Trends Cardiovasc. Med.* 2015, 25, 33–41.
59. Zikria, J.; Goldman, R.; Ansell, J. Cranberry juice and warfarin: When bad publicity trumps science. *Am. J. Med.* 2010, 123, 384–392.
60. Backman, J. Lack of correlation between in vitro and in vivo studies on the effects of tangeretin and tangerine juice on midazolam hydroxylation. *Clin. Pharmacol. Ther.* 2000, 67, 382–390.
61. Putnik, P.; Barba, F.J.; Lorenzo, J.M.; Gabrić, D.; Shpigelman, A.; Cravotto, G.; Bursać Kovačević, D. An integrated approach to mandarin processing: Food safety and nutritional quality, consumer preference, and nutrient bioaccessibility. *Compr. Rev. Food Sci. Food Saf.* 2017, 16, 1345–1358.
62. Obermeier, M.T.; White, R.E.; Yang, C.S. Effects of bioflavonoids on hepatic p450 activities. *Xenobiotica* 2009, 25, 575–584.

63. Kim, H.; Yoon, Y.-J.; Shon, J.-H.; Cha, I.-J.; Shin, J.-G.; Liu, K.-H. Inhibitory effects of fruit juices on cyp3a activity. *Drug Metab. Dispos.* 2006, 34, 521–523.
64. Zhang, H.; Ma, Z.; Luo, X.; Li, X. Effects of mulberry fruit (*morus alba* L.) consumption on health outcomes: A mini-review. *Antioxidants* 2018, 7, 69.
65. Rodríguez-Fragoso, L.; Reyes-Esparz, J. Fruit/vegetable-drug interactions: Effects on drug metabolizing enzymes and drug transporters. In *Drug Discovery*; IntechOpen, London, UK, 2013; pp. 1-33.
66. Piver, B.; Berthou, F.; Dreano, Y.; Lucas, D. Inhibition of cyp3a, cyp1a and cyp2e1 activities by resveratrol and other non volatile red wine components. *Toxicol. Lett.* 2001, 125, 83–91.
67. Hidaka, M.; Fujita, K.-i.; Ogikubo, T.; Yamasaki, K.; Iwakiri, T.; Okumura, M.; Kodama, H.; Arimori, K. Potent inhibition by star fruit of human cytochrome p450 3a (cyp3a) activity. *Drug Metab. Dispos.* 2004, 32, 581–583.
68. Eskra, J.N.; Schlicht, M.J.; Bosland, M.C. Effects of black raspberries and their ellagic acid and anthocyanin constituents on taxane chemotherapy of castration-resistant prostate cancer cells. *Sci. Rep.* 2019, 9, 1–12.
69. Gerber, W.; Steyn, J.; Kotzé, A.; Hamman, J. Beneficial pharmacokinetic drug interactions: A tool to improve the bioavailability of poorly permeable drugs. *Pharmaceutics* 2018, 10, 106.
70. Mallhi, T.H.; Sarriif, A.; Adnan, A.S.; Khan, Y.H.; Qadir, M.I.; Hamzah, A.A.; Khan, A.H. Effect of fruit/vegetable-drug interactions on cyp450, oatp and p-glycoprotein: A systematic review. *Trop. J. Pharm. Res.* 2015, 14, 1927–1935.
71. Taniguchi, S. Treatment of psoriasis by cyclosporine and grapefruit juice. *Arch. Dermatol.* 1996, 132.
72. van Agtmael, M.A.; Gupta, V.; van der Wösten, T.H.; Rutten, J.P.B.; van Boxtel, C.J. Grapefruit juice increases the bioavailability of artemether. *Eur. J. Clin. Pharmacol.* 1999, 55, 405–410.
73. Adegoke, S.A.; Oyelami, O.A.; Olatunya, O.S.; Adeyemi, L.A. Effects of lime juice on malaria parasite clearance. *Phytother. Res.* 2011, 25, 1547–1550.
74. Zhong, Y.; Wang, Y.; Guo, J.; Chu, H.; Gao, Y.; Pang, L. Blueberry improves the therapeutic effect of etanercept on patients with juvenile idiopathic arthritis: Phase iii study. *Tohoku J. Exp. Med.* 2015, 237, 183–191.
75. Blanco-Rojo, R.; Pérez-Granados, A.M.; Toxqui, L.; González-Vizcayno, C.; Delgado, M.A.; Vaquero, M.P. Efficacy of a microencapsulated iron pyrophosphate-fortified fruit juice: A randomised, double-blind, placebo-controlled study in spanish iron-deficient women. *Br. J. Nutr.* 2011, 105, 1652–1659.