

# PIM Kinases in Multiple Myeloma

Subjects: **Oncology**

Contributor: Jian Wu

Multiple myeloma is the second most common hematologic malignancy in the United States. Eventually, all myeloma patients will relapse and develop resistance to currently available agents. There is an unmet medical need to identify novel therapeutic targets. PIM kinases play an important role in myeloma pathogenesis and disease relapse.

PIM kinase

inhibitor

myeloma

resistance

PI3K/Akt/mTOR

## 1. Introduction

Multiple myeloma (MM) is a hematologic malignancy characterized by the proliferation of malignant plasma cells. Precision medicine has heralded an era of change and challenge in the treatment of patients with MM. Although targeted therapies and immunotherapies have made significant advances in the personalized treatment of MM, clinicians still face the persistence of disease recurrence and drug resistance. The acquisition of anti-cancer drug resistance is a major issue with therapies in MM. Cancer cells utilize multiple intercellular and intracellular signaling cascades mediated by oncogenes such as PIM kinases to maintain cell growth and survival. In normal cells, the activity of these kinases is tightly controlled, whereas their sustained activation promotes apoptotic resistance and uncontrolled proliferation in cancer cells <sup>[1]</sup>. The complexity of the kinase molecular signaling network along with its crosstalk with alternative oncogenic signaling pathways provides ample opportunities for MM to develop productive adaptive mechanisms. PIM kinase activation has been shown to play a significant role in this bypass signaling mechanism. A better understanding of PIM kinase synergism, in addition to other signaling pathways, is important to the development of PIM inhibitors and to provide the rationale of combination therapy to improve the treatment efficacy for patients with MM.

## 2. Background—Expression and Regulation of PIM Kinases

The PIM kinases (PIM1, PIM2, PIM3) are a family of serine/threonine kinases and were named for their mode of discovery as proviral common integration site in Moloney murine leukemia virus (mMuLV)-induced lymphomas. PIM1 is located on chromosome 17, PIM2 on the X chromosome, and PIM3 on chromosome 15. They share high sequence homology at the amino acid level; PIM1 and PIM2 are 61% identical and PIM1 and PIM3 are 71% identical. PIM kinase genes comprise 6 exons and are transcribed into mRNA transcripts by alternative splicing. Two isoforms of PIM1 with sizes 34 and 44 kDa with comparable kinase activities are generated by translation of its mRNA from alternative initiation sites. The three isoforms of PIM2 with sizes 34, 37, and 40 kDa are similarly generated. PIM3 has one isoform. PIM1 is highly expressed in human fetal hematopoietic tissues such as the liver,

spleen, and bone marrow <sup>[2][3]</sup>. PIM2 is mainly expressed in lymphoid and brain tissues. PIM3 is overexpressed in breast, kidney, and brain tissue <sup>[4]</sup>.

In contrast to the majority of other kinases, there have been no other regulatory post-translational modifications reported for PIM kinases <sup>[5]</sup>. PIM kinases constitutively adopt an active conformation due to the presence of an acidic residue in the A-loop (Asp 200 in PIM1, Asp 196, and possibly Asp 198 in PIM2) that forms a salt bridge with basic residues of the catalytic loop (C-loop) and thus mimics the phosphorylation of the serine or threonine residue <sup>[6]</sup>. This would suggest that PIM kinases are regulated predominantly at the transcriptional and translational levels.

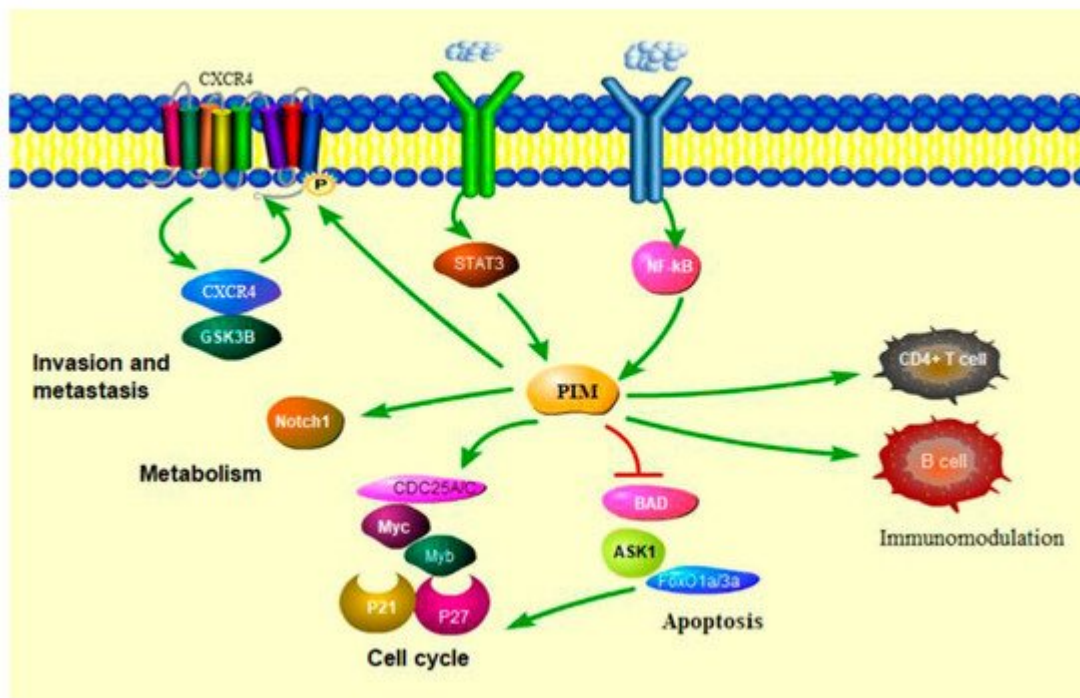
The abundance of the constitutively active PIM kinases is tightly regulated through the Janus kinase/signal transducer (JAK/STAT) pathway activator and the NF-κB pathway. STAT3 and STAT5 are known to bind to the PIM1 promoter, upregulating its transcription <sup>[7]</sup>. PIM1 binds to and activates the suppressor of cytokine signaling (SOCS) through phosphorylation to prevent activation of the JAK/STAT pathway, forming a classical negative feedback loop <sup>[8]</sup>. In addition, PIM kinases are critical downstream of ABL (Abelson) and FLT3 (FMS-related tyrosine kinase 3) oncogenes and are required in driving tumorigenesis. Constitutively active FLT3 signaling up-regulates PIM1 expression and PIM is a key contributor to FLT3-induced proliferative and anti-apoptotic pathways <sup>[9]</sup>. BCR-ABL plays an important role in mediated cell transformation through induced PIM1 expression via active STAT5.

It is noteworthy that PIM1 mRNA transcripts have a short half-life because of the presence of multiple copies of the destabilizing AUUU (A) sequence in the 3' untranslated region. In addition, PIM mRNAs contain long GC sequence-rich near the 5'UTR and hence are a "weak" transcript that requires cap-dependent translation <sup>[10]</sup>. It is also important to note that the phosphorylation of S6K and 4EBP1, substrates of mTORC1 signaling, increases after PIM2 signaling and facilitates cap-dependent translation <sup>[11]</sup>.

The stability of the transcribed PIM proteins is the key regulator of PIM activity. Its stability is largely controlled through ubiquitination and proteasomal degradation. Members of the heat shock protein family have opposite functions in stabilizing PIM activity. Binding to Hsp70 induces ubiquitylation and proteasomal degradation of PIM1, while Hsp90 protects PIM1 from proteasomal degradation <sup>[12]</sup>. On the other hand, ETK tyrosine kinase phosphorylates PIM1 at Y218, which is located in the activation loop, increasing its activity <sup>[13]</sup>. In addition, dephosphorylation of PIM kinases by the serine/threonine phosphatase, PP2A, promotes their ubiquitination and subsequent proteasomal degradation. Therefore, even though PIM kinases are constitutively active and do not depend on post-translational modifications for their activity, phosphorylation and dephosphorylation can still affect PIM's stability.

### 3. PIM Kinase and Cancers

PIM kinases are constitutively active serine/threonine kinases that are overexpressed in hematological malignancies [14]. Expression of both PIM1 and PIM2 are elevated in diffuse large B-cell lymphoma (DLBCL), while PIM2 alone is most highly expressed in B-cell chronic lymphocytic leukemia, acute myeloid leukemia (AML), and MM [15]. Increased PIM3 expression is typically observed in solid tumors [16]. Overexpression of PIM kinases is observed in MM and plays an important role in mediating survival and proliferation of MM cells, by inhibiting apoptosis and inducing cap-dependent translation, modulate immune cells, respectively [17] (Figure 1).



**Figure 1.** The regulation effect of PIM kinases in cancer. PIM kinase regulates many tumoral pathways by phosphorylating several target proteins, thereby activating or inactivating proteins involved in cell cycle progression, apoptosis, migration, or metabolism. PIM kinases promote the proliferation and survival of CD4+ T cells and B cells.

### 3.1. PIM Kinases in Cancer Cell Cycle

PIM kinases exert their oncogenic effects through the phosphorylation of different proteins that are involved in the cell cycle and proliferation. Proliferation is enhanced by PIM1 mediated phosphorylation of the cell cycle inhibitor p21waf1, leading to cytoplasmic sequestration of p21waf1 and its inability to interact with cyclin E/CDK2 in the nucleus [18]. Furthermore, PIM kinases mediate phosphorylation and inactivation of fork-head transcription factors, FOXO1a and FOXO3a, which are involved in p27kip1 transcriptional repression [19]. All three PIM kinases can phosphorylate p27 kip at Thr157 and Thr198, which allows binding to 14-3-3 protein resulting in p27 nuclear export and proteasome-dependent degradation [20]. Due to its activity in cell cycle regulation, PIM kinase plays an important role in regulating the proliferation of tumor cells.

PIM kinases also promote cell cycle progression via direct phosphorylation and activation of the CDC25A and CDC25C phosphatases, as well as inhibition of the CDC25A inhibitory kinase c-Tak1 [21]. Numerous studies have demonstrated that pharmacological inhibition of PIM induced cell cycle arrest in multiple tumor types, indicative of the overlapping activity of PIM kinases [22].

### 3.2. PIM Kinases in Cancer Cell Survival

One of the main mechanisms by which PIM kinases exert their anti-apoptotic effects is via regulation of Bcl-2 family members [23][24]. The Bcl-2 family is comprised of both pro-apoptotic proteins, such as BAD and BAX, and anti-apoptotic protein, such as Bcl-2 and Bcl-XL. PIM phosphorylates BAD at Ser112, which disrupts its association with Bcl-2 and promotes binding to 14-3-3 and retention in the cytosol. Eventually, the dissociation of BAD and Bcl-2 promotes anti-apoptotic activity. PIM is also implicated in the regulation of apoptosis via the c-Jun-N-terminal kinase (JNK) signaling pathway [1]. PIM1 directly phosphorylates Ask1 at Ser83, which decreases its ability to phosphorylate and activate its substrates JNK and p38 [25]. Other anti-apoptotic activities of PIM kinases include phosphorylation of murine double minute 2 homolog (MDM2) at serine 166 and 186 to prevent proteasomal degradation of p53 in mantle cell lymphoma [26].

### 3.3. PIM Kinases in Cancer Cell Metabolism

All three PIM kinases phosphorylate the intracellular domain of Notch1 (N1ICD) at Ser2152 and thereby stimulate the nuclear localization and transcriptional activity of N1ICD. In breast cancer cells, PIM-mediated phosphorylation of N1ICD balances cell metabolism, while its inhibition enforces glycolytic metabolism via interfering with the mitochondrial function [27]. Several studies describe a correlation between cellular glucose metabolism and tumorigenesis: to sustain energetic demands due to increased cell proliferation, cancer cells need to readjust their cellular metabolism [28]. Currently available studies have suggested that PIM1 expression is correlated with the enhanced metastatic potential of the tumor and can be predictive of tumor outcome following chemotherapy and surgery [29]. Knockout of PIM1 could lessen glucose consumption and decrease key enzymes of the glycolytic pathway in hepatocellular carcinoma [30]. These findings indicate the importance of PIM kinases in regulating cancer cell metabolism and promoting tumor progression.

### 3.4. PIM Kinases and Immune Modulation

The mechanisms by which PIM kinases modulate the immune microenvironment and regulate immune cells and the effects of PIM kinase inhibitors on immunity have not been systematically described. However, several studies have shown that PIM kinases can modulate the immune microenvironment and regulate immune cells [31][32].

PIM kinases positively regulate glycolysis in T cells and inhibition of PIM kinases leads to reduced glycolysis, increased T cell persistence, and enhanced tumor control. Moreover, PIM kinase inhibition in T cells led to higher FOXO1 activity, which translates to a T central memory phenotype (TCM, CD44+CD62L+) when compared with the control (vehicle-treated) T cells [33]. PIM1/3 inhibition prevented CD4+ T cell proliferation by inducing a G0/G1 cell cycle arrest without affecting cellular survival [34]. PIM1/PIM2 mRNAs are selectively up-or down-regulated in CD4+

cells, which subsequently affects the T cell differentiation into Th1 or Th2 cells by IL-2, IFN- $\alpha$ , and IL-4 [35]. T cell differentiation could be modulated by the upregulation of PIM1 and PIM2 expression. PIM2 induced by FOXP3 is essential for Treg cell expansion and, conversely, PIM2 also inhibits the suppressive function of Treg cells by phosphorylating FOXP3. These findings indicate the complex roles of PIM2 in the regulation of Treg cells [36]. Recently, studies have shown that PIM kinases promote survival and immune escape in primary mediastinal large-B cell lymphoma through modulation of JAK-STAT and NF- $\kappa$ B activity [37]. Given the potential role of PIM kinases in regulating tumor immunity, some cancer patients may benefit from combination strategies of PIM inhibition with checkpoint inhibitors.

## References

1. Warfel, N.A.; Kraft, A.S. PIM kinase (and Akt) biology and signaling in tumors. *Pharmacol. Ther.* 2015, 151, 41–49.
2. An, N.; Lin, Y.W.; Mahajan, S.; Kellner, J.N.; Wang, Y.; Li, Z.; Kraft, A.S.; Kang, Y. Pim1 serine/threonine kinase regulates the number and functions of murine hematopoietic stem cells. *Stem Cells* 2013, 31, 1202–1212.
3. An, N.; Kraft, A.S.; Kang, Y. Abnormal hematopoietic phenotypes in Pim kinase triple knockout mice. *J. Hematol. Oncol.* 2013, 6, 12.
4. Le, X.; Antony, R.; Razavi, P.; Treacy, D.J.; Luo, F.; Ghandi, M.; Castel, P.; Scaltriti, M.; Baselga, J.; Garraway, L.A. Systematic Functional Characterization of Resistance to PI3K Inhibition in Breast Cancer. *Cancer Discov.* 2016, 6, 1134–1147.
5. Iyer, R.S.; Chatham, L.; Sleight, R.; Meek, D.W. A functional SUMO-motif in the active site of PIM1 promotes its degradation via RNF4, and stimulates protein kinase activity. *Sci. Rep.* 2017, 7, 3598.
6. Adam, K.; Lambert, M.; Lestang, E.; Champenois, G.; Dusanter-Fourt, I.; Tamburini, J.; Bouscary, D.; Lacombe, C.; Zermati, Y.; Mayeux, P. Control of Pim2 kinase stability and expression in transformed human haematopoietic cells. *Biosci. Rep.* 2015, 35, e00274.
7. Mologni, L.; Magistroni, V.; Casuscelli, F.; Montemartini, M.; Gambacorti-Passerini, C. The Novel PIM1 Inhibitor NMS-P645 Reverses PIM1-Dependent Effects on TMPRSS2/ERG Positive Prostate Cancer Cells And Shows Anti-Proliferative Activity in Combination with PI3K Inhibition. *J. Cancer* 2017, 8, 140–145.
8. Hildebrand, D.; Heeg, K.; Kubatzky, K.F. *Pasteurella multocida* Toxin Manipulates T Cell Differentiation. *Front. Microbiol.* 2015, 6, 1273.
9. Darici, S.; Alkhaldi, H.; Horne, G.; Jorgensen, H.G.; Marmioli, S.; Huang, X. Targeting PI3K/Akt/mTOR in AML: Rationale and Clinical Evidence. *J. Clin. Med.* 2020, 9, 2934.

10. Hoover, D.S.; Wingett, D.G.; Zhang, J.; Reeves, R.; Magnuson, N.S. Pim-1 protein expression is regulated by its 5'-untranslated region and translation initiation factor eIF-4E. *Cell Growth Differ.* 1997, 8, 1371–1380.
11. Keane, N.A.; Reidy, M.; Natoni, A.; Raab, M.S.; O'Dwyer, M. Targeting the Pim kinases in multiple myeloma. *Blood Cancer J.* 2015, 5, e325.
12. Leung, C.O.; Wong, C.C.; Fan, D.N.; Kai, A.K.; Tung, E.K.; Xu, I.M.; Ng, I.O.; Lo, R.C. PIM1 regulates glycolysis and promotes tumor progression in hepatocellular carcinoma. *Oncotarget* 2015, 6, 10880–10892.
13. Kim, O.; Jiang, T.; Xie, Y.; Guo, Z.; Chen, H.; Qiu, Y. Synergism of cytoplasmic kinases in IL6-induced ligand-independent activation of androgen receptor in prostate cancer cells. *Oncogene* 2004, 23, 1838–1844.
14. Cervantes-Gomez, F.; Chen, L.S.; Orlowski, R.Z.; Gandhi, V. Biological effects of the Pim kinase inhibitor, SGI-1776, in multiple myeloma. *Clin. Lymphoma Myeloma Leuk.* 2013, 13, S317–S329.
15. Koblish, H.; Li, Y.L.; Shin, N.; Hall, L.; Wang, Q.; Wang, K.; Covington, M.; Marando, C.; Bowman, K.; Boer, J.; et al. Preclinical characterization of INCB053914, a novel pan-PIM kinase inhibitor, alone and in combination with anticancer agents, in models of hematologic malignancies. *PLoS ONE* 2018, 13, e0199108.
16. Motylewska, E.; Braun, M.; Stepień, H. High Expression of NEK2 and PIM1, but Not PIM3, Is Linked to an Aggressive Phenotype of Bronchopulmonary Neuroendocrine Neoplasms. *Endocr. Pathol.* 2020, 31, 264–273.
17. Saurabh, K.; Scherzer, M.T.; Shah, P.P.; Mims, A.S.; Lockwood, W.W.; Kraft, A.S.; Beverly, L.J. The PIM family of oncoproteins: Small kinases with huge implications in myeloid leukemogenesis and as therapeutic targets. *Oncotarget* 2014, 5, 8503–8514.
18. Cottage, C.T.; Bailey, B.; Fischer, K.M.; Avitabile, D.; Collins, B.; Tuck, S.; Quijada, P.; Gude, N.; Alvarez, R.; Muraski, J.; et al. Cardiac progenitor cell cycling stimulated by pim-1 kinase. *Circ. Res.* 2010, 106, 891–901.
19. Mondello, P.; Cuzzocrea, S.; Mian, M. Pim kinases in hematological malignancies: Where are we now and where are we going? *J. Hematol. Oncol.* 2014, 7, 95.
20. Chen, L.S.; Balakrishnan, K.; Gandhi, V. Inflammation and survival pathways: Chronic lymphocytic leukemia as a model system. *Biochem. Pharmacol.* 2010, 80, 1936–1945.
21. Bachmann, M.; Hennemann, H.; Xing, P.X.; Hoffmann, I.; Moroy, T. The oncogenic serine/threonine kinase Pim-1 phosphorylates and inhibits the activity of Cdc25C-associated kinase 1 (C-TAK1): A novel role for Pim-1 at the G2/M cell cycle checkpoint. *J. Biol. Chem.* 2004, 279, 48319–48328.

22. Mumenthaler, S.M.; Ng, P.Y.; Hodge, A.; Bearss, D.; Berk, G.; Kanekal, S.; Redkar, S.; Taverna, P.; Agus, D.B.; Jain, A. Pharmacologic inhibition of Pim kinases alters prostate cancer cell growth and resensitizes chemoresistant cells to taxanes. *Mol. Cancer Ther.* 2009, 8, 2882–2893.
23. Aho, T.L.; Sandholm, J.; Peltola, K.J.; Mankonen, H.P.; Lilly, M.; Koskinen, P.J. Pim-1 kinase promotes inactivation of the pro-apoptotic Bad protein by phosphorylating it on the Ser112 gatekeeper site. *FEBS Lett.* 2004, 571, 43–49.
24. Yan, B.; Zemskova, M.; Holder, S.; Chin, V.; Kraft, A.; Koskinen, P.J.; Lilly, M. The PIM-2 kinase phosphorylates BAD on serine 112 and reverses BAD-induced cell death. *J. Biol. Chem.* 2003, 278, 45358–45367.
25. Gu, J.J.; Wang, Z.; Reeves, R.; Magnuson, N.S. PIM1 phosphorylates and negatively regulates ASK1-mediated apoptosis. *Oncogene* 2009, 28, 4261–4271.
26. Hogan, C.; Hutchison, C.; Marcar, L.; Milne, D.; Saville, M.; Goodlad, J.; Kernohan, N.; Meek, D. Elevated levels of oncogenic protein kinase Pim-1 induce the p53 pathway in cultured cells and correlate with increased Mdm2 in mantle cell lymphoma. *J. Biol. Chem.* 2008, 283, 18012–18023.
27. Santio, N.M.; Landor, S.K.; Vahtera, L.; Yla-Pelto, J.; Paloniemi, E.; Imanishi, S.Y.; Corthals, G.; Varjosalo, M.; Manoharan, G.B.; Uri, A.; et al. Phosphorylation of Notch1 by Pim kinases promotes oncogenic signaling in breast and prostate cancer cells. *Oncotarget* 2016, 7, 43220–43238.
28. Carafa, V.; Altucci, L.; Nebbioso, A. Dual Tumor Suppressor and Tumor Promoter Action of Sirtuins in Determining Malignant Phenotype. *Front. Pharmacol.* 2019, 10, 38.
29. Guo, S.; Mao, X.; Chen, J.; Huang, B.; Jin, C.; Xu, Z.; Qiu, S. Overexpression of Pim-1 in bladder cancer. *J. Exp. Clin. Cancer Res.* 2010, 29, 161.
30. Xue, C.; He, Y.; Hu, Q.; Yu, Y.; Chen, X.; Chen, J.; Ren, F.; Li, J.; Ren, Z.; Cui, G.; et al. Downregulation of PIM1 regulates glycolysis and suppresses tumor progression in gallbladder cancer. *Cancer Manag. Res.* 2018, 10, 5101–5112.
31. Daenthanasamak, A.; Wu, Y.; Iamsawat, S.; Nguyen, H.D.; Bastian, D.; Zhang, M.; Sofi, M.H.; Chatterjee, S.; Hill, E.G.; Mehrotra, S.; et al. PIM-2 protein kinase negatively regulates T cell responses in transplantation and tumor immunity. *J. Clin. Investig.* 2018, 128, 2787–2801.
32. Knudson, K.M.; Pritzi, C.J.; Saxena, V.; Altman, A.; Daniels, M.A.; Teixeira, E. NFκB-Pim-1-Eomesodermin axis is critical for maintaining CD8 T-cell memory quality. *Proc. Natl. Acad. Sci. USA* 2017, 114, E1659–E1667.
33. Chatterjee, S.; Chakraborty, P.; Daenthanasamak, A.; Iamsawat, S.; Andrejeva, G.; Luevano, L.A.; Wolf, M.; Baliga, U.; Krieg, C.; Beeson, C.C.; et al. Targeting PIM Kinase with PD1 Inhibition Improves Immunotherapeutic Antitumor T-cell Response. *Clin. Cancer Res.* 2019, 25, 1036–1049.

34. Jackson, L.J.; Pheneger, J.A.; Pheneger, T.J.; Davis, G.; Wright, A.D.; Robinson, J.E.; Allen, S.; Munson, M.C.; Carter, L.L. The role of PIM kinases in human and mouse CD4+ T cell activation and inflammatory bowel disease. *Cell Immunol.* 2012, 272, 200–213.
35. Aho, T.L.; Lund, R.J.; Ylikoski, E.K.; Matikainen, S.; Lahesmaa, R.; Koskinen, P.J. Expression of human pim family genes is selectively up-regulated by cytokines promoting T helper type 1, but not T helper type 2, cell differentiation. *Immunology* 2005, 116, 82–88.
36. Du, W.; Chen, T.; Ni, Y.; Hou, X.; Yu, Y.; Zhou, Q.; Wu, F.; Tang, W.; Shi, G. Role of PIM2 in allergic asthma. *Mol. Med. Rep.* 2017, 16, 7504–7512.
37. Szydlowski, M.; Debek, S.; Prochorec-Sobieszek, M.; Szolkowska, M.; Tomirotti, A.M.; Juszczyński, P.; Szumera-Cieckiewicz, A. PIM Kinases Promote Survival and Immune Escape in Primary Mediastinal Large B-Cell Lymphoma through Modulation of JAK-STAT and NF-kappaB Activity. *Am. J. Pathol.* 2021, 191, 567–574.

---

Retrieved from <https://encyclopedia.pub/entry/history/show/33671>