Anthocyanins

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Anthocyanins are biologically active water-soluble plant pigments that are responsible for blue, purple, and red colors in various plant parts—especially in fruits and blooms. Anthocyanins have attracted attention as natural food colorants to be used in yogurts, juices, marmalades, and bakery products. Numerous studies have also indicated the beneficial health effects of anthocyanins and their metabolites on humans, including free-radical scavenging and antioxidant activity.

Keywords: Anthocyanin metabolites ; antioxidants ; cardioprotection ; hepatoprotection ; nephroprotection ; neuroprotection

1. Introduction

Anthocyanins are pigments belonging to the flavonoid group, which is widely distributed in plants. They are responsible for blue, purple, and red colors in flowers, fruits, and vegetables and protect plants from environmental stresses such as high sunlight irradiance ^[1] or low nitrogen ^[2]. Chemically, anthocyanins are produced when anthocyanidins are glycosylated. The most abundant anthocyanidin in plants is cyanidin. Other anthocyanidins are less abundant, and their frequency decreases in this order: delphinidin, peonidin, pelargonidin, petunidin, and malvidin^[3]. Anthocyanidins are flavylium ion derivatives that vary in terms of their substituent groups: -H, -OH, or -OCH₃. Usually, anthocyanidins are glycosylated at the C3 or C3 and C5 sites, but the glycosylation of other sites has also been reported [4]. The biological activity of anthocyanins depends on their structure; however, all samples, including those with different compositions and amounts of anthocyanins, extracted from various berries and vegetables, are biologically active ^[5]. Azevedo et al. ^[6] established that the radical scavenging activity and reducing properties of anthocyanins strongly depend on the chemical structures of particular anthocyanins; this effect increases with the presence of catechol and pyrogallol groups in ring B of cyanidin-3glucosides and the respective aglycones. Some studies have shown that delphinidin has the highest antioxidant activity compared with the other five anthocyanidins due to the three hydroxyl groups on the B-ring [5][2]. An increasing body of evidence shows that anthocyanin intake can have a protective effect on human and animal brains, hearts, livers, and kidneys, and many of the therapeutic effects may be purported to the antioxidant activities of anthocyanins and their metabolites ^{[8][9][10][11][12]}. The antioxidant activity of these compounds manifests through direct and indirect methods of action. Thus, anthocyanins can directly scavenge reactive oxygen species (ROS) [13][14], whereas the indirect pathways involve stimulation of the synthesis or activity of antioxidant enzymes (catalase, superoxide dismutase (SOD), glutathione peroxidase) [15]; inhibition of ROS-forming enzymes, such as nicotinamide adenine dinucleotide phosphate (NADPH) oxidase and others ^{[16][17]}; or even mild uncoupling of mitochondrial respiration preventing ROS generation ^{[9][18]}. It can also be assumed that for effective therapeutic action of anthocyanins, both the ROS scavenging activity and the modulation of cellular antioxidant systems are required $\frac{[14]}{}$.

2. Natural Sources of Anthocyanins

Anthocyanins are natural, water-soluble plant pigments that are responsible for blue, red, or purple colors in plants. Plant genotypes, agro-climatic conditions, and fruit or vegetable maturity are significant factors in the composition and quantity of anthocyanins ^[19]. Therefore, the main sources of anthocyanins in human diet are fruits and vegetables, which accumulate anthocyanins in both the peel and flesh; however, their content varies greatly (<u>Table 1</u>).

Table 1. Maximum amount of anthocyanins (mg 100 g^{-1} of fresh weight (FW)) in fruits and vegetables.

Source	Anthocyanin amount	Dominant Anthocyanins	References
	mg 100 g ⁻¹ FW		

Bilberry	772.4	Dp3gal, Dp3glc, Dp3ara, Mv3glc, Cy3gal, Cy3glc, Cy3ara	[20]
Blackcurrant	478.6	Dp3rut, Cy3rut, Dp3glc, Cy3glc	
Golden currant	615.5	Cy3rut, Cy3glc, Pn3rut	[21][22]
Redcurrant	66.7	Cy3glc, Cy3rut, Cy3sam	
Elderberry	580.0	Cy3sam, Cy3glc	[<u>19]</u>
Grapes	116.4	Mv3glc, Cy3glc, Dp3glc, Pt3glc, Pn3glc	[23]
Sour cherry	147.0	Cy3rut	[24]
Sweet cherry	244.0	Cy3rut, Pn3rut	[25]
Wild strawberry	10.0	Pg3glc, Cy3glc	[26]
Black carrot	126.4	Cy3xylglcgal, Cy3xylgal	[27]
Eggplant	8.7	Dp3glc, Dp3rut,	[<u>28][29]</u>
Red cabbage	23.4	Cy3glc, Cy3rut, Dp3glc, Dp3rut, Cy3diglc5glc	[<u>30][31]</u>
Red chicory	39.2	Cy3glc	[<u>29]</u>
Purple wheat	23.5	Cy3glc	[<u>32]</u>

Cy—cyanidin; Dp—delphinidin; Mv—malvidin; Pg—pelargonidin; Pn—peonidin; Pt—petunidin; ara—arabinoside; gal—galactoside; glc—glucoside; rut—rutinoside; sam—sambubioside; xyl—xyloside.

3. Stability of Anthocyanins in Foods and Beverage

Anthocyanins are natural plant compounds that are increasingly being used in the food and pharmaceutical industry due to their effects on human health. However, the low stability of anthocyanins is still an obstacle to their use ^[33]. The stability of anthocyanins depends on the pH, temperature, light, presence of solvents and oxygen, and other factors ^[34]. Anthocyanins are more stable under acidic conditions. At a pH of 1.0, flavylium cations are the predominant species and contribute to the development of purple and red pigments, while at pH 2.0–4.0, the blue quinoidal species predominates. When the pH value reaches 7.0, anthocyanins are usually degraded ^[34]. The storage temperature affects the concentration of anthocyanins in extracts; for instance, 11% of rosella anthocyanins were lost after storage for 60 days at 4 °C, while 99% of anthocyanins were degraded in the same extracts stored at 37 °C for the same period ^[35].

The stability of anthocyanins from various Ribes species was reported to depend on their composition and storage conditions ^[31]. Anthocyanins in redcurrant berry extract have been shown to be more stable at room temperature and in the presence of light than extracts from berries of golden currant and gooseberry. After the storage of anthocyanin extracts under dark and cold conditions (+4 °C) for 84 days, up to 90% of redcurrant, 80% of gooseberry and golden currant, and up to 50% of blackcurrant anthocyanins remained intact ^[31].

Thermal food processing negatively affects the nutritional value of anthocyanin-rich juices as it results in anthocyanin degradation ^[36]. Thermal processing is responsible for the loss of up to 35% of anthocyanins. Even short-term thermal treatment (5 s at 85 °C) resulted in a loss of 9% of anthocyanins from strawberry juice, while pasteurization for 15 min

resulted in a loss of 21% ^[37]. Boiling of red cabbage resulted in a loss of 41.2% of anthocyanins, while the anthocyanin concentration remained the same after steaming or stir-frying. Possibly, as highly water-soluble pigments, anthocyanins may be lost by leaching in the case of boiling ^[5].

Various anthocyanin stabilization methods are being developed. For instance, the improvement of anthocyanin thermal stability by yeast mannoproteins at pH 7.0 has been studied. The complexes were found to effectively protect anthocyanins from degradation during heating at 80 and 126 °C ^[38]. Mixing of clarified acerola juice with montmorillonite resulted in 50% more anthocyanins, regardless of time or pH changes ^[39]. Copigmentation is another natural tool that can be used to enhance anthocyanin stability. The most studied copigments are phenolic acids such as hydroxycinnamic and hydroxybenzoic acids ^[40]. Babaloo and Jamei ^[41] established that caffeic acid provides more stability for anthocyanins than benzoic, tannic, and coumaric acids. Encapsulation with polysaccharides, such as β -cyclodextrin, maltodextrin, or Arabic gum, is also important for the stabilization of anthocyanins. The protective effect of β -cyclodextrin was evident for all blackberry anthocyanins after thermal treatment at 90 °C for 2 h ^[42].

Novel techniques for phenolic compound isolation from natural products that avoid the degradation of these compounds have been developed. Block freeze concentration has been employed to extract anthocyanins from strawberries and enrich yogurt with the obtained concentrated strawberry juice. As a result, yogurt with a high anthocyanin content and greater antioxidant activity was produced; however, it had a short shelf life ^[43]. The foam mat drying technique is now considered to be an effective dehydration method to produce powder from juices or pulp. Only a small reduction in anthocyanins (7–9%) was observed after the storage of jambolana juice powder produced by foam mat drying for 150 days ^[44]. When foam-mat freeze-drying was used for powder production from blueberry juice, 80–100% of anthocyanins remained after processing; the most stable was Cy3glc ^[45].

Biological Effects Related to the Antioxidant Activity of Anthocyanins— In Vivo Studies in Humans

Epidemiological and clinical studies suggest that an anthocyanin-enriched diet may lower levels of certain oxidative stress biomarkers in humans, and this could be associated with reduced risk of cognitive decline and the development of neurodegenerative and cardiovascular diseases, as well as 358 having sustained hepatic function and kidney protecting activities [19][46][47][48][49][50][51][52][53][54][55][56][57].

A randomized clinical trial ^[58] evaluated the effects of a standardized maqui berry (Aristotelia chilensis (Mol.) Stuntz) extract (containing 162 mg of anthocyanins) on products of lipid peroxidation in healthy, overweight, and smoker adults. The results suggested that supplementation with the extract can be related to a limited term (max for 40 days) reduction in oxidized low-density lipoprotein (LDL) levels and a decrease in urinary F2-isoprostanes. Another study ^[59] concluded that the acute consumption of anthocyanin-rich red Vitis labrusca L. grape juices could be related to decreased levels of thiobarbituric acid reactive substances and lipid peroxides in the serum of healthy subjects. It has also been demonstrated that regular (for 30 days) anthocyanin-rich sour cherry consumption could suppress the formation of ROS by circulating phagocytes and decrease the risk of systemic imbalance between oxidants and antioxidants ^[60]. It is worth noting that a portion (300 g) of blueberries, the dietary source of anthocyanins provided to young volunteers involved in a randomized cross-over study, significantly reduced H₂O₂-induced DNA damage in blood mononuclear cells ^[61]. In another human pilot intervention study, the consumption of anthocyanin-rich bilberry (Vaccinium myrtillius L.) pomace extract was found to modulate transcription factor E2-related factor 2 (Nrf2)-dependent gene expression in peripheral blood mononuclear cells ^[62].

A single-blind randomized placebo-controlled intervention trial, which lasted for 8 weeks and involved 72 unmedicated subjects, revealed that the administration of various berries (including bilberries, chokeberries, and blackcurrants) increased both the concentration of high-density lipoprotein (HDL) cholesterol and the plasma antioxidant capacity ^[63]. Higher dietary anthocyanin and flavan-3-ol intake was associated with anti-inflammatory effects in 2375 Framingham Heart Study Offspring Cohort participants ^[64]. Interestingly, the consumption of 300 mL of red wine (a total dose of anthocyanins was 304 μ M, which was the highest amount among detected compounds) with a meal was shown to prevent the postprandial increases in plasma lipid hydroperoxides and cholesterol oxidation products and therefore protect against a potential pro-atherosclerogenic effect ^[65]. Similar findings were obtained in a randomized cross-over trial, in which it was concluded that moderate consumption of red wine decreases erythrocyte SOD activity ^[66]. In another randomized double-blind trial, 150 subjects with hypercholesterolemia consumed a purified anthocyanin mixture derived from bilberries and blackcurrants (320 mg/d) for 24 weeks ^[67]. It was found that anthocyanin consumption significantly decreased the levels of inflammatory biomarkers (C-reactive protein, soluble vascular cell adhesion molecule-1 and

plasma IL-1 β) and increased the HDL cholesterol level. Recently, it was shown that a daily intake of 150 g of anthocyaninrich blueberries resulted in clinically relevant improvements in endothelial function and systemic arterial stiffness; probably due to the improved nitric oxide bioactivity and HDL status ^[68].

Nonalcoholic fatty liver disease (NAFLD), defined by excessive lipid accumulation in the liver, is the hepatic manifestation of insulin resistance and metabolic syndrome. NAFLD encompasses a wide spectrum of liver diseases ranging from simple uncomplicated steatosis, to steatohepatitis, cirrhosis, and hepatocellular carcinoma ^[48]. Zhang et al. ^[69] reported that anthocyanins extracted from bilberry and blackcurrant (320 mg/day) and administered for 12 weeks ameliorated liver injury in patients with NAFLD. It was observed that a so called "anthocyanin group" exhibited significant decreases in the plasma alanine aminotransferase, cytokeratin-18 M30 fragment, and myeloperoxidase levels. It was also found that consumption of Myrica rubra Sieb. and Zucc. juice (250 mL for 4 weeks) protected young adults (18–25 years old) against NAFLD by improving the plasma antioxidant status and inhibiting the inflammatory and apoptotic responses involved in this disease ^[70].

In another study, red fruit juice (40% red grape juice, 20% blackberry juice, 15% sour cherry juice, 15% blackcurrant juice, and 10% elderberry juice) with high polyphenol and anthocyanin contents was tested for its preventive potential in hemodialysis patients ^[71]. For this purpose, 21 subjects consumed 200 mL/day of juice according to the following protocols: 3-week run-in; 4-week juice 411 uptake; 3-week wash-out. The results revealed a significant decrease in DNA oxidation damage and protein and lipid peroxidation and an increase in the reduced glutathione level; the effects were attributed to the high anthocyanin and polyphenol contents of the juice ^[71]. Another study ^[72] demonstrated that regular consumption of concentrated red grape juice by hemodialysis patients could be associated with the reduced neutrophil NADPH-oxidase activity and plasma concentrations of oxidized LDL and inflammatory biomarkers.

5. Conclution

In conclusion, anthocyanins are valuable biomolecules with a broad variety of biological effects on human health, and we suggest adding more anthocyanin rich fruits, vegetables, and their products to the daily diet. Numerous studies indicate that bilberry, blackcurrant, elderberry, and other berries have the highest total concentrations of anthocyanins; therefore, the consumption of fresh berries and their processed products may have greater beneficial effects on humans. Various anthocyanin stabilization methods have been developed e.g., copigmentation with other phenolic acids, encapsulation with polysaccharides, block freeze concentration, and powder production using the foam mat drying technique. All of them enable anthocyanins to be preserved during processing, thus increasing their bioavailability and delivery to target tissues in the human body. However, long-term studies on the impacts of anthocyanin-rich product consumption on human health are still rare. Further studies could focus on the identification and tracking of individual anthocyanin metabolites and on the determination of the exact dosage and delivery platforms sustaining the antioxidant properties of anthocyanins in vivo. In addition, as an alternative to natural sources, synthesis of the most bioactive anthocyanins in bioreactors should be considered.

References

- 1. Landi, M.; Tattini, M.; Gould, K.S. Multiple functional roles of anthocyanins in plant-environment interactions. Environ. Exp. Bot. 2015, 119, 4–17.
- 2. Liang, J.; He, J. Protective role of anthocyanins in plants under low nitrogen stress. Biochem. Biophys. Res. Commun. 2018, 498, 946–953.
- 3. Cody, R.B.; Tamura, J.; Downard, K.M. Quantitation of anthocyanins in elderberry fruit extracts and nutraceutical formulations with paper spray ionization mass spectrometry. J. Mass Spectrom. 2018, 53, 58–64.
- 4. Zhao, C.L.; Chen, Z.J.; Bai, X.S.; Ding, C.;•Long, T.J.; Wei, F.G.; Miao, K.R. Structure–activity relationships of anthocyanidin glycosylation. Mol. Divers. 2014, 18, 687–700.
- Blando, F.; Calabriso, N.; Berland, H.; Maiorano, G.; Gerardi, C.; Carluccio, M.A.; Andersen, Ø.M. Radical Scavenging and Anti-Inflammatory Activities of Representative Anthocyanin Groupings from Pigment-Rich Fruits and Vegetables. Int. J. Mol. Sci. 2018, 19, 169.
- Azevedo, J.; Fernandes, I.; Faria, A.; Oliveira, J.; Fernandes, A.; de Freitas, V.; Mateus, N. Antioxidant properties of anthocyanidins, anthocyanidins 3-glucosides and respective portisins. Food Chem. 2010, 119, 518–523.
- 7. Ali, M.H.; Almagribi, W.; Al-Rashidi, M.N. Antiradical and reductant activities of anthocyanidins and anthocyanins, structure-activity relationship and synthesis. Food Chem. 2016, 194, 1275–1282.

- 8. Pojer, E.; Mattivi, F.; Johnson, D.; Stockley, C.S. The Case for Anthocyanin Consumption to Promote Human Health: A Review. Compr. Rev. Food Sci. Food Saf. 2013, 12, 483–508.
- Liobikas, J.; Skemiene, K.; Trumbeckaite, S.; Borutaite, V. Anthocyanins in Cardioprotection: A Path Through Mitochondria. Pharmacol. Res. 2016, 113, 808–815.
- 10. Kalt, W. Anthocyanins and Their C6-C3-C6 Metabolites in Humans and Animals. Molecules 2019, 24, 4024.
- 11. Bendokas, V.; Skemiene, K.; Trumbeckaite, S.; Stanys, V.; Passamonti, S.; Borutaite, V.; Liobikas, J. Anthocyanins: From Plant Pigments to Health Benefits at Mitochondrial Level. Crit. Rev. Food Sci. Nutr. 2019, in press.
- 12. Kalt, W.; Cassidy, A.; Howard, L.R.; Krikorian, R.; Stull, A.J.; Tremblay, F.; Zamora-Ros, R. Recent Research on the Health Benefits of Blueberries and Their Anthocyanins. Adv. Nutr. 2020, 11, 224–236.
- 13. Miguel, M.G. Anthocyanins: Antioxidant and/or anti-inflammatory activities. J. Appl. Pharm. Sci. 2011, 1, 07–15.
- Ereminas, G.; Majiene, D.; Sidlauskas, K.; Jakstas, V.; Ivanauskas, L.; Vaitiekaitis, G.; Liobikas, J. Neuroprotective Properties of Anthocyanidin Glycosides Against H2O2-induced Glial Cell Death Are Modulated by Their Different Stability and Antioxidant Activity in Vitro. Biomed. Pharmacother. 2017, 94, 188–196.
- 15. Aboonabi, A.; Singh, I. Chemopreventive Role of Anthocyanins in Atherosclerosis via Activation of Nrf2-ARE as an Indicator and Modulator of Redox. Biomed. Pharmacother. 2015, 72, 30–36.
- 16. Lim, T.G.; Jung, S.K.; Kim, J.; Kim, Y.; Lee, H.J.; Jang, T.S.; Lee, K.W. NADPH Oxidase Is a Novel Target of Delphinidin for the Inhibition of UVB-induced MMP-1 Expression in Human Dermal Fibroblasts. Exp. Dermatol. 2013, 22, 428–430.
- 17. Reis, J.F.; Monteiro, V.V.S.; de Souza Gomes, R.; do Carmo, M.M.; da Costa, G.V.; Ribera, P.C.; Monteiro, M.Ch. Action Mechanism and Cardiovascular Effect of Anthocyanins: A Systematic Review of Animal and Human Studies. J. Transl. Med. 2016, 14, 315.
- Skemiene, K.; Rakauskaite, G.; Trumbeckaite, S.; Liobikas, J.; Brown, G.C.; Borutaite, V. Anthocyanins block ischemiainduced apoptosis in the perfused heart and support mitochondrial respiration potentially by reducing cytosolic cytochrome c. Int. J. Biochem. Cell Biol. 2013, 45, 23–29.
- 19. Mikulic-Petkovsek, M.; Schmitzer, V.; Slatnar, A.; Todorovic, B.; Veberic, R.; Stampar, F.; Ivancic, A. Investigation of anthocyanin profile of four elderberry species and interspecific hybrids. J. Agric. Food Chem. 2014, 62, 5573–5580.
- 20. Veberic, R.; Slatnar, A.; Bizjak, J.; Stampar, F.; Mikulic-Petkovsek, M. Anthocyanin composition of different wild and cultivated berry species. LWT 2015, 60, 509–517.
- 21. Stanys, V.; Bendokas, V.; Rugienius, R.; Sasnauskas, A.; Frercks, B.; Mažeikienė, I.; Šikšnianas, T. Management of anthocyanin amount and composition in genus Ribes using interspecific hybridisation. Sci. Hortic. 2019, 247, 123–129.
- 22. Siksnianas, T.; Bendokas, V.; Rugienius, R.; Sasnauskas, A.; Stepulaitiene, I.; Stanys, V. Anthocyanin content and stability in Ribes species and interspecific hybrids. Rural Dev. 2013, 6, 258–261.
- 23. Dimitrovska, M.; Bocevska, M.; Dimitrovski, D.; Murkovic, M. Anthocyanin composition of Vranec, Cabernet Sauvignon, Merlot and Pinot Noir grapes as indicator of their varietal differentiation. Eur. Food Res. Technol. 2011, 32, 591–600.
- 24. Bendokas, V.; Stepulaitiene, I.; Stanys, V.; Siksnianas, T.; Anisimoviene, N. Content of anthocyanin and other phenolic compounds in cherry species and interspecific hybrids. Acta Hortic. 2017, 1161, 587–592.
- 25. Blackhall, M.L.; Berry, R.; Davies, N.W.; Wallsa, J.T. Optimized extraction of anthocyanins from Reid Fruits' Prunus avium 'Lapins' cherries. Food Chem. 2018, 256, 280–285.
- Rugienius, R.; Bendokas, V.; Kazlauskaitė, E.; Siksnianas, T.; Stanys, V.; Kazanaviciute, V.; Sasnauskas, A. Anthocyanin content in cultivated Fragaria vesca berries under high temperature and water deficit stress. Acta Hortic. 2016, 1139, 639–644.
- Algarra, M.; Fernandes, A.; Mateus, N.; de Freitas, V.; da Silva, J.C.G.; Casado, E.J. Anthocyanin profile and antioxidant capacity of black carrots (Daucus carota L. ssp. sativus var. atrorubens Alef.) from Cuevas Bajas, Spain. J. Food Compost. Anal. 2014, 33, 71–76.
- 28. Liu, Y.; Tikunov, Y.; Schouten, R.E.; Marcelis, L.F.M.; Visser, R.G.F.; Bovy, A. Anthocyanin Biosynthesis and Degradation Mechanisms in Solanaceous Vegetables: A Review. Front. Chem. 2018, 6, 52.
- Frond, A.D.; Iuhas, C.I.; Stirbu, I.; Leopold, L.; Socaci, S.; Andreea, S.; Ayvaz, H.; Andreea, S.; Mihai, S.; Diaconeasa, Z.; et al. Phytochemical Characterization of Five Edible Purple-Reddish Vegetables: Anthocyanins, Flavonoids, and Phenolic Acid Derivatives. Molecules 2019, 24, 1536.
- Murador, D.C.; Mercadante, A.Z.; de Rosso, V.V. Cooking techniques improve the levels of bioactive compounds and antioxidant activity in kale and red cabbage. Food Chem. 2016, 196, 1101–1107.

- 31. Tong, T.; Niu, Y.H.; Yue, Y.; Wu, S.-C.; Ding, H. Beneficial effects of anthocyanins from red cabbage (Brassica oleracea L. var. capitata L.) administration to prevent irinotecaninduced mucositis. J. Funct. Foods 2017, 32, 9–17.
- 32. Gamel, T.H.; Wright, A.J.; Tucker, A.J.; Pickard, M.; Rabalski, I.; Podgorski, M.; Di Ilio, N.; O'Brien, C.; Abdel-Aal, E.M. Absorption and metabolites of anthocyanins and phenolic acids after consumption of purple wheat crackers and bars by healthy adults. J. Cereal. Sci. 2019, 86, 60–68.
- Tan, C.; Selig, M.J.; Lee, M.C.; Abbaspourrad, A. Polyelectrolyte microcapsules built on CaCO3 scaffolds for the integration, encapsulation, and controlled release of copigmented anthocyanins. Food Chem. 2018, 246, 305–312.
- 34. Castañeda-Ovando, A.; Pacheco-Hernández, M.L.; Páez-Hernández, M.E.; Rodríguez, J.A.; Galán-Vidal, C.A. Chemical studies of anthocyanins: A review. Food Chem. 2009, 113, 859–871.
- 35. Sinela, A.; Rawat, N.; Mertz, C.; Achir, N.; Fulcrand, H.; Dornier, M. Anthocyanins degradation during storage of Hibiscus sabdariffa extract and evolution of its degradation products. Food Chem. 2017, 214, 234–241.
- 36. Wang, Z.; Zhang, M.; Wu, Q. Effects of temperature, pH, and sunlight exposure on the color stability of strawberry juice during processing and storage. LWT 2015, 60, 1174–1178.
- 37. Weber, F.; Larsen, L.R. Influence of fruit juice processing on anthocyanin stability. Food Res. Int. 2017, 100, 354–365.
- 38. Wu, J.; Guan, Y.; Zhong, Q. Yeast mannoproteins improve thermal stability of anthocyanins at pH 7.0. Food Chem. 2015, 172, 121–128.
- 39. Ribeiro, H.L.; de Oliveira, A.V.; de Brito, E.S.; Ribeiro, P.R.V.; Filho, M.M.S.; Azeredo, H.M.C. Stabilizing effect of montmorillonite on acerola juice anthocyanins. Food Chem. 2018, 245, 966–973.
- 40. Bimpilas, A.; Panagopoulou, M.; Tsimogiannis, D.; Oreopoulou, V. Anthocyanin copigmentation and color of wine: The effect of naturally obtained hydroxycinnamic acids as cofactors. Food Chem. 2016, 197, 39–46.
- 41. Babaloo, F.; Jamei, R. Anthocyanin pigment stability of Cornus mas–Macrocarpa under treatment with pH and some organic acids. Food Sci. Nutr. 2018, 6, 168–173.
- Fernandes, A.; Rocha, M.A.A.; Santos, L.M.N.B.F.; Brás, J.; Oliveira, J.; Mateus, N.; de Freitas, V. Blackberry anthocyanins: β-Cyclodextrin fortification for thermal and gastrointestinal stabilization. Food Chem. 2018, 245, 426– 431.
- Jaster, H.; Arend, G.D.; Rezzadori, K.; Chaves, V.C.; Reginatto, F.H.; Petrus, J.C.C. Enhancement of antioxidant activity and physicochemical properties of yogurt enriched with concentrated strawberry pulp obtained by block freeze concentration. Food Res. Int. 2018, 104, 119–125.
- Tavares, I.M.deC.; Sumere, B.R.; Gómez-Alonso, S.; Gomes, E.; Hermosín-Gutiérrez, I.; Da-Silva, R.; Lago-Vanzela, E.S. Storage stability of the phenolic compounds, color and antioxidant activity of jambolan juice powder obtained by foam mat drying. Food Res. Int. 2020, 128, 108750.
- Darniadi, S.; Ifie, I.; Ho, P.; Murray, B.S. Evaluation of total monomeric anthocyanin, total phenolic content and individual anthocyanins of foam-mat freeze-dried and spray-dried blueberry powder. J. Food Meas. Charact. 2019, 13, 1599–1606.
- 46. Suda, I.; Ishikawa, F.; Hatakeyama, M.; Miyawaki, M.; Kudo, T.; Hirano, K.; Ito, A.; Yamakawa, O.; Horiuchi, S. Intake of Purple Sweet Potato Beverage Affects on Serum Hepatic Biomarker Levels of Healthy Adult Men With Borderline Hepatitis. Eur. J. Clin. Nutr. 2008, 62, 60–67.
- 47. Gao, X.; Cassidy, A.; Schwarzschild, M.A.; Rimm, E.B.; Ascherio, A. Habitual intake of dietary flavonoids and risk of Parkinson disease. Neurology 2012, 78, 1138–1145.
- 48. Valenti, L.; Riso, P.; Mazzocchi, A.; Porrini, M.; Fargion, S.; Agostoni, C. Dietary Anthocyanins as Nutritional Therapy for Nonalcoholic Fatty Liver Disease. Oxid. Med. Cell. Longev. 2013, 2013, 145421.
- 49. Vendrame, S.; Del Bo, C.; Ciappellano, S.; Riso, P.; Klimis-Zacas, D. Berry Fruit Consumption and Metabolic Syndrome. Antioxidants 2016, 5, 34.
- Oki, T.; Kano, M.; Ishikawa, F.; Goto, K.; Watanabe, O.; Suda, I. Double-blind, Placebo-Controlled Pilot Trial of Anthocyanin-Rich Purple Sweet Potato Beverage on Serum Hepatic Biomarker Levels in Healthy Caucasians With Borderline Hepatitis. Eur. J. Clin. Nutr. 2017, 71, 290–292.
- 51. Fairlie-Jones, L.; Davison, K.; Fromentin, E.; Hill, A.M. The Effect of Anthocyanin-Rich Foods or Extracts on Vascular Function in Adults: A Systematic Review and Meta-Analysis of Randomised Controlled Trials. Nutrients 2017, 9, 908.
- 52. Godos, J.; Vitale, M.; Micek, A.; Ray, S.; Martini, D.; Del Rio, D.; Riccardi, G.; Galvano, F.; Grosso, G. Dietary Polyphenol Intake, Blood Pressure, and Hypertension: A Systematic Review and Meta-Analysis of Observational Studies. Antioxidants 2019, 8, 152.

- 53. Ullah, R.; Khan, M.; Shah, S.A.; Saeed, K.; Kim, M.O. Natural Antioxidant Anthocyanins-A Hidden Therapeutic Candidate in Metabolic Disorders With Major Focus in Neurodegeneration. Nutrients 2019, 11, 1195.
- 54. Winter, A.N.; Bickford, P.C. Anthocyanins and Their Metabolites as Therapeutic Agents for Neurodegenerative Disease. Antioxidants 2019, 8, 333.
- 55. Krikorian, R.; Kalt, W.; McDonald, J.E.; Shidler, M.D.; Summer, S.S.; Stein, A.L. Cognitive performance in relation to urinary anthocyanins and their flavonoid-based products following blueberry supplementation in older adults at risk for dementia. J. Funct. Foods 2020, 64, 103667.
- 56. Cásedas, G.; Les, F.; López, V. Anthocyanins: Plant Pigments, Food Ingredients or Therapeutic Agents for the CNS? A Mini-Review Focused on Clinical Trials. Curr. Pharm. Des. 2020, 26, 1790–1798.
- 57. Danielewski, M.; Matuszewska, A.; Nowak, B.; Kucharska, A.Z.; Sozański, T. The Effects of Natural Iridoids and Anthocyanins on Selected Parameters of Liver and Cardiovascular System Functions. Oxid. Med. Cell. Longev. 2020, 2020, 2735790.
- 58. Davinelli, S.; Bertoglio, J.C.; Zarrelli, A.; Pina, R.; Scapagnini, G. A Randomized Clinical Trial Evaluating the Efficacy of an Anthocyanin-Maqui Berry Extract (Delphinol®) on Oxidative Stress Biomarkers. J. Am. Coll. Nutr. 2015, 34, 28–33.
- Toaldo, I.M.; Cruz, F.A.; de Lima Alves, T.; de Gois, J.S.; Borges, D.L.G.; Cunha, H.P.; da Silva, E.L.; Bordignon-Luiz, M.T. Bioactive Potential of Vitis Labrusca L. Grape Juices From the Southern Region of Brazil: Phenolic and Elemental Composition and Effect on Lipid Peroxidation in Healthy Subjects. Food Chem. 2015, 173, 527–535.
- 60. Bialasiewicz, P.; Prymont-Przyminska, A.; Zwolinska, A.; Sarniak, A.; Wlodarczyk, A.; Krol, M.; Markowski, J.; Rutkowski, K.P.; Nowak, D. Sour Cherries but Not Apples Added to the Regular Diet Decrease Resting and fMLP-Stimulated Chemiluminescence of Fasting Whole Blood in Healthy Subjects. J. Am. Coll. Nutr. 2018, 37, 24–33.
- Del Bó, C.; Riso, P.; Campolo, J.; Møller, P.; Loft, S.; Klimis-Zacas, D.; Brambilla, A.; Rizzolo, A.; Porrini, M. A Single Portion of Blueberry (Vaccinium Corymbosum L) Improves Protection Against DNA Damage but Not Vascular Function in Healthy Male Volunteers. Nutr. Res. 2013, 33, 220–227.
- Kropat, C.; Mueller, D.; Boettler, U.; Zimmermann, K.; Heiss, E.H.; Dirsch, V.M.; Rogoll, D.; Melcher, R.; Richling, E.; Marko, D. Modulation of Nrf2-dependent Gene Transcription by Bilberry Anthocyanins in vivo. Mol. Nutr. Food Res. 2013, 57, 545–550.
- 63. Erlund, I.; Koli, R.; Alfthan, G.; Marniemi, J.; Puukka, P.; Mustonen, P.; Mattila, P.; Jula, A. Favorable Effects of Berry Consumption on Platelet Function, Blood Pressure, and HDL Cholesterol. Am. J. Clin. Nutr. 2008, 87, 323–331.
- Cassidy, A.; Rogers, G.; Peterson, J.J.; Dwyer, J.T.; Lin, H.; Jacques, P.F. Higher Dietary Anthocyanin and Flavonol Intakes Are Associated With Anti-Inflammatory Effects in a Population of US Adults. Am. J. Clin. Nutr. 2015, 102, 172– 181.
- 65. Natella, F.; Macone, A.; Ramberti, A.; Forte, M.; Mattivi, F.; Matarese, R.M.; Scaccini, C. Red Wine Prevents the Postprandial Increase in Plasma Cholesterol Oxidation Products: A Pilot Study. Br. J. Nutr. 2011, 105, 1718–1723.
- 66. Estruch, R.; Sacanella, E.; Mota, F.; Chiva-Blanch, G.; Antúnez, E.; Casals, E.; Deulofeu, R.; Rotilio, D.; Andres-Lacueva, C.; Lamuela-Raventos, R.M.; et al. Moderate Consumption of Red Wine, but Not Gin, Decreases Erythrocyte Superoxide Dismutase Activity: A Randomised Cross-Over Trial. Nutr. Metab. Cardiovasc. Dis. 2011, 21, 46–53.
- 67. Zhu, Y.; Ling, W.; Guo, H.; Song, F.; Ye, Q.; Zou, T.; Li, D.; Zhang, Y.; Li, G.; Xiao, Y.; et al. Anti-inflammatory Effect of Purified Dietary Anthocyanin in Adults With Hypercholesterolemia: A Randomized Controlled Trial. Nutr. Metab. Cardiovasc. Dis. 2013, 23, 843–849.
- 68. Curtis, P.J.; van der Velpen, V.; Berends, L.; Jennings, A.; Feelisch, M.; Umpleby, A.M.; Evans, M.; Fernandez, B.O.; Meiss, M.S.; Minnion, M.; et al. Blueberries Improve Biomarkers of Cardiometabolic Function in Participants With Metabolic Syndrome-Results From a 6-month, Double-Blind, Randomized Controlled Trial. Am. J. Clin. Nutr. 2019, 109, 1535–1545.
- Zhang, P.-W.; Chen, F.-X.; Li, D.; Ling, W.-H.; Guo, H.-H. A CONSORT-compliant, Randomized, Double-Blind, Placebo-Controlled Pilot Trial of Purified Anthocyanin in Patients with Nonalcoholic Fatty Liver Disease. Medicine (Baltimore) 2015, 94, e758.
- 70. Guo, H.; Zhong, R.; Liu, Y.; Jiang, X.; Tang, X.; Li, Z.; Xia, M.; Ling, W. Effects of Bayberry Juice on Inflammatory and Apoptotic Markers in Young Adults With Features of Non-Alcoholic Fatty Liver Disease. Nutrition 2014, 30, 198–203.
- 71. Spormann, T.M.; Albert, F.W.; Rath, T.; Dietrich, H.; Will, F.; Stockis, J.-P.; Eisenbrand, G.; Janzowski, C. Anthocyanin/polyphenolic-rich Fruit Juice Reduces Oxidative Cell Damage in an Intervention Study With Patients on Hemodialysis. Cancer Epidemiol. Biomarkers Prev. 2008, 17, 3372–3380.
- 72. Castilla, P.; Dávalos, A.; Teruel, J.L.; Cerrato, F.; Fernández-Lucas, M.; Merino, J.L.; Sánchez-Martín, C.C.; Ortuño, J.; Lasunción, M.A. Comparative Effects of Dietary Supplementation With Red Grape Juice and Vitamin E on Production

of Superoxide by Circulating Neutrophil NADPH Oxidase in Hemodialysis Patients. Am. J. Clin. Nutr. 2008, 87, 1053–1061.

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