MicroRNA-361

Subjects: Medicine, General & Internal

Contributor: Peixin Dong

MicroRNA-361-5p (miR-361) expression frequently decreases or is lost in different types of cancers, and contributes to tumor suppression by repressing the expression of its target genes implicated in tumor growth, epithelial-to-mesenchymal transition (EMT), metastasis, drug resistance, glycolysis, angiogenesis, and inflammation.

Keywords: microRNA-361; EMT; angiogenesis; tumor microenvironment; cancer diagnosis; cancer treatment

1. Introduction

Large-scale transcriptional analysis reveals that more than 80% of the human genome is transcribed into RNA, whereas less than 2% of the human genome is used for protein translation ^[1], suggesting that the vast majority of the human transcriptome is composed of non-coding RNAs (ncRNAs). MicroRNAs (miRNAs) are a class of endogenous regulatory noncoding RNAs, typically 20–23 nucleotides in length, thereby exerting essential roles in a wide range of physiological processes ^[2]. Although some miRNAs can bind to the 5'-untranslated regions (5'-UTRs) or the coding regions of target messenger RNAs (mRNAs) ^{[3][4]}, miRNAs primarily suppress the expression of their target genes by targeting the 3'-UTRs of target mRNAs for mRNA degradation or translation inhibition ^[2]. A single miRNA can target many genes, and multiple miRNAs can regulate a single gene. Previous studies indicated that miRNAs may regulate as many as one third of human genes ^[2]. miRNAs are evolutionary highly conserved ncRNAs and are expressed in a tissue-specific or developmental stage-specific manner, thereby contributing to cell or tissue-specific protein expression profiles ^{[5][6][7]}.

In human cancer cells, miRNAs exert either pro- or anti-tumorigenic effects through tissue-dependent mechanisms. Some miRNAs that are amplified or overexpressed in cancer could act as oncogenes to either directly or indirectly downregulate the expression of tumor suppressors [8][9]. On the other hand, some miRNAs, such as miR-361-5p (miR-361), can target mRNAs encoding oncogenic proteins and therefore be categorized as tumor suppressors [8][9][10].

Dysregulation of miRNA expression was reported in most cancer types [3][9]. miRNA expression profiles can distinguish cancer tissues from normal tissues and separate different cancers subtypes [11]. miRNAs have already been described as non-invasive biomarkers useful for cancer diagnosis, patient stratification, and the prediction of patient prognosis, and treatment efficacy [12][13][14].

miRNAs mediate tumor initiation and progression by regulating a variety of biological processes, including cell proliferation, migration, invasion, metastasis, glycolysis, apoptosis, cancer stem cell (CSC)-like phenotype, chemoresistance, and epithelial-to-mesenchymal transition (EMT) [11][15]. The components of the tumor microenvironment, which includes the extracellular matrix (ECM), fibroblasts, immune cells, inflammatory cells, endothelial cells, lymphatic endothelial cells, growth factors, and cytokines, play an important role during tumor progression and metastasis [16]. Recent works demonstrated the importance of miRNAs in regulating complex signaling networks involved in multiple aspects of the microenvironment remodeling, including the hypoxic response, angiogenesis, anti-tumor immune response, inflammation, and ECM organization [16].

2. Dysregulation of miR-361 in Tumor

Previous studies of solid tumors showed that miR-361 is frequently downregulated in various tumor tissues, including cutaneous squamous cell carcinoma $^{[17]}$, osteosarcoma $^{[18]}$, breast cancer $^{[19]}$ $^{[20]}$ $^{[21]}$ glioma $^{[23]}$ papillary thyroid carcinoma $^{[25]}$, lung cancer $^{[26]}$ $^{[27]}$ gastric cancer $^{[30]}$ hepatocellular carcinoma $^{[33]}$, colorectal cancer $^{[32]}$ ovarian cancer $^{[34]}$, endometrial cancer $^{[10]}$, cervical cancer $^{[35]}$, and prostate cancer $^{[36]}$ However, increased expression of miR-361 was detected in acute myeloid leukemia $^{[39]}$, indicating the possibility that miR-361 dysregulation might be required to impair differentiation programs in leukemia, and miR-361 may regulate the expression of the hematopoietic differentiation-specific genes, which have a weak importance in solid tumors. Several works demonstrated

that low levels of miR-361 were associated with shorter overall survival in patients with breast cancer $\frac{[20][21]}{2}$, gastric cancer $\frac{[32]}{2}$, and colorectal cancer $\frac{[32]}{2}$.

3. Mechanisms of miR-361 Regulation in Tumor

Large-scale profiling studies have revealed that dysregulation of miRNA is a common event during cancer carcinogenesis and metastasis [11][12]. The molecular mechanisms regulating miRNA expression include genomic amplification or deletion of miRNA genes, abnormal transcriptional control of miRNAs, epigenetic silencing, and defects in miRNA biogenesis and processing machinery [11][40]. The downregulation of miR-361 in tumor tissues could be caused by several mechanisms (**Figure 1**A).

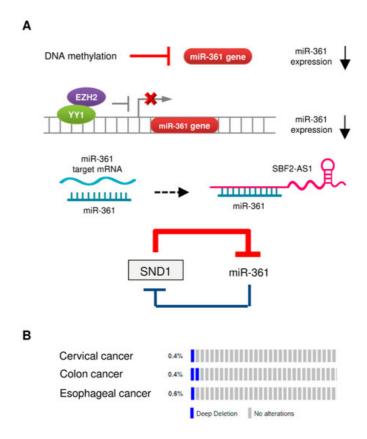


Figure 1. Mechanisms of miR-361 dysregulation in tumors. (**A**) Reported mechanisms responsible for miR-361 downregulation in tumors. (**B**) OncoPrint of cBioPortal showing the genetic alterations of *miR-361* (deep deletion) in tumor samples obtained from The Cancer Genome Atlas (TCGA)-cervical cancer, TCGA-colon cancer, and TCGA-esophageal cancer datasets. Each bar indicates the individual cases and % on the left indicates the percentage of cases altered in the human *miR-361* gene.

3.1. DNA Hypermethylation

Hypermethylation of tumor suppressor gene promoter regions leads to silencing of those genes. A previous study reported that the expression of miR-361 in hepatocellular carcinoma cell lines was restored upon treatment with an inhibitor of DNA methylation 5-azacytidine (5-AZA) [33]. Similar results were obtained from another study, where 5-AZA treatment significantly upregulated miR-361 expression in endometrial cancer cells [10].

3.2. Transcriptional Control of miR-361 Expression

We previously reported that enhancer of zeste homolog 2 (EZH2), which represses gene expression by methylation of histone H3 on lysine 27, acted as a co-suppressor of transcription factor YY1 to epigenetically suppress the transcription of miR-361 [10]. The use of GSK343 (a specific EZH2 inhibitor) was found to increase the levels of miR-361 in endometrial cancer cells [10].

3.3. Long Non-Coding RNA (IncRNA) SBF2-AS1 Acts as a Sponge for miR-361

LncRNAs are non-protein coding transcripts longer than 200 nucleotides that could serve as miRNA sponges to inhibit the interaction between miRNA-target mRNAs or suppressing the levels of miRNAs $^{[41]}$. For instance, in mouse cardiomyocytes, mitochondrial dynamic related lncRNA (MDRL) directly binds to miR-361 and acts as its sponge to promote the processing of pri-miR-484 $^{[42]}$. Another lncRNA, Maternally Expressed Gene 3 (MEG3), was shown to

facilitate cardiac hypertrophy by sponging miR-361 $^{[43]}$. In cervical cancer cells, IncRNA SBF2-AS1 (SBF2 Antisense RNA 1) was shown to function as an endogenous RNA sponge that interacted with miR-361 and suppressed its expression $^{[35]}$.

3.4. The SND1/miR-361 Feedback Loop Controls miR-361 Expression

miR-361 directly targeted SND1 (Staphylococcal nuclease and tudor domain containing 1), and SND1 conversely suppressed the expression of miR-361 by binding to pre-miR-361, thus creating a double-negative feedback loop, in which miR-361 and SND1 repress expression of each other in gastric and colon cancer cells [24].

3.5. Deletion of the Human miR-361 Gene

The reduced miRNA expression in tumor cells could arise from copy-number alterations and chromosomal aberrations (such as amplification, deletion, or translocation) [11]. However, whether genomic alterations of the human miR-361 gene can lead to decreased expression of miR-361 in cancer is poorly understood. We investigated copy-number alterations and nucleotide changes of the miR-361 gene in human cervical, colon and esophageal cancer samples from the cBioPortal database. As shown in **Figure 2**B, gene deletion is the most frequent alteration type in these cancers, supporting the notion that the loss of miR-361 expression plays an important role in the development of colon and cervical cancers [32][35].

4. The Impact of miR-361 on the Aggressive Properties of Tumor Cells and Tumor Microenvironment Remodeling

MiR-361 has been shown to act as a novel tumor suppressor that represses a large number of downstream target transcripts implicated in cellular proliferation, glycolysis, migration, invasion, EMT, chemoresistance, cancer stemness, angiogenesis, and inflammation (**Figure 2**).

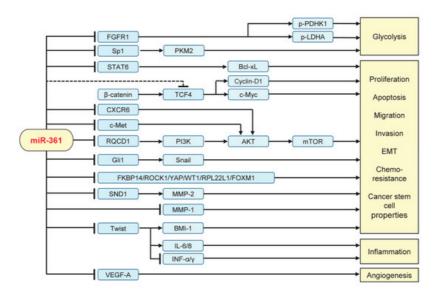


Figure 2. Validated targets and signaling pathways regulated by miR-361 in human tumor cells. FGFR1: Fibroblast growth factor receptor 1; Sp1: Transcription factor; STAT6: Signal transducer and activator of transcription 6; PKM2: pyruvate kinase M2; PDHK1: pyruvate dehydrogenase kinase 1; LDHA: lactate dehydrogenase A; TCF4: Transcription factor 4; CXCR6: C-X-C chemokine receptor type 6; RQCD1: CCR4-NOT transcription complex subunit 9; PI3K: phosphoinositide 3-kinase; mTOR: mammalian target of rapamycin; SND1: Staphylococcal nuclease and tudor domain containing 1; MMP: Matrix metallopeptidase; IL: interleukin; INF-α/y: interferon α/y; VEGF-A: vascular endothelial growth factor A.

4.1. Inhibiting Tumor Growth, Invasion, EMT, Metastasis, and Glycolysis

Restoration of miR-361 expression by transfection with miR-361 mimics inhibited the proliferation of osteosarcoma, breast cancer, thyroid papillary carcinoma, lung cancer, gastric cancer, colorectal cancer, hepatocellular carcinoma, cervical cancer, and prostate cancer cells [18][19][25][26][27][28][29][30][31][32][33][35][36]. Moreover, over-expression of miR-361 attenuated cell migration and invasion in endometrial cancer, breast cancer, glioma, papillary thyroid carcinoma, lung cancer, gastric cancer, ovarian cancer, and prostate cancer [10][19][23][24][25][26][28][30][32][34][36]. Conversely, knocking down miR-361 expression using anti-miR-361 inhibitor promoted cell migration and invasion in endometrial cancer, breast cancer, glioma, papillary thyroid carcinoma, lung cancer, gastric cancer, and ovarian cancer [10][19][22][23][24][25][26][28][30][32][34][36]. Mechanically, miR-361 impairs tumor cell proliferation, migration, and invasion by directly targeting and downregulating the expression of FKBP14 [18], MMP-1 [19], SND1 [24], ROCK1 [25], YAP [27], WT1 [28], RPL22L1 [34] and STAT6 [29][37].

The phosphoinositide 3-kinase (PI3K)/AKT pathway is activated in a wide range of cancers, and associated with cell growth, proliferation, survival, motility, tumor progression and resistance to cancer therapies. CCR4-NOT transcription complex subunit 9 (CNOT9/RQCD1) has been identified as a key activator of the PI3K/AKT pathway [44], and upregulation of miR-361 in breast cancer cells can suppress its direct target RQCD1, leading to the downregulation of PI3K, AKT, and MMP-9 [22]. The C-X-C motif chemokine receptor 6 (CXCR6), when bound with its ligand CXCL16, induced the activation of the PI3K/AKT signaling in cancer cells [45]. A previous study demonstrated that miR-361 inhibited the proliferation of hepatocellular carcinoma cells by directly targeting CXCR6 [33]. These results provided examples of miR-361-mediated repression of the PI3K/AKT signaling at different levels and illustrated the importance of miR-361 regulation in carcinogenesis and tumor progression. EMT encompasses a series of phenotypic and biochemical changes that enable epithelial cells to acquire a mesenchymal cell phenotype, which includes enhanced migration, invasion, metastasis, CSC-like features, resistance to conventional chemotherapy, radiotherapy, and small-molecule drugs [46][47][48].

EMT is mediated by a core set of key transcription factors, including Twist, Zinc finger E-box binding homeobox 1 (ZEB1), ZEB2, Snail and Slug, and the expression of these transcription factors are finely mediated at the transcriptional, translational and post-translational levels [46][47][48].

In accordance with its reported anti-tumor functions, ectopic expression of miR-361 was found to cause dramatic suppression of EMT process in various cancer cells. For example, experiments show that enforced overexpression of miR-361 greatly suppressed EMT, invasion and metastasis in many tumors, including endometrial cancer, glioma, lung cancer, gastric cancer, colorectal cancer, ovarian cancer, and prostate cancer [10][23][26][32][34][36]. By inhibiting the expression of Twist, miR-361 played a crucial role in suppressing EMT characteristics and cancer stem cell (CSC)-like properties of endometrial cancer cells [10].

In addition to targeting EMT-promoting transcription factors directly, miR-361 also modulated the expression of key mediators of the EMT program. For example, the loss of miR-361 expression activated Gli1 and its downstream effector Snail to promote EMT and prostate cancer cell invasion $^{[36]}$. In ovarian cancer cells, miR-361 targeted and reduced the levels of RPL22L1 and another target gene c-Met $^{[34]}$, which could serve as an upstream stimulator of the PI3K/AKT signaling and EMT-associated signaling pathways $^{[49]}$. Additionally, miR-361 attenuated EMT and chemoresistance in cancer cells by suppressing the expression of FOXM1 $^{[26][31]}$, an oncogenic transcription factor required for EMT and metastasis $^{[50]}$.

Activation of the Wingless-type MMTV integration site family (Wnt)/β-catenin signaling in cancer cells is responsible for EMT induction, metastasis, CSC self-renewal, increased resistance to chemotherapy or radiotherapy and immunosuppression $^{[51]}$. Although the introduction of miR-361 into gastric cancer cells downregulated the expression of Wnt/β-catenin pathway-related proteins (TCF4, Cyclin-D1 and c-Myc) $^{[30]}$, it remains unknown whether these genes are direct targets of miR-361.

Cancer cells are known to consume more glucose to produce lactate by glycolysis rather than oxidative phosphorylation, even in oxygen-rich conditions $^{[52]}$. Recent data suggested that miR-361 directly targeted the 3'-UTR of *FGFR1*, which promotes glycolysis through activation of two critical glycolytic enzymes lactate dehydrogenase A (LDHA) and pyruvate dehydrogenase kinase 1 (PDK1/PDHK1), thereby suppressing glucose consumption and lactate production of breast cancer cells $^{[19]}$. The glycolytic enzyme pyruvate kinase M2 (PKM2) is often highly expressed in cancer cells but is present at a very low level in normal cells $^{[52]}$. PKM2 catalyzes the rate-limiting ATP-generating step of glycolysis, controlling the conversion of phosphoenolpyruvate and ADP to pyruvate and ATP, respectively $^{[52]}$. Transfection of miR-361 mimic was shown to inhibit glucose metabolism by targeting Sp1 and subsequently downregulating the expression of PKM2 in prostate cancer $^{[38]}$. The role of miR-361 in the regulation of glucose metabolism in human cancers has not yet been fully investigated.

4.2. Suppressing Angiogenesis and Inflammation

Accumulated evidence showed that miRNAs participate in the remodeling of tumor microenvironments through several mechanisms, including the regulation of the expression of cell membrane proteins, secretion of cytokines, as well as transmission of mature miRNAs between different cell types via exosomes [53][54]. It has become apparent that miR-361 is able to regulate cancer progression through modulating tumor microenvironments (**Figure 2**).

Angiogenesis, an important hallmark of cancer, plays an essential role in providing tumor cells with oxygen and nutrients. Some miRNAs modulate the expression of regulatory molecules driving angiogenesis, including vascular endothelial growth factors (such as vascular endothelial growth factor A (VEGF-A)), cytokines, metalloproteinases, and growth factors [55]. MiR-361 targeted the 3'-UTR of VEGF-A to repress its expression in skin squamous cell carcinoma [17]. Moreover,

overexpression of miR-361 was shown to indirectly reduce the expression of VEGF-A through inhibiting the Wnt/ β -catenin pathway in gastric cancer cells [30]. Consistent with these data, transient transfection with miR-361 mimic significantly downregulated the expression of VEGF-A, whereas the silencing of miR-361 with miRNA inhibitor enhanced the levels of VEGF-A in endometrial cancer cells [10]. Collectively, these results suggest that reduced levels of miR-361 could be an important driving mechanism for the formation of a pro-angiogenic tumor microenvironment.

Numerous studies have indicated that chronic inflammation actively promotes tumor initiation, progression, and metastasis via multiple mechanisms, including generation of an immunosuppressive tumor microenvironment $^{[56]}$. Tumor cells undergoing EMT could modulate the surrounding microenvironment via enhanced secretion of inflammatory cytokines (including IL-6 and IL-8) $^{[57][58]}$. We reported that miR-361 could downregulate the mRNA expression of IL-6 and IL-8 in endometrial cancer cells through targeting Twist $^{[10]}$. Additionally, the activation of signal transducer and activator of transcription (STAT) family members (for example STAT6) is closely linked to tumor-promoting inflammation and the suppression of anti-tumor immunity in multiple cancer tissues $^{[59][60]}$. MiR-361 was shown to directly inhibit the expression of STAT6 by binding to its 3'-UTR region $^{[29][37]}$. These data support a novel function of miR-361 in exerting anti-angiogenesis and anti-inflammatory effects, at least by regulating the EMT-associated signaling and the production of pro-inflammatory cytokines.

References

- 1. ENCODE Project Consortium. An integrated encyclopedia of DNA elements in the human genome. Nature 2012, 489, 57–74.
- 2. Shivdasani, R.A. MicroRNAs: Regulators of gene expression and cell differentiation. Blood 2006, 108, 3646–3653.
- 3. Da Sacco, L.; Masotti, A. Recent Insights and Novel Bioinformatics Tools to Understand the Role of MicroRNAs Binding to 5' Untranslated Region. Int. J. Mol. Sci. 2012, 14, 480–495.
- 4. O'Brien, J.; Hayder, H.; Zayed, Y.; Peng, C. Overview of MicroRNA Biogenesis, Mechanisms of Actions, and Circulatio n. Front. Endocrinol. 2018, 9, 402.
- 5. Pasquinelli, A.E.; Reinhart, B.J.; Slack, F.; Martindale, M.Q.; Kuroda, M.I.; Maller, B.; Hayward, D.C.; Ball, E.E.; Degna n, B.; Müller, P.; et al. Conservation of the sequence and temporal expression of let-7 heterochronic regulatory RNA. N ature 2000, 408, 86–89.
- 6. Ruvkun, G. Glimpses of a Tiny RNA World. Science 2001, 294, 797–799.
- 7. Lee, R.C. An Extensive Class of Small RNAs in Caenorhabditis elegans. Science 2001, 294, 862-864.
- 8. Ebert, M.S.; Sharp, P.A. Roles for microRNAs in conferring robustness to biological processes. Cell 2012, 149, 515–52 4.
- 9. Król, J.; Loedige, I.; Filipowicz, W. The widespread regulation of microRNA biogenesis, function and decay. Nat. Rev. G enet. 2010, 11, 597–610.
- 10. Ihira, K.; Dong, P.; Xiong, Y.; Watari, H.; Konno, Y.; Hanley, S.J.; Noguchi, M.; Hirata, N.; Suizu, F.; Yamada, T.; et al. EZ H2 inhibition suppresses endometrial cancer progression via miR-361/Twist axis. Oncotarget 2017, 8, 13509–13520.
- 11. Peng, Y.; Croce, C.M. The role of MicroRNAs in human cancer. Signal. Transduct. Target. Ther. 2016, 1, 15004.
- 12. Hayes, J.; Peruzzi, P.P.; Lawler, S. MicroRNAs in cancer: Biomarkers, functions and therapy. Trends Mol. Med. 2014, 2 0, 460–469.
- 13. Schwarzenbach, H.; Nishida, N.; Calin, G.A.; Pantel, K. Clinical relevance of circulating cell-free microRNAs in cancer. Nat. Rev. Clin. Oncol. 2014, 11, 145–156.
- 14. Bouchie, A. First microRNA mimic enters clinic. Nat. Biotechnol. 2013, 31, 577.
- 15. Dong, P.; Xiong, Y.; Yu, J.; Chen, L.; Tao, T.; Yi, S.; Hanley, S.J.B.; Yue, J.; Watari, H.; Sakuragi, N. Control of PD-L1 ex pression by miR-140/142/340/383 and oncogenic activation of the OCT4–miR-18a pathway in cervical cancer. Oncoge ne 2018, 37, 5257–5268.
- 16. Rupaimoole, R.; Calin, G.A.; Lopez-Berestein, G.; Sood, A.K. MicroRNA deregulation in cancer cells and the tumor mic roenvironment. Cancer Discov. 2016, 6, 235–246.
- 17. Kanitz, A.; Imig, J.; Dziunycz, P.J.; Primorac, A.; Galgano, A.; Hofbauer, G.F.L.; Gerber, A.P.; Detmar, M. The Expressio n Levels of MicroRNA-361-5p and Its Target VEGFA Are Inversely Correlated in Human Cutaneous Squamous Cell Car cinoma. PLoS ONE 2012, 7, e49568.

- 18. Wang, K.; Qi, X.; Liu, H.; Su, H. MiR-361 inhibits osteosarcoma cell lines invasion and proliferation by targeting FKBP1 4. Eur. Rev. Med. Pharmacol. Sci. 2018, 22, 79–86.
- 19. Ma, F.; Zhang, L.; Ma, L.; Zhang, Y.; Zhang, J.; Guo, B. MiR-361-5p inhibits glycolytic metabolism, proliferation and invasion of breast cancer by targeting FGFR1 and MMP-1. J. Exp. Clin. Cancer Res. 2017, 36, 158.
- 20. Chang, J.T.H.; Wang, F.; Chapin, W.; Huang, R.S. Identification of MicroRNAs as Breast Cancer Prognosis Markers thr ough the Cancer Genome Atlas. PLoS ONE 2016, 11, e0168284.
- 21. Cao, Z.G.; Huang, Y.N.; Yao, L.; Liu, Y.R.; Hu, X.; Hou, Y.F.; Shao, Z.M. Positive expression of miR-361-5p indicates be tter prognosis for breast cancer patients. J. Thorac. Dis. 2016, 8, 1772–1779.
- 22. Han, J.; Yu, J.; Dai, Y.; Li, J.; Guo, M.; Song, J.; Zhou, X. Overexpression of miR-361-5p in triple-negative breast cance r (TNBC) inhibits migration and invasion by targeting RQCD1 and inhibiting the EGFR/PI3K/Akt pathway. Bosn. J. Basi c Med. Sci. 2019, 19, 52–59.
- 23. Wei, C.; Qu, J.; Zhang, X.; Li, J.; Liu, J. MicroRNA-361-5p inhibits epithelial-to-mesenchymal transition of glioma cells t hrough targeting Twist1. Oncol. Rep. 2017, 37, 1849–1856.
- 24. Liu, J.; Yang, J.; Yu, L.; Rao, C.; Wang, Q.; Sun, C.; Shi, C.; Hua, D.; Zhou, X.; Luo, W.; et al. miR-361-5p inhibits gliom a migration and invasion by targeting SND1. OncoTargets Ther. 2018, 11, 5239–5252.
- 25. Li, R.; Dong, B.; Wang, Z.; Jiang, T.; Chen, G. MicroRNA-361-5p inhibits papillary thyroid carcinoma progression by tar geting ROCK1. Biomed. Pharmacother. 2018, 102, 988–995.
- 26. Hou, X.W.; Sun, X.; Yu, Y.; Zhao, H.M.; Yang, Z.J.; Wang, X.; Cao, X.C. miR-361-5p suppresses lung cancer cell lines p rogression by targeting FOXM1. Neoplasma 2017, 64, 526–534.
- 27. Zhang, S.; Liu, Z.; Wu, L.; Wang, Y. MiR-361 targets Yes-associated protein (YAP) mRNA to suppress cell proliferation in lung cancer. Biochem. Biophys. Res. Commun. 2017, 492, 468–473.
- 28. Wang, J.; Shang, J.; Yang, S.; Zhang, Y.; Zhao, X. microRNA-361 targets Wilms' tumor 1 to inhibit the growth, migration and invasion of non-small-cell lung cancer cells. Mol. Med. Rep. 2016, 14, 5415–5421.
- 29. Ma, Y.; Bao, C.; Kong, R.; Xing, X.; Zhang, Y.; Li, S.; Zhang, W.; Jiang, J.; Zhang, J.; Qiao, Z.; et al. MicroRNA-361-5p s uppresses cancer progression by targeting signal transducer and activator of transcription 6 in non-small cell lung canc er. Mol. Med. Rep. 2015, 12, 7367–7373.
- 30. Tian, L.; Zhao, Z.; Xie, L.; Zhu, J. MiR-361-5p inhibits the mobility of gastric cancer cells through suppressing epithelial-mesenchymal transition via the Wnt/β-catenin pathway. Gene 2018, 675, 102–109.
- 31. Tian, L.; Zhao, Z.; Xie, L.; Zhu, J. MiR-361-5p suppresses chemoresistance of gastric cancer cells by targeting FOXM1 via the PI3K/Akt/mTOR pathway. Oncotarget 2017, 9, 4886–4896.
- 32. Ma, F.; Song, H.; Guo, B.; Zhang, Y.; Zheng, Y.; Lin, C.; Wu, Y.; Guan, G.; Sha, R.; Zhou, Q.; et al. MiR-361-5p inhibits colorectal and gastric cancer growth and metastasis by targeting staphylococcal nuclease domain containing-1. Oncota rget 2015, 6, 17404–17416.
- 33. Sun, J.J.; Chen, G.Y.; Xie, Z.T. MicroRNA-361-5p Inhibits Cancer Cell Growth by Targeting CXCR6 in Hepatocellular C arcinoma. Cell. Physiol. Biochem. 2016, 38, 777–785.
- 34. Ma, J.; Jing, X.; Chen, Z.; Duan, Z.; Zhang, Y. MiR-361-5p decreases the tumorigenicity of epithelial ovarian cancer cell s by targeting at RPL22L1 and c-Met signaling. Int. J. Clin. Exp. Pathol. 2018, 11, 2588–2596.
- 35. Gao, F.; Feng, J.; Yao, H.; Li, Y.; Xi, J.; Yang, J. LncRNA SBF2-AS1 promotes the progression of cervical cancer by regulating miR-361-5p/FOXM1 axis. Artif. Cells Nanomed. Biotechnol. 2019, 47, 776–782.
- 36. Chen, S.; Zhang, G.; Yu, Q.; Zhang, X.; Han, G. MicroRNA-361 inhibited prostate carcinoma cell invasion by targeting Gli1. Int. J. Clin. Exp. Pathol. 2017, 10, 6108–6116.
- 37. Liu, D.; Tao, T.; Xu, B.; Chen, S.; Liu, C.; Zhang, L.; Lu, K.; Huang, Y.; Jiang, L.; Zhang, X.; et al. MiR-361-5p acts as a t umor suppressor in prostate cancer by targeting signal transducer and activator of transcription-6(STAT6). Biochem. Bi ophys. Res. Commun. 2014, 445, 151–156.
- 38. Ling, Z.; Liu, D.; Zhang, G.; Liang, Q.; Xiang, P.; Xu, Y.; Han, C.; Tao, T. miR-361-5p modulates metabolism and autoph agy via the Sp1-mediated regulation of PKM2 in prostate cancer. Oncol. Rep. 2017, 38, 1621–1628.
- 39. Koutova, L.; Sterbova, M.; Pazourková, E.; Pospisilova, S.; Svobodová, I.; Horinek, A.; Lysak, D.; Korabečná, M. The i mpact of standard chemotherapy on miRNA signature in plasma in AML patients. Leuk. Res. 2015, 39, 1389–1395.
- 40. Ha, M.; Kim, V.N. Regulation of microRNA biogenesis. Nat. Rev. Mol. Cell Biol. 2014, 15, 509-524.
- 41. Dong, P.; Xiong, Y.; Yue, J.; Hanley, S.J.B.; Kobayashi, N.; Todo, Y.; Watari, H. Exploring IncRNA-Mediated Regulatory Networks in Endometrial Cancer Cells and the Tumor Microenvironment: Advances and Challenges. Cancers 2019, 11,

- 42. Wang, K.; Sun, T.; Li, N.; Wang, Y.; Wang, J.X.; Zhou, L.Y.; Long, B.; Liu, C.Y.; Liu, F.; Li, P.F. MDRL IncRNA Regulates the Processing of miR-484 Primary Transcript by Targeting miR-361. PLoS Genet. 2014, 10, e1004467.
- 43. Zhang, J.; Liang, Y.; Huang, X.; Guo, X.; Liu, Y.; Zhong, J.; Yuan, J. STAT3-induced upregulation of lncRNA MEG3 regulates the growth of cardiac hypertrophy through miR-361-5p/HDAC9 axis. Sci. Rep. 2019, 9, 460.
- 44. Ajiro, M.; Katagiri, T.; Ueda, K.; Nakagawa, H.; Fukukawa, C.; Lin, M.L.; Park, J.H.; Nishidate, T.; Daigo, Y.; Nakamura, Y. Involvement of RQCD1 overexpression, a novel cancer-testis antigen, in the Akt pathway in breast cancer cells. Int. J. Oncol. 2009, 35, 673–681.
- 45. Wang, J.; Lu, Y.; Wang, J.; Koch, A.E.; Zhang, J.; Taichman, R.S. CXCR6 Induces Prostate Cancer Progression by the AKT/Mammalian Target of Rapamycin Signaling Pathway. Cancer Res. 2008, 68, 10367–10376.
- 46. Nieto, M.A.; Huang, R.Y.J.; Jackson, R.A.; Thiery, J.P. EMT: 2016. Cell 2016, 166, 21-45.
- 47. Huo, W.; Zhao, G.; Yin, J.; Ouyang, X.; Wang, Y.; Yang, C.; Wang, B.; Dong, P.; Wang, Z.; Watari, H.; et al. Lentiviral C RISPR/Cas9 vector mediated miR-21 gene editing inhibits the epithelial to mesenchymal transition in ovarian cancer ce Ils. J. Cancer 2017, 8, 57–64.
- 48. Dong, P.; Xiong, Y.; Watari, H.; Hanley, S.J.; Konno, Y.; Ihira, K.; Suzuki, F.; Yamada, T.; Kudo, M.; Yue, J.; et al. Suppre ssion of iASPP-dependent aggressiveness in cervical cancer through reversal of methylation silencing of microRNA. Sc i. Rep. 2016, 6, 35480.
- 49. Zhang, Y.; Xia, M.; Jin, K.; Wang, S.; Wei, H.; Fan, C.; Wu, Y.; Li, X.; Li, X.; Li, G.; et al. Function of the c-Met receptor t yrosine kinase in carcinogenesis and associated therapeutic opportunities. Mol. Cancer 2018, 17, 45.
- 50. Liao, G.B.; Li, X.Z.; Zeng, S.; Liu, C.; Yang, S.M.; Yang, L.; Hu, C.J.; Bai, J.Y. Regulation of the master regulator FOXM 1 in cancer. Cell Commun. Signal. 2018, 16, 57.
- 51. Zhan, T.; Rindtorff, N.; Boutros, M. Wnt signaling in cancer. Oncogene 2017, 36, 1461-1473.
- 52. Jang, M.; Kim, S.S.; Lee, J. Cancer cell metabolism: Implications for therapeutic targets. Exp. Mol. Med. 2013, 45, e45.
- 53. Zhang, Y.; Yang, P.; Wang, X.-F. Microenvironmental regulation of cancer metastasis by miRNAs. Trends Cell Biol. 201 4, 24, 153–160.
- 54. Yang, N.; Zhu, S.; Lv, X.; Qiao, Y.; Liu, Y.J.; Chen, J. MicroRNAs: Pleiotropic Regulators in the Tumor Microenvironmen t. Front. Immunol. 2018, 9, 2491.
- 55. Salinas-Vera, Y.M.; Marchat, L.A.; Gallardo-Rincón, D.; Ruiz-García, E.; Astudillo-De La Vega, H.; Echavarría-Zepeda, R.; López-Camarillo, C. AngiomiRs: MicroRNAs driving angiogenesis in cancer (Review). Int. J. Mol. Med. 2019, 43, 65 7–670.
- 56. Wang, D.; Dubois, R.N. Immunosuppression associated with chronic inflammation in the tumor microenvironment. Carc inogenesis 2015, 36, 1085–1093.
- 57. Palena, C.; Hamilton, D.H.; Fernando, I.R. Influence of IL-8 on the epithelial—mesenchymal transition and the tumor mic roenvironment. Futur. Oncol. 2012, 8, 713–722.
- 58. Landskron, G.; De La Fuente, M.; Thuwajit, P.; Thuwajit, C.; Hermoso, M.A. Chronic Inflammation and Cytokines in the Tumor Microenvironment. J. Immunol. Res. 2014, 2014, 1–19.
- 59. Yu, H.; Pardoll, D.; Jove, R. STATs in cancer inflammation and immunity: A leading role for STAT3. Nat. Rev. Cancer 20 09, 9, 798–809.
- 60. León-Cabrera, S.; Molina-Guzman, E.; Delgado-Ramírez, Y.; Vazquez-Sandoval, A.; Ledesma-Soto, Y.; Perez-Plasenci a, C.; Chirino, I.Y.; Delgado-Buenrostro, N.L.; Rodriguez-Sosa, M.; Vaca-Paniagua, F.; et al. Lack of STAT6 Attenuates I nflammation and Drives Protection against Early Steps of Colitis-Associated Colon Cancer. Cancer Immunol. Res. 201 7, 5, 385–396.