

Cancer Immunotherapy

Subjects: Oncology

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Cancer treatment has evolved tremendously in the last few decades. Immunotherapy has been considered to be the forth pillar in cancer treatment in addition to conventional surgery, radiotherapy, and chemotherapy. Though immunotherapy has resulted in impressive response, it is generally limited to a small subset of patients. Understanding the mechanisms of resistance toward cancer immunotherapy may shed new light to counter that resistance. In this entry, we highlighted and summarized two major hurdles (recognition and attack) of cancer elimination by the immune system. The mechanisms of failure of some available immunotherapy strategies were also described. Moreover, the significance role of immune compartment for various established cancer treatments were also elucidated. Then, the mechanisms of combinatorial treatment of various conventional cancer treatment with immunotherapy were discussed. Finally, a strategy to improve immune cancer killing by characterizing cancer immune landscape, then devising treatment based on that cancer immune landscape was put forward.

Keywords: immune escape ; cancer ; immunotherapy ; T cell ; immunosuppression ; antigen presentation ; immune checkpoint ; resistance ; treatment

1. Introduction

The immune system not only functions to keep foreign pathogens away from the host, it also has a role in suppressing cancer^[1]. The precursor of cancerous cells is initially a normal cell that becomes malignant due to multiple mutations within its genome. The mutated genes, when expressed, will result in the production of non-self-antigens. Theoretically, those mutated tumor neo antigens will be recognized and eliminated by our immune system because there has been no tolerance toward those tumor neo antigens. However, cancer cells could develop mechanisms to trick that immune system.

The mechanism of immune escape is one of the major mechanisms of mutated cells to gain potential to grow and eventually metastasize^[2]. Understanding the mechanism of immune escape by cancer cells will provide us with the required insight to potentially develop treatment to reverse that mechanism. In general, immune escape is due to tolerance or failure of two main functions of the immune system: failure of immune recognition and failure of immune attack.

2. Major Histocompatibility Complex (MHC) Are Core Molecules in Immune Recognition

Generally, there are two main mechanisms for cancer cell recognition by immune cells. The first mechanism is self-presentation of tumor antigen through MHC or human leukocyte antigen (HLA) Class I. The second mechanism is tumor antigen presentation through professional antigen presenting cells (APCs) via MHC or HLA Class II^[3]. Both methods of tumor antigen presentation are essential in immune surveillance and recognition^{[3][4]}. Cancer cells generally harbor multiple mutations within their genome. Some parts of the mutated genes may express foreign antigens^[5]. These mutated genes are not previously known by the host immune system, therefore, they should be able to be recognized by immune cells^{[5][6]}.

The process of antigen presentation of those cancer cells to the immune system requires a properly functioning MHC Class I molecule. MHC Class I molecule is expressed in all nucleated cells as part of the physiologic cellular defense apparatus from foreign antigens. The process of tumor antigen presentation starts with cleavage of tumor antigen in the cytoplasm of the tumor until the tumor antigen becomes a small sequence of amino acids^{[3][6]}. These cleaved segments will combine with MHC Class I molecule in endoplasmic reticulum. These complexes will later be transported to the plasma membrane^[6]. The T cells will be able to bind and recognize the tumor antigen-MHC Class I complex on the plasma membrane through a specific T cell receptor.

Many cancers circumvent the process of immune recognition or tumor antigen presentation from the cancer cells to T cells by downregulating MHC Class I within tumor cells [7][8][9][10] (Figure 1a). Various methods were thought to be employed by cancer cells to suppress the expression of MHC Class I. Epigenetic silencing within tumor cells resulted in direct methylation of MHC Class I gene by DNA methyltransferase. The methylation process has been observed in various types of cancers [7][10][11]. The methylation of various essential genes resulted in suppression and downregulation of MHC Class I expression, thus preventing those tumor cells to be recognized by the immune system.

Furthermore, tumor cells can also downregulate MHC Class I molecule by suppressing various MHC Class I transcriptor activators. Loss of Interferon regulatory factor 2 (IRF2) and NOD-like receptor (NLR) family and caspase recruitment (CARD) domain containing 5 (NLRC5) expression have been shown to be correlated with lower expression of MHC Class I and higher expression of immune exhaustion marker [8][9]. These transcriptor activators in cancer cells were suppressed by various means including methylation, copy number loss, or somatic mutation [8][9]. These are some of the known and most common mechanisms of cancers to fail the process of immune recognition via MHC Class I.

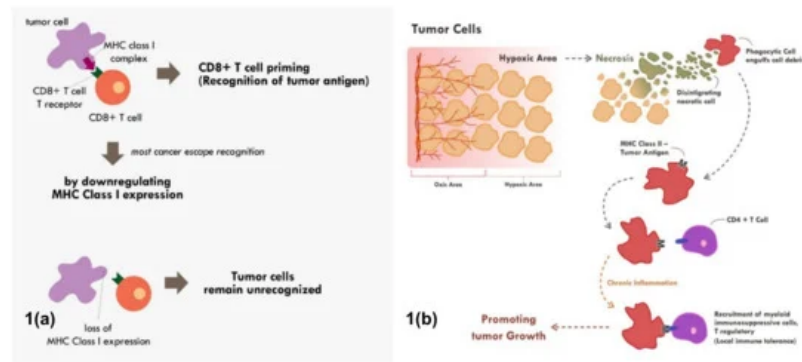


Figure 1. Mechanisms of tumor escape immune recognition. (a) Through downregulation of MHC Class I in tumor cells, rendering tumor cells unrecognized. (b) Through engulfment of tumor debris and further presentation through MHC Class II and APC cells (chronic process of that event leads to chronic inflammation resulting in sensitization and differentiation of CD4+ T cells toward immune suppressive CD4+ T regulatory cells phenotype).

Tumor cell recognition through professional antigen presenting cells and the MHC Class II molecule is also an important process in cancer immune surveillance and recognition [12][13]. Though MHC Class II is only expressed in professional antigen presenting cells, its role has been shown to be as critical as MHC Class I in cancer immunotherapy [4]. Tumor cells will continuously proliferate until a point in which the supply of nutrients and oxygen through neo-vasculature becomes inadequate. At that point, the tumor becomes hypoxic and later dies. This process of cell death is commonly immunogenic in nature. This immunogenic tumor death will result in the release of various damage-associated molecular patterns (DAMPs) such as adenosine triphosphate (ATP), high-mobility group protein B1 (HMGB1), and heat-shock proteins that will recruit various antigen presenting cells [14][15].

3. Functional T Cells for Effective Immune Attack

Even though the administration of immune checkpoint inhibitors described in the previous section have successfully reversed the immunosuppressive mechanism by cancer cells in some cases, however, a substantial number of cases still did not show tumor regression or, at most, only a temporary tumor regression. This persistent immune tolerance is due to the proliferation of dysfunctional to exhausted T cells within the tumor microenvironment [16][17][18]. An in-depth study analyzing the transcriptome of every single immune infiltrating lymphocytes in melanoma samples revealed that majority of CD8+ T cells were indeed lacking a complete cytotoxic gene expression, thus making them dysfunctional [16]. Furthermore, those dysfunctional CD8+ T cells were clonal and very proliferative within the tumor microenvironment [16] (Figure 2).

Local Tumor Microenvironment

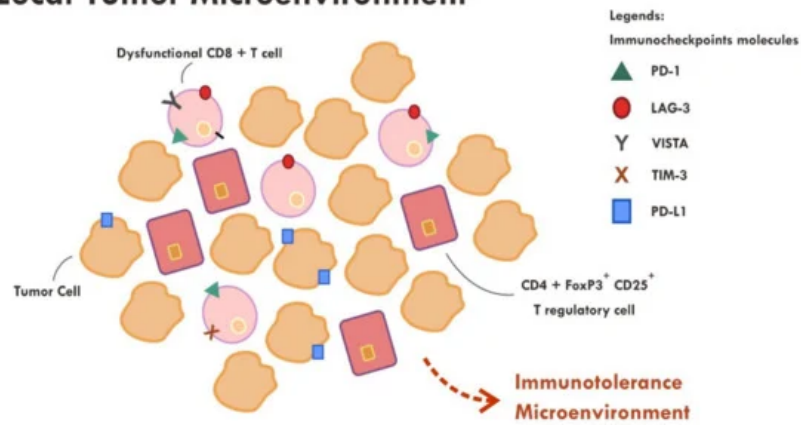


Figure 2. Abundance of dysfunctional CD8+ T cells expressing various immune inhibitory molecules (PD-1, LAG-3, VISTA, TIM-3, PD-L1) within the tumor microenvironment resulted in failure of tumor attack.

Apart from the presence of highly proliferative dysfunctional CD8+ T cells, tumor rejection is also hampered by the presence of a highly immunosuppressive microenvironment elicited by T regulatory cells. T regulatory cells that exhibit an immunosuppressive phenotype generally express CD45RA-, FOXP3+high, CD4+ and CD25+high markers [19]. A study revealed that those T regulatory cells were found to be abundant in the tumor sample compared to systemic circulation (10–50% vs. 2–5%) [19][20]. Various chemokines and cytokines secreted by tumor cells are thought to be the main perpetrator of T regulatory cells recruitment [19]. The T regulatory cells also express immune checkpoint molecules such as CTLA-4 and PD-1 [19][21]. The administration of anti CTLA-4 monoclonal antibody resulted in ADCC, thus reducing the number of T regulatory cells [19][22], while the anti PD-1 effect on T regulatory was not very clear. Some evidence suggests that anti PD-1, contrary to that expected, promotes an enhanced immunosuppressive T regulatory activity [19][21].

4. Hurdles of Immunotherapy

There are various forms of immunotherapy available as cancer treatment. The oldest form of immunotherapy is in the form of pro-inflammatory cytokine administration such as interferon. Later, immune checkpoint inhibitors were developed based on monoclonal antibody. These immune checkpoint inhibitors are the most commonly utilized immunotherapy today. The other forms of immunotherapy are adoptive cell transfer and oncolytic virus vaccine [23][24]. Adoptive cell transfer is based on isolation or engineering of immune cells such as cytotoxic T cells or NK cells that are able to recognize tumor neo antigens [23][24]. Then, these adoptive cells are expanded ex vivo and then infused into the patients [23]. It is expected that those infused killer cells would recognize and attack the cancer cells. Oncolytic virus vaccine is based on engineered virus with its virulence gene deleted [25]. The administration of oncolytic virus into the tumor will trigger tumor infection, which later leads to cell lysis with positive inflammatory response [25].

The effectiveness of all modes of immunotherapy is very dependent on the intrinsic and dynamic tumor microenvironment of each patient. Any immunotherapy will only be effective if strong response of tumor recognition and tumor attack are elicited. In cytokine based immunotherapy, the interferon increases dendritic cell maturation, favors T helper 1 differentiation, increases cytotoxic function of Natural Killer (NK) cells, and increases tumor MHC class 1 expression [26][27]. However, interferon has also been shown to induce upregulation of immune checkpoints such as PD-L1 [28][29]. Therefore, immune tumor attack becomes suboptimal. Combining interferon and immune checkpoint inhibitor has a sound rationale. Phase I and II trials have shown the safety and initial efficacy of these combinations [30][31]. Nevertheless, complete response was only observed in a small subset of patients [30][31]. This observation underscores that some other mechanisms were in place that rendered either tumor recognition or tumor attack suboptimal.

Various immune checkpoint inhibitors have been approved and used in clinic. A phase III trial comparing chemotherapy and a single agent pembrolizumab, an anti PD-1, has shown a remarkable survival benefit of pembrolizumab over chemotherapy in metastatic non-small cell lung cancer [32]. Furthermore, an analysis of a KEYNOTE-001 study showed that 16% of patients receiving pembrolizumab for metastatic melanoma had a durable complete response [33][32]. These significant findings were clinically very meaningful, but unfortunately was only observed in small subsets of patients. Immune checkpoint inhibitors generally act to re-invigorate the tumor immune attack, as discussed in the previous section. In a condition where there is a lack of tumor immune recognition per se, then immune checkpoint inhibitors are unlikely to be clinically beneficial.

The hurdles of immunotherapy have to be viewed in a more comprehensive perspective. The hurdles are due to the establishment of inherent subversion mechanisms of immune recognition and immune attack of tumor cells. Immunotherapy alone in most cases would not be able to counteract all of these mechanisms. One way to counteract these mechanisms is by combination therapy. Various commonly employed anti-cancer treatments are able to tackle some of these mechanisms. When combined with immunotherapy, multiple mechanisms could be reversed simultaneously and result in better tumor control. Next, in the following section, we will discuss various commonly employed anti-cancer treatment and its relationship with tumor immunity.

References

1. Alberto Mantovani; Andrea Ponzetta; Antonio Inforzato; Sébastien Jaillon; Innate immunity, inflammation and tumour progression: double-edged swords. *Journal of Internal Medicine* **2019**, 285, 524-532, [10.1111/joim.12886](https://doi.org/10.1111/joim.12886).
2. Douglas Hanahan; Robert A. Weinberg; Hallmarks of Cancer: The Next Generation. *Cell* **2011**, 144, 646-674, [10.1016/j.cell.2011.02.013](https://doi.org/10.1016/j.cell.2011.02.013).
3. Kenneth L. Rock; Eric Reits; Jacques Neefjes; Present Yourself! By MHC Class I and MHC Class II Molecules. *Trends in Immunology* **2016**, 37, 724-737, [10.1016/j.it.2016.08.010](https://doi.org/10.1016/j.it.2016.08.010).
4. Roberto S. Accolla; Elise Ramia; Alessandra Tedeschi; Greta Forlani; CIITA-Driven MHC Class II Expressing Tumor Cells as Antigen Presenting Cell Performers: Toward the Construction of an Optimal Anti-tumor Vaccine.. *Frontiers in Immunology* **2019**, 10, 1806, [10.3389/fimmu.2019.01806](https://doi.org/10.3389/fimmu.2019.01806).
5. Tao Jiang; Tao Shi; Henghui Zhang; Jie Hu; Yuanlin Song; Jia Wei; Shengxiang Ren; Caicun Zhou; Tumor neoantigens: from basic research to clinical applications. *Journal of Hematology & Oncology* **2019**, 12, 1-13, [10.1186/s13045-019-0787-5](https://doi.org/10.1186/s13045-019-0787-5).
6. Mark Yarchoan; Burles A. Johnson; Eric R. Lutz; Daniel A. Laheru; Mark Yarchoan Burles A. Johnson Iii Eric R. Lutz Daniel A. Laheru Elizabeth M. Jaffee; Targeting neoantigens to augment antitumour immunity. *Nature Reviews Cancer* **2017**, 17, 209-222, [10.1038/nrc.2016.154](https://doi.org/10.1038/nrc.2016.154).
7. Luo, N.; Nixon, M.J.; Gonzalez-Ericsson, P.I.; Sanchez, V.; Opalenik, S.R.; Li, H.; Zahnow, C.A.; Nickels, M.L.; Liu, F.; Tantawy, M.N.; et al. DNA Methyltransferase Inhibition Upregulates MHC-I to Potentiate Cytotoxic T Lymphocyte Responses in Breast Cancer. *Nat. Commun.* 2018, 9, 248.
8. Kriegsman, B.A.; Vangala, P.; Chen, B.J.; Meraner, P.; Brass, A.L.; Garber, M.; Rock, K.L. Frequent Loss of IRF2 in Cancers Leads to Immune Evasion through Decreased MHC Class I Antigen Presentation and Increased PD-L1 Expression. *J. Immunol.* 2019, 203, 1999–2010.
9. Yoshihama, S.; Roszik, J.; Downs, I.; Meissner, T.B.; Vijayan, S.; Chapuy, B.; Sidiq, T.; Shipp, M.A.; Lizée, G.A.; Kobayashi, K.S. NLRC5/MHC Class I Transactivator Is a Target for Immune Evasion in Cancer. *Proc. Natl. Acad. Sci. USA* **2016**, 113, 5999–6004.
10. van den Elsen, P.J.; Holling, T.M.; van der Stoep, N.; Boss, J.M. DNA Methylation and Expression of Major Histocompatibility Complex Class I and Class II Transactivator Genes in Human Developmental Tumor Cells and in T Cell Malignancies. *Clin. Immunol.* 2003, 109, 46–52.
11. Huimin Dan; Shanshan Zhang; Yongning Zhou; Quanlin Guan; DNA Methyltransferase Inhibitors: Catalysts For Antitumor Immune Responses. *OncoTargets and Therapy* **2019**, 12, 10903-10916, [10.2147/ott.s217767](https://doi.org/10.2147/ott.s217767).
12. Johnson, D.B.; Estrada, M.V.; Salgado, R.; Sanchez, V.; Doxie, D.B.; Opalenik, S.R.; Vilgelm, A.E.; Feld, E.; Johnson, A.S.; Greenplate, A.R.; et al. Melanoma-Specific MHC-II Expression Represents a Tumour-Autonomous Phenotype and Predicts Response to Anti-PD-1/PD-L1 Therapy. *Nat. Commun.* 2016, 7, 10582.
13. Park, I.A.; Hwang, S.-H.; Song, I.H.; Heo, S.-H.; Kim, Y.-A.; Bang, W.S.; Park, H.S.; Lee, M.; Gong, G.; Lee, H.J. Expression of the MHC Class II in Triple-Negative Breast Cancer is Associated with Tumor-Infiltrating Lymphocytes and Interferon Signaling. *PLoS ONE* 2017, 12, e0182786.
14. Zhou, J.; Wang, G.; Chen, Y.; Wang, H.; Hua, Y.; Cai, Z. Immunogenic Cell Death in Cancer Therapy: Present and Emerging Inducers. *J. Cell. Mol. Med.* 2019, 23, 4854–4865.
15. Inoue, H.; Tani, K. Multimodal Immunogenic Cancer Cell Death as a Consequence of Anticancer Cytotoxic Treatments. *Cell Death Differ.* 2014, 21, 39–49.
16. Li, H.; van der Leun, A.M.; Yofe, I.; Lubling, Y.; Gelbard-Solodkin, D.; van Akkooi, A.C.J.; van den Braber, M.; Rozeman, E.A.; Haanen, J.B.A.G.; Blank, C.U.; et al. Dysfunctional CD8 T Cells Form a Proliferative, Dynamically Regulated Compartment within Human Melanoma. *Cell* 2019, 176, 775–789.e18.

17. Im, S.J.; Ha, S.-J. Re-Defining T-Cell Exhaustion: Subset, Function, and Regulation. *Immune Netw.* 2020, 20, e2.
18. McGovern, I.; Dunn, J.; Batham, J.; Tu, W.J.; Chrisp, J.; Rao, S. Epitherapy and Immune Checkpoint Blockade: Using Epigenetic Reinvigoration of Exhausted and Dysfunctional T Cells to Reimburse Immunotherapy Response. *BMC Immunol.* 2020, 21, 22.
19. Yosuke Togashi; Kohei Shitara; Hiroyoshi Nishikawa; Regulatory T cells in cancer immunosuppression — implications for anticancer therapy. *Nature Reviews Clinical Oncology* **2019**, 16, 356-371, [10.1038/s41571-019-0175-7](https://doi.org/10.1038/s41571-019-0175-7).
20. Yasuko Tada; Yosuke Togashi; Daisuke Kotani; Takeshi Kuwata; Eichi Sato; Akihito Kawazoe; Toshihiko Doi; Hisashi Wada; Hiroyoshi Nishikawa; Kohei Shitara; et al. Targeting VEGFR2 with Ramucirumab strongly impacts effector/activated regulatory T cells and CD8+ T cells in the tumor microenvironment. *Journal for Immunotherapy of Cancer* **2018**, 6, 106, [10.1186/s40425-018-0403-1](https://doi.org/10.1186/s40425-018-0403-1).
21. Yosuke Togashi; Takahiro Kamada; Akinori Sasaki; Yoshiaki Nakamura; Shota Fukuoka; Yasuko Tada; Akihito Kawazoe; Kohei Shitara; Hiroyoshi Nishikawa; Clinicopathological, genomic and immunological features of hyperprogressive disease during PD-1 blockade in gastric cancer patients.. *Journal of Clinical Oncology* **2018**, 36, 4106-4106, [10.1200/jco.2018.36.15_suppl.4106](https://doi.org/10.1200/jco.2018.36.15_suppl.4106).
22. Xuexiang Du; Fei Tang; Mingyue Liu; Juanjuan Su; Yan Zhang; Wei Wu; Martin Devenport; Christopher A Lazarski; Peng Zhang; Xu Wang; et al. A reappraisal of CTLA-4 checkpoint blockade in cancer immunotherapy. *Cell Research* **2018**, 28, 416-432, [10.1038/s41422-018-0011-0](https://doi.org/10.1038/s41422-018-0011-0).
23. Kruger, S.; Ilmer, M.; Kobold, S.; Cadilha, B.L.; Endres, S.; Ormanns, S.; Schuebbe, G.; Renz, B.W.; D'Haese, J.G.; Schloesser, H.; et al. Advances in Cancer Immunotherapy 2019—Latest Trends. *J. Exp. Clin. Cancer Res.* 2019, 38, 268.
24. Farkona, S.; Diamandis, E.P.; Blasutig, I.M. Cancer Immunotherapy: The Beginning of the End of Cancer? *BMC Med.* 2016, 14, 73.
25. Robert M. Conry; Brian Westbrook; Svetlana McKee; Timothy Graham Norwood; Talimogene laherparepvec: First in class oncolytic virotherapy. *Human Vaccines & Immunotherapeutics* **2018**, 14, 839-846, [10.1080/21645515.2017.1412896](https://doi.org/10.1080/21645515.2017.1412896).
26. Medrano, R.F.V.; Hunger, A.; Mendonça, S.A.; Barbuto, J.A.M.; Strauss, B.E. Immunomodulatory and Antitumor Effects of Type I Interferons and Their Application in Cancer Therapy. *Oncotarget* 2017, 8, 71249–71284.
27. Parker, B.S.; Rautela, J.; Hertzog, P.J. Antitumour Actions of Interferons: Implications for Cancer Therapy. *Nat. Rev. Cancer* 2016, 16, 131–144.
28. Minn, A.J. Interferons and the Immunogenic Effects of Cancer Therapy. *Trends Immunol.* 2015, 36, 725–737.
29. Yang, X.; Zhang, X.; Fu, M.L.; Weichselbaum, R.R.; Gajewski, T.F.; Guo, Y.; Fu, Y.-X. Targeting the Tumor Microenvironment with Interferon- β Bridges Innate and Adaptive Immune Responses. *Cancer Cell* 2014, 25, 37–48.
30. Fujimura, T.; Hidaka, T.; Kambayashi, Y.; Furudate, S.; Kakizaki, A.; Tono, H.; Tsukada, A.; Haga, T.; Hashimoto, A.; Morimoto, R.; et al. Phase I Study of Nivolumab Combined with IFN- β for Patients with Advanced Melanoma. *Oncotarget* 2017, 8, 71181–71187.
31. Davar, D.; Wang, H.; Chauvin, J.-M.; Pagliano, O.; Fourcade, J.J.; Ka, M.; Menna, C.; Rose, A.; Sander, C.; Borhani, A. A.; et al. Phase Ib/II Study of Pembrolizumab and Pegylated-Interferon Alfa-2b in Advanced Melanoma. *J. Clin. Oncol. Off. J. Am. Soc. Clin. Oncol.* 2018, 36, JCO1800632.
32. Mok, T.S.K.; Wu, Y.-L.; Kudaba, I.; Kowalski, D.M.; Cho, B.C.; Turna, H.Z.; Castro, G.J.; Srimuninnimit, V.; Laktionov, K.; Bondarenko, I.; et al. Pembrolizumab versus chemotherapy for previously untreated, PD-L1-expressing, locally advanced or metastatic non-small-cell lung cancer (KEYNOTE-042): a randomised, open-label, controlled, phase 3 trial. *The Lancet* **2019**, 393, 1819-1830, [10.1016/s0140-6736\(18\)32409-7](https://doi.org/10.1016/s0140-6736(18)32409-7).
33. Caroline Robert; Antoni Ribas; Omid Hamid; Adil Daud; Jedd D. Wolchok; Anthony M. Joshua; Wen-Jen Hwu; Jeffrey S. Weber; Tara C. Gangadhar; Richard W. Joseph; et al. Durable Complete Response After Discontinuation of Pembrolizumab in Patients With Metastatic Melanoma. *Journal of Clinical Oncology* **2018**, 36, 1668-1674, [10.1200/jco.2017.75.6270](https://doi.org/10.1200/jco.2017.75.6270).