COVID-19 Vaccine Candidates

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Abstract

COVID-19 vaccines are indispensable, with the number of cases and mortality still rising, and currently no medicines are routinely available for reducing morbidity and mortality, apart from dexamethasone, although others are being trialed and launched.

1. Introduction

While many candidates have been proposed to prevent and treat patients with COVID-19, including hydroxychloroquine, lopinavir-ritonavir, molnupiravir, and remdesivir, to date, only dexamethasone has shown a reduction in mortality in hospitalized patients receiving respiratory support; however, there is increasing evidence for medicines such as tocilizumab as well as casirivimab and imdevimab [1][2][3][4][5][6]. Whilst vaccines are being developed and administered, the recommended approach to reduce morbidity and mortality due to COVID-19 is the instigation of lockdown and social distancing measures [7][8][9][10]. However, lockdown measures have unintended consequences. Transport restrictions, closure of clinics, and concerns among patients attending hospital clinics have resulted in increases in non-communicable diseases as well as increased morbidity and mortality among unvaccinated children [11][12][13][14][15]. Lockdown measures also have economic consequences, especially among developing countries [16]. Consequently, there is increased urgency for an effective vaccine to combat COVID-19.

2. Current Vaccine Candidates

Recent advances in bioprocess technology, genomics, structural biology, and immunopathology have significantly contributed to the speed of COVID-19 vaccine development. Researchers have used accumulative knowledge from previous vaccine candidates, and within twenty-two months of the emergence of COVID-19, developed 155 preclinical vaccine candidates with 23 emergency use authorized ones (Figure 1).

Figure 1. Types of COVID-19 vaccine developed based on different technologies.
(1) inactivated,
(2) mRNA,
(3) viral vector, and
(4) nanoparticle-based peptide vaccines.

2.1. Inactivated Vaccine

Inactivated vaccines are formulated by inactivating virulent particles of viruses by treating the virus particle with chemicals, including formaldehyde, β-propiolactone, ethylenimine, phenol, ascorbic acid, β-aminophenylketone, and diethylpyrocarbonate \[^{17}\]. Among these inactivating chemicals, formaldehyde is currently not used to reduce the risk of incomplete inactivation \[^{18}\]. Since the virus particles are inactivated, they cannot multiply after entering the human body. Consequently, they are safe for administration but need to be introduced in large amounts compared to live attenuated vaccine.

BBIBP-CorV is a whole virion inactivated vaccine manufactured by Sinopharm (Beijing, China) and formulated by inactivating the novel coronavirus strain HB02, isolated from a patient admitted to the hospital. The reason behind the selection was its replication efficiency in Vero cells \[^{19}\][\(^{20}\)]. CoronaVac, another inactivated SARS-CoV-2 whole virion vaccine manufactured by Sinovac Life Sciences Co., was assembled by propagating the virus in Vero cells, followed by the inactivation using β-propiolactone. Aluminum hydroxide was coupled to the vaccine formulation as an adjuvant \[^{21}\]. During the phase 1/2 trial (human model; age 18-59 years), the production process of the vaccine was slightly different. The cell factory process was used to generate 50L Vero cell culture for the preclinical and Phase 1 trials, respectively, while for the Phase 2 trial, a bioreactor process was used for vaccine production \[^{21}\].

Interestingly, the bioreactor production process was more appropriate, as the control of the environment was easier and accurate during vaccine production. Moreover, it was suitable for bulk production and ensured the biosafety requirements. The bioreactor batch of the vaccine contained a higher spike antigen than the vaccine used in the phase 1 trial, which was unexpected. Fortunately, it did not change the vaccine’s safety profile. Instead, it increased the immunogenicity of the vaccine candidate \[^{21}\]. A separate study was conducted to evaluate the vaccine’s immunogenicity and safety in people aged 60 and older with CoronaVac vaccine. This study showed that the vaccine was suitable for older people as well \[^{22}\]. A Phase 3 trial of CoronaVac was conducted in Turkey. It was a double-blind, randomized, placebo-controlled trial with 10,218 volunteers aged 18-59 years. Among the participants, nine cases of COVID-19 were seen in the vaccine group, whereas thirty-two cases were reported in the placebo group during a follow-up period of 43 days. The overall efficacy of the vaccine was 83.5% after the second dose. A total of 18.9% of the population in the vaccine group and 16.9% in the placebo group experienced minor adverse events; injection site pain was the most frequent adverse event \[^{23}\].

BBV152 (Covaxin), another whole virion inactivated vaccine developed by Bharat Biotech (India), was produced by inactivating the virus and then formulating it with a toll-like receptor 7/8 agonist molecule, which was absorbed to alum (Algel-IMDG) \[^{24}\]. The formulation of this vaccine was decided after its preclinical trial in BALB/c mice, New Zealand white rabbits, and the Syrian hamster model. Three types of formulations, BBV152A (0.3 μg Ag + Algel-IMDG), BBV152B (0.6 μg Ag + Algel-IMDG), and BBV152C (0.6 μg Ag + Algel), were assessed. The BBV152B formulation showed a 10-fold better immune response in mice, whereas BBV152A showed better results in the Syrian hamster. Confirmation of the safety and reactogenicity of the vaccine formulation enabled it to receive approval for trials in humans \[^{25}\][\(^{26}\]. The other inactivated vaccine developed by Sinopharm (Beijing, China) (ChiCTP2000031809) was also made by isolating a SARS-CoV-2 strain WIV04 from a patient. The virus was cultivated in Vero cells, and the supernatant of the infected cell was treated with β-propiolactone to inactivate the virus. Alum was used as an adjuvant to the vaccine formulation \[^{27}\].
After administering the inactivated vaccine formulations, mild side-effects such as pain in the injection site, fever, fatigue, headache, nausea, and vomiting were observed. Nevertheless, no profound negative result was reported, confirming that they were safe and immunogenic [24][19][21][22][27].

Human trials with the above-discussed inactivated vaccine formulations induced considerable immune responses. Seroconversion was reported in all participants of these vaccine studies. Overall, the efficacies of these inactivated vaccines, such as BBIBP-CorV (82.50%), BBV152 (81%), and CoronaVac (83.5%), were found to be almost similar to each other (Figure 2). The neutralizing antibody (Nab) titer was seen to be increasing with the increased dosage of the vaccine (Figure 3). However, the increased dosage caused unfavorable events in the trial population in each trial, as mentioned above, other than the CoronaVac trial (Figure 4) [24][19][27]. For CoronaVac, Nab titer and minor unusual reactions to the vaccine were the same in the higher (6 µg) and lower (3 µg) groups. Thus, 3 µg dose of the vaccine was selected for the Phase 3 trial [21]. The cellular and humoral response and T cell memory response were generated by BBV152, although the B cell memory response is yet to be assessed [24].

Figure 2. Efficacy of different SARS-CoV-2 vaccine candidates. Here, the small circles imply the reported efficacy after vaccination. All the data were extracted from the included articles, which were selected for this systematic review only. As all the vaccines did not have the same response levels, the 95% CIs were not evenly distributed. For BBIBP-CorV, we were not able to find the upper and lower limits of 95% CI; thus, it was not reported in the figure.
Figure 3. IgG seroconversion of several SARS-CoV-2 vaccines by trial Phase (i.e., Phase I/II/III), dose number (i.e., 1st or 2nd dose), or days after vaccination (i.e., day 14/28/29/42/56). Data were extracted from the included articles which were selected for this systematic review only.

Figure 4. Adverse effects (AE) of several SARS-CoV-2 vaccines by trial phase (i.e., Phase I/II/III), dose number (i.e., 1st or 2nd dose). Data were extracted from the included articles which were selected for this systematic review only.
The trial of the BBIBP-CorV inactivated vaccine was limited by the absence of a longitudinal follow-up study and the assessment of the safety and immunogenicity in children. The vaccine trial was not designed to measure the efficacy of the vaccine. The study of CoronaVac only evaluated humoral response in the Phase 2 trial and did not report immune response data on immunocompromised or more susceptible populations. The comparison of the Nab titer determined from the trial data with the titer observed in COVID-19 patients was not reported. Only the neutralizing antibody assay was undertaken in older people, which excluded T cell response observation. The study subjects of the aged group were healthy. Consequently, there were no assessment results about the safety and immunogenicity of the vaccine in immunocompromised people. The longitudinal follow-up result of the participants is yet to be observed.

The trial of BBV152 was conducted when the number of COVID-19 cases was rapidly increasing. However, the results of the trial did not confirm the evaluation of core vaccine efficacy. Besides, the study population lacked multi-ethnicity and longitudinal re-examination. Several participants reported minor unfortunate adverse reactions to the vaccine during the Phase 2 trial compared to the Phase 1 trial (Figure 4). However, the positive aspect was that it was designed with participants from diverse geographic sites and different age ranges. Regardless of their age, no differences were reported in the immune response. After completing the Phase 1 trial, 6 µg Ag + Algel-IMDG formulation was selected for the Phase 3 trial. Because of the current pandemic situation, it received approval for emergency use in India. In the Phase 3 trial of BBV152, 24,419 participants enrolled. It was a double-blind, randomized, and multicenter trial. A total of 24 and 106 cases of COVID-19 were reported in the vaccine and placebo groups, respectively. Overall, the vaccine’s efficacy was 77.8%, whereas the vaccine’s efficacy against severe symptomatic and asymptomatic COVID-19 cases was 93.4% and 63.6%, respectively.

The trial design of the inactivated vaccine developed by Sinopharm did not include a plan for interim analysis in the original protocol. It was included while conducting the study, which was necessary for designing a Phase 3 trial. Despite producing a robust antibody response, whether the vaccine can provide protective immunity against COVID-19 is still unknown. The study result was based on a few groups of patients; hence, the study was unable to provide a comprehensive profile of suitability, immunity, and immune persistence. These ongoing vaccine studies need further evaluation to assess if this vaccine provides long-lived immunity against SARS-CoV-2.

### 2.2. mRNA Vaccine

mRNA vaccine generally consists of the elements essential for the encoded protein to be expressed. In the mRNA vaccine, 1-methyl-pseudouridine modification is incorporated in mRNA molecules, enhancing mRNA translation in the body. The antigen is initially identified from the target pathogen. After sequencing and synthesizing, the gene is usually cloned into a plasmid. Before being delivered into the host, the mRNA is transcribed in vitro. After its injection into the body, it uses the host cellular machinery to translate the mRNA into the target antigen. Commonly, both humoral and cellular immunity are induced as the mRNA vaccine mimics the initial viral infection. Chemokines and cytokines (i.e., IL-12, TNF) are produced at the injection site, generating robust innate immunity.

Compared to subunit, killed, live attenuated, and DNA-based vaccines, mRNA vaccines are preferred as they are safe and hardly have any harmful risk of infection. Besides, the mRNA vaccine is more stable, easily translatable, rapidly producible, and usually economical. The easy availability of mRNA’s printing facility plays a crucial role in producing considerable quantities of mRNA that facilitate mRNA vaccine production. The mRNA vaccine’s good adequacy and self-adjuvant properties elicit adaptive solid immune responses by releasing TNF-α, IFN-α, and other chemokines, by immune cells. Polypeptide and protein-based vaccines require additional adjuvants, whereas the mRNA vaccine does not require these. Again, mRNA vaccines express target proteins in the cytoplasm instead of entering the nucleus, making them more efficient than DNA vaccines.
mRNA-based vaccines against HIV, CMV, Chikungunya, and Zika were developed before the COVID-19 pandemic [42]. However, none of these have been approved by the FDA to date. Due to advancements in nucleic acid technologies, their performance has improved in humans in recent years because of their new formulations and modifications [40]. During the current COVID-19 pandemic, Moderna, BioNTech, and Pfizer have launched mRNA-based vaccine candidates.

Moderna and Pfizer/BioNTech developed COVID-19 vaccines by implementing nanotechnology [43][44][45][46]. The human body has ribonuclease enzymes present in every tissue and ready to destroy scattered RNA. Ribonuclease creates a restricted environment for foreign RNA, which might originate from plants or animals. Moreover, negatively charged mRNA cannot cross the negatively charged cell membrane, making it challenging to restore mRNA integrity and enter the host cell. Researchers designed lipid nanoparticles to carry siRNA or mRNA into host cells. Lipid nanoparticles can be created to encapsulate RNA and shield it from ribonuclease. The lipid nanoparticles also allow passage through the cell membrane. Those nanoparticles can be decorated with ligands that allow targeting certain immune cell types.

Moreover, adjuvants can be added to the interior of the lipid nanoparticles for further upregulation of the immune system’s response to the vaccine. Lipid nanoparticle-based technology can be a beneficial solution to challenge RNA delivery to cells [42][49][50][51][52][53][54]. Vaccine development by nanotechnology improves nucleic acid delivery and conformation-stabilized subunit vaccines to lymph nodes. It triggers cellular and humoral immunity, preventing viral infection and disease severity [55].

The mRNA-1273 vaccine, manufactured by Moderna, encodes SARS-CoV-2’s Spike glycoprotein (S-2P antigen). An intact S1-S2 cleavage site and SARS-CoV-2 glycoprotein, along with the transmembrane, were anchored, making up the vaccine. Two successive proline substitutions at amino acid positions 986 and 987 stabilize S-2P on its prefusion conformation. These empowered the assurance of an atomic-level structure for the prefusion adaptation of spikes from endemic and pandemic strains counting HKU1, SARS-CoV, and MERS-CoV. Innovative structure-based vaccine design, modified nucleotides, and delivery methods by lipid nanoparticles are the principal reasons for mRNA-1273’s rapid immunogenicity.

The Phase 1 trial of mRNA-1273 on humans was conducted at the Kaiser Permanente Washington Health Research Institute in Seattle and the Emory University School of Medicine in Atlanta. After the first immunization, the day 29 enzyme-linked immunosorbent assay anti–S-2P antibody geometric mean titer (GMT) was found as 40,227 (25 μg group), 109,209 (100 μg group), and 213,526 (250 μg group), where a positive correlation was observed between dose and GMT. After the second immunization, day 57 GMT enhanced up to 299,751 (25 μg group), 782,719 (100 μg group), and 1,192,154 (250 μg group), respectively maintaining the initial response (Figure 3). However, approximately 21% of the 250 μg dose group had one or more severe unfortunate events (Figure 4) [56].

Significant immune responses to SARS-CoV-2 were reported in patients 18 years and older from the Phase 2 trials, which confirmed immunogenicity and safety of 50 and 100 μg of the mRNA-1273 vaccine. Within 28 days after the first vaccination, anti-SARS-CoV-2 spike binding and neutralizing antibodies were elicited by both doses of the mRNA-1273 vaccine. After the second vaccination, titers peaked by 14 days, which exceeded the COVID-19 patient’s convalescent sera level (Figure 3). Nevertheless, the Phase 2 trial had some limitations. The study population was not outlined at the time to be representative of those likely to get COVID-19. Statistical comparison of superiority or proportionality between doses was also not included in the study [57].

The Phase 3 trial showed 94.1% efficacy of the mRNA-1273 vaccine in preventing COVID-19 illness (Figure 2). The reactogenicity profile was similar to the Phase 1 data, and no unexpected concerns were detected. After the first dose of the vaccine, the level of reactogenicity was less than the zoster vaccine that was recently approved. The short duration of follow-up to investigate the safety and efficacy, and lack of any specified correlation of protection, were the drawbacks of the Phase 3 trial [58].
BioNTech and Pfizer manufactured both the BNT162b2 and BNT162b1, where a full-length spike of SARS-CoV-2 was encoded by BNT162b2. Two proline mutations were carried out to lock its prefusion conformation. As a result, this vaccine mimicked the intact virus and elicited immunity. On the other hand, the receptor-binding domain (RBD) of spike protein was encoded by BNT162b1. Trimerization was carried out by adding a T4 fibritin-foldon domain. Consequently, the immunogenicity was enhanced by the multivalent display. Lipids were used to formulate this vaccine and supplied as a buffered liquid solution.

The Phase 2/3 trial with the BNT162b1 vaccine was performed among 45 healthy adults, which assessed safety, tolerability, and immunogenicity of BNT162b1 at three dose levels (i.e., 10 µg, 30 µg, or 100 µg). BNT162b1 generated robust immunogenicity after vaccination, where dose levels had a positive correlation with the antibody titer. Moreover, the second dose increased SARS-CoV-2 neutralizing antibody titers and RBD-binding IgG concentrations. The second immunization with 100 µg was not administered because of the expanded reactogenicity and a need for seriously increased immunogenicity after a single dose compared with the 30 µg dosage. This trial proposed that dose levels between 10 µg and 30 µg with BNT162b1 might be well-tolerated and immunogenic.

However, several limitations were noticed. These included the kind, i.e., T cells versus B cells or both, and the level of immunity required to ensure protection from COVID-19 was unknown. In addition, the immune responses or safety were not assessed beyond two weeks after the 2nd vaccination. Again, as the study population was only up to 55 years old, the trial could not evaluate the plausible risk factors in the people beyond that particular age range.

A randomized, placebo-controlled, double-blind Phase 1 study of the BNT162b1 mRNA vaccine was conducted in younger and older Chinese adults to assess the preliminary safety and immunogenicity. This study showed an acceptable safety profile of BNT162b1.

The safety and immunogenicity of three dose levels of BNT162b1 and BNT162b2 were also assessed in a Phase 1 trial on 145 healthy adults. In this trial, BNT162b2 showed less severity and incidence of adverse effects than BNT162b1 while eliciting a similar dose-dependent antibody titer, parallel to the GMT of SARS-CoV-2 convalescent patients, or even more in some cases. The reason behind the high reactogenic profile of BNT162b1 compared to BNT162b2 was the difference in their nucleotide sequences by which vaccine antigens were encoded and the overall RNA construct size. As a result, RNA molecules of BNT162b1 were five times higher than the same concentration of BNT162b2, which elicited high immune stimulation and a reactogenic profile. Besides, the lack of knowledge regarding how effective it would be in protecting COVID-19 in a real-world sense was a major limitation of the Phase 1 trial along with the assessment of humoral and cellular immunity in protecting COVID-19. Although the Phase 1 part of this trial evaluated several hypotheses, it was not large enough to allow systematic statistical comparisons and standardized among laboratories. Regarding all these issues, BNT162b2 was subsequently selected for Phase 2/3 safety and efficacy trials.

Phase 2/3 trials evaluated the safety, immunogenicity, and efficacy of BNT162b2 in preventing COVID-19. Two doses of 30 µg BNT162b2 conferred 95% effectiveness in preventing COVID-19 in patients aged 16 years of age or older and showed safety over a median of 2 months (Figure 2). However, this trial also had several limitations. Unexpected events such as right axillary lymphadenopathy, right leg paresthesia, paroxysmal ventricular arrhythmia, and shoulder injury were reported among BNT162b2 recipients (Figure 4). The vaccine also required freezing temperature for shipping and more extended storage.

A Phase 3 trial was also conducted to assess the safety, efficacy, and immunogenicity of BNT162b2 in 12- to 15-year-old participants. This study showed that a two-dose regimen of 30 µg of BNT162b2 was highly safe and immunogenic for adolescents (12 to 15 years of age) with an efficacy of 100%.

An exciting observation reported in BNT162b2 recipients were those who previously had SARS-CoV-2...
infection; the anti-Spike titer increased approximately 140-fold within 19–29 days compared to those who had not been infected \[63\]. A single dose of this mRNA vaccine enhanced spike protein-specific antibody IgG level, ACE2 receptor binding inhibition reactions, and post-vaccine symptoms in individuals who were previously infected with SARS-CoV-2, similar to the second dose in individuals who were not infected \[66\]. Another study suggested that antibodies against SARS-CoV-2 nucleocapsid (N), RBD, and spike proteins (i.e., S1 and S2) were raised after the single dose of mRNA vaccine in those who were already seropositive or had a recent history of infection as compared to individuals with no history \[68\].

### 2.3. Viral Vector-Based Vaccine

In a viral vector-based vaccine, a gene/cDNA coding for a pathogen-derived antigen is incorporated into a non-pathogenic or attenuated viral species \[69\]. These non-pathogenic species serve as a vector. The recombinant vector immunizes against the pathogen while the gene product is expressed on its surface. For this purpose, a few sites are removed from the vector genome where the targeted pathogen's foreign DNA can be integrated. After injecting, that foreign DNA can replicate within the host and express the following pathogen's gene product, eliciting cell-mediated and humoral immunity. Different viruses, including adenovirus, vaccinia virus, canarypox virus, and attenuated poliovirus, act as viral vectors. Among them all, the adenovirus is relatively conventional in vaccine formulation \[69\]–\[70\].

Adenoviruses contain a distinctive icosahedral protein structure that encapsulates a linear double-stranded DNA genome of 36k base pairs. The viral genome has approximately a dozen capsid proteins without any lipid envelop and encodes nearly 35 proteins elicited in two phases, “early” and “late,” related to viral DNA replication \[71\]. Different types of adenoviral serotypes are separated from a variety of mammalian species. There are approximately 51 serotypes from humans, 27 from simians, and 7 from chimpanzees. Human serotypes are subdivided into six subgroups \[72\]. They are responsible for various clinical diseases that commonly infect the gastrointestinal and respiratory systems \[72\]. More than one serotype could be accountable for mild infection in most immunocompetent individuals, which is supposed to render lifelong immunity \[71\].

There are several distinctive features reflected by adenoviruses, making them a suitable option for vaccine formulation. Firstly, recombinant adenoviruses have a better safety record when used as a vector for gene therapy in humans. Because of its extensive tissue tropism ability, adenoviral vectors are supposed to exploit vaccine development against M. tuberculosis and influenza. Secondly, adenoviruses are highly immunogenic, driving up a strong and long-lasting immune response. For vaccine design purposes, human serotype 5 adenoviruses (AdHu5) are used broadly, while other non-human adenoviruses are also used in modern times. The adenoviral genome encodes five early proteins (E1a, E1b, E2, E3, and E4) responsible for DNA replication and evasion and a single late protein responsible for structural conformation. Removing the E1a gene (critical regulator of viral replication) from the adenoviral genome eliminates the virus’s ability to replicate, simultaneously increasing the potency to accommodate transgene cassettes up to 5000 bp in size. Deleting multiple genome units can augment the vector capacity for yielding multivalent vaccines against deadly pathogens \[74\].

Recombinant adenoviral vector vaccine trials were undertaken during the MERS-CoV epidemic. Dromedary camels were an animal reservoir of MERS-CoV. Two recombinant adenoviral vector vaccines, Ad5.MERS-S and Ad5.MERS-S1, were designed to generate an immune response against dromedary camels to eradicate MERS-CoV transmission from the reservoir to humans. Ad5.MERS-S encoded full-length S protein of MERS-CoV, and other Ad5.MERS-S1 encoded S protein’s S1 extracellular domain. Both vaccines immunized BALB/c mice and generated effective immunity \[75\].

BVRS-GamVac-Combi vaccine was manufactured by NF Gamaleya Research Institute of Epidemiology and Microbiology against MERS-CoV, where recombinant human adenoviruses 26 and 5 serotypes were used. Both vectors encoded the glycoprotein of the MERS-CoV. This vaccine immunized mice of C57BL/6 strain and common marmoset \[76\].
During the COVID-19 pandemic, the Beijing Institute of Biotechnology and CanSino Biologics manufactured the Ad5 vector COVID-19 vaccine. The replication-defective Ad5 vector vaccine encodes spike glycoprotein. Based on Wuhan-Hu-1 (GenBank accession number YP_009724390), an optimized full-length spike gene was cloned with tissue plasminogen activator signal peptide gene and E1 and E3 deleted Ad5 vector, resulting in an Ad5 vector COVID-19 vaccine developed utilizing the Admax system. It was manufactured as the liquid formulation in a vial containing $5 \times 10^{10}$ viral particles/0.5 mL. The Phase 1 trial found that the vaccine is tolerable and immunogenic after 28 days of vaccination. From day 14 of immunization, rapid T cell responses were remarkable. Humoral immune responses peaked at day 28 of immunization. The Phase 2 trial showed a $5 \times 10^{10}$ viral particles dose as safe for vaccination. Significant immune responses were induced after a single immunization. The short duration of follow-up, small cohort size, and absence of randomized control groups limited the findings from the Phase 1 trial. The study was also not statistically powered to assess the level of any side-effects. The Phase 2 trial, too, also presented some limitations. The problems began as the study started before complete data analysis from the Phase 1 study was accessible. Consequently, it failed to show the difference between different dosage groups. Children were also not included in this trial.

The Institute of Biotechnology, Academy of Military Medical Sciences, PLA of China, developed an aerosolized adenovirus type-5 vector-based COVID-19 vaccine (Ad5-nCoV). The Phase 1 trial was performed at Zhongnan Hospital to evaluate the vaccine’s safety, tolerability, and immunogenicity administered via inhalation. This study showed that aerosolized Ad5-nCoV is painless, simple, well-tolerable, and an aerosolized dose-induced antibody and cellular immune responses are equal to a fifth of the usual injected dose.

Among several candidate vaccines, the three most promising adenoviral vector-based vaccines have been going through clinical trials to ensure the host’s safety, efficacy, and immunogenicity while inducing an immune response. Adenoviral vector combined with DNA and poxviral vector to induce immunogenicity showed cellular and humoral response enhancement. The Oxford/AstraZeneca vaccine contains a homologous adenoviral vector that could mitigate the efficiency of the second dose due to anti-vector immunity. The “chimpanzee adenovirus” was an excellent vector for vaccine formulation from previous vaccine preparations against MERS-CoV. The Phase 1 clinical trial showed promising results, maintaining potency and protecting non-human primates against MERS-CoV. ChAdOx1 MERS exhibited safety and efficacy, which expressed cellular and humoral immune responses at the highest dose ($5 \times 10^9$ viral particles).

The Oxford/AstraZeneca vaccine is an adenoviral vector-based vaccine comprising DNA that encodes surface glycoprotein protein embedded in a capsid from a modified chimpanzee adenovirus, ChAdOx1 (replication-deficient simian adenovirus vector). A single vaccination with ChAdOx1 nCoV-19 into rhesus macaques showed potent humoral and cellular responses in a preclinical trial stage. A high-dose vaccination into a non-human primate showed protection against lower respiratory tract infections. After the Phase 1/2 clinical trial, the ChadOx1 nCoV-19 vaccine showed anti-spike IgG response, early T-cell response, and neutralizing antibody response, which illustrated an admissible safety profile with an enhanced humoral and cellular response. Vaccinated participants presented different amounts of immune cell clusters of B-cells, T-cells, and NK cells. Strong activated B-cells are found, and anti-spike IgG and IgA antibodies against SARS-CoV-2 spike protein were identified from vaccinated individuals. Identical CD4+ and CD8+ T-cell patterns were observed, responsible for the expression of CD69 and Ki-67, between days at 7 and 28 after vaccination. Production of cytokines (TNF-α and IFN-γ) by CD4+ T-cells was also identified on day 14 after immunization. On the other hand, NK cells expressed cytotoxic activity against viral infection at the highest rate at day 28.

A published report showed that a booster dose is more efficient in inducing multifunctional antibody titers with different types of effector mechanisms, including antibody-dependent neutrophil/monocyte phagocytosis, natural killer (NK) cell degranulation, complement activation, and cellular phagocytosis against SARS-CoV-2 infection. Strong T-cell responses have also been delineated, in which highly
activated cytotoxic T-cells destroy virus-infected cells to stop the further cell-to-cell spread of the virus. On the other hand, helper T-cells play a supporting role in activating B-cells for frequent antibody production. The booster dose is less reactogenic compared with the priming dose. Local and systemic reactions were consistently reduced after the second dose (Figure 4). A booster dose necessitates its importance in the restoration of sustainable immunity [83].

The Phase 3 trial of the ChadOx1 nCoV-19 vaccine has shown improved findings, with the vaccine being more well-tolerated and efficient (70.4%) in older than younger adults and similarly immunogenic for all age groups [83]. A well-accepted safety and efficacy profile of AZD1222 (ChadOx1 nCoV-19) has been established after analyzing ongoing trials in Brazil, South Africa, and the UK, making it an efficient and robust candidate vaccine against SARS-CoV-2 globally (Figure 2 and Figure 4) [84]. Epidemiologic efficacy of the Oxford/AstraZeneca ChAdOx1 nCoV-19 vaccine against the B.1.351 variant was 10.4% (95% CI, −76.8 to 54.8). However, the vaccine effectiveness against the B.1.1.7 variant was comparatively higher than the exhibited efficacy of 70.4% (95% CI 43.6–84.5) for B.1.1.7 and 81.5% (67.9–89.4) for non-B.1.1.7 lineages. However, the neutralization activity of ChAdOx1 nCoV-1 decreased in the B.1.1.7 variant compared to the non-B.1.1.7 variant [85][86].

Mutations in the RBD and N-terminal domain (NTD) of the SARS-CoV-2 spike gene are the major concern for vaccine development in recent pandemics [86]. RBD mutations (N501Y mutation) are responsible for increasing affinity to the ACE-2 receptor. On the other hand, E484K and K417N RBD mutations and mutations in NTD are accountable for escaping from the neutralizing antibody response. The B.1.1.7 lineage, first identified in the UK, contains the N501Y mutation with 53% increased transmissibility. Another mutated clan, named B.1.351, identified in South Africa, includes three RBD mutations and five NTD mutations. An independent lineage found in Brazil also adopted E484K, K417N, and some B.1.351 mutations [86].

An individual analysis of the Oxford/AstraZeneca vaccine against B.1.351 (South African variant) was undertaken between 24 June and 9 November 2020, where 2026 participants were enrolled. The T-cell response was not effective. Thereby, significant portions of viral antigens of B.1.351 variants remained flawlessly active. Furthermore, the vaccine failed to show a protective immune response against the B.1.351 variant, whereby the vaccine efficacy against the variant was 10.4% (95% CI, −76.8 to 54.8) [86].

Sputnik V, a heterologous adenoviral vector-based vaccine manufactured by Gamaleya Research Institute, was designed with two recombinant adenovirus vectors, adenovirus type 26 (rAd26) and adenovirus type 5 (rAd5), and both contain full-length glycoprotein S [87]. rAd26 vector was previously used for different vaccine candidates, such as Ad26.ZE.BOV against Ebola virus; Ad26.Mos.HIV, Ad26.Mos4.HIV, and Ad26.ENVA.01 against HIV; Ad26.CS.01 against Malaria; and Ad26.ZIKV.001 against Zika. These candidate vaccines are being tested, and clinical studies are still ongoing [88][89][90][91][92].

Sputnik V vaccine was developed in two versions, i.e., frozen (Gam-COVID-Vac) and lyophilized (Gam-COVID-Vac-Lyo). The Gam-COVID-Vac study took place at the branch of Burdenko Hospital, and the volunteers who took part in this study were military personnel and civilians. The study of Gam-COVID-Vac-Lyo was undertaken at Sechenow University, and all volunteers were civilians [87]. The vaccine showed strong immune response and protection in non-human primates against SARS-CoV-2 and displayed 100% defensive measure against a lethal version of SARS-CoV-2 in a preclinical study with immunosuppressed hamsters.

After the Phase 1/2 trials, Sputnik V showed T cell responses in healthy adults and decent titers of neutralizing antibodies. Collective data exhibit higher immunogenicity with robust cellular and humoral immune responses, resulting in higher antibody titers in vaccinated participants than the individuals with convalescent plasma [87].

The Phase 3 trial took place at 25 hospitals and polyclinics in Russia. The Gam-COVID-Vac trial showed
91.6% efficacy against COVID-19 and 100% protection against severe COVID-19 (Figure 2). In this trial, people 60 years or older were given importance in the vaccine-inducing immune response’s protection measures and efficiency. Results showed the ability to induce a virus-neutralizing humoral reaction in 60-year-old individuals. Vaccine efficacy did not alter significantly in young adults and old-age vaccinated participants. However, all risk groups, including children and pregnant women, were not enrolled in the Phase 3 trial. The vaccine is developed in liquid form (stored at −18 °C), and the freeze-dried (held at 2–8 °C) formulation is helpful in the distribution of vaccines in different weather conditions globally. The Phase 3 clinical trial showed a compatible safety profile and robust immune responses in all age groups from young to old participants (Figure 3 and Figure 4) [93]. The Phase 3 trial showed the sputnik vaccine required two doses to reach 91.6% efficacy, with a 79.4% efficacy after one dose as emergency administration [87][93].

FINLAY-FR-1A, a recombinant dimeric RBD-based vaccine manufactured by a Cuban epidemiological research institute, Finlay Institute, against COVID-19, showed a safe and reactogenic outcome in the Phase 1 trial (Figure 4). Secondary outcomes evaluated vaccine immunogenicity. One week after vaccination with a single dose, antibody response enhanced more than 20-fold compared to the Cuban convalescent serum panel’s median [94].

VXA-CoV2-1, developed by Vaxart, is an attractive recombinant vaccine candidate against SARS-CoV-2 and is an oral vaccine formulation. The preclinical trial of this vaccine was conducted in 6- to 8-week-old female Balb/c mice. Due to the mice’s inability to swallow pills, they were injected with 20 µL of the vaccine formulation. Two types of recombinant vaccine formulations, rAd-S-N (vector that expresses S and N protein) and rAd-S1-N (vector that expresses a fusion protein of S1 domain and N protein), were assessed. The former was selected for GMP manufacturing, as it induced a higher immune response, including vaccine-induced T cell response and the production of IFN-γ, TNF-α, and IL-2 CD4+ T cells. This vaccine candidate presents several advantages. Because it is an oral formulation, this makes it more accessible, as it is easier to administer. It is also easier to store, being an oral tablet vaccine, eliminating the need for cold storage transport and holding it in a refrigerator once delivered. Another advantage of this vaccine candidate is it is safe [95].

Another promising candidate vaccine is Ad26.COV2.S, manufactured by Janssen Pharmaceuticals. Ad26.COV2.S utilizes a recombinant, replication-deficient adenovirus serotype 26 (Ad26) vector encoding a stabilized SARS-CoV-2 spike (S) protein educated from the first clinical isolate of the Wuhan strain (Wuhan 2019; whole-genome sequence, NC_045512). Ad26 vector-based vaccines are usually safe and highly efficient and are also being used in the Sputnik V vaccine [87][96].

The Ad26.COV2.S vaccine has been tested in adult and aged rhesus macaques dividing into one- and two-dose regimens to evaluate the protective immune response and efficacy. A two-dose Ad26.COV2.S regimen promoted an ascending neutralizing antibody response compared to a single-dose regimen. However, neutralizing antibody responses were stable for a minimum of 14 weeks in one-dose regimens of the Ad26.COV2.S vaccine, and it also upregulated the humoral immunity and Th1 cellular responses in aged NHP [97].

Eight hundred five healthy adults have been assigned for participating in a Phase 1-2a trial of the Ad26.COV2.S vaccine. In the trial, cohort 1 belongs to participants aged 18–55 years, and cohort 3 belongs to 65 years or older-aged participants. A single dose of the Ad26.COV2.S vaccine elicited both neutralizing antibody and spike-binding antibody responses in 90% of participants on day 29 and reached 100% at day 57 with an increase in the titer. In addition, the CD4+ T-cell responses were found in 76–83% of participants in cohort 1 and 60–67% of those in cohort 3. On the other side, strong CD8+ T-cell responses were detected in all participants but at a comparatively lower level in older individuals than in younger [96].

The Phase 3 trial demonstrated the efficacy of a single dose of the Ad26.COV2.S vaccine. A total of 67%
and 66% efficacy was shown in participants against moderate to severe-critical COVID-19 disease with an onset at least 14 and 28 days after vaccine administration, respectively. The vaccine efficacy was 76.7% and 85.4% for severe-critical COVID-19 with onset at days 14 and 28, respectively. Reactogenicity was higher with the vaccine group, but a casualty of adverse events was not serious in the vaccine group. Overall, a single dose of Ad26.COV2.S is protective against symptomatic and asymptomatic SARS-CoV-2 infections and effective against severe/critical disease to reduce hospitalization and death. The Janssen adenovirus virus vaccine against B.1.351 variant has shown a 57% efficacy.

2.4. Nanoparticle-Based Peptide Vaccine

Nanotechnology has played an influential role in vaccine development with variations based on nanoparticles’ different compositions, shapes, sizes, and surface properties. Nanoparticles, being smaller in size, can quickly enter into living cells through endocytosis. Different types of nanoparticles are being used in vaccine development, including polymeric nanoparticles, inorganic nanoparticles, liposomes, immune-stimulating complex (ISCOM), virus-like particles, self-assembled proteins, and emulsions. Nanoparticles are most commonly used as immunostimulants or delivery materials. In vaccine formulation, the association between nanoparticles and antigens is essential. Nanoparticles act as a temporary carrier and protector of the antigen, which needs to reach the desired location. By interacting with the antigen, nanoparticles enhance immunogenicity and antigen processing, which activate immune responsive pathways.

Nanotechnology-based vaccine mechanisms are highly efficient, whereas solid nanocarriers transport the core antigen portion of vaccines into the gut-associated lymphoid tissues and mucosa-associated lymphoid tissues, ensuring proper delivery through oral or mucosal routes. Core particles are taken up by the dendritic cells and macrophages, which improve the cellular uptake of antigens and upregulate the antigen recognition and presentation. Nanoparticles are coated with immune cell-targeting molecules that bind with the cellular receptors to stimulate the specific and appropriate immune response. However, no contextual and relevant results have yet been published regarding nanotechnology-based vaccines since the outbreak of severe acute respiratory syndrome (SARS-CoV) and middle-east respiratory syndrome (MERS-CoV) other than the current COVID-19 pandemic.

In this ongoing SARS-CoV-2 pandemic, a subunit vaccine (NVX-CoV2373) has been developed using full-length glycoprotein S and administered with Matrix-M adjuvant into non-human primates and mice models, spurring Th1-dependent B- and T-cell responses, production of hACE2 receptor blocking antibodies, and SARS-CoV-2 neutralizing antibodies. No vaccine-related adverse effects were reported in mice models, which encouraged further clinical development of NVX-CoV2373 against COVID-19.

Researchers fabricated a modified “spike gene” of SARS-CoV-2 and installed it into baculovirus, which can only infect insects. Hence, selective moth cells were chosen and infected with the recombinant baculovirus. Consequently, the infected cell started to produce spike proteins assembled to form full-length spike protein similar to SARS-CoV-2. After that, spike proteins were purified and fixed with nanoparticles, which were used as a vaccine. Before being mixed with adjuvant distilled from soapbark plants, this vaccine attracted the immune cells to the injection site and activated the solid immune response to nanoparticles. Antigen-presenting cells (APC) uptake and present the spike nanoparticles on its membrane to T lymphocytes via major histocompatibility complex (MHC). T lymphocytes activate the antibody-producing B cells. A different type can be started by APC, called a killer T cell, which can recognize coronavirus-infected cells and destroy them before the further proliferation of new viruses.

In Phase 1/2 trials, 131 healthy adults of different age groups participated in two-dose regimens and were administered with 5 μg and 25 μg of rSARS-CoV-2 with or without the Matrix-M1 adjuvant. Considerable safety results and the ability to induce immune responses with higher amounts of neutralizing antibodies
were found in groups with adjuvant compared to groups without adjuvant. After the second vaccination with a similar dosage, the antibody response in participants had surpassed compared to the convalescent serum from COVID-19 patients. This report expresses the advantage of Matrix-M1 in the case of accelerating the functional antibody and T-cell response. No serious local or systemic adverse events occurred with the vaccinated groups. Body pain, joint pain, and fatigue were the most common systemic events that have been reported after the Phase 1/2 trial (Figure 4) [112].

A total of 15,187 participants were included in the Phase 3 trial of NVX-CoV2373, which was found to be 89.7% effective against both B.1.1.7 and non-B.1.1.7 variants. This B.1.1.7 variant is more transmissible and infectious than the previous strains. The vaccine efficacy of NVX-CoV2373 was higher than that of the ChAdOx1 nCoV-19 vaccine (Oxford/AstraZeneca) (70.4%) after the Phase 2-3 trial. The NVX-CoV2373 vaccine exhibited a lower efficacy level (51.0%) against the B.1.351 variant [89][111] (Figure 2). Scientists suggested that the NovaVax can be effective for a long time and prevent future coronavirus infections. It is easy to store for a long time at 4 °C, making it convenient to transport [111].

References

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