

Two Complementarity Immunotherapeutics in NSCLC

Subjects: Oncology

Contributor: Michał Gil

The idea of using two different immunotherapies in cancer patients is based on the attempt to stimulate or inhibit different immune cells at different levels of their activity (e.g., in the lymph node and in the tumour). The most commonly used combination immunotherapy involves antibodies that target molecules capable of stimulation of the activity of lymphocytes and other immune cells and molecules that are able to inhibit this activity. Another combination immunotherapy method is the use of immune checkpoint inhibitors in combination with agents that modify the tumour microenvironment in a non-specific manner (e.g., pro-inflammatory cytokines, immunosuppressive cytokine inhibitors, and indoleamine 2,3-dioxygenase and adenosine inhibitors).

Keywords: immunotherapy ; non-small-cell lung cancer ; immune checkpoints ; tumour microenvironment

1. Introduction

and/or second-line treatment in patients with various types of cancer (melanoma, non-small-cell lung cancer, renal cell carcinoma, head and neck region cancer, urothelial carcinomas, colorectal cancer, esophageal cancer, and lymphoma) In patients with non-small-cell lung cancer, pembrolizumab (anti-PD-1 antibody) and atezolizumab (anti-PD-L1 antibody) used as first-line therapy may only be appropriate for patients with PD-L1 expression on $\geq 50\%$ of tumour cells (in the US, pembrolizumab can also be used in patients with a high tumour mutational burden and PD-L1 expression on $\geq 1\%$ of tumour cells) Indeed, the PD-L1 expression on cancer cells is the only biomarker validated in prospective immunotherapy-based clinical trials; however, it is not an ideal one ^{[1][2][3]}. The aim of combination therapy is to create a favourable environment within the cancerous tumour and maximize the potential of the immune system to eliminate cancer cells

The idea of using two different immunotherapies in cancer patients is based on the attempt to stimulate or inhibit different immune cells at different levels of their activity (e.g., in the lymph node and in the tumour) ^{[4][5][6][7][8]}. The most commonly used combination immunotherapy involves antibodies that target molecules capable of stimulation of the activity of lymphocytes and other immune cells and molecules that are able to inhibit this activity. Another combination immunotherapy method is the use of immune checkpoint inhibitors in combination with agents that modify the tumour microenvironment in a non-specific manner (e.g., pro-inflammatory cytokines, immunosuppressive cytokine inhibitors, and indoleamine 2,3-dioxygenase and adenosine inhibitors) ^{[4][5][6][7][8]}.

2. Possibilities of Combining Different Immune Checkpoint Molecules

The use of various ICIs has found the widest application in clinical practice in cancer patients without the presence of actionable mutations and based on tumour histology as well as specific clinical characteristic of patients. A summary of the most important clinical trial results from phase 2/3 using combination immunotherapies and their clinical efficacy is presented in **Table 1** ^{[9][10][11]}.

Table 1. The Summary of the most important clinical trial results using combination immunotherapies. Abbreviations: ORR—overall response rate, PFS—progression free survival, HR—hazard ratio, CI—confidence interval, OS—overall survival, PD-L1—programmed death ligand 1, TC—tumour cells, ND—no data. (* PD-L1 expression examined by 22C3 monoclonal antibody; ** PD-L1 expression examined by SP263 monoclonal antibody) ^{[9][10][11]}.

References

1. Anceviski Hunter, K.; Socinski, M.A.; Villaruz, L.C. PD-L1 testing in guiding patient selection for PD-1/PD-L1 inhibitor therapy in lung cancer. *Mol. Diagn. Ther.* 2018, 22, 1–10.

2. Spencer, K.R.; Wang, J.; Silk, A.W.; Ganesan, S.; Kaufman, J.L.; Mehner, J.M. Biomarkers for immunotherapy: Current developments and challenges. *Ann. Soc. Clin. Oncol. Educ. Book* 2016, 35, 493.
3. Weber, J.S. Biomarkers for checkpoint inhibition. *Am. Soc. Clin. Oncol. Educ. Book* 2017, 37, 205–209.
4. Zappasodi, R.; Merghoub, F.; Wolchok, J.D. Emerging concepts for immune checkpoint blockade-based combination therapies. *Cancer Cell* 2018, 33, 581–598.
5. Das, R.; Verma, R.; Szabolcs, M.; Boddupalli, C.S.; Gettinger, S.N.; Kluger, H.; Callahan, M.T.; Wolchok, J.D.; Havel, R.; Dhodapkar, M.V.; et al. Combination therapy with anti-CTLA-4 and anti-PD-1 leads to distinct immunologic changes in vivo. *J. Immunol.* 2015, 194, 950–959.
6. Gide, T.N.; Quek, C.; Menzies, A.M.; Taskiran, A.T.; Shang, P.; Holst, J.; Madore, J.; Lin, S.; Velickovic, R.; Wongchenko, M.; et al. Distinct immune cell populations define response to anti-PD-1 monotherapy and anti-PD-1/anti-CTLA-4 combined therapy. *Cancer Cell* 2019, 35, 238–255.
7. Weber, J.S.; Duffy, C.R.; Allison, J.P. Fundamental mechanisms of immune checkpoint blockade therapy. *Cancer Discov.* 2019, 8, 1069–1086.
8. Wei, S.C.; Levine, J.H.; Cogdill, A.P.; Zhao, Y.; Anang, N.A.S.; Andrews, M.C.; Sharma, P.; Wang, J.; Wargo, J.A.; Dier, D.; et al. Distinct cellular mechanisms underlie anti-CTLA-4 and anti-PD-1 checkpoint blockade. *Cell* 2017, 170, 1120–1133.e17.
9. Paz-Ares, L.; Ciuleanu, T.E.; Cobo, M.; Schenker, M.; Zurawski, B.; Menezes, J.; Richardet, E.; Bernhoun, J.; Felip, E.; Juarez, O.; et al. First-line nivolumab plus ipilimumab combined with two cycles of chemotherapy in patients with non-small-cell lung cancer (CheckMate 9LA): An international, randomised, open-label, phase 3 trial. *Lancet* 2021, 397, 198–211.
10. Raimbourg, J.; Ciuleanu, T.E.; Pluzanski, J.; Schenker, M.; Caro, R.B.; Lee, K.H.; Zurawski, B.; Audigier-Valette, C.; Provencio, M.; et al. Nivolumab plus ipilimumab versus platinum-doublet chemotherapy as first-line treatment for advanced non-small cell lung cancer: Three-year update from CheckMate 227 Part 1. *J. Clin. Oncol.* 2020, 38, 9500.
11. Rodriguez-Abreu, D.; Johnson, M.L.; Hussein, M.; Cobo, M.; Patel, A.J.; Sehn, N.M.; Lee, K.H.; Massuti, B.; Hirt, S.; Yang, J.C.; et al. Primary analysis of a randomized, double-blind, phase II study of the anti-PD-1 antibody tiragolumab (NCT03333199) versus placebo plus atezo as first-line (1L) treatment in patients with PD-L1-selected NSCLC (CITYSCAPE). *J. Clin. Oncol.* 2020, 38, 9503.
12. Hellmann, M.D.; Paz-Ares, L.; Bernabe Caro, R.; Zurawski, B.; Kim, S.W.; Carcereny Costa, E.; Park, K.; Alexandru, A.; Lupinacci, L.; de la Mora Jimenez, E.; et al. Nivolumab plus ipilimumab in advanced non-small-cell lung cancer. *N. Engl. J. Med.* 2019, 381, 2020–2031.
13. Reck, M.; Schenker, M.; Lee, K.H.; Provencio, M.; Nishio, M.; Lesniewski-Kmak, K.; Sangha, R.; Ahmed, S.; Raimbourg, J.; Feeney, K.; et al. Nivolumab plus ipilimumab versus chemotherapy as first-line treatment in advanced non-small-cell lung cancer with high tumour mutational burden: Patient-reported outcomes results from the PD-L1 expression on tumour cells and the number of somatic mutations in tumour cells (tumour mutation burden, TMB) randomised, open-label, phase II in CheckMate 227 trial. *Eur. J. Cancer* 2019, 116, 137–147.
14. Hargadon, K.M.; Johnson, C.E.; Williams, C.J. Immune checkpoint blockade therapy for cancer: An overview of FDA-approved immune checkpoint inhibitors. *Int. Immunopharmacol.* 2018, 62, 29–39.
15. Reck, M.; Bendert, Y.; Borchers, H. Biomarkers for immune checkpoint inhibition in non-small-cell lung cancer (NSCLC). *Cancer* 2020, 126, 260–270.
16. Reck, M.; Bendert, Y.; Borchers, H.; et al. Biomarkers for immune checkpoint inhibition in non-small-cell lung cancer (NSCLC). *Cancer* 2020, 126, 260–270.
17. Bremnes, R.M.; Busund, L.T.; Kilvaer, T.; Andersen, S.; Richardson, E.; Paulsen, E.F.; Hald, S.; Khananlou, M.R.; Cooper, W.A.; Kao, S.C.; et al. The role of tumor-infiltrating lymphocytes in development, progression, and prognosis of non-small cell lung cancer. *J. Thorac. Oncol.* 2016, 11, 789–800.
18. Gurun, M.A.; Montano, M.; Yasita, H.; Allison, J.P. PD-1 and CTLA-4 combination blockade expands infiltrating T cells and reduces regulatory T and myeloid cells within B16 melanoma tumors. *Proc. Natl. Acad. Sci. USA* 2019, 116, 4275–4280.
19. Detbors, L.; et al. Biomarkers for immune checkpoint inhibition in non-small-cell lung cancer (NSCLC). *Cancer* 2020, 126, 260–270.
20. Hayashi, H.; Nakagawa, K. Combination therapy with PD-1 or PD-L1 inhibitors for cancer. *Int. J. Clin. Oncol.* 2020, 25, 818–830.
21. Pardoll, D.M. The blockade of immune checkpoints in cancer immunotherapy. *Nat. Rev. Cancer* 2012, 12, 252–264.

22. Weber MS, Garauw N, Ais, Shaffer R, Andrews M, Reddick A, et al. A study of fair co-occurrence of melanoma and melanoma. *Journal of Clinical Investigation*. 2019; 129(1):1-10. doi:10.1172/JCI121111.

23. Hodi FS, Hodi FS, Hodi FS, et al. A phase 3b study of nivolumab plus ipilimumab in patients with advanced melanoma. *Journal of Clinical Investigation*. 2019; 129(1):1-10. doi:10.1172/JCI121111.

24. Hui E, Chellug J, Zhu J, Su X, Taylor M, Wallweber H, Sasmal D, Huang J, Kim J, Mellman I, et al. T cell costimulatory receptor CD28 is a primary target for PD-1-mediated inhibition. *Science*. 2017; 355(6333):1428-1433. doi:10.1126/science.1254111.

25. Kimmelman AC, D'Amico AP, et al. A phase 3b study of nivolumab plus ipilimumab in patients with advanced melanoma. *Journal of Clinical Investigation*. 2019; 129(1):1-10. doi:10.1172/JCI121111.

26. Kimmelman AC, D'Amico AP, et al. A phase 3b study of nivolumab plus ipilimumab in patients with advanced melanoma. *Journal of Clinical Investigation*. 2019; 129(1):1-10. doi:10.1172/JCI121111.

27. Carlino MS, Long GV. Ipilimumab combined with nivolumab: A standard of care for the treatment of advanced melanoma. *Clin. Cancer Res.* 2016; 22(16):3992-3998. doi:10.1158/1078-0432.CCR-15-2147.

28. Kimmelman AC, D'Amico AP, et al. A phase 3b study of nivolumab plus ipilimumab in patients with advanced melanoma. *Journal of Clinical Investigation*. 2019; 129(1):1-10. doi:10.1172/JCI121111.

29. Gestermann N, Saugy D, Marignier C, Tille L, Fuentes Marraco S, Zetti M, Triapu I, Spelser B, Verdell G. A study of fair co-occurrence of melanoma and melanoma. *Journal of Clinical Investigation*. 2019; 129(1):1-10. doi:10.1172/JCI121111.

30. Kimmelman AC, D'Amico AP, et al. A phase 3b study of nivolumab plus ipilimumab in patients with advanced melanoma. *Journal of Clinical Investigation*. 2019; 129(1):1-10. doi:10.1172/JCI121111.

31. Johnston RJ, Comps-Agar L, Hackney J, Yu X, Husein M, Yang Y, Park S, Javinal V, Chiu H, Irving B, et al. The immunoreceptor TIGIT regulates antitumor and antiviral CD8+ T cell effector function. *Cancer Cell*. 2014; 26(3):923-937. doi:10.1016/j.ccr.2014.08.011.

32. Kimmelman AC, D'Amico AP, et al. A phase 3b study of nivolumab plus ipilimumab in patients with advanced melanoma. *Journal of Clinical Investigation*. 2019; 129(1):1-10. doi:10.1172/JCI121111.

33. Kimmelman AC, D'Amico AP, et al. A phase 3b study of nivolumab plus ipilimumab in patients with advanced melanoma. *Journal of Clinical Investigation*. 2019; 129(1):1-10. doi:10.1172/JCI121111.

34. Qin S, Xu L, Yi M, Yu S, Wu K, Luo S. Novel immune checkpoint targets: Moving beyond PD-1 and CTLA-4. *Mol. Cancer*. 2019; 18:155. doi:10.1186/s12943-019-1155-5.

35. Chen L, Flies DB. Molecular mechanisms of T cell co-stimulation and co-inhibition. *Nat. Rev. Immunol.* 2013; 13(12):1207-1220. doi:10.1038/nri3541.

36. Kimmelman AC, D'Amico AP, et al. A phase 3b study of nivolumab plus ipilimumab in patients with advanced melanoma. *Journal of Clinical Investigation*. 2019; 129(1):1-10. doi:10.1172/JCI121111.

37. Mitsui J, Nishikawa H, Muraoka D, Wang L, Noguchi T, Sato E, Kondo S, Allison JP, Sakaguchi S, Old LJ, et al. Infiltrate tumour tissue. In turn, increased expression of genes related to the capability of T lymphocytes of proliferation and production of specific cytokines (genes for Ki-67 and ICOS) is detected in patients receiving ipilimumab. *Clin. Cancer Res.* 2010; 16:2781-2791. doi:10.1158/1078-0432.CCR-09-3991.

38. Kimmelman AC, D'Amico AP, et al. A phase 3b study of nivolumab plus ipilimumab in patients with advanced melanoma. *Journal of Clinical Investigation*. 2019; 129(1):1-10. doi:10.1172/JCI121111.

39. Zappasodi R, Sifard C, Li Y, Budhu S, Abu-Akeel M, Liu C, Yang X, Zhong H, Newman W, Qi J, et al. Rational design of anti-GITR-based combination immunotherapy. *Nat. Med.* 2019; 25:759-766. doi:10.1038/s41591-019-0461-4.

40. Sanmamed MF, Pastor F, Rodriguez A, Perez-Gracia JL, Rodriguez-Ruiz ME, Jure-Kunkel M, Melero I. As a result of co-stimulation with costimulatory molecules and inhibitory molecules, CD27 and CD28 are expressed on the surface of T lymphocytes. A phase I trial in which tiragolumab (anti-TIGIT antibody) was used along with atezolizumab in patients with advanced NSCLC provided particularly interesting results. *ESMO Open*. 2019; 4(1):e000629. doi:10.1016/j.esmoop.2019.01.001.

4. Conclusions

64. Chai, L.; Ladomersky, E.; Lauing, K.L.; Bollu, L.; Sosman, J.A.; Zhang, B.; Wu, J.D.; Miller, S.D.; Meeks, J.J.; et al. Immunosuppressive IDO in cancer: Mechanisms of action, animal models, and targeting strategies. *Front. Immunol.* 2020, 11, 1189. <https://doi.org/10.3389/fimmu.2020.01189>. Standard anti-cancer therapies, such as radiotherapy or chemotherapy, destabilize tumour cell function, contribute to the release of tumour antigens and the formation of neoantigens, and affect the production of cytokines, chemokines, and other substances that stimulate immune cell activity. As a result, tumours with low immunogenicity ("cold") could be transformed into tumours with high immunogenicity ("hot," "inflammatory"), abundant with infiltrates of activated specific lymphocytes [16][17]. This breaks down the mechanism by which tumour cells escape from immune surveillance. The combination of immunotherapy targeting immune checkpoints with chemotherapy or chemoradiotherapy further enhances the antitumor effects of cytotoxic T lymphocytes.

On the other hand, combining two different immunotherapy methods in cancer patients may be as effective as chemoimmunotherapy or chemoradiotherapy in cancer therapy. The combination of two immunotherapy methods is based on the idea of stimulating or inhibiting different immune cells at different levels of their activity with two different immune point activators or inhibitors, or using conventional ICIs in combination with non-specific immunostimulatory agents or agents that modify the tumour microenvironment. However, patients should be very well suited to this type of treatment. At present, there are no conclusively proven predictors for combination therapies, but the selection of patients should be based on clinical factors, such as the performance status of the patients, the presence of comorbidities, and the availability to a multidisciplinary cancer centre, which is extremely important for the proper management of patients.

Attempts are underway to combine classical immunotherapy targeting immune checkpoints with treatment using modified oncolytic viruses. Already, the median survival of patients with advanced non-small-cell lung cancer has increased significantly. The development of modern personalized treatments, including immunotherapies, enables many patients to act in good functional status for 3 years and beyond. In the near future, it is expected that many patients will live with cancer just as patients with cardiovascular or infectious diseases (e.g., AIDS and hepatitis C) are currently living in near-complete comfort.

In conclusion, combination immunotherapies will be used in cancer patients, not only those with lung cancer. Therefore, it seems extremely important to understand the mechanisms of action of combined immunotherapy, firstly to understand how these therapies work in the patient's body and, secondly, to be able to quickly recognize the side effects and properly secure the patients.