Biotechnology of Amaryllidaceae Alkaloids

Subjects: Plant Sciences

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Plants belonging to the monocotyledonous Amaryllidaceae family include about 1100 species divided among 75 genera. They are well known as medicinal and ornamental plants, producing pharmaceutically important alkaloids, the most intensively investigated of which are galanthamine and lycorine. Amaryllidaceae alkaloids possess various biological activities, the most important one being their anti-acetylcholinesterase activity, used for the treatment of Alzheimer's disease. Due to increased demand for Amaryllidaceae alkaloids (mainly galanthamine) and the limited availability of plant sources, in vitro culture technology has attracted the attention of researchers as a prospective alternative for their sustainable production. Plant in vitro systems have been extensively used for continuous, sustainable, and economically viable production of bioactive plant secondary metabolites. Over the past two decades, a significant success has been demonstrated in the development of in vitro systems synthesizing Amaryllidaceae alkaloids. The present material discusses the state of the art of in vitro Amaryllidaceae alkaloids production, summarizing the authors' point of view on the development of biotechnological production processes with a focus on the future prospects of in vitro culture technology for the commercial production of these valuable alkaloids.

alkaloids Amaryllidaceae bioreactors galanthamine plant in vitro systems

1. Introduction

Plants belonging to the monocotyledonous Amaryllidaceae family include about 1100 species divided among 75 genera (*Amaryllis*, *Galanthus*, *Leucojum*, *Narcissus*, *Haemanthus*, *Nerine*, *Hippeastrum*, *Sternbergia*, *Clivia*, *Rhodophiala*, *Pancratium*, *Hymenocallis*, *Crinum*, *Lycoris*, etc.), which are well known, mainly as medicinal and ornamental plants [1][2]. The Amaryllidaceae family is one of the most important alkaloid-producing plant families [3], as the most intensively investigated alkaloids are galanthamine and lycorine [4]. Due to the increased demand for Amaryllidaceae alkaloids (mainly galanthamine) and the limited availability of plant sources, in vitro culture technology has attracted the attention of researchers as a prospective alternative for their sustainable production of bioactive plant secondary metabolites [6]. A significant success has been demonstrated in the development of in vitro systems for production of Amaryllidaceae alkaloids [7][8][9][10][11][12]. Optimization of the production process is also well documented, starting with nutrient media optimization [13], development of appropriate bioreactor design [8] [14][15], and optimization of the cultivation conditions [11][16][17].

2. Biotechnological Production of Amaryllidaceae Alkaloids

The development of biotechnologies for the production of biologically active substances based on plant cells and tissues cultured under in vitro conditions is a complex and multi-stage process. Algorithms for optimizing and controlling the process of biosynthesis of the target metabolite in the in vitro systems under study should be based on the bioengineering, physiological and phytochemical peculiarities of the particular in vitro culture, as well as on the subsequent analysis of the relationships in the "in vitro system—product" biological system^{[18][19]}. A preliminary requirement for industrially significant yields of target biologically active substances from plant in vitro systems is the development of an efficient cultivation system with appropriately designed bioreactors, as well as unconventional strategies for optimization of the biological systems^{[20][21]}.

Based on the summarized results of extensive research in this field over a period of more than 20 years, we propose an integrated approach to process development for production of Amaryllidaceae alkaloids by plant in vitro systems (Figure 1).

Figure 1. Integrated approach to biotechnological production of Amaryllidaceae alkaloids.

It is postulated that the genetic potential of an intact plant from which explants are extracted to produce in vitro systems directly affects the biosynthetic potential of in vitro systems. This fact is often underestimated by plant biotechnologists, although it is the basis for the successful development of technologies for bioactive secondary metabolite production. It is clear that the selection of the plant individuals further used as primary material for development of a plant in vitro system should be based on systematic screening procedures. Extensive botanical and phytochemical evaluation of the wild populations and/or clone origin of the field crop is required to select the most suitable plant individuals for further in vitro work. In our opinion, there is a need of more thorough investigations on the intrapolulation variations of alkaloids in wild habitats in order to develop and validate procedures for selecting the most productive plant individuals for further introduction in vitro.

Data available in the scientific literature clearly show that undifferentiated in vitro systems (callus) derived from Amaryllidacea species exhibit lower biosynthetic potential compare to differentiated plant in vitro systems [6][22][23][24] [25][26][27]. This fact leaves open many questions regarding the scale-up of cultivation processes, because the difficulties associated with bioreactor cultivation of differentiated plant in vitro systems are well known [28].

Recently, in attempts to respond to the demand of Amaryllidaceae alkaloids, substantial efforts have been made to improve the yields and biosynthesis efficiency of target molecules, including the application of metabolic engineering technologies. In contrast to the fast-growing knowledge about Amaryllidaceae alkaloids chemistry and pharmacological activities, very little is known about their biosynthesis, the responsible functional and regulatory genes, and the molecular mechanisms underlying its regulation [1][29][30]. Although rapid advances in omics technologies over the past few years have led to significant discoveries in Amaryllidaceae alkaloid biosynthesis, there are still many unknown details about their biosynthetic pathway. The biosynthesis of Amaryllidaceae alkaloids is a complex process and is a result of coordinated action of enzymes from different biosynthetic pathways of primary, secondary and specialized cell metabolism. According to Desgagne'-Penix 30, the biosynthetic pathway of Amaryllidaceae alkaloids could be provisionally divided into five stages: 1) Biosynthesis of the two aromatic amino acid L-phenylalanine and L-tyrosine, which are the building blocks of Amaryllidaceae alkaloids; 2) Formation of 3,4dihydroxybenzaldehyde from phenylalanine, which is the aldehyde moiety of Amaryllidaceae alkaloids; 3) Formation of 4'-O-methylnorbelladine from norbelladine (a condensation product of tyramine with 3,4dihydroxybenzaldehyde); 4) Formation of unstable intermediate products by specific phenol coupling of 4'-Omethylnorbelladine followed by a reduction step; 5) Biosynthesis of the different types of Amaryllidaceae alkaloids (galanthamine, lycorine, homolycorine, galasine, haemanthamine, plicamine, secoplicamine, narciclasine, pretazettine, crinine, cripowelline, graciline, montanine, ismine, cherylline and norbelladine types, as well as galanthindole, maritinamine and elwesine). The last stage involves very specialized enzymes which could be found in different plant species producing specific Amaryllidaceae alkaloids [30][31].

Obviously, a better understanding of Amaryllidaceae alkaloid biosynthesis is crucial so that the advantages of metabolic engineering can be applied in constructing effective plant or microbial systems able to produce a desirable molecule. The development of such expression systems will have a great impact on the cost of Amaryllidaceae alkaloids, their availability and sustainable production. Till now, there is no enough information about the structural and regulatory genes involved into fifth stage of Amaryllidaceae alkaloids biosynthesis, and thus, we cannot suggest the optimal strategy for engineering this part of the pathway. Nowadays, there is no report on engineered plant or microbial system expressing target Amaryllidaceae alkaloids.

As stated in the Introduction, the optimization of the production process of Amaryllidaceae alkaloids is well documented, especially in the case of galanthamine [8][11][13][14][15][16][17]. It is clear that the main stages of such optimization are: optimization of the nutrient medium for maximal yields of the target alkaloids; selecting the appropriate cultivation system and improving its design; optimization of the environmental conditions of cultivation and process control, management and modeling. In our opinion, elicitation is one of the most effective approaches to optimizing the biosynthesis of Amaryllidaceae alkaloids and it is a mandatory step in a future technology for their production [16]. Important steps for effective elicitation are the experimental determination of the type of elicitor, its

concentration and time of addition to the cultivation system, and these experiments should be carried out after the cultivation system is selected, as well as the composition of the nutrient medium and environmental conditions of cultivation are optimized. Elicitation is also a powerful tool for directing biosynthesis to one or another group of Amaryllidaceae alkaloids depending on the specific aim of the production process being developed^[16]. As we stated above, the differentiated shoot type cultures are most suitable for the biosynthesis of Amaryllidaceae alkaloids, which creates problems with the development and optimization of cultivation systems^[28]. For the production of Amaryllidaceae alkaloids it seems that temporary immersion cultivation systems and modified bubble column bioreactors are the most appropriate^[15].

3. Conclusions and Future Prospects

Many of the known Amaryllidaceae alkaloids show remarkable biological activities and some of them, galanthamine for example, are already used in medicine. However, the limited amounts found in plants, in combination with the wide diversity in the chemical structures of these alkaloids render their production, isolation and purification very expensive.

Plant in vitro systems are an alternative approach to Amaryllidaceae alkaloid bioproduction. Nowadays, alkaloid yields are still too low and not attractive for commercial production despite the huge progress in the development of an integrated approach for their biotechnological production. It seems this disadvantage could be solved using metabolic engineering for constructing effective plant or microbial systems able to produce a desirable molecule. Possible solutions could be: 1) a combination of chemical synthesis of norcraugsodine as precursor and its conversion into nornarwedine by engineered microbial system; or 2) developing hybrid biosynthetic pathway for Amaryllidaceae alkaloids synthesis by yeasts or in vitro plant systems; or 3) developing hybrid bioprocess systems.

References

- 1. Takos, A.M.; Rook, F. Towards a molecular understanding of the biosynthesis of amaryllidaceae alkaloids in support of their expanding medical use. Int. J. Mol. Sci. 2013, 14, 11713–11741.
- 2. Jin, Z.; Xu, X.H. Amaryllidaceae alkaloids. In Natural products: Phytochemistry, Botany and Metabolism of Alkaloids, Phenolics and Terpenes; Ramawat, K.G., Mérillon, J.-M., Eds.; Springer-Verlag: Berlin, Germany, 2013; pp. 479–522.
- 3. Jin, Z. Amaryllidaceae and Sceletium alkaloids. Nat. Prod. Rep. 2005, 22, 111–126.
- 4. Bastida, J.; Lavilla, R.; Viladomat, F. Chemical and biological aspects of Narcissus alkaloids. In Alkaloids: Chemistry and Biology; Geoffrey, A.C., Ed.; Academic Press: New York, NY, USA, 2006; Volume 63, pp. 87–179.
- 5. Pavlov, A.; Berkov, S.; Courot, E.;;Gocheva, T., Tuneva, D.; Pandova, B.; Georgiev, M.; Georgiev, V.; Yanev, S.; Burrus, M. Galanthamine production by Leucojum aestivum in vitro systems. Proc.

- Biochem. 2007, 42, 734-739.
- 6. Ferdausi, A.; Chang, X.; Hall, A.; Jones, M. Galanthamine production in tissue culture and metabolomics study on Amaryllidaceae alkaloids in Narcissus pseudonarcissus cv. Carlton. Ind. Crop. Prod. 2020, 144, 112058.
- 7. Bogdanova, Y.; Pandova, B.; Yanev, S.; Stanilova, M.; Biosynthesis of lycorine by in vitro cultures of Pancratium maritimum L. (Amaryllidaceae). Biotechnol. Biotechnol. Eq. 2009, 23, 919–922.
- 8. Georgiev, V.; Ivanov, I.; Berkov, S.; Ilieva, M.; Georgiev, M.; Gocheva, T.; Pavlov, A. Galanthamine production by Leucojum aestivum L. shoot culture in a modified bubble column bioreactor with internal sections. Eng. Life Sci. 2012, 12, 534–543.
- 9. Schumann, A.; Berkov, S.; Claus, D.; Gerth, A.; Bastida, J.; Codina, C. Production of galanthamine by Leucojum aestivum shoots grown in different bioreactor systems. Appl. Biochem. Biotechnol. 2012, 167, 1907–1920.
- 10. Stanilova, M.I.; Molle, E.D.; Yanev, S.G. Galanthamine production by Leucojum aestivum cultures in vitro. In The Alkaloids; Cordell, G.A., Ed.; Elsevier: Amsterdam, The Netherlands, 2010; Volume. 68, pp. 167–270.
- 11. Berkov, S.; Ivanov. I.; Georgiev, V.; Codina, C.; Pavlov, A. Galanthamine biosynthesis in plant in vitro systems. Eng. Life Sci. 2014, 14, 643–650.
- 12. Laurain-Mattar, D.; Ptak, A. Amaryllidaceae Alkaloid accumulation by plant in vitro systems. In Bioprocessing of Plant In Vitro Systems, Reference Series in Phytochemistry; Pavlov, A., Bley, Th., Eds.; Springer International Publishing: Geneva, Switzerland, 2018; pp. 203–224.
- 13. Georgiev, V.; Berkov, S.; Georgiev, M.; Burrus, M.; Codina, C.; Bastida, J.; Ilieva, M.; Pavlov. A. Optimized nutrient nedium for galanthamine production in Leucojum aestivum L. in vitro shoot system. Z. Naturforsch. 2009, 64C, 219–224.
- 14. Ivanov, I.; Georgiev, V.; Georgiev, M.; Ilieva, M.; Pavlov, A. Galanthamine and related alkaloids production by Leucojum aestivum L. shoot culture using a temporary immersion technology. Appl. Biochem. Biotechnol. 2011, 163, 268–277.
- 15. Ivanov, I.; Georgiev, V.; Berkov, S.; Pavlov, A. Alkaloid patterns in Leucojum aestivum shoot culture cultivated at temporary immersion conditions. J. Plant. Physiol. 2012, 169, 206–211.
- 16. Ivanov, I.; Georgiev, V.; Pavlov, A. Elicitation of galanthamine biosynthesis by Leucojum aestivum liquid shoot cultures. J. Plant. Physiol. 2013, 170, 1122–1129.
- 17. Ivanov, I.; Berkov, S.; Pavlov, A.; Georgiev, V. In sito galanthamine extraction during the cultivation of Leucojum aestivum L. shoot culture in two—Phase bubble column cultivation system. Eng. Life Sci. 2019, 19, 1000–1005.
- 18. Pavlov, A. Plant cells and algae in bioreactors. Eng. Life Sci. 2009, 9, 154–155.

- 19. Pavlov, A. Plant cells and algae in bioreactors II. Eng. Life Sci. 2014, 14, 548–549.
- 20. Pavlov, A. Plant cell and algae in bioreactors III. Eng. Life Sci. 2019, 19, 828–829.
- 21. Pavlov, A.; Bley, T. Bioprocessing of Plant In Vitro Systems, Reference Series in Phytochemistry; Springer International Publishing: Geneva, Switzerland, 2018; ISBN:978-3-319-54599-8.
- 22. Berkov, S.; Pavlov, A.; Georgiev, V.; Bastida, J.; Burrus, M. Ilieva, M.; Codina, C. Alkaloid synthesis and accumulation in Leucojum aestivum in vitro cultures. Nat. Prod. Comm. 2009, 4, 359–364.
- 23. Tarakemeh, A.; Azizi, M.; Rowshan, V.; Salehi, H.; Spina, R.; Dupire, F.; Arouie, H.; Laurain-Mattar, D. Screening of Amaryllidaceae alkaloids in bulbs and tissue cultures of Narcissus papyraceus and four varieties of Narcissus tazetta. J. Pharm. Biomed. Anal. 2019, 172, 230–237.
- 24. 24 Tarakemeh, A.; Azizi, M.; Rowshan, V.; Salehi, H.; Spina, R.; Dupire, F.; Arouie, H.; Laurain-Mattar, D. Quantitative determination of lycorine and galanthamine in different in vitro tissues of Narcissus tazetta by GC-MS. Int. J. Hort. Sci. Technol. 2019, 6, 151–157.
- 25. Subramaniam, S.; Sundarasekar, J.; Sahgal, G.; Murugaiyah, V. Comparative analysis of lycorine in wild plant and callus culture samples of Hymenocallis littoralis by HPLC-UV method. Sci. World J. 2014, 408306.
- 26. Ptak, A.; El Tahchy, A.; Dupire, F.; Boisbrun, M.; Henry, M.; Chapleur, Y.; Mos, M.; Laurain-Mattar, D. LCMS and GCMS for the screening of alkaloids in natural and in vitro extracts of Leucojum aestivum. J. Nat. Prod. 2009, 72, 142–147.
- 27. Ptak, A.; El Tahchy, A.; Skrzypek, E.; Wójtowicz, T.; Laurain-Mattar, D. Influence of auxins on somatic embryogenesis and alkaloid accumulation in Leucojum aestivum callus. Cent. Eur. J. Biol. 2013, 8, 591–599.
- 28. Steingroewer, J.; Bley, Th.; Georgiev, V.; Ivanov, I.; Lenk, F.; Marchev, A.; Pavlov, A. Bioprocessing of differentiated plant in vitro systems. Eng. Life Sci. 2013, 13, 26–38.
- 29. Kilgore, M.B.; Kutchan, T.M. The amaryllidaceae alkaloids: Biosynthesis and methods for enzyme discovery. Phytochem. Rev. 2016, 15, 317–337.
- 30. Desgagné-Penix, I. Biosynthesis of alkaloids in Amaryllidaceae plants: A review. Phytochem. Rev. 2020, doi:10.1007/s11101-020-09678-5.
- 31. Berkov, S.; Osorio, E.; Viladomat, F.; Bastida, J. Chapter two—Chemodiversity, chemotaxonomy and chemoecology of amaryllidaceae alkaloids. In The alkaloids: Chemistry and Biology, Knölker, H.-J., Ed. Academic Press: New York, NY, USA, 2020; Volume 83, pp 113–185.

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