Oxaliplatin-Induced Peripheral Neuropathy

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Oxaliplatin-induced peripheral neurotoxicity (OIPN) is a severe and potentially permanent side effect of cancer treatment affecting the majority of oxaliplatin-treated patients, mostly with the onset of acute symptoms, but also with the establishment of a chronic sensory loss that is supposed to be due to dorsal root ganglia neuron damage. The pathogenesis of OIPN is still largely unknown. This lack of information is a limitation in the identification of effective strategies to prevent or limit OIPN incidence and severity. So far, no treatment is available for the prevention of OIPN, although duloxetine showed moderate evidence of efficacy in the reduction of OIPN symptoms. On this background, several neuroprotection clinical trials are ongoing in oxaliplatin-treated patients, although only part of them relies on solid preclinical evidence supporting the study hypothesis. Based on the available literature it can be concluded that dose and schedule modification is currently the most effective approach to limit the severity of OIPN since pharmacological prevention and treatment of OIPN still remains an unmet clinical need.

Keywords: oxaliplatin ; neurotoxicity ; acute ; chronic ; prevention ; treatment ; pain ; neuropathy

1. Introduction

Oxaliplatin (OHP) is a platinum-based antineoplastic drug used for colorectal cancer treatment whose main dose-limiting side effect is peripheral neurotoxicity, a disabling and potentially permanent consequence which could impair life quality of patients during and after therapy ^{[1][2]}. The main target of peripheral nervous system (PNS) damage due to OHP antineoplastic therapy is believed to be dorsal root ganglia (DRG), as a result of platinum accumulation ^{[3][4][5]}.

Oxaliplatin (OHP)-induced peripheral neurotoxicity (OIPN) produces two distinct manifestations of peripheral neuropathy: an acute cold-enhanced form and a chronic distal sensory neuropathy ^{[4][5]}.

The acute neurotoxicity, affecting the majority (80-90%) of OHP-treated patients ^[6], is a transient form ensuing hours after OHP administration and lasting 1-3 days, characterized by cold-induced paresthesias/dysesthesias, with predominant oropharyngeal and limbs distribution, frequently associated with cramps and fasciculations. Although of short duration, acute OIPN is disturbing for the patients. Moreover, it has been reported that patients with more severe acute OIPN are more likely to develop severe chronic neurotoxicity ^[2]. Although this observation does not certainly imply a direct causal relationship, it might however be considered an evidence of higher susceptibility of some individuals to PNS injury.

The incidence of chronic OIPN is variable, also because of the different assessment methods for its diagnosis, however it can be considered a frequent side effect, which might seriously impair the quality of life of the most severely affected patients. The first symptoms are generally numbness and tingling in hands and feet, with progressive distal-to-proximal limb involvement after increasing OHP exposition. Peripheral neurotoxicity may then progress toward sensory ataxia, with difficulty in manipulating small objects and necessity to base widening in standing; these symptoms are more evident in all the situations where defective proprioceptive input cannot be compensated by visual input, such as in poorly lit environments. Cranial and motor nerve impairment or autonomic disorders have very rarely been reported as manifestations of chronic OIPN ^[8].

The clinical features of acute and chronic manifestations of OHP neurotoxicity may be helpful in understanding the basic mechanisms of their onset. Acute OIPN is a typical feature of OHP treatment, while chronic OIPN, a cumulative and dose-dependent peripheral neurotoxicity, is common to other platinum-based drugs, in particular cisplatin.

2. Basic mechanisms: acute OIPN.

The time course of acute OIPN, with rapidly onset and reversal of symptoms, clearly suggests a functional, rather than structural, impairment.

A possible mechanism to explain acute OIPN is reversible neuronal plasma membrane ion channels dysfunction in the dorsal root ganglia (DRG). In fact, OHP can slow the inactivation of voltage-gated Na+ channels, an effect that may be enhanced by exposure to cold ^{[9][10][11][12][13]}, and cooling may also slow the activation of axonal slow K+ (Kv7) channels, ensuing axonal excitability modification ^[14]. This pathogenetic hypothesis involving ion channel-interference has been demonstrated in animal studies ^[15], and validated in small cohorts of OHP-treated patients through a non-standard neurophysiological technique, the Nerve Excitability Test, which gives indirect information about ion-channel activity ^{[16][17]}.

However, other hypotheses have been raised to explain the mechanism at the basis of acute OIPN. It has been recently reported, both in mouse DRG neuron culture and *in vivo*, that OHP causes an acidification of the cytosol and this in turn is responsible for sensitization of TRPA1 channels ^[18]. In the *in vitro* study a concentration of OHP like those found in plasma of treated patients has been employed.

Investigating OHP effects on the PNS, additional and unexpected targets were discovered. For instance, it has been demonstrated that OHP leads to a reduction of intracellular pH by forming adducts with neuronal hemoglobin. This observation is in agreement with the concept that hemoglobin in non-hematopoietic cells has non-iron-related activities. For instance, in neurons it can act as a proton buffer, an activity impaired by OHP binding. Moreover, it has been demonstrated that OHP neurotoxicity is sensitive to the activity of carbonic anhydrase, an enzyme that is linked to hemoglobin in intracellular pH homeostasis. The potential relevance of this latter observation is highlighted by the report that drugs such as topiramate and acetazolamide, able to inhibit carbonic anhydrase activity, can revert OHP-induced cytosolic acidification of DRG of treated animals and acute OIPN, without affecting OHP-induced cytotoxicity on cancer cells ^[19].

3. Basic mechanisms: chronic OIPN

The anticancer effect of OHP is mainly due to its binding and subsequent damage of nuclear DNA. In particular, the cytotoxicity is thought to result from platinum derivatives entering the cell nucleus with formation of both inter- and intrastrand cross-links, which prevent DNA replication and transcription; the inhibition of DNA synthesis leads to cell death, similarly to other platinum compounds (e.g., cisplatin, carboplatin).

The lack of blood-brain barrier in DRG allows platinum accumulation in DRG sensory neurons. The first descriptions of OIPN in rodent animal models evidenced DRG nucleolar, nuclear, and somatic size changes ^[20], and since then, many other preclinical studies investigated the effect of OHP administration and the related mechanisms of OIPN onset. Recently it was also demonstrated that OHP, although unable to cross the blood-brain barrier, could also affect the spinal cord, since wide dynamic range neurons hyperexcitability ^[21] and central glial activation ^[22] have been reported in OIPN rodent models.

Other studies suggest that also mitochondrial dysfunctions and oxidative stress could represent important pathogenetic mechanisms, since platinum metabolites can cause a significant reactive oxygen species (ROS) generation. The consequent oxidative stress can impair metabolic activity, both in cancer and healthy cells. Podratz and colleagues ^[23] reported in an *in vitro* study that cisplatin binds mitochondrial DNA with the same binding affinity observed for nuclear DNA in DRG neurons. However, since mitochondrial DNA is not provided with systems of DNA repair like those of nuclear DNA (e.g., base excision repair, nucleotide excision repair pathways), platinum adducts cannot be removed, leading to problems in mitochondrial DNA replication and transcription and protein synthesis, dysfunction of respiratory chain and, eventually, energy failure of neurons and oxidative stress. Other authors confirmed the presence of dysfunction in mitochondrial respiration and a decrease of ATP production in the sciatic nerves of rats treated with OHP ^[25]. In agreement with these data, Xiao and Bennet ^[24] pathological observations demonstrated swollen and vacuolated mitochondria in saphenous nerves of rats treated with OHP compared to controls. Therefore, the mitotoxicity hypothesis has gained a prominent position to explain the pathogenesis of chronic OIPN.

Other pathological events which may play an important role in chronic OIPN onset are neuroinflammation, either related to or independent from oxidative stress ^{[26][27]}, and modulation of voltage-gated Na+ channels. Finally, the mechanisms allowing intracellular accumulation of OHP must be considered. Under this perspective, the DRG neurons selective expression of plasma membrane transporters' (e.g., copper transporters such as CTR1, organic cation transporters such as OCT1 and 2) and their activity might explain their peculiar sensitivity to the toxic effects of OHP ^{[28][29]}.

A clinically relevant aspect of OIPN is the marked difference in its severity in different individuals with the same oncological and neurological conditions and treated with the same OHP schedule. Evidence of different susceptibility to OIPN in different mice strains suggests a possible role of genetic background, and this hypothesis has been investigated

in several clinical trials in OHP-treated patients, but they failed so far to provide firm confirmation, also in view of several methodological flaws in their design ^[30]

4. Conclusion

The lack of conclusive pathophysiologic evidence to explain OIPN clinical features must be clearly acknowledged, and this is one of the main reasons at the basis of the current incapacity to effectively treat this very serious condition. Currently, the only recommended treatment for OHP-induced painful peripheral neuropathy is the serotonin-norepinephrine reuptake inhibitor duloxetine ^[31], even though this drug is not effective for everyone and its treatment schedule is still under investigation in clinical trials.

Based on the available data, it can be stated that OHP dose and schedule modification is currently the most effective approach to limit the severity of OIPN, and that pharmacological prevention and treatment of OIPN still remains an unmet clinical need. This leads to the conclusion that more accurate and reliable preclinical investigation of the causes of OIPN is mandatory.

References

- 1. Cavaletti, G.; Marmiroli, P. Chemotherapy-induced peripheral neurotoxicity. Curr. Opin. Neurol. 2015, 28, 500–507.
- 2. Grisold, W.; Cavaletti, G.; Windebank, A.J. Peripheral neuropathies from chemotherapeutics and targeted agents: Diag nosis, treatment, and prevention. Neuro Oncol. 2012, 14 (Suppl. 40), 45–54.
- Cavaletti, G.; Tredici, G.; Petruccioli, M.G.; Dondè, E.; Tredici, P.; Marmiroli, P.; Minoia, C.; Ronchi, A.; Bayssas, M.; Eti enne, G.G. Effects of different schedules of oxaliplatin treatment on the peripheral nervous system of the rat. Eur. J. Ca ncer 2001, 37, 2457–2463.
- Marmiroli, P.; Riva, B.; Pozzi, E.; Ballarini, E.; Lim, D.; Chiorazzi, A.; Meregalli, C.; Distasi, C.; Renn, C.L.; Semperboni, S; et al. Susceptibility of different mouse strains to oxaliplatin peripheral neurotoxicity: Phenotypic and genotypic insight s. PLoS ONE 2017, 12, e0186250.
- Avan, A.; Postma, T.J.; Ceresa, C.; Cavaletti, G.; Giovannetti, E.; Peters, G.J. Platinum-induced neurotoxicity and preve ntive strategies: Past; Present; and Future. Oncologist 2015, 20, 411–432.
- Argyriou, A.A.; Velasco, R.; Briani, C.; Cavaletti, G.; Bruna, J.; Alberti, P.; Cacciavillani, M.; Lonardi, S.; Santos, C.; Cort inovis, D.; et al. Peripheral neurotoxicity of oxaliplatin in combination with 5-fluorouracil (FOLFOX) or capecitabine (XEL OX): A prospective evaluation of 150 colorectal cancer patients. Ann. Oncol. 2012, 23, 3116–3122.
- Argyriou, A.A.; Cavaletti, G.; Briani, C.; Velasco, R.; Bruna, J.; Campagnolo, M.; Alberti, P.; Bergamo, F.; Cortinovis, D.; Cazzaniga, M.; et al. Clinical pattern and associations of oxaliplatin acute neurotoxicity: A prospective study in 170 pati ents with colorectal cancer. Cancer 2013, 119, 438–444.
- 8. Grisold, W.; Grisold, A.; Löscher, W.N. Neuromuscular complications in cancer. J. Neurol. Sci. 2016, 367, 184–202.
- Kawashiri, T.; Egashira, N.; Kurobe, K.; Tsutsumi, K.; Yamashita, Y.; Ushio, S.; Yamashita, Y.; Ushio, S.; Yano, T.; Oishi, R. L type Ca²⁺ channel blockers prevent oxaliplatin-induced cold hyperalgesia and TRPM8 overexpression in rats. Mol. Pain 2012, 8, 7.
- Park, S.B.; Lin, C.S.; Kiernan, M.C. Nerve excitability assessment in chemotherapy-induced neurotoxicity. J. Vis. Exp. 2 012, 26, 3439.
- 11. Krishnan, A.V.; Goldstein, D.; Friedlander, M.; Kiernan, M.C. Oxaliplatin and axonal Na+ channel function in vivo. Clin. Cancer Res. 2006, 12, 4481–4484.
- 12. Gamelin, E.; Gamelin, L.; Bossi, L.; Quasthoff, S. Clinical aspects and molecular basis of oxaliplatin neurotoxicity: Curr ent management and development of preventive measures. Semin. Oncol. 2002, 29 (Suppl. 15), 21–33.
- 13. Adelsberger, H.; Quasthoff, S.; Grosskreutz, J.; Lepier, A.; Eckel, F.; Lersch, C. The chemotherapeutic oxaliplatin alters voltage-gated Na(+) channel kinetics on rat sensory neurons. Eur. J. Pharmacol. 2000, 406, 25–32.
- 14. Benoit, E.; Brienza, S.; Dubois, J.M. Oxaliplatin; an anticancer agent that affects both Na+ and K+ channels in frog peri pheral myelinated axons. Gen. Physiol. Biophys. 2006, 25, 263–276.
- 15. Alberti, P.; Canta, A.; Chiorazzi, A.; Fumagalli, G.; Meregalli, C.; Monza, L.; Pozzi, E.; Ballarini, E.; Rodriguez-Menende z, V.; Oggioni, N.; et al. Topiramate prevents oxaliplatin-related axonal hyperexcitability and oxaliplatin induced peripher al neurotoxicity. Neuropharmacology 2020, 164, 107905.

- Bennedsgaard, K.; Ventzel, L.; Grafe, P.; Tigerholm, J.; Themistocleous, A.C.; Bennett, D.L.; Tankisi, H.; Finnerup, N.B. Cold aggravates abnormal excitability of motor axons in oxaliplatin-treated patients. Muscle Nerve 2020, doi:10.1002/m us.26852. [Epub ahead of print].
- 17. Heide, R.; Bostock, H.; Ventzel, L.; Grafe, P.; Bergmans, J.; Fuglsang-Frederiksen, A.; Finnerup, N.B.; Tankisi, H. Axon al excitability changes and acute symptoms of oxaliplatin treatment: In vivo evidence for slowed sodium channel inactiv ation. Clin. Neurophysiol. 2018, 129, 694–706.
- Riva, B.; Dionisi, M.; Potenzieri, A.; Chiorazzi, A.; Cordero-Sanchez, C.; Rigolio, R.; Carozzi, V.A.; Lim, D.; Cavaletti, G.; Marmiroli, P.; et al. Oxaliplatin induces pH acidification in dorsal root ganglia neurons. Sci. Rep. 2018, 8, 15084.
- 19. Potenzieri, A.; Riva, B.; Rigolio, R.; Chiorazzi, A.; Pozzi, E.; Ballarini, E.; Cavaletti, G.; Genazzani, A.A. Oxaliplatin-indu ced neuropathy occurs through impairment of haemoglobin proton buffering and is reversed by carbonic anhydrase inhi bitors. Pain 2020, 161, 405–415.
- Cavaletti, G.; Tredici, G.; Marmiroli, P.; Petruccioli, MG.; Barajon, I.; Fabbrica, D. Morphometric study of the sensory ne uron and peripheral nerve changes induced by chronic cisplatin (DDP) administration in rats. Acta Neuropathol. 1992, 8 4, 364–371.
- 21. Renn, C.L.; Carozzi, V.A.; Rhee, P.; Gallop, D.; Dorsey, S.G.; Cavaletti, G. Multimodal assessment of painful peripheral neuropathy induced by chronic oxaliplatin-based chemotherapy in mice. Mol. Pain 2011, 7, 29.
- 22. Di Cesare Mannelli, L.; Pacini, A.; Micheli, L.; Tani, A.; Zanardelli, M.; Ghelardini, C. Glial role in oxaliplatin-induced neu ropathic pain. Exp. Neurol. 2014, 261, 22–33.
- 23. Podratz, J.L.; Knight, A.M.; Ta, L.E.; Staff, N.P.; Gass, J.M.; Genelin, K.; Schlattau, A.; Lathroum, L.; Windebank, A.J. Ci splatin induced mitochondrial DNA damage in dorsal root ganglion neurons. Neurobiol. Dis. 2011, 41, 661–668.
- 24. Xiao, W.H.; Bennett, G.J. Effects of mitochondrial poisons on the neuropathic pain produced by the chemotherapeutic a gents, paclitaxel and oxaliplatin. Pain 2012, 153, 704–709.
- 25. Zheng, H.; Xiao, W.H.; Bennett, G.J. Functional deficits in peripheral nerve mitochondria in rats with paclitaxel- and oxa liplatin-evoked painful peripheral neuropathy. Exp. Neurol. 2011, 232, 154–161.
- Makker, P.G.; Duffy, S.S.; Lees, J.G.; Perera, C.J.; Tonkin, R.S.; Butovsky, O.; Park, S.B.;Goldstein, D.;Moalem-Taylor, G. Characterisation of immune and neuroinflammatory changes associated with chemotherapy-induced peripheral neur opathy. PLoS ONE 2017, 12, e0170814.
- 27. Starobova, H.; Vetter, I. Pathophysiology of chemotherapy-induced peripheral neuropathy. Front. Mol. Neurosci. 2017, 10, 174.
- 28. Sprowl, J.A.; Ong, S.S.; Gibson, A.A.; Hu, S.; Du, G.; Lin, W.; Li, L.; Bharill, B.; Ness, R.A.; Stecula, A.; et al. A phospho tyrosine switch regulates organic cation transporters. Nat. Commun. 2016, 7, 10880.
- 29. Harrach, S.; Ciarimboli, G. Role of transporters in the distribution of platinum-based drugs. Front. Pharmacol. 2015, 6, 85.
- 30. Argyriou, A.A.; Bruna, J.; Genazzani, A.A.; Cavaletti, G. Chemotherapy-induced peripheral neurotoxicity: management i nformed by pharmacogenetics. Nat. Rev. Neurol. 2017, 13, 492–504.
- Hershman, D.L.; Lacchetti, C.; Dworkin, R.H.; Lavoie Smith, E.M.; Bleeker, J.; Cavaletti, G.; Chauhan, C.; Gavin, P.; La vino, A.; Lustberg, M.B.; et al. Prevention and management of chemotherapy-induced peripheral neuropathy in survivor s of adult cancers: American Society of Clinical Oncology clinical practice guideline. J. Clin. Oncol. 2014, 32, 1941–196 7.

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