

GABAAR-Mediated Currents

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Propofol or barbiturate application at low concentrations increases desensitization and slows deactivation of GABA-induced current and propofol/barbiturate at high concentrations directly elicits after-responses upon their washout in hippocampal or sensory neurons. It is postulated that the generation of such after-responses is caused by removal of the blockade by anesthetic agents as partial antagonists. However, the increased desensitization was invariably followed by slowdown of deactivation of GABA-induced current, and the after-response may arise as a consequence of extreme slowdown of deactivation following strong desensitization. It is thus possible that propofol and barbiturate can facilitate resensitization of GABA responses. Propofol and barbiturate are useful to treat the alcohol/benzodiazepine withdrawal syndrome. Considering that the slowdown of deactivation following desensitization and the after-response induced by propofol or barbiturate application, the regulatory mechanisms of desensitization/resensitization of GABA_AR-mediated currents might be important for understanding the treatment of the alcohol/benzodiazepine withdrawal syndrome.

GABA

Desensitization

Resensitization

1.Alcohol/Benzodiazepine Withdrawal Syndrome and its Treatment by Modulating Desensitization/Resensitization Kinetics of GABA_AR-Mediated Currents

Alcohol and benzodiazepine are useful to mitigate anxiety through enhancing GABA_AR-mediated inhibition. However, alcohol and benzodiazepine are known as abused drugs. The alcohol or benzodiazepine withdrawal syndrome appears following a reduction on alcohol or benzodiazepine use after a period of excessive use [\[1\]\[2\]\[3\]\[4\]](#). The alcohol or benzodiazepine withdrawal symptoms typically include anxiety, sweating, hand tremor and sleep disturbance. The underlying mechanisms involve neuronal adaptations, which are revealed as decreased GABAergic responses [\[5\]](#) and enhancement of NMDA responses [\[6\]\[7\]\[8\]\[9\]](#). Although the exact mechanism for the reduced responsiveness of GABA_ARs remains uncertain, changes in surface GABA_AR protein level and subunit composition, changes in turnover, recycling, and production rates, degree of phosphorylation and decreased coupling mechanisms between GABA and alcohol/benzodiazepine sites are thought to be involved in the reduced responsiveness [\[10\]\[11\]\[12\]\[13\]](#). It has recently been demonstrated that the benzodiazepine diazepam caused downregulation of GABAergic inhibition through phospholipase C (PLC δ)/Ca²⁺/calcineurin signaling pathway [\[14\]](#). The study showed that overexpression of PRIP-1 suppressed diazepam-dependent activation of PLC δ and diazepam-dependent downregulation of GABA_ARs in HEK293 cells [\[14\]](#), indicating that PRIP-1 acts as an inhibitor by outcompeting the PLC δ binding to GABA_ARs. Because intracellular Ca²⁺ and calcineurin activity are increased in PRIP-DKO mice [\[15\]](#), these findings suggest that the diazepam-induced long-term downregulation of GABAergic inhibition is mediated by PLC δ /Ca²⁺/calcineurin signaling pathway. Nevertheless, it is also true that calcineurin

causes resensitization of GABA_AR-mediated currents by facilitating their desensitization [15][16]. Given the apparently contradictory behaviors of GABA_AR-mediated currents by calcineurin activation, the two different behaviors of GABA_AR-mediated currents may depend on whether calcineurin activation occurs before or after activation of GABA_ARs.

As for the treatment of the alcohol/benzodiazepine withdrawal syndrome, propofol and barbiturate which enhance GABA_AR-mediated inhibition are useful. Indeed, it was demonstrated that propofol and barbiturates (pentobarbital and phenobarbital) were effective for the treatment of alcohol withdrawal syndrome [17][18], and barbiturate (pentobarbital) was effective for the treatment of benzodiazepine withdrawal syndrome [19]. However, it remains unclear how propofol and barbiturate ameliorate reduced GABA responsiveness in patients with alcohol/benzodiazepine withdrawal syndrome. Although the concentrations of propofol and barbiturates that generated the hump-like current are very high [20][21][22] compared to the dose used for treatment of the withdrawal syndrome [17][18], the generation of hump-like GABA_AR currents itself may suggest the occurrence of resensitization of GABA_AR-mediated currents. Indeed, the desensitization and deactivation of GABA_AR-mediated currents are facilitated and slowed, respectively, by propofol/barbiturate at much lower concentrations [21][22]. Then, propofol and barbiturate may improve the reduced GABA responsiveness through the resensitization of GABA_AR-mediated currents. Therefore, the regulatory mechanisms of desensitization/resensitization of GABA_AR-mediated currents are important to better understand the benzodiazepine/alcohol withdrawal syndrome and to develop the treatment method.

2. Drugs that Cause Desensitization and Resensitization of GABA_AR-Mediated Currents

The drugs that cause desensitization and resensitization of GABA_AR-mediated currents are summarized in Table 1. Hump-like GABA_AR currents after a strong desensitization were seen at the offset of propofol application at a high concentration (600 μ M) in hippocampal pyramidal neurons [22], etomidate application at a high concentration (1 mM) in rat spinal dorsal horn neurons [23], pentobarbital application at high concentrations (1–3 mM) in frog sensory neurons [21][22], rat hippocampal neurons [24] and recombinant GABA_ARs [25][26][27][28][29][30] or phenobarbital application at a high concentration (10 mM) in rat hippocampal neurons [24], although these were not seen at the offset of GABA application. It is believed that the generation of hump-like currents may be caused by removal of the blockade by anesthetic agents as partial antagonists [25], although their mechanisms remain unclear and the involvement of desensitization is not necessarily denied. To better understand the actions of these anesthetic drugs, the molecular and regulatory mechanisms of desensitization/resensitization of GABA_AR-mediated currents might be important.

Table 1. Drugs that modulate GABA responses and directly activate GABA_ARs at higher concentrations.

Drugs	Neurons/ Recombinant GABA _A Rs	Effects	Refs.
Anesthetics			
Propofol	Mouse hippocampal neurons	slows deactivation and increases apparent desensitization of GABA responses at low concentrations and directly elicits after-responses upon washout at high concentrations	[22]
Etomidate	Rat spinal dorsal horn neurons	slows deactivation of GABA responses at low concentrations while directly elicits tail currents upon washout at high concentrations	[23]
Barbiturate			
Pentobarbital	Frog sensory neurons	slows deactivation and increases apparent desensitization of GABA responses at low concentrations, and directly elicits hump currents upon washout at high concentrations	[20, 21]
	Rat hippocampal neurons	slows deactivation and increases apparent desensitization of GABA responses at low concentrations, and directly elicits rebound currents upon washout at high concentrations	[24]
	$\alpha 1\beta 2\gamma 2L$	directly elicits tail currents upon washout at high concentrations	[25, 27]
	$\alpha 1\beta 3\gamma 2L$	slows deactivation and increases apparent desensitization of GABA responses at low concentrations, and directly elicits rebound currents upon washout at high concentrations	[26]
	$\alpha 1\beta 2\gamma 2S$, $\alpha 6\beta 2\gamma 2S$	directly elicits hump currents upon washout at high concentrations	[28]
	$\beta 3$	increases apparent desensitization of GABA responses and directly elicits rebound currents upon washout at high concentrations	[29]
Phenobarbital	$\alpha 1\beta 3\gamma 2L$	directly elicits tail currents upon washout at high concentrations	[30]
	Rat hippocampal neurons	slows deactivation and increases apparent desensitization of GABA responses at low concentrations, and directly elicits rebound currents upon washout at high concentrations	[24]

3. A possible Kinetic Mechanism for Resensitization of GABA_A Currents

In layer II/III pyramidal cells of the mouse barrel cortex, the deletion of PRIP-1/2 enhanced the desensitization of GABA_AR-mediated currents but paradoxically induced a hump-like tail-current at the offset of the GABA puff [15], and also resulted in the prolongation of the decay phase of eIPSCs [31]. To understand the kinetic mechanisms

underlying the generation of the hump-like tail-currents and the prolongation of eIPSCs, these currents were simulated using a model previously proposed [32] (Figure 1). It was examined whether the possible increase in the fast desensitization rate (d_2) and the possible decrease in the unbinding rate (k_{off}) can lead to the generation of the hump-like tail-current at the offset of the GABA puff. In the simulated wild-type pyramidal cell, GABA_AR-mediated currents were induced without a hump-like tail-current in response to 2-sec GABA puff at 50 μM [15]. In contrast, in the simulated PRIP-DKO pyramidal cell, GABA_AR-mediated currents displayed a prominent desensitization and was followed by a prominent hump-like tail-current [15]. Thus, a slowdown of k_{off} and an acceleration of d_2 resulted in a generation of a hump-like tail-current. Following a sharp decrease in [GABA] at the offset of GABA puff, a sharp decrease in d_2 to a level smaller than the fast de-desensitization (i.e. resensitization) rate constant (r_2) occurred to subsequently induce a hump-like tail-current. It can be concluded that a higher calcineurin activity in PRIP-DKO layer III pyramidal cells might have caused a slowdown of k_{off} and an acceleration of d_2 through the modulation of its GABA concentration dependency, leading to a generation of hump-like tail-currents in PRIP-DKO pyramidal cells. It was also investigated whether the increase in d_2 and the decrease in k_{off} can also lead to the prolongation of eIPSCs. Simulated IPSCs in PRIP-DKO and the wild-type pyramidal cells that have the half-durations similar to those obtained in the real experiments [15] revealed that a prolongation of eIPSCs/eIPSPs in PRIP-DKO pyramidal cells results from resensitization of GABA_AR-mediated currents, which is brought about by an acceleration of d_2 through the modulation of its [GABA] dependency together with a slowdown of k_{off} .

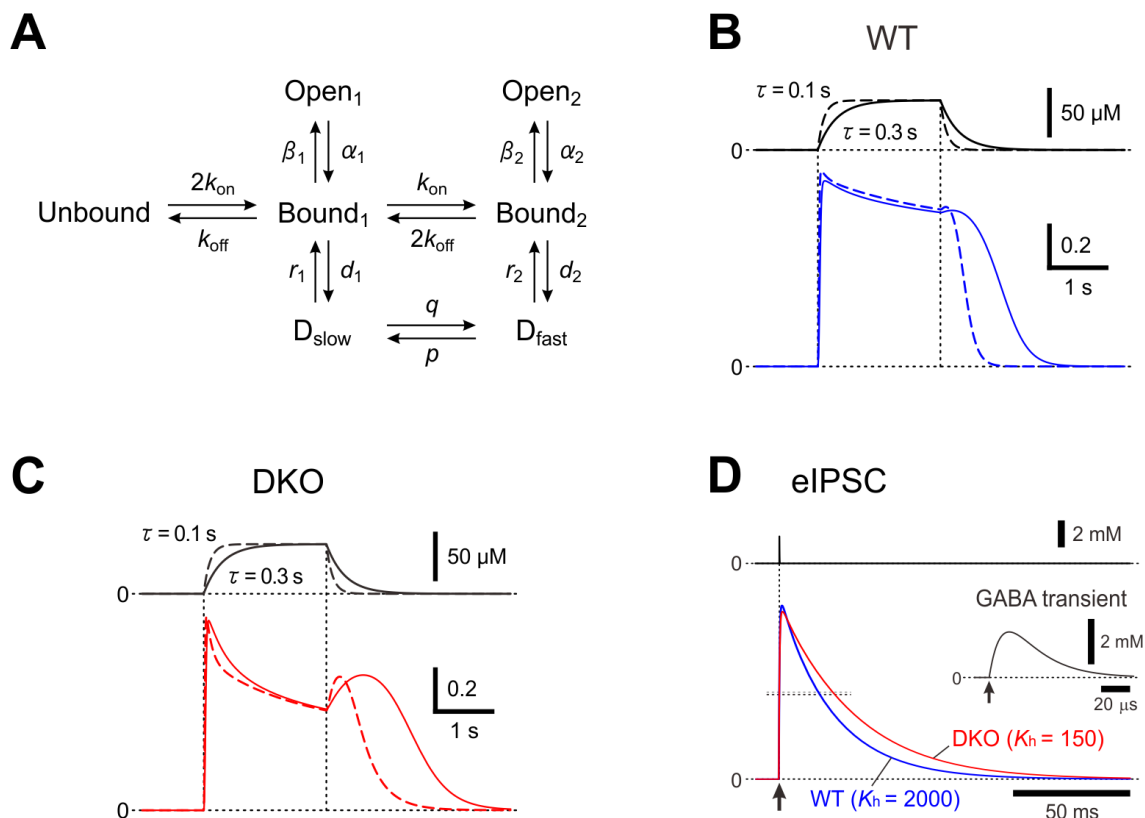


Figure 1. A kinetic model for a hump-like tail-current. (A) A kinetic model of GABA_ARs representing mono- and double-liganded states, each providing access to open and desensitized states. (B and C) Top; Presumed [GABA] changes created by puff application of GABA with a rectangular pressure pulse through a puff pipette containing 200 μM GABA in the extracellular medium was assumed to be diluted 4 times, and the onset and offset of the puff

application were assumed to be attenuated with a time constant ranging between 0.1 and 0.3 sec. Bottom; superimposed traces of the simulated GABA_AR-mediated currents under the condition that the attenuation time constant is 0.3 and 0.1 sec (solid and interrupted traces, respectively) in simulated wild-type (B) and PRIP-DKO (C) pyramidal cells. (D) Superimposed traces of a simulated wild-type and PRIP-DKO eIPSC induced by a GABA transient shown on an expanded time scale (inset) with a small maximum conductance. Adopted from [15] and [31].

Based on the experimental and simulation studies, the regulatory mechanisms of GABA_ARs are schematically depicted (Figure 2).

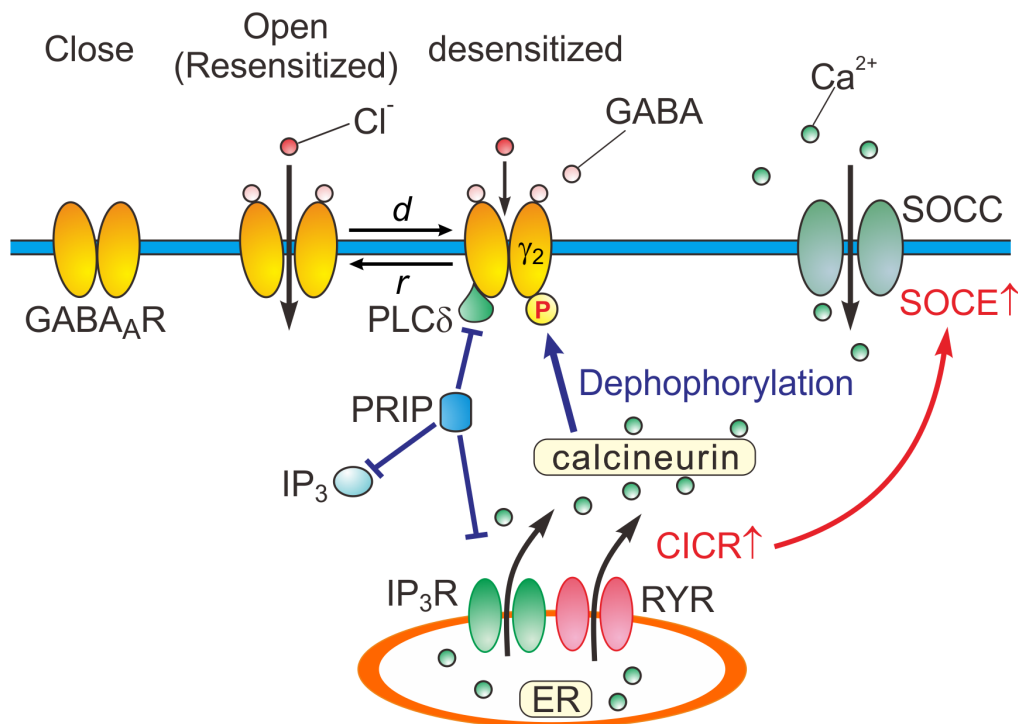


Figure 2. Close, open (resensitized) and desensitized states of GABA_ARs. When GABA binds to GABA_ARs, the receptors open the pore, and consequently increasing the permeability of the ion pore to Cl⁻. In response to a prolonged application of GABA, GABA_ARs are desensitized (*d*) by increased calcineurin activity due to potentiated Ca²⁺-induced Ca²⁺ release (CICR) followed by store-operated Ca²⁺ entry (SOCE) [15]. GABA_ARs are resensitized through the de-desensitization (*r*) at the offset of the GABA puff. PRIP outcompetes the PLCδ in binding to GABA_A β subunits [14]. *d*: desensitization, *r*: resensitization, RYR: ryanodine receptor, SOCC: store-operated Ca²⁺ channel, IP₃R: inositol trisphosphate receptor.

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