

EWSR1

Subjects: **Pathology**

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soft tissue tumors

bone tumors

pathology

molecular

1. Introduction

Ewing sarcoma was molecularly defined by Delattre et al. in 1992 upon the identification of the Ewing sarcoma breakpoint region 1 (*EWSR1*) located on chromosome 22q12.2 and the term for this gene was coined [1]. *EWSR1* is a multifunctional protein ubiquitously expressed in most cell types, indicating diverse roles in physiological cellular processes, including organ development and aging. Genetic and epigenetic pathways are modulated by *EWSR1* but the exact mechanisms are still poorly understood [2].

EWSR1 belongs to the FET (also known as TET) family of RNA-binding proteins that also includes Fused in Sarcoma (FUS), and TATA-box binding protein Associated Factor 15 (TAF15) [2]. As a consequence of the multifunctional role of *EWSR1* leading to a high frequency of transcription of the chromosomal region where the gene is located, *EWSR1* is exposed to aberrations such as rearrangements. Consecutive binding to other genes leads to chimeric proteins inducing oncogenesis. These various somatic genetic rearrangements involving *EWSR1* result in a fusion of its N-terminal coding region to the C-terminal DNA binding domain of one of several transcription factors. They are reported to act as aberrant transcription factors with the N-terminal domain of *EWSR1* as a strong transactivator. The other TET family members are homologous and are involved in strikingly similar translocation events giving rise to the production of structurally similar oncoproteins [3][4].

With the advent of widely used modern molecular techniques during the last decades, it has become obvious that *EWSR1* is involved in development of diverse benign and malignant tumors with mesenchymal, neuroectodermal, and epithelial/myoepithelial features [5]. As oncogenic transformation mediated by *EWSR1*-fusion proteins leads to such diverse tumor types, there must be a selection on a multipotent stem cell level [2].

2. Ewing Sarcoma

Arthur Purdy Stout and James Ewing were the first to describe this aggressive small, blue round-cell entity in 1918 and 1921, respectively [6][7][8]. Later on, the chromosomal translocation (11;22) was found by Aurias et al. and Turc-Carel et al. in 1983, the second breakthrough of translocation/fusion-gene associated sarcomas following alveolar

rhabdomyosarcoma (ARMS) [9][10][11]. Subsequently, the fusion gene has been detected as mentioned in the introduction [1], being the genetic hallmark by an otherwise aspecific small blue, round-cell tumor.

Ewing sarcoma, the prototypic round-cell sarcoma, is relatively common in comparison to other small blue round-cell sarcomas. It arises in soft tissue and bone of children, adolescents, and young adults. Exceptionally, older patients are affected. The mean age is in the second to third decade. White males have the highest incidence and black females the lowest due to ethnic genetic preposition differences. Tumors can originate anywhere in the body, and around 80% of the neoplasms arise in the bone with preference sites in decreasing order of frequency: lower extremities, pelvis, upper extremities, ribs, spine, and craniofacial. Distribution in the soft tissue is extremities, chest wall, retroperitoneum, paravertebral, pelvis, and head and neck. Visceral organs, skin, and epidural spaces are rarely involved [12][13]. The origin of the peripheral nerve as reported by Stout in 1918 can clinically be confused with malignant peripheral nerve sheath tumor [7].

Macroscopically, these infiltrative lesions are (multi)nodular, fleshy, and often necrotic. A pseudocapsule can be present in soft tissue neoplasms. Post-therapy specimens show fibrosis, necrosis, and hemorrhage, often without visible tumor [12][13].

Histologically, Ewing sarcoma is composed of cellular sheets of relatively featureless small cells with round dark nuclei and inconspicuous cytoplasm (Figure 1). In some cases, cells are larger displaying more nuclear variability. The cytoplasm can appear clear due to retraction artefacts. Homer-Wright rosettes may be numerous in a subset of cases initially called peripheral primitive neuroectodermal tumors [6][13]. Adamantinoma-like Ewing sarcoma shows more cohesive sheets and nests of cells with peripheral palisading, prominent desmoplastic stroma with production of hyaline membrane collagen, presence of keratin pearl formation, and comedo-like necrosis. These lesions are predestinated for misinterpretation as carcinoma, since keratins, including high molecular keratins, p40, and p63, are commonly positive [6][14].

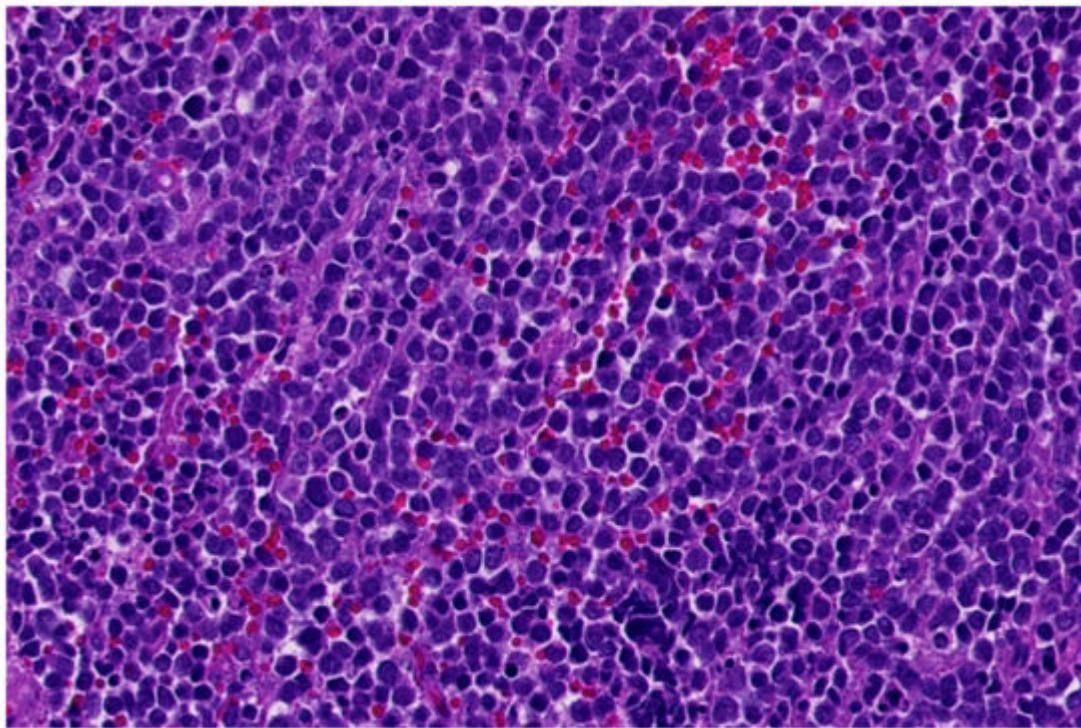


Figure 1. Classical morphology of Ewing sarcoma (HE; 40× magnification).

Immunohistochemically, CD99 is specific in its distinct staining pattern of the cell-membrane. Nuclear FLI and ERG expression is commonly observed in the cases with corresponding fusion genes. Neuroendocrine markers may be expressed. Keratin-expression, often dot-like, was found in 1/3 of the cases; it can be confused with small-cell carcinoma, especially when combined with the expression of p40 and p63 [6][14]. This is of particular importance in the head and neck area [14]. Expression of NKX2-2 in Ewing sarcoma seems to be highly sensitive, with imperfect specificity in comparison to other small, blue round-cell tumors [15][16][17][18][19]. Expression of desmin is reported in a few cases, and can be confused with ARMS or desmoplastic small round-cell tumor (DSRCT) [6][20].

Ewing sarcoma is genetically characterized by binding of *EWSR1* or other members of the TET/FET family to members of the ETS family [5]. Approximately 85–90% of the Ewing's sarcomas display the translocation t(11;22) (q24;q12) resulting in the *EWS/FLI1* fusion gene, and approximately 5–10% harbor a *EWSR1-ERG* fusion gene [6]. The remaining cases show rare gene partners, such as *ETV1*, *ETV4*, and *FEV*, and *EWSR1* can be substituted by *FUS* [21].

Although prognosis has improved markedly for patients with primary disease (5-year survival rate around 65%), presence of metastatic disease at time of diagnosis or early relapse leads to an adverse prognosis (5-year survival rate around 25–30%), with adequate surgical resection, aggressive multimodal chemotherapy, and adjuvant local radiotherapy being the optimal treatments.

Differential diagnoses are listed in Table 1.

Table 1. Differential diagnoses of Ewing sarcoma.

Entity	Morphology	IHC	Common Genetic Alterations
CIC-sarcoma	Sheets of undifferentiated round/spindle/epithelioid cells; mild nuclear pleomorphism; and necrosis	CD99 (mostly patchy), WT1, ETV4, DUX4, and NUT (<i>CIC-NUTM1</i>)	<i>CIC-DUX4/FOXO4/LEUTX/NUTM1/2A</i> fusions
BCOR-sarcoma	Sheets/nests/short fascicles of uniform; bland round-oval-spindle cells; rich capillary network; and myxoid matrix (variable)	BCOR, SATB2, cyclin D1, TLE1, CCNB3 (<i>BCOR-CCNB3</i>), and CD99 (50%)	<i>BCOR-CCNB3/MAML3/ZC3H7B, KMT2D</i>); <i>BCOR</i> ITD*; and <i>YWHAE-NUTM2B</i> ; *ITD, internal tandem duplication
<i>EWSR1</i> -nonETS round-cell sarcomas	Cords/nests/pseudoacinar pattern of round-spindle cells; bland-pleomorphic spectrum; and fibro-/myxohyaline stroma	CD99, NKX2.2, and CKAE1/3 (focal, dot-like)	<i>EWSR1/FUS-NFATc2</i>
	Diverse morphology: round-spindle cells; fibrous stroma	Co-expression of myogenic markers (desmin/myogenin/MyoD1), neurogenic markers (S100/SOX10/MITF/GFAP) and keratins (AE1/3)	<i>EWSR1-PATZ1</i> or <i>EWSR1-VEZF1</i>
Desmoplastic small round-cell tumor	Sheets/nests/cords of uniform; bland round cells; and desmoplastic stroma	Desmin (dot-like), keratin, EMA, and WT1 (C-terminus)	<i>EWSR1-WT1</i>
Lymphoblastic lymphoma	Small-medium blastoid cells; minimal cytoplasm	CD99, TdT, CD45, CD34, CD1a, and B- and T-cell markers	Diverse
Small-cell carcinoma	Small-medium round-oval cells; salt and pepper chromatin; indistinct nucleoli; molding; and apoptosis	Keratins, CD56, synaptophysin, chromogranin, and TTF1	Diverse; <i>TP53, PTEN</i> mutations; <i>RB1</i> , 3p loss; and <i>MYC</i> amplification
NUT carcinoma	Poorly cohesive sheets of primitive/basaloid cells; abrupt keratinization; and coagulative necrosis	CK5/6, P40, P63, and NUT	<i>NUT-BRD3/BRD4/NSD3/CIC/BCORL1/MGA/MXD4</i>
Myoepithelial carcinoma	Solid sheets/nests of cell with high nuclear grade or undifferentiated round-cell morphology; facultatively glandular component;	Pankeratins, S100, EMA, GFAP, SOX10, P63, SMA, calponin, desmin (focal); and INI1 loss (subset)	<i>EWSR1</i> rearrangements (various fusion partners); <i>PLAG1</i> rearrangements (mixed tumors)

Entity	Morphology	IHC	Common Genetic Alterations
	necrosis; and high mitotic count		
ARMS	Nests with central discohesion-solid nests; monomorphic primitive round cells; and multinucleated wreath-like giant cells	Desmin, myogenin (strong, diffuse), MyoD1, keratin, neuro-endocrine markers (CD56, synaptophysin, and chromogranin)	<i>PAX3/PAX7-FOXO1</i>
Sinonasal glomangiopericytoma	Solid-fascicular pattern; spindle-round cells with minimal atypia; arranged around staghorn vessels; and perivascular hyalinization	Beta-catenin (nuclear), SMA	<i>CTNNB1</i> mutations
Glomus tumor	Solid-nested pattern; small, uniform round cells with round nucleus, amphophilic-slightly eosinophilic cytoplasm and sharply defined cell borders; and variable vascular pattern	SMA with membranous accentuation, caldesmon, and collagen IV	<i>MIR143-NOTCH1/2/3</i> , and <i>BRAF/KRAS</i> mutations
Rhabdoid tumor	Solid pattern; rounded polygonal cells with vesicular nuclei and prominent nucleoli; and eosinophil hyaline-like cytoplasmic inclusions	Diverse; keratins, EMA, CD99, synaptophysin, SALL4, glypican-3, and INI1 loss	<i>SMARCB1</i> biallelic loss, <i>SMARCB1</i> or <i>SMARCA4</i> (germline) mutations
Mesenchymal chondrosarcoma	Biphasic: poorly differentiated round cells and islands of hyaline cartilage; staghorn-like vessels	S100, CD99, SOX9, EMA, desmin, myogenin, and MyoD1	<i>HEY1-NCOA2</i>
Synovial sarcoma with round-cell features	Fascicles or sheets of small round hyperchromatic cells; high N/C ratio; staghorn vessels; necrosis; and thin fibrovascular septa	CD99, BCL2, CD56, TLE1, S100 (focal), EMA, and keratins (variable)	SS18-SSX1/2/4

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