Estrogen Sulfotransferase

Subjects: Medicine, General & Internal

Contributor: Su-Jun Lee

Estrogen sulfotransferase (SULT1E1) is a phase II enzyme that sulfates estrogens to inactivate them and regulate their homeostasis. This enzyme is also involved in the sulfation of thyroid hormones and several marketed medicines. Though the profound action of SULT1E1 in molecular/pathological biology has been extensively studied, its genetic variants and functional studies have been comparatively rarely studied.

Keywords: estrogen sulfotransferase; SULT1E1

1. Introduction

Human SULT1E1 cDNA was first isolated, cloned, and characterized from the liver, and its localization was mapped to human chromosome 4 $^{[1]}$. SULT1E1 is expressed in the human embryo, and is also highly expressed in a wide range of fetal tissues, such as the liver, lung, kidney, and hormone-dependent tissues—such as the testis or endometrium—but its expression in adults with normal status is much lower than in the fetus and placenta $^{[2][3]}$. The expression of SULT1E1 varies widely in the human population, although it is not known whether this is under genetic control or not $^{[4]}$. Thus, it is possible that the variability in SULT1E1 expression results from different chemical influences.

Two agonists of peroxisome-proliferator-activated receptor α (PPAR α), WY14643 and IGF-1, show different regulatory effects on the SULT1E1 promoter activity. While WY14643 suppressed SULT1E1 activity, IGF-1 upregulated it, as measured by estrogen levels in endothelial cells and smooth muscle cells [5]. Interestingly, SULT1E1 was attenuated by both transfection with PPARy small interfering RNA (siRNA) and exposure to GW9662, the PPARy antagonist [6].

SULT1E1 regulation was observed when hepatocyte nuclear factor 4α (HNF4 α) was silenced. The significant suppression of both mRNA and protein levels of SULT1E1 occurred via Farnesoid X receptor (FXR) agonists in HepG2 cells ^[Z]. This finding confirmed that the effect of FXR on E₂ was SULT1E1-dependent. In patients with obstructive cholestasis, the accumulation of bile acids (activator of FXR) led to reduced mRNA and protein expression of hepatic SULT1E1, increased serum E₂ levels, and decreased serum estrone sulfate concentration ^[8]. Phosphorylated ROR α takes a regulatory signal to HNF4 α , and then activates the *SULT1E1* promoter in human liver cells ^[9].

Basal expression of SULT1E1 in the liver is relatively low $^{[\underline{10}]}$, but its expression and role could be impacted in response to ligands/substrates for nuclear receptors, such as the liver X receptor (LXR) $^{[\underline{10}]}$, the glucocorticoid receptor (GR) $^{[\underline{11}]}$, the constitutive androstane receptor (CAR) $^{[\underline{12}]}$, the estrogen receptor α (ER α) $^{[\underline{13}]}$, the pregnane X receptor (PXR) $^{[\underline{14}]}$, and the RAR-related orphan receptor α (ROR α) $^{[\underline{15}]}$ (Table 1).

Table 1. The nuclear receptors associated with *Sult1e1* regulation.

Gene ID ¹	Locus ²	Alias ¹	Number of Amino Acids ³	Number of Exons ¹
SULT1A1	Chr 16p11.2	HAST1/HAST2, P-PST, PST, ST1A1, ST1A3, STP, STP1, TSPST1	295 (isoform a)	13
			217 (isoform b)	13
SULT1A2	Chr 16p11.2	HAST4, P-PST, P-PST 2, ST1A2, STP2, TSPST2	295 (isoform 1)	8
			262 (isoform 2)	8
SULT1A3	Chr 16p11.2	HAST, HAST3, M-PST, ST1A3, ST1A3/ST1A4, ST1A4, ST1A5, STM, TL-PST	295	8
SULT1A4	Chr 16p11.2	HAST3, M-PST, ST1A3, ST1A3/ST1A4, ST1A4, STM, TL- PST	295	8
SULT1B1	Chr 4q13.3	ST1B1, ST1B2, SULT1B2	296	10

Gene ID ¹	Locus ²	Alias ¹	Number of Amino Acids ³	Number of Exons ¹
SULT1C2	Chr 2q12.3	ST1C1, ST1C2, SULT1C1, humSULTC2	296 (isoform a)	9
			307 (isoform b)	-
SULT1C3	Chr 2q12.3	ST1C3	304 (isoform 1)	10
			304 (isoform 2)	10
SULT1C4	Chr 2q12.3	SULT1C, SULT1C2	302 (isoform 1)	7
			227 (isoform 2)	,
SULT1E1	Chr 4q13.3	EST, EST-1, ST1E1, STE	294	9
SULT2A1	Chr 19q13.33	DHEA-ST, DHEA-ST8, DHEAS, HST, ST2, ST2A1, ST2A3, STD, SULT2A3, hSTa	285	6
SULT2B1	Chr 19q13.33	ARCI14, HSST2	350 (isoform a)	7
			365 (isoform b)	,
SULT4A1	Chr 22q13.31	BR-STL-1, BRSTL1, DJ388M5.3, NST, SULTX3, hBR-STL-1	284	11
SULT6B1	Chr 2p22.2	ST6B1	304 (isoform 1)	9
			265 (isoform 2)	3
SUPI1C2P1	Chr 2q12.3	SULT1C1P	pseudogene	4
SULT1C2P2	Chr 2q12.3		pseudogene	
SULT1D1P	Chr 4q13.3	SULT1D1	pseudogene	
SULT6B2P	Chr 12p12.1		pseudogene	

2. Sulfation of Estrogens and Thyroid Hormones by SULT1E1

2.1. Sulfation of Estrogens

Estrogens play fundamental roles in a variety of physiological systems. It has been widely established that estradiol (E2) exposure is one of the risk factors for breast carcinogenesis. One of the critical pathways for E2 inactivation is sulfation by SULT1E1. Estrone (E1) is synthesized by aromatization of androstenedione and is subsequently sulfated. After E1 is desulfated and subsequently turned into E2 by the 17β -hydroxysteroid dehydrogenases (17β -HSD), E2 can then be sulfated through SULT1E1 [16]. As previously mentioned, SULT1E1 is a cytosolic enzyme that catalyzes estrogen sulfation at the 3-hydroxyl site while using PAPS as a sulfate donor (Figure 1). Moreover, this enzyme has high affinity for its substrate E2, indicating its crucial role in modulating estrogen's action and homeostasis [17].

Figure 1. A schematic sulfation pathway of estrogens. STS, steroid sulfatase; PAPS, 3'-phosphoadenosine 5'-phosphosulfate; PAP, 3'-phosphoadenosine 5'-phosphote.

SULT1E1 has shown the distinct characteristic of having a high sulfating affinity for not only E_2 , but also other estrogens, such as E_1 and ethinylestradiol (EE_2), with nanomolar K_m values (<u>Table 2</u>). Due to its high affinity for sulfate estrogens, SULT1E1 exhibits inhibition of substrate with increasing E_2 and E_1 concentrations. SULT1E1 is also used to sulfate other compounds, namely dehydroepiandrosterone (DHEA), pregnenolone, diethylstilbestrol (DES), and equilenin [18][19].

Table 2. Substrates of SULT1E1.

Species	RefSeq ¹	RefSeq mRNA ²	RefSeq Protein ³	Number of Exons ¹
Homo sapiens (human)	NC_000004.12	NM_005420.3	NP_005411.1	9
Mus musculus (mouse)	NC_000071.7	NM_023135.2	NP_075624.2	8
Rattus norvegicus (rat)	NC_005113.4	NM_012883.2	NP_037015.2	10
Bos taurus (cow)	NC_037333.1	NM_177488.3	NP_803454.2	9
Oryctolagus cuniculus (rabbit)	NC_013683.1	XM_002717123.2	XP_002717169.1	8
Sus scrofa (pig)	NC_010450.4	NM_213992.1	NP_999157.1	9
Equus caballus (horse)	NC_009146.3	NM_001081918.1	NP_001075387.1	8

SULT1E1 is also expressed in hormone-dependent tissues, such as endometrium $^{[3][20]}$ and placenta $^{[2]}$. SULT1E1 is specifically expressed during the secretory phase of the menstrual cycle in human endometrium $^{[21]}$. Upregulated SULT1E1 activity in the endometrium may result in sulfating E₂ after ovulation $^{[22]}$. In addition, SULT1E1 can be induced by progestins in human Ishikawa endometrial adenocarcinoma cells $^{[23]}$.

As an interesting effect, estrogens inhibit expression of the potent growth factor repressor transforming growth factor (TGF)- β 1. In addition, it was observed that MCF-7 cells expressing SULT1E1 activity did not show a decrease in ER α levels, an increase in progesterone receptor, or a decrease in transforming growth factor- β expression, suggesting the rapid sulfoconjugation of E₂ by SULT1E1.

It is possible that SULT1E1 contributes to EE_2 sulfation during hepatic-mediated first-pass metabolism. SULT1E1 is the high-affinity enzyme responsible for EE_2 sulfation at nanomolar concentrations, so SULT1E1 plays a predominant role in the sulfation of EE_2 in the intestine and liver.

2.2. Sulfation of Thyroid Hormones

Many factors serve as regulators for the effectiveness and bioavailability of receptor active thyroid hormone (T_3) [24][25]. The prohormone thyroxine (T_4) is predominantly secreted to regulate metabolism [26][27]. Deiodination is one of the principal and major pathways to degrade active compounds, and there are three types of deiodinase selenoproteins—iodothyronine deiodinases (D1, D2, and D3) [28]. These deiodinases are promotive of the reductive T_4 deiodination and its metabolites (Figure 2).

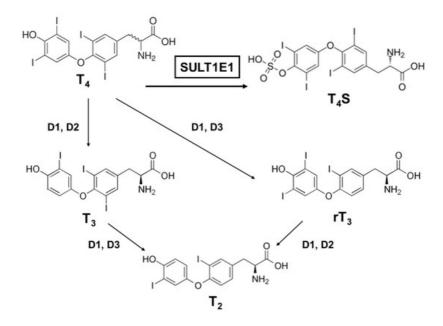


Figure 2. A schematic metabolic pathway of thyroid hormones. T_4 , thyroxine (prohormone); T_4S , thyroxine sulfate (sulfoconjugated metabolite); T_3 , 3.3',5-triiodothyronine (receptor active iodothyronine); T_3 , 3.3',5'-triiodothyronine (receptor inactive iodothyronine); T_2 , 3.3'-diiodothyronine.

One major modification thyroid hormones receive is sulfation, which deactivates them. Thyroxine sulfate can be detected in human fetal blood and amniotic fluid, indicating that the production of sulfoconjugates is critical in utero [29]. lodothyronine sulfates (T_4S , T_3S , rT_3S , and T_2S) are generated by SULT enzymes, which are located in a variety of different tissues, and catalyze the sulfation and substitution of the hydroxyl groups of various compounds using PAPS as the sulfate donor [30]. Interestingly, among the SULTs, most of the SULT1 enzymes catalyze the sulfation of iodothyronines [31][32][33][34]. SULT1E1 is highly effective at catalyzing rT_3 sulfation and has sulfating activity for all iodothyronines (Table 4). For rT_3 sulfation especially, SULT1E1 has the highest activity among the SULT1 subfamily, even compared to SULT1A1 [32]. Moreover, SULT1E1 was reported as the most active enzyme that exhibited catalyzing activity for T_4 sulfation [35].

SULT1E1 can be detected in the human endometrium and in the mouse uterus, so it might be possible that the uterus could protect the fetus from excessive thyroid hormone by inactivating pathways via SULT1E1 or D3. It is notable that the metabolites derived from D3 (rT_3 and T_2) are also favorable substrates of SULT1E1, suggesting that T_4 and T_3 are metabolized in the uterus by consecutive sulfation. The physiological roles of each iodothyronine SULT still remain too complex to be comprehended in full. Although SULT1E1 has been proven to be a potent iodothyronine SULT along with SULT1A1, it is probable that the other SULT1 enzymes contribute to iodothyronine sulfation in a tissue- or growth-dependent way $\frac{[36][37][38]}{[38]}$.

References

- 1. Her, C.; Aksoy, I.A.; Kimura, S.; Brandriff, B.F.; Wasmuth, J.J.; Weinshilboum, R.M. Human estrogen sulfotransferase gene (STE): Cloning, structure, and chromosomal localization. Genomics 1995, 29, 16–23.
- 2. Stanley, E.L.; Hume, R.; Visser, T.J.; Coughtrie, M.W. Differential expression of sulfotransferase enzymes involved in thyroid hormone metabolism during human placental development. J. Clin. Endocrinol. Metab. 2001, 86, 5944–5955.
- 3. Rubin, G.L.; Harrold, A.J.; Mills, J.A.; Falany, C.N.; Coughtrie, M.W. Regulation of sulphotransferase expression in the endometrium during the menstrual cycle, by oral contraceptives and during early pregnancy. Mol. Hum. Reprod. 1999, 5, 995–1002.
- 4. Song, W.C.; Qian, Y.; Li, A.P. Estrogen sulfotransferase expression in the human liver: Marked interindividual variation and lack of gender specificity. J. Pharmacol. Exp. Ther. 1998, 284, 1197–1202.
- 5. Li, Y.; Xu, Y.; Li, X.; Qin, Y.; Hu, R. Effects of PPAR-α agonist and IGF-1 on estrogen sulfotransferase in human vascular endothelial and smooth muscle cells. Mol. Med. Rep. 2013, 8, 133–139.
- 6. Xu, Y.; Yang, X.; Wang, Z.; Li, M.; Ning, Y.; Chen, S.; Yin, L.; Li, X. Estrogen sulfotransferase (SULT1E1) regulates inflammatory response and lipid metabolism of human endothelial cells via PPARy. Mol. Cell Endocrinol. 2013, 369, 140–149.

- 7. Wang, S.; Yuan, X.; Lu, D.; Guo, L.; Wu, B. Farnesoid X receptor regulates SULT1E1 expression through inhibition of PGC1α binding to HNF4α. Biochem. Pharmacol. 2017, 145, 202–209.
- 8. Liu, X.; Xue, R.; Yang, C.; Gu, J.; Chen, S.; Zhang, S. Cholestasis-induced bile acid elevates estrogen level via farnesoid X receptor-mediated suppression of the estrogen sulfotransferase SULT1E1. J. Biol. Chem. 2018, 293, 12759–12769.
- 9. Hu, H.; Negishi, M. ROR α phosphorylation by casein kinase 1 α as glucose signal to regulate estrogen sulfation in human liver cells. Biochem. J. 2020, 477, 3583–3598.
- 10. Gong, H.; Guo, P.; Zhai, Y.; Zhou, J.; Uppal, H.; Jarzynka, M.J.; Song, W.C.; Cheng, S.Y.; Xie, W. Estrogen deprivation and inhibition of breast cancer growth in vivo through activation of the orphan nuclear receptor liver X receptor. Mol. Endocrinol. 2007, 21, 1781–1790.
- 11. Gong, H.; Jarzynka, M.J.; Cole, T.J.; Lee, J.H.; Wada, T.; Zhang, B.; Gao, J.; Song, W.C.; DeFranco, D.B.; Cheng, S.Y.; et al. Glucocorticoids antagonize estrogens by glucocorticoid receptor-mediated activation of estrogen sulfotransferase. Cancer Res. 2008, 68, 7386–7393.
- 12. Sueyoshi, T.; Green, W.D.; Vinal, K.; Woodrum, T.S.; Moore, R.; Negishi, M. Garlic extract diallyl sulfide (DAS) activates nuclear receptor CAR to induce the Sult1e1 gene in mouse liver. PLoS ONE 2011, 6, e21229.
- 13. Yi, M.; Fashe, M.; Arakawa, S.; Moore, R.; Sueyoshi, T.; Negishi, M. Nuclear receptor CAR-ERalpha signaling regulates the estrogen sulfotransferase gene in the liver. Sci. Rep. 2020, 10, 5001.
- 14. Hu, H.; Yokobori, K.; Negishi, M. PXR phosphorylated at Ser350 transduces a glucose signal to repress the estrogen sulfotransferase gene in human liver cells and fasting signal in mouse livers. Biochem. Pharmacol. 2020, 180, 114197.
- 15. Fashe, M.; Hashiguchi, T.; Yi, M.; Moore, R.; Negishi, M. Phenobarbital-induced phosphorylation converts nuclear receptor RORalpha from a repressor to an activator of the estrogen sulfotransferase gene Sult1e1 in mouse livers. FEBS Lett. 2018, 592, 2760–2768.
- 16. Poisson Paré, D.; Song, D.; Luu-The, V.; Han, B.; Li, S.; Liu, G.; Labrie, F.; Pelletier, G. Expression of Estrogen Sulfotransferase 1E1 and Steroid Sulfatase in Breast Cancer: A Immunohistochemical Study. Breast Cancer 2009, 3, 9–21.
- 17. Song, W.C.; Melner, M.H. Steroid transformation enzymes as critical regulators of steroid action in vivo. Endocrinology 2000, 141, 1587–1589.
- 18. Falany, C.N.; Wheeler, J.; Oh, T.S.; Falany, J.L. Steroid sulfation by expressed human cytosolic sulfotransferases. J. Steroid Biochem. Mol. Biol. 1994, 48, 369–375.
- 19. Falany, C.N.; Comer, K.A.; Dooley, T.P.; Glatt, H. Human dehydroepiandrosterone sulfotransferase. Purification, molecular cloning, and characterization. Ann. N. Y. Acad. Sci. 1995, 774, 59–72.
- 20. Buirchell, B.J.; Hahnel, R. Metabolism of estradiol-17beta in human endometrium during the menstrual cycle. J. Steroid Biochem. 1975, 6, 1489–1494.
- 21. Falany, J.L.; Azziz, R.; Falany, C.N. Identification and characterization of cytosolic sulfotransferases in normal human endometrium. Chem. Biol. Interact. 1998, 109, 329–339.
- 22. Kotov, A.; Falany, J.L.; Wang, J.; Falany, C.N. Regulation of estrogen activity by sulfation in human Ishikawa endometrial adenocarcinoma cells. J. Steroid Biochem. Mol. Biol. 1999, 68, 137–144.
- 23. Falany, J.L.; Falany, C.N. Regulation of estrogen sulfotransferase in human endometrial adenocarcinoma cells by progesterone. Endocrinology 1996, 137, 1395–1401.
- 24. Moreau, X.; Lejeune, P.J.; Jeanningros, R. Kinetics of red blood cell T3 uptake in hypothyroidism with or without hormonal replacement, in the rat. J. Endocrinol. Investig. 1999, 22, 257–261.
- 25. Kim, S.Y.; Seo, S.; Choi, K.H.; Yun, J. Evaluation of phototoxicity of tattoo pigments using the 3 T3 neutral red uptake phototoxicity test and a 3D human reconstructed skin model. Toxicol. In Vitro 2020, 65, 104813.
- 26. Leiner, K.A.; Mackenzie, D.S. Central regulation of thyroidal status in a teleost fish: Nutrient stimulation of T4 secretion and negative feedback of T3. J. Exp. Zool. Comp. Exp. Biol. 2003, 298, 32–43.
- 27. Mihasan, M.; Brandsch, R. A predicted T4 secretion system and conserved DNA-repeats identified in a subset of related Arthrobacter plasmids. Microbiol. Res. 2016, 191, 32–37.
- 28. St Germain, D.L.; Galton, V.A.; Hernandez, A. Minireview: Defining the roles of the iodothyronine deiodinases: Current concepts and challenges. Endocrinology 2009, 150, 1097–1107.
- 29. Wu, S.Y.; Huang, W.S.; Polk, D.; Florsheim, W.H.; Green, W.L.; Fisher, D.A. Identification of thyroxine-sulfate (T4S) in human serum and amniotic fluid by a novel T4S radioimmunoassay. Thyroid 1992, 2, 101–105.

- 30. Chatterjee, B.; Song, C.S.; Kim, J.M.; Roy, A.K. Androgen and estrogen sulfotransferases of the rat liver: Physiological function, molecular cloning, and in vitro expression. Chem. Biol. Interact. 1994, 92, 273–279.
- 31. Kester, M.H.; van Dijk, C.H.; Tibboel, D.; Hood, A.M.; Rose, N.J.; Meinl, W.; Pabel, U.; Glatt, H.; Falany, C.N.; Coughtrie, M.W.; et al. Sulfation of thyroid hormone by estrogen sulfotransferase. J. Clin. Endocrinol. Metab. 1999, 84, 2577–2580.
- 32. Kester, M.H.; Kaptein, E.; Roest, T.J.; van Dijk, C.H.; Tibboel, D.; Meinl, W.; Glatt, H.; Coughtrie, M.W.; Visser, T.J. Characterization of human iodothyronine sulfotransferases. J. Clin. Endocrinol. Metab. 1999, 84, 1357–1364.
- 33. Wang, J.; Falany, J.L.; Falany, C.N. Expression and characterization of a novel thyroid hormone-sulfating form of cytosolic sulfotransferase from human liver. Mol. Pharmacol. 1998, 53, 274–282.
- 34. Li, X.; Clemens, D.L.; Anderson, R.J. Sulfation of iodothyronines by human sulfotransferase 1C1 (SULT1C1). Biochem. Pharmacol. 2000, 60, 1713–1716.
- 35. Kester, M.; Coughtrie, M.W.; Visser, T.J. Sulfation of Thyroid Hormones. In Human Cytosolic Sulfotransferases; Pacifici, G.M., Coughtrie, M.W., Eds.; CRC Press/Taylor & Francis Group: Boca Raton, FL, USA, 2005; pp. 121–134.
- 36. Glatt, H.; Boeing, H.; Engelke, C.E.; Ma, L.; Kuhlow, A.; Pabel, U.; Pomplun, D.; Teubner, W.; Meinl, W. Human cytosolic sulphotransferases: Genetics, characteristics, toxicological aspects. Mutat. Res. 2001, 482, 27–40.
- 37. Weinshilboum, R.M.; Otterness, D.M.; Aksoy, I.A.; Wood, T.C.; Her, C.; Raftogianis, R.B. Sulfation and sulfotransferases 1: Sulfotransferase molecular biology: cDNAs and genes. FASEB J. 1997, 11, 3–14.
- 38. Kiehlbauch, C.C.; Lam, Y.F.; Ringer, D.P. Homodimeric and heterodimeric aryl sulfotransferases catalyze the sulfuric acid esterification of N-hydroxy-2-acetylaminofluorene. J. Biol. Chem. 1995, 270, 18941–18947.

Retrieved from https://encyclopedia.pub/entry/history/show/27707