Flavonoids in Skin Senescence Prevention

Skin aging is associated with the accumulation of senescent cells and is related to many pathological changes, including decreased protection against pathogens, increased susceptibility to irritation, delayed wound healing, and increased cancer susceptibility. Senescent cells secrete a specific set of pro-inflammatory mediators, referred to as a senescence-associated secretory phenotype (SASP), which can cause profound changes in tissue structure and function. Thus, drugs that selectively eliminate senescent cells (senolytics) or neutralize SASP (senostatics) represent an attractive therapeutic strategy for age-associated skin deterioration. There is growing evidence that plant-derived compounds (flavonoids) can slow down or even prevent aging-associated deterioration of skin appearance and function by targeting cellular pathways crucial for regulating cellular senescence and SASP. This review summarizes the senostatic and senolytic potential of flavonoids in the context of preventing skin aging.

1. Introduction

Besides being an economic and social problem, aging is predominantly a medical issue. Thus, there is an increasing need to understand the mechanisms underlying this highly complex process[1], which inevitably leads to impaired body homeostasis and function, an increased risk of complex diseases, and, finally, death.

Cellular senescence contributes to age-related tissue and organ dysfunction and diseases through mechanisms that perturb stem cell niches, induce aberrant cell differentiation, disrupt the extracellular matrix, stimulate tissue inflammation, and induce senescence in neighboring cells [2][3]. It is believed that the accumulation of senescent cells in tissues contributes to the impairment of their homeostasis and increases the risk of many age-related diseases[4]. Therefore, eliminating senescent cells or neutralizing SASP components may provide beneficial effects not only for the affected tissue but also the whole organism. Drugs that selectively eliminate senescent cells (senolytics) or neutralize SASP (senostatics) represent an attractive therapeutic strategy for delaying aging and age-related diseases[5].

Skin aging is associated with an increasing number of senescent cells and is related to many pathological changes, including decreased protection against pathogens, increased susceptibility to irritation, delayed wound healing, and increased cancer susceptibility[6]. Therefore, therapies that reduce senescent cell numbers or block SASP may be an effective treatment for aging-associated skin deterioration[7]. The senolytic and senostatic activities of several drugs (e.g., metformin and rapamycin) have already been demonstrated in preliminary clinical trials[8][9]. However, in vitro and in vivo data show that different flavonoids have similar properties; therefore, they can be considered a therapeutic option for skin aging prevention and treatment.
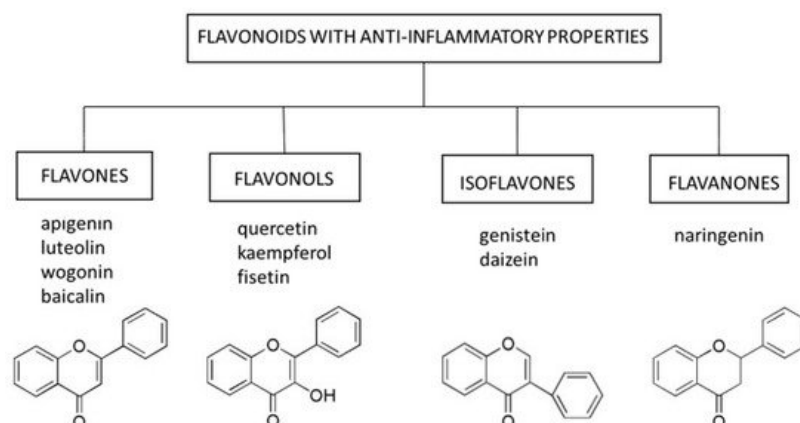
2. Therapeutic Strategies Targeting Skin Senescence

Due to the harmful effects of senescent cells and SASP components on many tissues, strategies aimed at selective induction of senescent cell death or inhibiting SASP without affecting the selective induction of death of surrounding cells are currently being investigated[10]. Removal of senescent cells from aging tissues is considered a promising anti-aging therapy. However, under certain circumstances, such skin cells can also play a positive role[11]. Therefore, SASP modification and maintaining the beneficial features of cell senescence seem to be a more rational therapeutic approach than senescent cell removal.

Complex signaling pathways control SASP production. Nuclear factor κ-light-chain enhancer of activated B cells (NF-κB) is a crucial transcription factor for SASP induction. However, the DNA damage response (DDR), p38 mitogen-activated protein kinase (MAPK), CCAAT/enhancer-binding protein b (C/EBPb), mechanistic target of rapamycin (mTOR), phosphoinositide-3-kinase (PI3K), Janus kinase/signal transducer and activator of transcription (JAK/STAT), protein kinase D1, and several other factors are also involved in regulating SASP production by senescent cells[12].

Different drugs specifically block the signals associated with senescent cell secretion. For example, glucocorticosteroids can reduce SASP secretion and inflammation induced by senescent cells and SASP due to their ability to decrease the transcriptional activity of NF-κB However, several adverse side effects of glucocorticoid treatment (e.g., skin thinning and impaired wound healing) limit their application as skin senolytics[13]. There is growing evidence that flavonoids can prevent skin from aging by targeting cellular pathways crucial for regulating cellular senescence and SASP production.

3. Flavonoids as a Senostatic and Senolytic Strategy

Flavonoids are natural substances with variable phenolic structures containing 15 carbon atoms. They consist of two benzene rings connected by a short three-carbon chain. One of the carbons in this chain is connected to carbon in one of the benzene rings, either through an oxygen bridge or directly yielding a third middle ring[14], **Figure1**. To date, over 8000 different flavonoids have been identified[15].

Flavonoids are divided into different subtypes: flavones, flavonols, isoflavones, flavanones, anthoxanthins, anthocyanins, and chalcones. They are present in fruits, vegetables, grains, flowers, tea, and wine, and are well-known for their beneficial effects on health. Flavonoids are an indispensable component of various pharmaceutical, medical, and cosmetic applications due to their antioxidative, anti-inflammatory, anti-mutagenic, and anti-carcinogenic properties coupled with their capacity to modulate critical enzyme functions. All these features make flavonoids excellent candidates for anti-aging therapies.

The enhanced binding of NF-κB to nuclear DNA is one of the hallmarks of aging and is observed in several tissues. NF-κB is a critical transcription factor involved in the production of SASP and the pathogenesis of many age-related disorders, including inflammatory and metabolic diseases[16]. Several flavonoids can disrupt the activation of NF-κB and related pathways, including the kinase 1 signaling pathway associated with the IL-1 receptor (IRAK1)/IκBα and IκBζ, which blocks SASP in vitro[17]. Structural analyses using synthetic flavones revealed that hydroxyl substitutions at C-2′, 3′, 4′, 5′, and 7' are essential in inhibiting SASP production[18]. Furthermore, flavonoids have a protective effect in animal models of age-related disorders by preventing increased production of IL-1β and tumor necrosis factor (TNF)-α[19].

In this review, we focused on select representatives of flavones, flavonols, isoflavones, and flavanones, whose anti-inflammatory potential in the context of skin cell senescence has been demonstrated in vitro or in vivo (**Figure 1**). However, it should be mentioned that several other compounds from the group of flavonoids (e.g., curcumin) are being tested for their senolytic and senostatic properties in the context of skin disorders[20].

3.1. Flavones

Flavones occur in a wide variety of fruits, vegetables, and cereal grains in the form of glycosides. As with other flavonoid glycosides in foods, flavones must be hydrolyzed to aglycones to be absorbed. They are then metabolized to glucuronidated or sulfated forms before reaching systemic circulation. The main flavones in the diet are apigenin and luteolin; however, some other compounds (e.g., baicalin and wogonin) are also worth mentioning[21].

3.1.1. Apigenin

Apigenin, a flavone present in select fruits, vegetables, and herbs, can induce apoptosis and inhibit proliferation and angiogenesis in several cancer cell lines[22]. The anti-cancer activities of apigenin result from its ability to interact with the PI3K/protein kinase B (ERK)/mTOR, JAK/STAT, NF-κB, MAPK, and Wnt/β-catenin pathways[23]. Interference with mTOR signaling is a dominant mechanism by which apigenin inhibits skin cancer development and progression[24]. Moreover, apigenin has antioxidant and anti-inflammatory properties and can restore the proper function of the skin (e.g., DNA repair

and viability of human keratinocytes and dermal fibroblasts) after damage caused by exposure to UVA and UVB radiation[25][26][27]. The molecular mechanisms underlying these phenomena involve the ability of apigenin to inhibit the expression of cyclooxygenase-2 (COX-2) and the NF-κB pathway, which controls the inflammation caused by UVA and UVB radiation[22]. The interaction between apigenin and the NF-κB pathway also seems to be a key mechanism for reducing the secretion of several SASP factors (e.g., IL-6 and IL-8) in human fibroblasts induced to undergo senescence by bleomycin[18]. Moreover, topical administration of apigenin to mice exposed to UVB radiation reduced cutaneous inflammation by inducing thrombospondin 1 (TSP-1) expression and repressing IL-6 and IL-12 levels and inflammatory infiltrates[28].

Aging is associated with increased interferon-γ-inducible protein 10 (IP10) levels that can elicit abnormal immune responses in the elderly[29]. Interestingly, apigenin inhibits the production of IP10, a component of SASP secreted by senescent fibroblasts. IP10 and other chemokines (CXCL9 and CXCL11) promote a Th1 response to cellular damage. Apigenin protects the skin against UVA and UVB radiation-induced destruction of the collagen matrix, which causes loss of elasticity and skin dryness, by decreasing the activity of MMP-1. It also induces collagen type I and III de novo synthesis in dermal fibroblasts in vitro and increases dermal thickness and collagen deposition in the dermis in vivo in mice[30][31]. These anti-aging effects of apigenin were confirmed in clinical trials; its topical application improves markers of skin aging, such as firmness, elasticity, and fine wrinkling and maintains hydration[26][32].

3.1.2. Baicalin

Baicalin is a flavone isolated from the roots of *Scutellaria lateriflora Georgi* (Huang Qin In China) that plays a role in skin protection against UVB-induced photodamage[33]. This function is related to its anti-inflammatory and antioxidative properties through modulating NF-κB, COX-1, and inducible nitric oxide synthase (iNOS) activity[34]. By inhibiting the UV-induced generation of ROS in fibroblasts, baicalin prevents the activation of transcription factors (e.g., activator protein 1, AP-1) responsible for the transcription of MMP-encoding genes and subsequent collagen degradation. The senolytic properties of baicalin are not limited to its effects on SASP. This flavone can also decrease the percentage of β-galactosidase-positive cells and p16, p21, and p53 expression in UVB-treated fibroblast cultures[35]. Moreover, treatment of skin fibroblasts with baicalin reduces the number of DNA double-strand breaks induced by UVB[35]. Anti-mutagenic properties of baicalin were also demonstrated in keratinocytes, where this flavone prevented the formation of oxidative adducts induced by UVC[36]. However, it should be stressed that baicalin does not affect cells that have not been exposed to UV irradiation.

3.1.3. Luteolin

The flavone luteolin is a glycoside found in flowers, herbs, vegetables, and spices. After consumption, it is metabolized to the active aglycone, which has antioxidative properties due to the unique luteolin chemical structure. The C2–C3 double bond donates a hydrogen/electron and stabilizes the radical species and the oxo group at C4 that binds transitional metal ions (e.g., iron and copper) to prevent oxidative damage. By decreasing ROS production, luteolin modulates several cellular pathways, including MAPK and NF-κB, and several downstream genes (e.g., COX-2, IL-6, IL-1β, TNF-α), producing an anti-inflammatory effect[37]. These properties are of particular importance in the context of skin photoaging. Luteolin reduces UV-induced ROS production and subsequent release of pro-inflammatory cytokines (e.g., IL-6 and IL-20) from keratinocytes and MMP-1 from fibroblasts[38][39]. By decreasing ROS production, luteolin prevents increased degradation of hyaluronic acid, which, together with collagen, is the major non-fibrous component of the dermis and epidermis extracellular matrix[40]. Moreover, luteolin alone or in combination with apigenin can directly inhibit UVB-induced MMP-1 production in fibroblasts by inhibiting the Ca^{2+} influx that prevents the phosphorylation of Ca^{2+}/calmodulin-dependent MAPKs and binding of the AP-1 transcription factor to the promoter of the MMP-1 gene[41][42].

3.1.4. Wogonin

Wogonin is a flavone extracted from *Scutellaria baicalensis* with proven efficacy as a SASP regulator in cancer[43]. By inactivating the MAPK/AP-1 and NF-κB/IκBα signaling pathways, wogonin downregulates COX-2 and iNOS expression in skin fibroblasts and MMP-1 and IL-6 in UVB-induced keratinocytes[44]. Moreover, treatment with wogonin effectively restores procollagen type I levels and increases the expression of cytoprotective antioxidants (e.g., heme oxygenase-1 [HO-1] and NAD(P)H dehydrogenase [quinone] 1 [NQ-O1]) in keratinocytes by activating the tumor growth factor β (TGF-β)/Smad pathway[45]. Wogonin also reduces dermis prostaglandin E2 (PGE2), TNF-α, intercellular adhesion molecule-1 (ICAM1), and IL-1β levels in an animal model of skin inflammation when applied topically[44][46][47].

3.2. Flavonols

Flavonols are the most ubiquitous flavonoids in foods, including fruits, vegetables, red wine, and tea, and are represented by quercetin, kaempferol, and fisetin. Like other flavonoids, flavonols accumulate in plant tissue in glycosylated forms linked to mono-, di-, and tri-saccharides. Due to their antioxidative, anti-inflammatory, anti-cancerogenic, and vasodilating properties, flavonols have many benefits to human health, including their effects on senescence[48].

3.2.1. Quercetin

Quercetin is present in red wine, fruits, and vegetables. It can interact with protein kinase C (PKC) δ and Janus kinase 2 (JAK2) to block UV-induced expression of COX-2 and MMP-1 and collagen degradation in human skin and skin fibroblasts[49]. JAK2 kinase is an upstream regulator of STAT3. The STAT3 pathway is involved in stimulating inflammatory responses. In turn, PKCδ is a regulator of the MAPK and Akt signaling pathways and modulates the expression of collagen genes in skin cells[50]. Similar findings came from the study with quercetin surface-functionalized Fe_3O_4 nanoparticles (MNPQ). MNPQ-stimulated 5′ AMP-activated protein kinase (AMPK) activity in skin fibroblasts is accompanied by a decrease in the number of stress-induced senescent cells and suppression of senescence-associated secretion of the inflammatory mediators IL-8 and interferon-β[51]. In keratinocytes, quercetin decreases UV-induced activation of NF-κB, resulting in suppressed expression of IL-1β, IL-6, IL-8, and TNF-α. It did not affect UV-mediated activation of ERK, JNK, or p38. Moreover, the induction of AP-1 target genes (e.g., MMP-1 and MMP-3) is not suppressed by quercetin[52]. Apart from being senostatic, quercetin also has senolytic properties. The combination of dasatinib and quercetin effectively eliminates senescent fibroblasts in vitro and reduces senescence of primary mouse embryonic fibroblasts (MEFs) in vivo in chronologically aged or radiation-exposed mice and also progeroid mice model[5] .

3.2.2. Kaempferol

The flavonol kaempferol is found in many edible or traditional medicine plants and has antioxidant and anti-inflammatory properties by inhibiting the iNOS, COX-2, and NF-κB pathways[53]. Administration of kaempferol to aged (24-week-old) rats reduces the accumulation of advanced glycation endproducts (AGE) in different organs and decreases the expression of AGE receptor (RAGE) and AGE-induced reactive species (RS). Because RS are potent activators of NF-κB, both kaempferol-treated fibroblasts and animals have lower expression of MMP-9, adhesion molecules (e.g., ICAM-1), and several pro-inflammatory genes. Accordingly, in bleomycin-induced senescent fibroblasts and aged rats, kaempferol inhibits the induction of a subset of SASP mRNA and the activation of the NF-κB pathway[18].

3.2.3. Fisetin

Fisetin is a flavonol with a chemical structure similar to quercetin. It is present in many fruits and vegetables (e.g., apples, persimmon, grapes, onions, and cucumbers) at relatively low concentrations and at high concentrations in strawberries. Fisetin has demonstrated potent senolytic and senostatic properties in vitro and in vivo. Administration of fisetin to progeroid and old wild-type mice reduces senescence markers (i.e., p16 and p21), modifies the SASP composition in multiple tissues, and restores tissue homeostasis by inhibiting the PI3K/AKT/mTOR and NF-κB pathways and antioxidative activity[54].

In the context of skin aging, fisetin can inhibit TNF-α-induced inflammation and hydrogen peroxide-induced oxidative damage in human keratinocytes[55]. It can also decrease UVB-induced damage by inhibiting ROS generation and the MAPK/AP-1/MMP signaling pathway and decreasing collagen degradation and the inflammatory response in human skin fibroblasts[56]. When applied topically to hairless mice, fisetin inhibits iNOS, MMP-1, MMP-2, and COX-2 and increases the skin expression of filaggrin and aquaporins, protecting the animals from photo-inflammation and skin-drying[57]. Clinical trials are currently underway to evaluate the benefits of fisetin treatment on several aspects of aging[58].

3.3. Isoflavones

Isoflavones are non-active hydrophilic glycosides (e.g., daidzin and genistein in soybean) or methylated lipophilic derivatives (e.g., formononetin and biochanin A in red clover) in the plants from the *Leguminosae* family that are hydrolyzed by β-glucosidases in the gastrointestinal tract. These bioactive aglycones (e.g., daidzein and genistein formed from daidzin and genistin, respectively) are absorbed across the intestinal epithelium and metabolized to β-glucuronides and sulfate esters in the intestinal mucosa cells. These metabolites are subsequently excreted into plasma and bile[59].

The pleiotropic effects of isoflavones depend on their ability to interact with several nuclear receptors, including estrogen receptors (ER) α and β; peroxisome proliferator-activated receptors (PPARs) α, δ and γ; retinoid acid receptor (RAR); and aryl hydrocarbon receptor (AhR). However, the isoflavones also act by nuclear receptor-independent mechanisms,

including inhibition of protein tyrosine kinases (e.g., ERK1/2, crucial for regulating cell proliferation and differentiation), reduction of ROS levels, induction of antioxidant enzymes, and inhibition of COX-1 and NF-κB activity and thromboxane A_2 (TXA$_2$) synthesis. All these functions contribute to the anti-inflammatory properties of the isoflavones[16].

3.3.1. Daidzein and Genistein

Daidzein alone or in combination with genistein inhibits UV-induced MMP-1 and MMP-2 expression and collagen degradation in human skin fibroblasts in vitro and in hairless mice in vivo[60]. UV radiation can disrupt the skin collagen matrix by inhibiting the TGF-β pathway[51]. Daidzein increases TGF-β expression and activates its receptors (signal transducer and activator of transcription 2/3—Smad2/3) in skin fibroblasts. Importantly, daidzein does not affect skin cell viability[61]. Moreover, through its interaction with RAR in human keratinocytes, daidzein can inhibit the expression of MMP-9, a metalloproteinase involved in the development of chronic ulcers in diabetic patients[62].

Genistein prevents UV-dependent COX-2 expression in human keratinocytes in vitro and the release of pro-inflammatory mediators[63]. Moreover, topical genistein or its metabolite equol protects against UVB-induced oxidative DNA damage (DNA pyrimidine dimer formation) and ROS production in the skin of hairless mice[64]. Like daidzein, genistein increases the thickness of skin collagen fibers by inducing TGF-β expression and increasing tissue inhibitor of metalloproteinase (TIMP) protein levels[65]. Both genistein and daidzein have significant anti-inflammatory effects and promote genomic and mitochondrial DNA repair in human skin fibroblasts exposed to UVB radiation (REF). They also work synergistically to produce a photoprotective effect[66][67]. Moreover, daidzein and genistein stimulate the production of hyaluronic acid in transformed human keratinocyte culture and hairless mouse skin[68].

There are studies suggesting that the administration of isoflavones can reverse the symptoms of skin aging in humans. For instance, 12-week systemic treatment with 40 mg of soy isoflavone aglycones improved fine wrinkles and skin elasticity in middle-aged Japanese women[69]. However, 24-week topical genistein administration had no superiority over estradiol and was less effective than this hormone in improving epidermal thickness, the number of dermal papillae, fibroblasts, and vessels in post-menopausal women[70].

3.4. Flavanones

Flavanones are found chiefly in citrus fruits; the most abundant flavanone is naringenin present in grapefruits, lemons, tangerines, and oranges. Naringenin has many pharmacological properties, including anti-atherogenic, anti-cancer, antioxidant, and anti-inflammatory. In the context of skin aging, naringenin can protect human keratinocytes against UVB-induced carcinogenesis and aging in vitro and UVB-generated oxidative stress and inflammation in vivo[71][72]. Topical naringenin protects hairless mice from UVB-induced skin damage by inhibiting the production of SASP components (TNF-α, IL-1β, IL-6, and IL-10) and lipid hydroperoxides, while maintaining the expression of antioxidant genes, including glutathione peroxidase 1, glutathione reductase, and the nuclear factor erythroid 2-related factor 2 (Nrf2) transcription factor[73]. These effects are partly due to the ability of naringenin to decrease NF-κB, MMP-1, and MMP-3 levels[74].

4. Summary and Conclusions

Targeting senescent cells has become an alternative therapy for treating various age-related conditions and diseases. This targeting can be achieved on two levels: specific elimination of senescent cells and inhibition of their secretory phenotype. Because senescent cells play a significant role in skin physiology and pathophysiology, their elimination may have unpredictable adverse effects. Therefore, modulation of SASP may be a safer strategy to counteract the senescence of skin cells. In vitro and in vivo studies suggest that the administration of flavonoids both topically and systemically has many benefits in this regard. However, due to the heterogeneity of study protocols, these pre-clinical findings cannot be translated directly into clinical practice. Therefore, we still lack convincing clinical studies to confirm the effectiveness and safety of flavonoids in treating age-related skin changes and lesions. Additional research is needed to optimize the appropriate treatment and assess the potential adverse effects of flavonoid applications. Clinical trials must be supported by solid pre-clinical results obtained in appropriate cellular and animal models. It is also necessary to develop a treatment scheme and appropriate cell markers to assess the effectiveness of the therapy. Moreover, research protocols should be unified so that the results obtained with different research models are comparable and translatable to clinical practice.

Taking into account the potential beneficial effect of flavonoids on skin aging, a diet rich in vegetables, fruits, and cereals, which are a natural source of these compounds, should be recommended in general anti-aging management. Importantly, natural products constitute a mixture of various flavonoids that can act comprehensively and synergistically and, therefore, are more effective than compounds evaluated in experimental settings. Furthermore, since flavonoids in natural products are present in mild/moderate concentrations, they can be safely administered without the risk of overdosage. Moreover,

pre-clinical trials demonstrated a wide safe therapeutic range of flavonoids. Therefore, nutraceuticals and dietary supplements containing both natural flavonoids as well as semi-synthetic and synthetic compounds with a variety of substituents and proven activity may be considered as a rational method of preventing skin aging.

References

1. Van Deursen, J.M.; The role of senescent cells in ageing. *Nature* **2014**, *509*, 439–446, 10.1038/nature13193.

2. Muñoz-Espín, D.; Serrano, M.; Cellular senescence: From physiology to pathology.. *Nat. Rev. Mol. Cell Biol.* **2014**, *15*, 482–496, 10.1038/nrm3823.

3. Jesus, B.B.; De Blasco, M.A.; Europe PMC Funders Group Assessing Cell and Organ Senescence Biomarkers.. *Circ. Res.* **2016**, *111*, 97–109, 10.1161/CIRCRESAHA.111.247866..

4. Kaur, J.; Farr, J.N.; Cellular senescence in age-related disorders. . *Transl. Res.* **2020**, *226*, 96–104., .

5. Zhu, Y.; Tchkonia, T.; Pirtskhalava, T.; Gower, A.C.; Ding, H.; Giorgadze, N.; Palmer, A.; Ikeno, Y.; Hubbard, G.B.; Lenburg, M.E.; et al. The Achilles' heel of senescent cells: From transcriptome to senolytic drugs.. *Aging Cell* **2015**, *14*, 644–658., .

6. Laberge, R.-M.; Sun, Y.; Orjalo, A.V.; Patil, C.K.; Freund, A.; Zhou, L.; Curran, S.C.; Davalos, A.R.; Wilson-Edell, K.A.; Liu, S.; et al. MTOR regulates the pro-tumorigenic senescence-associated secretory phenotype by promoting IL1A translation.. *Nat. Cell Biol.* **2015**, *17*, 1049–1061, .

7. Kirkland, J.L.; Tchkonia, T.; Cellular Senescence: A Translational Perspective. . *EBioMedicine* **2017**, *21*, 21–28., .

8. De Kreutzenberg, S.V.; Ceolotto, G.; Cattelan, A.; Pagnin, E.; Mazzucato, M.; Garagnani, P.; Borelli, V.; Bacalini, M.G.; Franceschi, C.; Fadini, G.P.; et al. Metformin improves putative longevity effectors in peripheral mononuclear cells from subjects with prediabetes. A randomized controlled trial. . *Nutr. Metab. Cardiovasc. Dis.* **2015**, *25*, 686–693, .

9. Kraig, E.; Linehan, L.A.; Liang, H.; Romo, T.Q.; Liu, Q.; Wu, Y.; Benavides, A.D.; Curiel, T.J.; Javors, M.A.; Musi, N.; et al. A randomized control trial to establish the feasibility and safety of rapamycin treatment in an older human cohort: Immunological, physical performance, and cognitive effects. . *Exp. Gerontol.* **2018**, *105*, 53–69., .

10. Velarde, M.; De Maria, M.; Targeting Senescent Cells: Possible Implications for Delaying Skin Aging: A Mini-Review. . *Gerontology* **2016**, *62*, 513–518, .

11. Regulski, M.J.; Cellular Senescence: What, Why and How. . *Wounds* **2017**, *29*, 168–174, .

12. Salminen, A.; Kauppinen, A.; Kaarniranta, K.; Emerging role of NF-κB signaling in the induction of senescence-associated secretory phenotype (SASP). . *Cell. Signal.* **2012**, *24*, 835–845, .

13. Abraham, A.; Roga, G.; Topical steroid-damaged skin.. *Indian J. Dermatol.* **2014**, *59*, 456–459, .

14. Kumar, S.; Pandey, A.K.; Chemistry and Biological Activities of Flavonoids: An Overview. . *Sci. World J.* **2013**, *2013*, 162750., .

15. Panche, A.N.; Diwan, A.D.; Chandra, S.R.; Flavonoids: An overview.. *J. Nutr. Sci.* **2016**, *5*, e47, .

16. Kuryłowicz, A.; The Role of Isoflavones in Type 2 Diabetes Prevention and Treatment—A Narrative Review.. *Int. J. Mol. Sci.* **2020**, *22*, 218, .

17. Mansuri, M.L.; Parihar, P.; Solanki, I.; Parihar, M.S.; Flavonoids in modulation of cell survival signalling pathways.. *Genes Nutr* **2014**, *9*, 1–9, .

18. Lim, H.; Park, H.; Kim, H.P.; Effects of flavonoids on senescence-associated secretory phenotype formation from bleomycin-induced senescence in BJ fibroblasts. . *Biochem. Pharmacol.* **2015**, *96*, 337–348, .

19. Leyva-López, N.; Gutierrez-Grijalva, E.P.; Ambriz-Perez, D.L.; Heredia, J.B.; Flavonoids as Cytokine Modulators: A Possible Therapy for Inflammation-Related Diseases.. *Int. J. Mol. Sci.* **2016**, *17*, 921, .

20. Laura, V.; Mattia, F.; Roberta, G.; Federico, I.; Emi, D.; Chiara, T.; Luca, B.; Elena, C.; Potential of Curcumin in Skin Disorders. . *Nutrients* **2019**, *11*, 2169, .

21. Hostetler, G.L.; Ralston, R.A.; Schwartz, S.J.; Flavones: Food Sources, Bioavailability. . *Adv Nutr* **2017**, *8*, 423–435, .

22. Salehi, B.; Venditti, A.; Sharifi-Rad, M.; Kręgiel, D.; Sharifi-Rad, J.; Durazzo, A.; Lucarini, M.; Santini, A.; Souto, E.B.; Novellino, E.; et al. The Therapeutic Potential of Apigenin.. *Int. J. Mol. Sci.* **2019**, *20*, 1305, .

23. Ahmed, S.A.; Parama, D.; Daimari, E.; Girisa, S.; Banik, K.; Harsha, C.; Dutta, U.; Kunnumakkara, A.B.; Rationalizing the therapeutic potential of apigenin against cancer.. *Life Sci.* **2021**, *167*, 118814, .

24. Bridgeman, B.B.; Wang, P.; Ye, B.; Pelling, J.C.; Volpert, O.V.; Tong, X.; Inhibition of mTOR by apigenin in UVB-irradiated keratinocytes: A new implication of skin cancer prevention.. *Cell. Signal.* **2016**, *28*, 460–468, .

25. Tong, X.; Mirzoeva, S.; Veliceasa, D.; Bridgeman, B.B.; Fitchev, P.; Cornwell, M.L.; Crawford, S.E.; Pelling, J.C.; Volpert, O.V.; Chemopreventive apigenin controls UVB-induced cutaneous proliferation and angiogenesis through HuR and thrombospondin-1. . *Oncotarget* **2014**, *5*, 11413–11427, .

26. Choi, S.; Youn, J.; Kim, K.; Joo, D.H.; Shin, S.; Lee, J.; Lee, H.K.; An, I.-S.; Kwon, S.; Youn, H.J.; et al. Apigenin inhibits UVA-induced cytotoxicity in vitro and prevents signs of skin aging in vivo.. *Int. J. Mol. Med.* **2016**, *38*, 627–634, .

27. Britto, S.M.; Shanthakumari, D.; Agilan, B.; Radhiga, T.; Kanimozhi, G.; Prasad, N.R.; Apigenin prevents ultraviolet-B radiation induced cyclobutane pyrimidine dimers formation in human dermal fibroblasts.. *Mutat. Res. Genet. Toxicol. Environ. Mutagen.* **2017**, *821*, 28–35, .

28. Mirzoeva, S.; Tong, X.; Bridgeman, B.B.; Plebanek, M.P.; Volpert, O.V.; Apigenin Inhibits UVB-Induced Skin Carcinogenesis: The Role of Thrombospondin-1 as an Anti-Inflammatory Factor. . *Neoplasia* **2018**, *20*, 930–942, .

29. Perrott, K.M.; Wiley, C.D.; Desprez, P.-Y.; Campisi, J.; Apigenin suppresses the senescence-associated secretory phenotype and paracrine effects on breast cancer cells.. *GeroScience* **2017**, *39*, 161–173, .

30. Das, S.; Das, J.; Paul, A.; Samadder, A.; Khuda-Bukhsh, A.R.; Apigenin, a Bioactive Flavonoid from Lycopodium clavatum, Stimulates Nucleotide Excision Repair Genes to Protect Skin Keratinocytes from Ultraviolet B-Induced Reactive Oxygen Species and DNA Damage.. *J. Acupunct. Meridian Stud.* **2013**, *6*, 252–262, .

31. Zhang, Y.; Wang, J.; Cheng, X.; Yi, B.; Zhang, X.; Li, Q.; Apigenin induces dermal collagen synthesis via smad2/3 signaling pathway. . *Eur. J. Histochem* **2015**, *59*, 2467, .

32. Ve, A.S.G.; Apigenin as an Anti-Aging Skin Treatment.. *J. Clin. Cosmet. Dermatol.* **2018**, *2*, 1–8, .

33. Bing-Rong, Z.; Song-Liang, J.; Xiao, E.C.; Xiang-Fei, L.; Bao-Xiang, C.; Jie, G.; Dan, L.; Protective effect of the Baicalin against DNA damage induced by ultraviolet B irradiation to mouse epidermis. . *Photodermatol. Photoimmunol. Photomed.* **2008**, *24*, 175–182, .

34. Kim, D.H.; Kim, H.K.; Park, S.; Kim, J.Y.; Zou, Y.; Cho, K.H.; Kim, Y.S.; Kim, D.H.; Yu, B.P.; Choi, J.S.; et al. Short-term feeding of baicalin inhibits age-associated NF-κB activation. . *Mech. Ageing Dev.* **2006**, *127*, 719–725, .

35. Zhang, J.-A.; Yin, Z.; Ma, L.-W.; Yin, Z.-Q.; Hu, Y.-Y.; Xu, Y.; Wu, D.; Permatasari, F.; Luo, D.; Zhou, B.-R.; et al. The Protective Effect of Baicalin against UVB Irradiation Induced Photoaging: An In Vitro and In Vivo Study.. *PLoS ONE* **2014**, *9*, e99703, .

36. Wang, S.-C.; Chen, S.-F.; Lee, Y.-M.; Chuang, C.-L.; Bau, D.-T.; Lin, S.-S.; Baicalin scavenges reactive oxygen species and protects human keratinocytes against UVC-induced cytotoxicity.. *In Vivo* **2013**, *27*, 707–714, .

37. Gendrisch, F.; Esser, P.R.; Schempp, C.M.; Wölfle, U.; Luteolin as a modulator of skin aging and inflammation.. *BioFactors* **2020**, *47*, 170–180, .

38. Wölfle, U.; Haarhaus, B.; Schempp, C.M.; The Photoprotective and Antioxidative Properties of Luteolin are Synergistically Augmented by Tocopherol and Ubiquinone.. *Planta Med.* **2013**, *79*, 963–965, .

39. Wölfle, U.; Heinemann, A.; Esser, P.R.; Haarhaus, B.; Martin, S.F.; Schempp, C.M.; Luteolin Prevents Solar Radiation-Induced Matrix Metalloproteinase-1 Activation in Human Fibroblasts: A Role for p38 Mitogen-Activated Protein Kinase and Interleukin-20 Released from Keratinocytes.. *Rejuvenat. Res.* **2012**, *15*, 466–475, .

40. Averbeck, M.; Gebhardt, C.A.; Voigt, S.; Beilharz, S.; Anderegg, U.; Termeer, C.C.; Sleeman, J.P.; Simon, J.C.; Differential Regulation of Hyaluronan Metabolism in the Epidermal and Dermal Compartments of Human Skin by UVB Irradiation.. *J. Investig. Dermatol.* **2007**, *127*, 687–697, .

41. Hwang, Y.P.; Oh, K.N.; Yun, H.J.; Jeong, H.G.; The flavonoids apigenin and luteolin suppress ultraviolet A-induced matrix metalloproteinase-1 expression via MAPKs and AP-1-dependent signaling in HaCaT cells.. *J. Dermatol. Sci.* **2011**, *61*, 23–31, .

42. Hwang, Y.P.; Choi, J.H.; Kim, H.G.; Choi, J.M.; Hwang, S.K.; Chung, Y.C.; Jeong, H.G.; Cultivated ginseng suppresses ultraviolet B–induced collagenase activation via mitogen-activated protein kinases and nuclear factor κB/activator protein-1–dependent signaling in human dermal fibroblasts. . *Nutr. Res.* **2012**, *32*, 428–438, .

43. Yang, D.; Guo, Q.; Liang, Y.; Zhao, Y.; Tian, X.; Ye, Y.; Tian, J.; Wu, T.; Lu, N.; Wogonin induces cellular senescence in breast cancer via suppressing TXNRD2 expression. . *Arch. Toxicol.* **2020**, *94*, 3433–3447, .

44. Chi, Y.; Kim, H.; Suppression of cyclooxygenase-2 expression of skin fibroblasts by wogonin, a plant flavone from Scutellaria radix.. *Prostaglandins Leukot. Essent. Fat. Acids* **2005**, *72*, 59–66, .

45. Wang, Y.-S.; Cho, J.-G.; Hwang, E.-S.; Yang, J.-E.; Gao, W.; Fang, M.-Z.; Zheng, S.-D.; Yi, T.-H.; Enhancement of Protective Effects of Radix Scutellariae on UVB-induced Photo Damage in Human HaCaT Keratinocytes.. *Appl.*

Biochem. Biotechnol. **2018**, *184*, 1073–1093, .

46. Park, B.K.; Heo, M.Y.; Park, H.; Kim, H.P.; Inhibition of TPA-induced cyclooxygenase-2 expression and skin inflammation in mice by wogonin, a plant flavone from Scutellaria radix. . *Eur. J. Pharmacol.* **2001**, *425*, 153–157, .

47. Chi, Y.S.; Lim, H.; Park, H.; Kim, H.P.; Effects of wogonin, a plant flavone from Scutellaria radix, on skin inflammation: In vivo regulation of inflammation-associated gene expression. . *Biochem. Pharmacol.* **2003**, *66*, 1271–1278, .

48. Company, A.; Lloret-Fillol, J.; Costas, M. . Small Molecule Models for Nonporphyrinic Iron and Manganese Oxygenases; Company, A.; Lloret-Fillol, J.; Costas, M. , Eds.; Elsevier BV: Amsterdam, The Nethrelands, , 2013; pp. 487–564..

49. Shin, E.J.; Lee, J.S.; Hong, S.; Lim, T.-G.; Byun, S.; Quercetin Directly Targets JAK2 and PKCδ and Prevents UV-Induced Photoaging in Human Skin.. *Int. J. Mol. Sci.* **2019**, *20*, 5262, .

50. Wermuth, P.; Addya, S.; Jimenez, S.A.; Effect of Protein Kinase C delta (PKC-δ) Inhibition on the Transcriptome of Normal and Systemic Sclerosis Human Dermal Fibroblasts In Vitro. . *PLoS ONE* **2011**, *6*, e27110, .

51. Lewinska, A.; Adamczyk-Grochala, J.; Bloniarz, D.; Olszowka, J.; Kulpa-Greszta, M.; Litwinienko, G.; Tomaszewska, A.; Wnuk, M.; Pazik, R.; AMPK-mediated senolytic and senostatic activity of quercetin surface functionalized Fe3O4 nanoparticles during oxidant-induced senescence in human fibroblasts. . *Redox Biol* **2020**, *28*, 101337, .

52. Vicentini, F.; He, T.; Shao, Y.; Fonseca, M.J.; Verri, W.; Fisher, G.J.; Xu, Y.; Quercetin inhibits UV irradiation-induced inflammatory cytokine production in primary human keratinocytes by suppressing NF-κB pathway. . *J. Dermatol. Sci.* **2011**, *61*, 162–168, .

53. García-Mediavilla, M.V.; Crespo, I.; Collado, P.S.; Esteller, A.; Sánchez-Campos, S.; Tuñón, M.J.; González-Gallego, J.; The anti-inflammatory flavones quercetin and kaempferol cause inhibition of inducible nitric oxide synthase, cyclooxygenase-2 and reactive C-protein, and down-regulation of the nuclear factor kappaB pathway in Chang Liver cells. . *Eur. J. Pharmacol.* **2007**, *557*, 221–229, .

54. Yousefzadeh, M.J.; Zhu, Y.; McGowan, S.J.; Angelini, L.; Fuhrmann-Stroissnigg, H.; Xu, M.; Ling, Y.Y.; Melos, K.I.; Pirtskhalava, T.; Inman, C.L.; et al. Fisetin is a senotherapeutic that extends health and lifespan. . *EBioMedicine* **2018**, *36*, 18–28, .

55. Seo, S.-H.; Jeong, G.-S.; Fisetin inhibits TNF-α-induced inflammatory action and hydrogen peroxide-induced oxidative damage in human keratinocyte HaCaT cells through PI3K/AKT/Nrf-2-mediated heme oxygenase-1 expression. . *Int. Immunopharmacol* **2015**, *29*, 246–253, .

56. Chiang, H.-M.; Chan, S.-Y.; Chu, Y.; Wen, K.-C.; Fisetin Ameliorated Photodamage by Suppressing the Mitogen-Activated Protein Kinase/Matrix Metalloproteinase Pathway and Nuclear Factor-κB Pathways.. *J. Agric. Food Chem.* **2015**, *63*, 4551–4560, .

57. Wu, P.-Y.; Lyu, J.-L.; Liu, Y.-J.; Chien, T.-Y.; Hsu, H.-C.; Wen, K.-C.; Chiang, H.-M.; Fisetin Regulates Nrf2 Expression and the Inflammation-Related Signaling Pathway to Prevent UVB-Induced Skin Damage in Hairless Mice. . *Int. J. Mol. Sci.* **2017**, *18*, 2118, .

58. Alleviation by Fisetin of Frailty, Inflammation, and Related Measures in Older Adults—Full Text View—ClinicalTrials.gov. Available online: https://clinicaltrials.gov/ct2/show/NCT03675724

59. Barnes, S.; The Biochemistry, Chemistry and Physiology of the Isoflavones in Soybeans and their Food Products.. *Lymphat. Res. Biol.* **2010**, *8*, 89–98, .

60. Kim, S.-Y.; Kim, S.-J.; Lee, J.-Y.; Kim, W.-G.; Park, W.-S.; Sim, Y.-C.; Lee, S.-J.; Protective effects of dietary soy isoflavones against UV-induced skin-aging in hairless mouse model.. *J. Am. Coll. Nutr.* **2004**, *23*, 157–162, .

61. Zhao, D.; Shi, Y.; Dang, Y.; Zhai, Y.; Ye, X.; Daidzein stimulates collagen synthesis by activating the TGF-β/smad signal pathway. . *Australas. J. Dermatol.* **2014**, *56*, e7–e14, .

62. Oh, H.-J.; Kang, Y.-G.; Na, T.-Y.; Kim, H.-J.; Park, J.S.; Cho, W.-J.; Lee, M.-O.; Identification of daidzein as a ligand of retinoic acid receptor that suppresses expression of matrix metalloproteinase-9 in HaCaT cells. . *Mol. Cell. Endocrinol.* **2013**, *376*, 107–113, .

63. Widyarini, S.; Spinks, N.; Husband, A.J.; Reeve, V.E.; Isoflavonoid Compounds from Red Clover (Trifolium pratense) Protect from Inflammation and Immune Suppression Induced by UV Radiation.. *Photochem. Photobiol.* **2001**, *74*, 465, .

64. Reeve, V.E.; Widyarini, S.; Domanski, D.; Chew, E.; Barnes, K.; Protection Against Photoaging in the Hairless Mouse by the Isoflavone Equol.. *Photochem. Photobiol.* **2005**, *81*, 1548–1553, .

65. Polito, F.; Marini, H.R.; Bitto, A.; Irrera, N.; Vaccaro, M.; Adamo, E.B.; Micali, A.; Squadrito, F.; Minutoli, L.; Altavilla, D.; et al. Genistein aglycone, a soy-derived isoflavone, improves skin changes induced by ovariectomy in rats. . *Br. J. Pharmacol.* **2011**, *165*, 994–1005, .

66. Na Wang, Y.; Wu, W.; Chen, H.C.; Fang, H.; Genistein protects against UVB-induced senescence-like characteristics in human dermal fibroblast by p66Shc down-regulation.. *J. Dermatol. Sci.* **2010**, *58*, 19–27, .

67. Iovine, B.; Iannella, M.L.; Gasparri, F.; Monfrecola, G.; Bevilacqua, M.A.; Synergic Effect of Genistein and Daidzein on UVB-Induced DNA Damage: An Effective Photoprotective Combination.. *J. Biomed. Biotechnol.* **2011**, *2011*, 1–8, .

68. Miyazaki, K.; Hanamizu, T.; Iizuka, R.; Chiba, K.; Genistein and Daidzein Stimulate Hyaluronic Acid Production in Transformed Human Keratinocyte Culture and Hairless Mouse Skin. . *Skin. Pharmacol. Physiol.* **2002**, *15*, 175–183, .

69. Izumi, T.; Saito, M.; Obata, A.; Arii, M.; Yamaguchi, H.; Matsuyama, A.; Oral intake of soy isoflavone aglycone improves the aged skin of adult women.. *J. Nutr. Sci. Vitaminol.* **2007**, *53*, 57–62, .

70. Moraes, A.B.; Haidar, M.A.; Soares, J.M.; Simões, M.J.; Baracat, E.C.; Patriarca, M.T.; The effects of topical isoflavones on postmenopausal skin: Double-blind and randomized clinical trial of efficacy. . *Eur. J. Obstet. Gynecol. Reprod. Biol.* **2009**, *146*, 188–192, .

71. El-Mahdy, M.A.; Zhu, Q.; Wang, Q.-E.; Wani, G.; Patnaik, S.; Zhao, Q.; Arafa, E.-S.; Barakat, B.; Mir, S.N.; Wani, A.A.; et al. Naringenin Protects HaCaT Human Keratinocytes Against UVB-induced Apoptosis and Enhances the Removal of Cyclobutane Pyrimidine Dimers from the Genome.. *Photochem. Photobiol.* **2007**, *84*, 307–316, .

72. Martinez, R.M.; Pinho-Ribeiro, F.A.; Steffen, V.S.; Caviglione, C.V.; Vignoli, J.A.; Barbosa, D.S.; Baracat, M.M.; Georgetti, S.R.; Verri, W.A.; Casagrande, R.; et al. Naringenin Inhibits UVB Irradiation-Induced Inflammation and Oxidative Stress in the Skin of Hairless Mice. . *J. Nat. Prod.* **2015**, *78*, 1647–1655, .

73. Martinez, R.M.; Pinho-Ribeiro, F.A.; Steffen, V.S.; Silva, T.C.C.; Caviglione, C.V.; Bottura, C.; Fonseca, M.J.V.; Vicentini, F.T.M.C.; Vignoli, J.A.; Baracat, M.M.; et al. Topical Formulation Containing Naringenin: Efficacy against Ultraviolet B Irradiation-Induced Skin Inflammation and Oxidative Stress in Mice.. *PLoS ONE* **2016**, *11*, e0146296, .

74. Lim, K.H.; Kim, G.R.; Inhibitory effect of naringenin on LPS-induced skin senescence by SIRT1 regulation in HDFs. . *Biomed. Dermatol.* **2018**, *2*, 26, .