Polyphenols and Visual Health

Subjects: Chemistry, Medicinal

Contributor: Pere Garriga, Pol Fernandez Gonzalez, Aina Mas-Sanchez

Dietary polyphenols are a group of natural compounds that have been proposed to have beneficial effects on human health. They were first known for their antioxidant properties, but several studies over the years have shown that these compounds can exert protective effects against chronic diseases. Nonetheless, the mechanisms underlying these potential benefits are still uncertain and contradictory effects have been reported. The effect of these compounds on visual health, and particularly on retinal degenerative diseases, is a matter of renewed interest and recent studies show promising results for the use of these compounds to improve visual function.

Keywords: flavonoids; retinal degenerative diseases; retinitis pigmentosa; protein folding; ligand binding; rhodopsin

1. Introduction

Different studies have reported that dietary polyphenols exert protective and beneficial effects against chronic diseases such as neurodegenerative and cardiovascular diseases, cancer, and diabetes [1]; however, the mechanisms underlying these benefits are far from being completely understood and more research is needed in order to define them. Despite all the benefits polyphenols can provide, they have low oral bioavailability, and other properties like their physicochemical stability, gastrointestinal absorption, and metabolism are important to ensure an effective action [2][3].

Despite the existent gap of knowledge in their action mechanism, the World Health Organization has recommended to increase the intake of fruit, vegetables, and fiber due to the high number of plant-derived components [4] with polyphenols having an important role, since they may confer health benefits related to non-communicable diseases (NCDs) [5][6]. Although associating polyphenols with specific diseases is challenging [7], some promising results have been obtained in different observational studies regarding polyphenols and certain NCDs [8][9], including some visual diseases. For this reason, the implication of polyphenols in health and disease states needs to be studied and better defined because of the expected positive impact on human health.

2. Polyphenols as Repurposed Drugs

Polyphenols or dietary phenolic compounds are known as the largest group of phytochemicals [10] and are a group of natural compounds sharing common structural features (Figure 1). Different lines of evidence, derived from sustained work in the last several years, provide support for an important role for polyphenols both in helping maintain a healthy lifestyle and in the prevention of prevalent diseases like cancer, cardiovascular and neurodegenerative diseases [11][12][13]. Specifically, several studies have suggested that the consumption of different polyphenols from natural sources such as fruit and vegetables can contribute to preserving vision and can even reverse visual impairment in certain visual disorders [14][15]

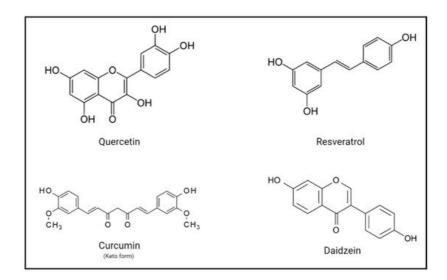


Figure 1. Structures of representative and abundant polyphenols from different subfamilies. Quercetin is a typical flavonoid found in many products. Resveratrol is a natural polyphenolic phytoalexin. Curcumin is derived from the rhizome of turmeric and is usually found in its keto form. Finally, daidzein is one of the most common isoflavones.

The polyphenol superfamily includes a large number of sub-families, among which we can find flavonoids, phenolic acids, stilbenes, and lignans $^{[16]}$. One of the most studied groups, from these different sub-classes, is that of flavonoids, comprising over 4000 members $^{[17]}$. Flavonoids have a characteristic structure of a 15-carbon skeleton of a chromane ring attached to another aromatic ring $^{[18]}$. The biosynthesis of these complex polyphenols is linked to primary metabolism $^{[10]}$. Flavonoids are stored, in their native state, in plants as glycoside and non-glycosylated conjugates and can be absorbed by the small intestine and readily metabolized, once ingested, by phase II enzymes. After this biochemical process, the resulting moieties can enter systemic circulation $^{[19][20]}$.

One of the main proposed biological actions of polyphenols is associated with their antioxidant power within living cells; however, detailed investigations indicate that these effects, in many tissues, may not be as relevant as previously suggested. This is due to the fact that in many tissues it is difficult for these compounds to reach the threshold concentration needed to exert any significant biological effect [21][22]. Nonetheless, recent studies have suggested that polyphenols may have significant effects on human health, such as anti-inflammatory, anti-microbial, and tumor-suppressing properties [23][24][25].

The diversity of polyphenolic compounds of natural origin, their chemical lability, and their complex bioavailability patterns consequently necessitates stringent evaluation of the physiological effects of these compounds, and such evaluations are not always available. These evaluations are absolutely needed for later use in therapeutic applications.

3. Implications and Potential Benefits of Polyphenols on Human Health

As already discussed, polyphenols have been well characterized for their antioxidant effects, but their physiological relevance has been questioned due to the limited bioavailability that renders relatively low concentrations which may hamper achieving significant in vivo effects $\frac{[21][22]}{[21]}$; however, different alternative molecular mechanisms in which polyphenols appear to have a role have been identified and this gives these compounds another set of properties that may represent benefits for human health. These include different actions both at the intra- and inter-cellular signaling pathways levels, like, for example, regulating nuclear transcription factors and fat metabolism and modulating the synthesis of inflammatory mediators like cytokines, tumor necrosis factor α , interleukin-1 β , and interleukin-6 $\frac{[26][27]}{[27]}$. As a general overview, different studied flavonoids have been shown to play different roles in cellular processes, such as increasing insulin secretion, reducing apoptosis, promoting β -cell proliferation, and reducing inflammation and oxidative stress in some cells $\frac{[28]}{[28]}$. All these effects play a role in different processes, such as glucoregulation, and show that flavonoids can have favorable effects on diabetes and obesity prevention and control $\frac{[29][30][31]}{[29][30][31]}$.

Despite all the potential beneficial effects of polyphenols proposed to date, a key aspect to consider concerns the effective concentration in the human body of these compounds and the amount of natural food that needs to be consumed to reach such a concentration. One of the problems appears if the physiologically active concentration cannot be reached with common food ingestion, in this case, dietary supplementation or pharmacological approaches may be necessary and this can lead to an increase in adverse secondary effects that would require a strict follow-up of the intake and a proper control of the dose regime [30].

Polyphenols are mainly provided by the intake of different food sources, such as coffee, tea, cocoa, and apples and they have been associated with several potential health benefits [5][6][11][14][32][33][34][35][36]. In fact, polyphenols have been mechanistically involved in glucose metabolism, platelet function, endothelial function, blood pressure, inflammation, and cholesterol levels, among others [37][38]. This variety of cellular functions that may be affected by the action of polyphenols provides an excellent platform for the development of effective health prevention strategies as well as novel therapeutic approaches not only for prevalent NCDs but even for genetic hereditary rare pathological conditions [5][6].

Some of the evidence regarding the beneficial effect of polyphenols on human health comes from observational studies and this implies taking several factors into consideration when extracting meaningful conclusions when interpreting experimental data. In fact, observational studies regarding this matter should be complemented and supported by rigorous and wide clinical studies that evaluate the hypothesis that dietary phenolics have a positive role in improving human health and preventing disease states [28].

In addition to all the effects on cardiometabolic health, polyphenols are also thought to have a beneficial role on cognitive function. For example, some longitudinal studies show that regular dietary chocolate consumption can reduce the risk of cognitive decline [39][40]. Studies of other food sources like tea show that its consumption can help lowering the risk of cognitive impairment, reduce the risk of depression, and have protective effects against some diseases like Parkinson's disease [41][42][43][44].

Dietary flavonoids can also have beneficial effects in retinal degenerative diseases like retinitis pigmentosa, where mutations in retinal proteins can cause photoreceptor cell death and vision loss, eventually leading to blindness. In fact, the flavonoid quercetin was found to have an effect on the conformational stability and function of the visual G protein-coupled receptor (GPCR) rhodopsin (Rho) $^{[45]}$, suggesting that quercetin can have a positive effect on the stability and conformational properties of Rho mutants. This effect on retinal Rho suggests an effect at the receptor level that deserves further investigation. These results open a new frame of possibilities to use this and other flavonoids, possibly in combination with specific retinoids, in order to treat retinal degeneration associated with RP. This strategy could also be used to overcome the mutational effect associated with different pathological conditions in other members of the GPCR superfamily $^{[45]}$.

Polyphenolic compounds, and particularly flavonoids, are good prospects for treating or ameliorating the progression of human diseases, in addition to their established antioxidant potential that is considered important as part of a healthy lifestyle.

4. Vertebrate Rho and Retinal Degeneration

Photoreceptor cells are primary sensory neurons in the retina that detect light and convert this energy into nerve impulses that lead to visual perception in the brain. Light absorption occurs in the two types of photoreceptor cells present in the retina, rods, and cones, and this process is mediated by the visual pigments contained in them. The main photoreceptor protein present in the retina is Rho [46][47]. Both Rho and cone opsins belong to the family of GPCRs and are composed of an apoprotein, opsin, and a chromophore, namely, 11-cis-retinal (11CR) [48][49][50].

4.1. Rho as a GPCR

GPCRs are membrane proteins consisting of a single polypeptide chain structured in a helical architecture. All members of the GPCRs superfamily share a common structure with seven hydrophobic helical segments connected by three extracellular and three intracellular loops. A fourth loop is formed by joining the C-terminal segment and the lipid bilayer through cysteine palmitoylation [51]. The *N*-terminal part and extracellular loops recognize a wide variety of ligands and modulate their binding to the receptor. The seven transmembrane segments form the structural nucleus and transduce extracellular signals into the internal domain by conformational changes. The intracellular part interacts with cytosolic G proteins, arrestins, GPCR kinases, and other signaling effectors [52][53][54].

GPCRs respond to a large number of endogenous allosteric modulators which regulate receptor function by binding to alternative regions of the conventional orthosteric site. While also permitting the binding of orthosteric ligands, they can modulate the affinity and efficacy of the orthosteric ligand [55]. Orthosteric and allosteric ligands that act on the same GPCR can participate in different regulatory and signaling pathways by interacting with effectors and regulatory proteins. Both can select different parts of these signaling and regulation pathways by establishing different receptor conformations, a phenomenon called functional selectivity. The pathways where the ligand—receptor complex is involved will determine the physiological effects of the ligand [56][57][58].

There is a growing interest in ligands that bind to allosteric sites, as they may be potentially more selective than orthosteric ligands. This is because their binding occurs in less conserved regions, making them promising therapeutic substances with a lower risk of overdose and fewer adverse effects [51].

Rho is the prototypical GPCR of the human retina that mediates dim light vision. It absorbs a quantum of light and converts it into an electrical signal that is transmitted to the brain by means of the visual phototransduction process. This process involves a specific G protein, transducin, and Rho kinase and arrestin, among other proteins that are key players in the correct functioning of the visual system [59].

4.2. Visual Phototransduction

Rho converts photons into chemical signals that can trigger biological processes by allowing the brain to perceive light stimuli $^{[59]}$. Rho is bound to 11CR in its dark-adapted (ground-state inactive) conformation. 11CR is a derivative of vitamin A that has a very fast response and a high quantum yield upon light absorption in its isomerization reaction to all-*trans*-retinal (ATR). The 11CR chromophore is covalently attached via a protonated bond from the Schiff base to K296 at the seventh transmembrane helix of Rho $^{[60]}$. The transduction of signals in the visual system comprises two processes: (i) the activation of Rho by a photon of light that leads to a conformational change and (ii) a deactivation step, or signal shut-off, that involves Rho kinase and arrestin to eventually regenerate Rho to its original inactive dark state $^{[61][62]}$.

In vertebrates, the visual signal begins with the absorption of photons by 11CR that cause the isomerization of the 11-12 double bond to yield the ATR stereochemical configuration $^{[49][63]}$. Complete chromophore isomerization causes a change in the conformation of the protein, making the coupling to opsin less energetically favorable and promoting ATR release from the retinal binding pocket. This active photoilluminated conformation, termed metarhodopsin II (meta II), activates the signal transduction process by binding to the heterotrimeric G-protein transducin (Gt) and activating it by promoting the dissociation of α from the $\beta\gamma$ subunits. This, in turn, activates a cyclic guanosine monophosphate (cGMP) phosphodiesterase that hydrolyzes cGMP causing the closure of the ion channels of the membrane and the subsequent hyperpolarization of the cell (Figure 2) $^{[62][64][65]}$. The potential difference in photoreceptor cells is transferred through the synaptic terminal to second-order neurons of the retina $^{[66]}$.

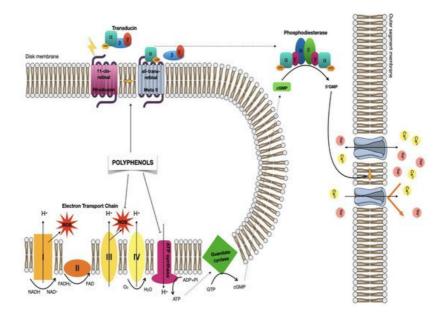


Figure 2. Visual phototransduction. Rho activates light causing isomerization of 11CR to ATR. The active form of Rho (meta II) interacts with Gt (composed of the α , β , and γ subunits) activating it and causing the exchange of GDP for GTP and the dissociation of the α subunit. This, in turn, activates cyclic guanosine monophosphate phosphodiesterase (cGMP), promoting the hydrolysis of cGMP and its conversion to 5'-GMP. Reduction in cytoplasmic cGMP concentration leads to closure of transmembrane channels by blocking the internal flow of Na⁺ and Ca²⁺ and leading to the hyperpolarization of the cell [62][64][65]. The electron transport chain and the ATP synthase present in the rod photoreceptors disks produce energy needed for the process, this energy production results in the production of reactive oxygen species (ROS) which can lead to cell damage. [67]. Polyphenols can act in different ways at the cellular level, they seem to be able to stabilize mutated Rho, can inhibit the ATP synthase or help prevent the ROS damage with their antioxidative effect.

After the activation of Rho, a constant supply of 11CR is required. This is obtained from the retinoid cycle, which is an enzymatic pathway occurring at the photoreceptors and the retinal pigment epithelium (RPE). The process re-isomerizes

Both the function and integrity of photoreceptors are crucial to vision. Mutations that affect the function of these receptors, or other factors that can alter the phototransduction process, can cause visual dysfunction or a loss of vision. Defects in other types of retinal cells (such as RPE) can also cause visual cycle dysfunction [66][69][70]. There is a high concentration of Rho in the retina, so intense light can cause a local concentration of free ATR that is toxic to cells, which can cause severe retinal degeneration and even eventually lead to complete blindness [46][69][71]. In addition, when 11CR is not effectively recombined with opsin, high concentrations of non-regenerated opsin can promote and enhance retinal degeneration processes [72][73][74][75][76].

The death of photoreceptors caused by persistent exposure to high light intensities is associated with changes in cellular metabolism and the overproduction of reactive oxygen species (ROS) that can cause cell damage [77][78]. As a result, apoptotic pathways are activated, resulting in the death of photoreceptor cells [78]. Furthermore, the toxic effect produced by light results in the expression of pro-inflammatory chemokines, thus stimulating the migration of macrophages and microglia towards photoreceptor cells [79][80]. Unbalanced homeostasis is the main mechanism that contributes to degenerative retinal disorders [81]. Furthermore, photoreceptors have been shown to present machinery for oxidative phosphorylation in their outer segments, including the electron transport chain and ATP synthetase [67][82][83]. It is believed that this machinery, typical of mitochondria, would be used for the energy needs of visual phototransduction. The increase in the demand for ATP increases the consumption of oxygen, thus increasing ROS production, which in turn causes oxidative stress. The retina is sensitive to oxidative stress and such stress can contribute to diseases such as AMD [83][84]. Polyphenolic compounds can help prevent photoreceptor cell damage caused by ROS, and thus they can have beneficial effects on visual function in retinal degenerative diseases.

The correct function of Rho depends on the correct expression, folding, trafficking, and integration into the lipid bilayer of the cell membrane [85]. Attempts are currently being made to find new ligands that can offset the effects of Rho mutations that can cause retinal diseases, particularly RP.

4.3. Mutations in Rho Associated with Retinal Degenerative Diseases

In the mammalian genome, the ciliary opsin family is made up of different genes, including the Rho gene (RHO), which consists of 5 exons that codify a 348-amino acid protein with a molecular weight of approximately 39 kDa [86].

There are two diseases associated with mutations in RHO: congenital stationary night blindness (CSNB) and RP. In the case of CSNB, it is inherited in a dominant way and the term stationary in its name has been questioned because it appears that night blindness could be the first step of a very slowly progressing RP. RP can be inherited both in dominant and recessive ways, although most of the diseases causing mutations are dominant and the recessive phenotype is rare [86]

The first mutation causing RP in the RHO gene was reported at position 23, involving a change from a proline to a histidine (P23H) $^{[87]}$ (a Rho model indicating the site of RP mutations is shown on <u>Figure 3</u>).

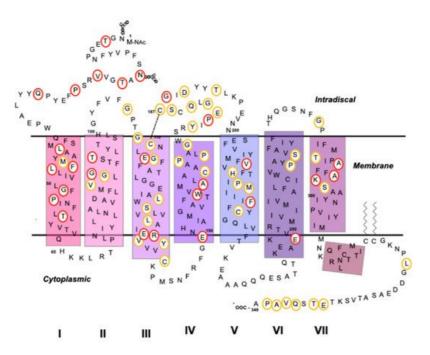


Figure 3. Secondary structure schematic model of Rho, showing amino acids that are sites where mutations associated with RP in patients are found. Sites of RP mutations are circled and those specific positions corresponding to mutations mentioned in the text are circled in red.

Five CSNB-associated missense RHO mutations have been identified: G90D [88][89], T94] [90] E113K [91], A292E [92], and A295V [93]. They are thought to produce a constitutive activation of Rho (except for the E113K mutation) [94]. CSNB mutants have been studied by X-ray crystallography and it has been found that a new salt bridge is formed between the aspartate residue of the G90D mutant and K296 at the retinal binding site. This bridge, and the concomitant breakage of the native salt bridge between E113 and K297, could be the reason for the increased basal activation of this mutant [95]. The constitutive activation of mutants means that they present activity in the absence of the retinal chromophore. Constitutive activity is also referred to Rho activation in the dark, although, in this case, the more precise term of dark activity should be used. The constitutive persistent activation of the phototransduction cascade has been considered a mechanism of cell death in RP [94]. The mutant G90D, which is the cause of CSNB and is a constitutively active mutant, activates the visual cascade without chromophore and in the dark. Interestingly, another mutation at position G90 (G90V) is associated with RP [71][89]. It is an unsolved puzzle why mutations at the same amino acid site cause such distinct clinical phenotypes. The molecular basis of such a striking difference could be related to the stability of quasi-native conformations of opsin (caused by mutations) that would not reflect protein misfolding but rather would affect the conformation equilibrium between active and inactive conformational states [94].

Some mutations that can constitutively activate transducin are the K296E (for RP) [96][97] along with A292E and T94I (for CSNB) [90][91]. They can do this by disrupting the salt bridge between E113 and K296. Moreover, the T94I CSNB mutant has a hydrophobic side chain that establishes contact with K296 and prolongs the useful lifetime of the active conformation by showing a longer-lived meta II compared to other mutants and the wild-type receptor (WT) [94][98].

The recessive form of RP presents two missense mutations of RHO, namely, E150K $\frac{[99][100][101]}{[99][100][101]}$ and M253I $\frac{[102]}{[102]}$, and two mutations with a premature stop codon, namely, W161ter $\frac{[103]}{[103]}$ and E249ter $\frac{[104]}{[104]}$. In contrast, the dominant autosomal inheritance of RP has more than 150 documented mutations, representing 20–30% of all cases, with the P23H mutation being the most studied $\frac{[105][106]}{[105]}$. The biochemical and functional phenotypes of several specific mutations in Rho associated with RP have been previously reported (Table 1).

Table 1. Summary of the biochemical phenotypes of selected mutations in Rho associated with retinal degenerative diseases, namely, RP and CSNB.

Mutation	Behavior/Effect	Class/Misfolds	References
G90X	Causes thermal instability and/or abnormal photoproduct formation in inducing a RP phenotype.	VI/No	[<u>45]</u>
T94I	Induces constitutive activation of the opsin in the absence of chromophore and in the dark.	VI/No	<u>[94]</u>
E113K	Associated with the two distinct phenotypes of RP and CSNB in independent members of the same family.	Unclassified	[<u>91]</u>
A292E	Anomalously activates transducin when the chromophore is missing.	Unclassified	[<u>92]</u>
P23H	Destabilizes outer rod segments via the formation of aggregates due to retention in the ER.	II/Yes	[87]
E150	No observed biochemical or cellular defects or not studied in detail.	Unclassified	[<u>101</u>]
W161X	No observed biochemical or cellular defects or not studied in detail.	Unclassified	[103]
G114V	No observed biochemical or cellular defects or not studied in detail.	Unclassified	[<u>107</u>]
Q184P	No observed biochemical or cellular defects or not studied in detail.	Unclassified	[<u>107</u>]
R135X	Affects endocytosis	III/No	[108]
G188R	Forms aggregates due to retention in the ER and cannot be easily constituted with 11CR.	II/Yes	[109]

Mutations found in the *N*-terminal segment of Rho are often associated with mild disease, which develops late and with slowly advancing symptoms. These include P23H, T4K, P23A/L, N15K, T17M, V20G, and Q28H [106][110][111]. The *N*-terminal segment is important because it helps stabilize the retinal-bound conformation of the receptor [86]. Many mutations in the seven transmembrane segments of Rho have been described that can cause different effects on the

protein. Many mutations may represent the introduction of a charged amino acid into the membrane domain. These include L40R, L46R, G51R, P53R, and T58R in the first transmembrane helix. The presence of a charged residue may prevent insertion of the domain into the membrane of the endoplasmic reticulum resulting in incorrect folding of the protein [86]. In other cases, RP mutations in the transmembrane helices could result in a loss of side chains necessary for conformational stability and/or functioning or otherwise introduce bulky side chains that may result in steric clashes in densely packed regions of the protein, such as the case of the A164V mutation, which causes an incorrect fold of the protein [112].

An interesting aspect of GPCR functioning is the relevance of receptor–receptor interaction and particularly dimerization and higher-order oligomerization. Rho has the ability to form oligomers [113][114][115], but the functional relevance of such complexes remains to be fully established. In this regard, it has been reported that some mutations have a marked influence on Rho oligomer formation capacity [116]. In the case of the F45, V209, and F220 amino acid positions, found in transmembrane helices 1 and 5, these are the sites of the F45L, V209, and F220C mutations that cannot form dimers or multimers as seen in the case of the WT protein [107][116][117][118].

Two different mutations affect codon 135, where arginine is replaced either by a tryptophan (R135W) or by a leucine (R135L) [119]. These mutations affect an amino acid of the third transmembrane helix at the cytoplasmic membrane boundary. Mutations in codon 135 involve a change in charge and size, a large and basic amino acid is replaced by a non-polar and smaller one in the R135L mutation, and a non-polar, large, and aromatic one in the R135W mutant [119]. E134 and R135 residues are part of the highly conserved D/ERY motif, a site of interaction with the G protein transducin [48][120]. Studies have shown that the R135L and R135W mutations can perform binding in the retina with almost with the same efficiency as in WT cases in reconstituted and purified systems, but they are functionally defective and are not able to efficiently activate transducin [108].

Two other interesting changes occur in the opposite extracellular domain. At the second intradiscal loop, one mutation affects codon 180 and the other affects codon 188, resulting in the P180A and G188R mutations, respectively [109][119]. The substitution of P180A results in the change of a medium-sized hydrophobic residue to a smaller hydrophobic one. The G188R mutation implies the replacement of a small, non-polar amino acid by a large, basic, and positively charged one [121]. These changes can involve both steric and electrostatic effects that can disturb the intradiscal domain packing and the overall conformational stability of the receptor.

A detailed analysis of the structural effects of RP mutations on Rho, as well as the study of genotype-phenotype correlations, is very relevant for elucidating the fine details of the photoreceptor degeneration process. This information is essential to investigate the effects of selected compounds, like polyphenols or specifically flavonoids, on the conformational properties of RP mutant proteins and the subsequent potential clinical benefits of some of these compounds.

5. Polyphenols Effects in Retinal Degenerative Diseases

Therapies for retinal degenerative diseases are currently limited, so there is a need to develop new strategies for more effective and safer therapies. As we have seen, polyphenols, especially flavonoids, could be viable drug candidates as they may be involved in visual signal transduction and visual pigment regeneration. Flavonoid-rich vegetables and fruits appear to have effects in improving eyesight in eye-related diseases [61][123].

We will focus on the effects of flavonoids in three different retinal-related diseases: RP, CSNB and age-related macular degeneration (AMD).

RP has already been described previously. Additionally, CSNB is a group of heterogeneous genetic disorders of the retina that manifest as non-progressive nyctalopia [124]. Finally, AMD is a complex disease that exhibits several different pathological mechanisms including degeneration of photoreceptors and RPE cells causing visual impairment [125].

Flavonoids such as quercetin and myricetin have been shown to improve the stability of opsin present in rods, increase the binding rate of ligand-free opsin, and facilitate its expression and integration into the membrane in vitro $^{[126]}$. In spite of the studies presenting beneficial effects of flavonoids, the mechanisms of their protective effects against light-induced retinal damage are not entirely known $^{[123]}$. Some studies suggest that flavonoids interact directly with Rho, increasing their rates of regeneration, stability, folding, and membrane orientation in vitro and have an effect on retinal degenerative diseases (Table 2) $^{[46][127][128]}$.

Table 2. Summary of different polyphenols effects on retinal physiology.

Compound	Condition/Cell Lines	Effect	References
Quercetin	Oxidative stress conditions. Assay in vitro in human hepatoma HepG2 cells.	Activates the Nrf2-ARE signaling pathway and exhibits anti-oxidative stress activity alone and together with kaempferol and pterostilbene.	[123]
	Oxidative stress conditions. Assay in vitro in human RPE cells and in <i>Ccl2/Cx3cr1</i> double knock-out mice.	Protects RPE cells from oxidative stress via inhibiting pro-inflammatory molecules and the intrinsic apoptosis pathway.	[129]
	VEGF-treated mouse photoreceptor- derived 661W cells.	Inhibits the production of inflammatory proteins in VEGF-stimulated 661W cells.	[130]
	Oxidative stress conditions. ARPE- 19 human retinal pigment epithelial cells.	Protects ARPE-19 cells from H2O2-induced cytotoxicity by activating the Nrf2 pathway, inhibiting ER stress and targeting antiapoptotic proteins.	[131]
	Oxidative stress conditions. Assay in vitro in human RPE cells.	Protects RPE cells from oxidative damage and cellular senescence in a dose-dependent manner.	[132]
	Oxidative stress conditions. Assay in vitro and in vivo in human RPE cells.	Protects against blue light-induced retinal damage.	[133]
Myricetin	Human MCF-7 breast cancer cells.	Reduces and scavenges intracellular ROS.	[134]
Apigenin	Bright light-exposed BALB/c mice.	Confers retinal protection by inhibiting retinal oxidative stress and retinal inflammatory responses.	[<u>135]</u>
Tannic acid	Assay in vitro in human RPE cells (ARPE-19).	Protects RPE against ultraviolet B radiation via the inhibition of the inflammatory response.	[136]
Fisetin/Luteolin	Assay in vitro in human RPE cells (ARPE-19).	Anti-inflammatory and cytoprotective effects when used as dietary supplements.	[137]

Flavonoids have been found to stimulate Rho expression, where specifically Rho and cone opsins expression have been improved upon treatment with quercetin and myricetin $\frac{[127][138][139]}{[127][138][139]}$.

The antioxidant effect of polyphenols can be invoked as a factor that may delay the progression of AMD. A particular compound, namely, stilbenoid resveratrol, a dietary compound with a wide range of effects on cell function, has been shown to effectively reduce ROS production, thus protecting against retinal damage [84][140].

Flavonoids can also inhibit inflammatory reactions by suppressing the expression of pro-inflammatory genes and molecules involved in retinal degeneration. In addition, they can also limit ROS levels by sequestering oxidative radicals. In this regard, RPE cells treated with quercetin could be protected from oxidative stress by inhibiting apoptosis pathways

and pro-inflammatory markers [134][141]. Flavonoids enhance the expression of photoreceptor-specific genes by also attenuating the expression of oxidative stress and inflammation-related malignancies and altering the balance between anti-apoptotic and pro-apoptotic genes [129][130][131][135].

Another polyphenol, tannic acid, has also been reported to inhibit the production of interleukin-6 and to down-regulate the expression of complement factor B in ARPE-19 cells, a factor that is believed to be related to AMD [136].

In ARPE-19 cells, quercetin protects against stress induced by lipid peroxidation $\frac{[142]}{}$. Quercetin was observed to reduce mitochondrial function protecting against hydrogen peroxide-induced oxidative stress in RPE cells of human donor eyes thus increasing its viability $\frac{[132]}{}$. Other studies have shown that this polyphenol can improve oxidative stress and its consequences in different regions of the eye $\frac{[133][143][144][145]}{}$.

Quercetin has also produced a protective effect against oxidative stress and its consequences on photoreceptor cells resulting from the reaction of ATR with phosphatidylethanolamine producing bis-retinoid photoreactive species $\frac{[146]}{}$.

Mechanisms involved in the antioxidant activity of polyphenols include suppression of ROS formation $^{[147][148]}$, thus reducing oxidative damage $^{[149]}$. The mechanism by which ROS formation is reduced involves phosphorylation of Nrf2 residues resulting in nuclear accumulation $^{[150]}$.

Although the implication of flavonoids in vision and vision diseases is still uncertain, some studies with dietary flavonoids like quercetin have suggested potential beneficial effects in some forms of RP $^{[45]}$. Mutations in Rho are associated with this disease and they can cause protein misfolding that leads to a progressive loss of rod and cone cells, further resulting in vision loss $^{[151][152][153]}$. These results should be analyzed in the context of research in the RP field, where several strategies based on pharmacological rescue have been proposed for RP treatment. The basic principle of this approach is that chemical or pharmacological chaperones bind to misfolded opsins and are able to stabilize them $^{[154]}$.

The dietary flavonoid quercetin, one of the most studied and widely known for its potential beneficial effects on health $\frac{[155]}{1}$, has been used in some experiments with the recombinant G90V mutant associated with RP and has shown satisfactory effects when combined with 9-*cis*-retinal (9CR), a retinal analog that is usually employed in vision studies. Over the past years, different investigations have focused on describing the pharmaceutical application of 9-*cis* retinoids to remedy the retinal dysfunction caused by deficient regeneration with 11CR $\frac{[156][157][158]}{1}$ and have shown that this retinal analog can increase the stability of the RP mutant G90V $\frac{[70]}{1}$.

In summary, the use of polyphenols, like quercetin, alone or in combination with other small ligands, like retinoids, opens new possibilities for the treatment of retinal degeneration associated with RP. Moreover, the new effect attributed to quercetin may also be applicable to other members of the GPCR superfamily [45]. In spite of these encouraging results, there is clearly a need to further investigate the in vivo potential of such strategies and particularly to increase the number of clinical studies being performed. This is essential to fully determine the exact reach of these newly proposed mechanisms and the potential physiological effects of specific compounds.

References

- 1. Rasouli, H.; Farzaei, M.H.; Khodarahmi, R. Polyphenols and their benefits: A review. Int. J. Food Prop. 2017, 20, 1700–1741.
- 2. Xiao, J.B. Stability of dietary polyphenols: It's never too late to mend? Food Chem. Toxicol. 2018, 119, 3–5.
- 3. Sies, H. Polyphenols and health: Update and perspectives. Arch. Biochem. Biophys. 2010, 501, 2-5.
- 4. World Health Organization. Diet, Nutrition and the Prevention of Chronic Diseases; World Health Organization: Geneva, Switzerland, 2003.
- 5. Hollman, P.C.; Geelen, A.; Kromhout, D. Dietary flavonol intake may lower stroke risk in men and women. J. Nutr. 2010, 140, 600–604.
- 6. Hooper, L.; Kroon, P.A.; Rimm, E.B.; Cohn, J.S.; Harvey, I.; Le Cornu, K.A.; Ryder, J.J.; Hall, W.L.; Cassidy, A. Flavonoids, flavonoid-rich foods, and cardiovascular risk: A meta-analysis of randomized controlled trials. Am. J. Clin. Nutr. 2008, 88, 38–50.
- 7. Lupton, J.R.; Atkinson, S.A.; Chang, N.; Fraga, C.G.; Levy, J.; Messina, M.; Richardson, D.P.; van Ommen, B.; Yang, Y.; Griffiths, J.C.; et al. Exploring the benefits and challenges of establishing a DRI-like process for bioactives. Eur. J. Nutr. 2014, 53, 1–9.

- 8. Kennedy, D.O. Polyphenols and the human brain: Plant "secondary metabolite" ecologic roles and endogenous signaling functions drive benefits. Adv. Nutr. 2014, 5, 515–533.
- 9. Marin, L.; Miguelez, E.M.; Villar, C.J.; Lombo, F. Bioavailability of dietary polyphenols and gut microbiota metabolism: Antimicrobial properties. Biomed. Res. Int. 2015, 2015, 1–18.
- 10. Tsao, R. Chemistry and biochemistry of dietary polyphenols. Nutrients 2010, 2, 1231–1246.
- 11. Miyata, Y.; Shida, Y.; Hakariya, T.; Sakai, H. Anti-cancer effects of green tea polyphenols against prostate cancer. Molecules 2019, 24, 193.
- 12. Serino, A.; Salazar, G. Protective role of polyphenols against vascular inflammation, aging and cardiovascular disease. Nutrients 2018, 11, 53.
- 13. Leri, M.; Scuto, M.; Ontario, M.L.; Calabrese, V.; Calabrese, E.J.; Bucciantini, M.; Stefani, M. Healthy effects of plant polyphenols: Molecular mechanisms. Int. J. Mol. Sci. 2020, 21, 1250.
- 14. Cho, E.; Seddon, J.M.; Rosner, B.; Willett, W.C.; Hankinson, S.E. Prospective study of intake of fruits, vegetables, vitamins, and carotenoids and risk of age-related maculopathy. Arch. Ophthalmol. 2004, 122, 883.
- 15. Tang, L.; Zhang, Y.; Jiang, Y.; Willard, L.; Ortiz, E.; Wark, L.; Medeiros, D.; Lin, D. Dietary wolfberry ameliorates retinal structure abnormalities in db/db mice at the early stage of diabetes. Exp. Biol. Med. 2011, 236, 1051–1063.
- 16. Manach, C.; Williamson, G.; Morand, C.; Scalbert, A.; Rémésy, C. Bioavailability and bioefficacy of polyphenols in humans. I. Review of 97 bioavailability studies. Am. J. Clin. Nutr. 2005, 81, 230S–242S.
- 17. Cheynier, V. Polyphenols in foods are more complex than often thought. Am. J. Clin. Nutr. 2005, 81, 223S-229S.
- 18. Wen, L.; Jiang, Y.; Yang, J.; Zhao, Y.; Tian, M.; Yang, B. Structure, bioactivity, and synthesis of methylated flavonoids. Ann. N. Y. Acad. Sci. 2017, 1398, 120–129.
- 19. Selma, M.V.; Espín, J.C.; Tomás-Barberán, F.A. Interaction between phenolics and gut microbiota: Role in human health. J. Agric. Food Chem. 2009, 57, 6485–6501.
- 20. Crozier, A.; Del Rio, D.; Clifford, M.N. Bioavailability of dietary flavonoids and phenolic compounds. Mol. Aspects. Med. 2010, 31, 446–467.
- 21. Galleano, M.; Verstraeten, S.V.; Oteiza, P.I.; Fraga, C.G. Antioxidant actions of flavonoids: Thermodynamic and kinetic analysis. Arch. Biochem. Biophys. 2010, 501, 23–30.
- 22. Fraga, C.G. Plant polyphenols: How to translate their in vitro antioxidant actions to in vivo conditions. IUBMB Life 2007, 59, 308–315.
- 23. Focaccetti, C.; Izzi, V.; Benvenuto, M.; Fazi, S.; Ciuffa, S.; Giganti, M.G.; Potenza, V.; Manzari, V.; Modesti, A.; Bei, R. Polyphenols as immunomodulatory compounds in the tumor microenvironment: Friends or foes? Int. J. Mol. Sci. 2019, 20, 1714.
- 24. Maleki, S.J.; Crespo, J.F.; Cabanillas, B. Anti-inflammatory effects of flavonoids. Food Chem. 2019, 299, 125124.
- 25. Dudnik, A.; Gaspar, P.; Neves, A.R.; Forster, J. Engineering of microbial cell factories for the production of plant polyphenols with health-beneficial properties. Curr. Pharm. Des. 2018, 24, 2208–2225.
- 26. Fraga, C.G.; Oteiza, P.I.; Galleano, M. Plant bioactives and redox signaling: (–)-Epicatechin as a paradigm. Mol. Aspects. Med. 2018, 61, 31–40.
- 27. Kim, H.-S.; Quon, M.J.; Kim, J. New insights into the mechanisms of polyphenols beyond antioxidant properties; lessons from the green tea polyphenol, epigallocatechin 3-gallate. Redox Biol. 2014, 2, 187–195.
- 28. Fraga, C.G.; Croft, K.D.; Kennedy, D.O.; Tomás-Barberán, F.A. The effects of polyphenols and other bioactives on human health. Food Funct. 2019, 10, 514–528.
- 29. Cremonini, E.; Bettaieb, A.; Haj, F.G.; Fraga, C.G.; Oteiza, P.I. (–)-Epicatechin improves insulin sensitivity in high fat diet-fed mice. Arch. Biochem. Biophys. 2016, 599, 13–21.
- 30. Vazquez-Prieto, M.A.; Bettaieb, A.; Haj, F.G.; Fraga, C.G.; Oteiza, P.I. (–)-Epicatechin prevents TNFα-induced activation of signaling cascades involved in inflammation and insulin sensitivity in 3T3-L1 adipocytes. Arch. Biochem. Biophys. 2012, 527, 113–118.
- 31. Bettaieb, A.; Cremonini, E.; Kang, H.; Kang, J.; Haj, F.G.; Oteiza, P.I. Anti-inflammatory actions of (–)-epicatechin in the adipose tissue of obese mice. Int. J. Biochem. Cell Biol. 2016, 81, 383–392.
- 32. Crichton, G.E.; Elias, M.F.; Dearborn, P.; Robbins, M. Habitual chocolate intake and type 2 diabetes mellitus in the Maine-Syracuse Longitudinal Study: (1975–2010): Prospective observations. Appetite 2017, 108, 263–269.

- 33. Buitrago-Lopez, A.; Sanderson, J.; Johnson, L.; Warnakula, S.; Wood, A.; Di Angelantonio, E.; Franco, O.H. Chocolate consumption and cardiometabolic disorders: Systematic review and meta-analysis. BMJ 2011, 343, d4488.
- 34. Khan, N.; Mukhtar, H. Tea polyphenols in promotion of human health. Nutrients 2018, 11, 39.
- 35. Bondonno, N.P.; Bondonno, C.P.; Blekkenhorst, L.C.; Considine, M.J.; Maghzal, G.; Stocker, R.; Woodman, R.J.; Ward, N.C.; Hodgson, J.M.; Croft, K.D. Flavonoid-rich apple improves endothelial function in individuals at risk for cardiovascular disease: A randomized controlled clinical trial. Mol. Nutr. Food Res. 2017, 62, 1700674.
- 36. Huang, H.; Chen, G.; Liao, D.; Zhu, Y.; Xue, X. Effects of berries consumption on cardiovascular risk factors: A Metaanalysis with trial sequential analysis of randomized controlled trials. Sci. Rep. 2016, 6, 23625.
- 37. Tomás-Barberán, F.A.; Selma, M.V.; Espín, J.C. Interactions of gut microbiota with dietary polyphenols and consequences to human health. Curr. Opin. Clin. Nutr. Metab. Care 2016, 19, 471–476.
- 38. Oteiza, P.I.; Fraga, C.G.; Mills, D.A.; Taft, D.H. Flavonoids and the gastrointestinal tract: Local and systemic effects. Mol. Aspects Med. 2018, 61, 41–49.
- 39. Crichton, G.E.; Elias, M.F.; Alkerwi, A. Chocolate intake is associated with better cognitive function: The Maine-Syracuse Longitudinal Study. Appetite 2016, 100, 126–132.
- 40. Moreira, A.; Diógenes, M.J.; de Mendonça, A.; Lunet, N.; Barros, H. Chocolate consumption is associated with a lower risk of cognitive decline. J. Alzheimers Dis. 2016, 53, 85–93.
- 41. Ng, T.P.; Feng, L.; Niti, M.; Kua, E.H.; Yap, K.B. Tea consumption and cognitive impairment and decline in older Chinese adults. Am. J. Clin. Nutr. 2008, 88, 224–231.
- 42. Kuriyama, S.; Hozawa, A.; Ohmori, K.; Shimazu, T.; Matsui, T.; Ebihara, S.; Awata, S.; Nagatomi, R.; Arai, H.; Tsuji, I. Green tea consumption and cognitive function: A cross-sectional study from the Tsurugaya Project. Am. J. Clin. Nutr. 2006, 83, 355–361.
- 43. Dong, X.; Yang, C.; Cao, S.; Gan, Y.; Sun, H.; Gong, Y.; Yang, H.; Yin, X.; Lu, Z. Tea consumption and the risk of depression: A meta-analysis of observational studies. Aust. N. Z. J. Psychiatry 2015, 49, 334–345.
- 44. Li, F.-J.; Ji, H.-F.; Shen, L. A Meta-analysis of tea drinking and risk of Parkinson's disease. Sci. World J. 2012, 2012, 1–6
- 45. Herrera-Hernández, M.G.; Ramon, E.; Lupala, C.S.; Tena-Campos, M.; Pérez, J.J.; Garriga, P. Flavonoid allosteric modulation of mutated visual rhodopsin associated with retinitis pigmentosa. Sci. Rep. 2017, 7, 11167.
- 46. Bourne, H.R.; Meng, E.C. Structure. Rhodopsin sees the light. Science 2000, 289, 733-734.
- 47. Nickell, S.; Park, P.S.-H.; Baumeister, W.; Palczewski, K. Three-dimensional architecture of murine rod outer segments determined by cryoelectron tomography. J. Cell Biol. 2007, 177, 917–925.
- 48. Palczewski, K.; Kumasaka, T.; Hori, T.; Behnke, C.A.; Motoshima, H.; Fox, B.A.; Le Trong, I.; Teller, D.C.; Okada, T.; Stenkamp, R.E.; et al. Crystal structure of rhodopsin: A G protein-coupled receptor. Science 2000, 289, 739–745.
- 49. Jastrzebska, B. GPCR: G protein complexes—The fundamental signaling assembly. Amino Acids 2013, 45, 1303–1314.
- 50. Katayama, K.; Gulati, S.; Ortega, J.T.; Alexander, N.S.; Sun, W.; Shenouda, M.M.; Palczewski, K.; Jastrzebska, B. Specificity of the chromophore-binding site in human cone opsins. J. Biol. Chem. 2019, 294, 6082–6093.
- 51. Zhang, D.; Zhao, Q.; Wu, B. Structural studies of G protein-coupled receptors. Mol. Cells 2015, 38, 836-842.
- 52. Alexander, S.P.H.; Benson, H.E.; Faccenda, E.; Pawson, A.J.; Sharman, J.L.; Spedding, M.; Peters, J.A.; Harmar, A.J. The concise guide to PHARMACOLOGY 2013/14: G proteincoupled receptors. Br. J. Pharmacol. 2013, 170, 1459–1581.
- 53. Fredriksson, R.; Lagerström, M.C.; Lundin, L.-G.; Schiöth, H.B. The G-protein-coupled receptors in the human genome form five main families. Phylogenetic analysis, paralogon groups, and fingerprints. Mol. Pharmacol. 2003, 63, 1256–1272.
- 54. Stevens, R.C.; Cherezov, V.; Katritch, V.; Abagyan, R.; Kuhn, P.; Rosen, H.; Wüthrich, K. The GPCR network: A large-scale collaboration to determine human GPCR structure and function. Nat. Rev. Drug Discov. 2013, 12, 25–34.
- 55. Lindsley, C.W.; Emmitte, K.A.; Hopkins, C.R.; Bridges, T.M.; Gregory, K.J.; Niswender, C.M.; Conn, P.J. Practical strategies and concepts in GPCR allosteric modulator discovery: Recent advances with metabotropic glutamate receptors. Chem. Rev. 2016, 116, 6707–6741.
- 56. Khoury, E.; Clément, S.; Laporte, S.A. Allosteric and biased g protein-coupled receptor signaling regulation: Potentials for new therapeutics. Front. Endocrinol. 2014, 5, 68.
- 57. Sato, J.; Makita, N.; Iiri, T. Inverse agonism: The classic concept of GPCRs revisited. Endocr. J. 2016, 63, 507-514.

- 58. Lane, J.R.; Abdul-Ridha, A.; Canals, M. Regulation of G protein-coupled receptors by allosteric ligands. ACS Chem. Neurosci. 2013, 4, 527–534.
- 59. Hubbard, R.; Kropf, A. The action of light on rhodopsin. Proc. Natl. Acad. Sci. USA 1958, 44, 130-139.
- 60. Nakamichi, H.; Okada, T. X-ray crystallographic analysis of 9-cis-rhodopsin, a model analogue visual pigment. J. Photochem. Photobiol. 2007, 83, 232–235.
- 61. Kalt, W.; Hanneken, A.; Milbury, P.; Tremblay, F. Recent research on polyphenolics in vision and eye health. J. Agric. Food Chem. 2010, 58, 4001–4007.
- 62. Zhong, M.; Kawaguchi, R.; Kassai, M.; Sun, H. Retina, retinol, retinal and the natural history of vitamin A as a light sensor. Nutrients 2012, 4, 2069–2096.
- 63. Park, J.H.; Scheerer, P.; Hofmann, K.P.; Choe, H.-W.; Ernst, O.P. Crystal structure of the ligand-free G-protein-coupled receptor opsin. Nature 2008, 454, 183–187.
- 64. Garriga, P.; Manyosa, J. The eye photoreceptor protein rhodopsin. Structural implications for retinal disease. FEBS Lett. 2002, 528, 17–22.
- 65. Ridge, K.D.; Abdulaev, N.G.; Sousa, M.; Palczewski, K. Phototransduction: Crystal clear. Trends Biochem. Sci. 2003, 28, 479–487.
- 66. Travis, G.H.; Golczak, M.; Moise, A.R.; Palczewski, K. Diseases caused by defects in the visual cycle: Retinoids as potential therapeutic agents. Annu. Rev. Pharmacol. Toxicol. 2007, 47, 469–512.
- 67. Calzia, D.; Barabino, S.; Bianchini, P.; Garbarino, G.; Oneto, M.; Caicci, F.; Diaspro, A.; Tacchetti, C.; Manni, L.; Candiani, S.; et al. New findings in ATP supply in rod outer segments: Insights for retinopathies. Biol. Cell. 2013, 105, 345–358.
- 68. Kiser, P.D.; Golczak, M.; Palczewski, K. Chemistry of the Retinoid (Visual) Cycle. Chem. Rev. 2014, 114, 194–232.
- 69. Fan, J.; Woodruff, M.L.; Cilluffo, M.C.; Crouch, R.K.; Fain, G.L. Opsin activation of transduction in the rods of dark-reared Rpe65 knockout mice. J. Physiol. 2005, 568, 83–95.
- 70. Toledo, D.; Ramon, E.; Aguilà, M.; Cordomí, A.; Pérez, J.J.; Mendes, H.F.; Cheetham, M.E.; Garriga, P. Molecular mechanisms of disease for mutations at Gly-90 in rhodopsin. J. Biol. Chem. 2011, 286, 39993–40001.
- 71. Palczewski, K. G protein-coupled receptor rhodopsin. Annu. Rev. Biochem. 2006, 75, 743-767.
- 72. Veleri, S.; Lazar, C.H.; Chang, B.; Sieving, P.A.; Banin, E.; Swaroop, A. Biology and therapy of inherited retinal degenerative disease: Insights from mouse models. Dis. Models Mech. 2015, 8, 109–129.
- 73. Chen, Y.; Okano, K.; Maeda, T.; Chauhan, V.; Golczak, M.; Maeda, A.; Palczewski, K. Mechanism of all-trans-retinal toxicity with implications for stargardt disease and age-related macular degeneration. J. Biol. Chem. 2012, 287, 5059–5069.
- 74. Kim, S.R.; Jang, Y.P.; Jockusch, S.; Fishkin, N.E.; Turro, N.J.; Sparrow, J.R. The all-trans- retinal dimer series of lipofuscin pigments in retinal pigment epithelial cells in a recessive Stargardt disease model. Proc. Natl. Acad. Sci. USA 2007, 104, 19273–19278.
- 75. Sparrow, J.R.; Wu, Y.; Kim, C.Y.; Zhou, J. Phospholipid meets all-trans-retinal: The making of RPE bisretinoids. J. Lipid. Res. 2010, 51, 247–261.
- 76. Gao, S.; Parmar, T.; Palczewska, G.; Dong, Z.; Golczak, M.; Palczewski, K.; Jastrzebska, B. Protective effect of a locked retinal chromophore analog against light-induced retinal degeneration. Mol. Pharmacol. 2018, 94, 1132–1144.
- 77. Kaarniranta, K.; Pawlowska, E.; Szczepanska, J.; Jablkowska, A.; Blasiak, J. Role of mitochondrial DNA damage in ROS-mediated pathogenesis of Age-related Macular Degeneration (AMD). Int. J. Mol. Sci. 2019, 20, 2374.
- 78. Sawada, O.; Perusek, L.; Kohno, H.; Howell, S.J.; Maeda, A.; Matsuyama, S.; Maeda, T. All-trans-retinal induces Bax activation via DNA damage to mediate retinal cell apoptosis. Exp. Eye Res. 2014, 123, 27–36.
- 79. Kohno, H.; Maeda, T.; Perusek, L.; Pearlman, E.; Maeda, A. CCL3 production by microglial cells modulates disease severity in murine models of retinal degeneration. J. Immunol. 2014, 192, 3816–3827.
- 80. Rashid, K.; Wolf, A.; Langmann, T. Microglia activation and immunomodulatory therapies for retinal degenerations. Front, Cell. Neurosci. 2018, 12, 176.
- 81. Rashid, K.; Akhtar-Schaefer, I.; Langmann, T. Microglia in Retinal Degeneration. Front. Immunol. 2019, 10, 1975.
- 82. Bruschi, M.; Bartolucci, M.; Peteretto, A.; Calzia, D.; Caicci, F.; Manni, L.; Traverso, C.E.; Candiano, G.; Panfoli, I. Differential expression of the five redox complexes in the retinal mitochondria or rod outer segment disks is consistent with their different functionality. FASEB BioAdv. 2020, 2, 315–324.

- 83. Bruschi, M.; Petretto, A.; Caicci, F.; Bartolucci, M.; Calzia, D.; Santucci, L.; Manni, L.; Ramenghi, L.A.; Ghiggeri, G.; Traverso, C.E.; et al. Proteome of bovine mitochondria and rod outer segments disks: Commonalities and differences. J. Proteome Res. 2018, 17, 918–925.
- 84. Ravera, S.; Esposito, A.; Degan, P.; Caicci, F.; Calzia, D.; Perrotta, E.; Manni, L.; Bisio, A.; Iobbi, V.; Schito, A.; et al. Sclareol modulates free radical production in the retinal rod outer segment by inhibiting the ectopic f1fo-atp synthase. Free Radic. Biol. Med. 2020, 60, 368–375.
- 85. Goldberg, A.F.; Moritz, O.L.; Williams, D.S. Molecular basis for photoreceptor outer segment architecture. Prog. Retin. Eye Res. 2016, 55, 52–81.
- 86. Athanasiou, D.; Aquila, M.; Bellingham, J.; Li, W.; McCulley, C.; Reeves, P.J.; Cheetham, M.E. The molecular and cellular basis of rhodopsin retinitis pigmentosa reveals potential strategies for therapy. Prog. Retin. Eye Res. 2018, 62, 1–23
- 87. Dryja, T.P.; McGee, T.L.; Hahn, L.B.; Cowley, G.S.; Olsson, J.E.; Reichel, E.; Sandberg, M.A.; Berson, E.L. Mutations within the rhodopsin gene in patients with autosomal dominant retinitis pigmentosa. N. Engl. J. Med. 1990, 323, 1302–1307.
- 88. Rao, V.R.; Cohen, G.B.; Oprian, D.D. Rhodopsin mutation G90D and a molecular mechanism for congenital night blindness. Nature 1994, 367, 639–642.
- 89. Sieving, P.A.; Richards, J.E.; Naarendorp, F.; Bingham, E.L.; Scott, K.; Alpern, M. Dark-light: Model for night blindness from the human rhodopsin Gly-90-->Asp mutation. Proc. Natl. Acad. Sci. USA 1995, 92, 880–884.
- 90. Al-Jandal, N.; Farrar, G.J.; Kiang, A.S.; Humphries, M.M.; Bannon, N.; Findlay, J.B.; Humphries, P.; Kenna, P.F. A novel mutation within the rhodopsin gene (Thr-94-IIe) causing autosomal dominant congenital stationary night blindness. Hum. Mutat. 1999, 13, 75–81.
- 91. Reiff, C.; Owczarek-Lipska, M.; Spital, G.; Roger, C.; Hinz, H.; Juschke, C.; Thiele, H.; Altmuller, J.; Nurnberg, P.; Da Costa, R.; et al. The mutation p.E113K in the Schiff base counterion of rhodopsin is associated with two distinct retinal phenotypes within the same family. Sci. Rep. 2016, 6, 36208.
- 92. Dryja, T.P.; Berson, E.L.; Rao, V.R.; Oprian, D.D. Heterozygous missense mutation in the rhodopsin gene as a cause of congenital stationary night blindness. Nat. Genet. 1993, 4, 280–283.
- 93. Zeitz, C.; Gross, A.K.; Leifert, D.; Kloeckener-Gruissem, B.; McAlear, S.D.; Lemke, J.; Neidhardt, J.; Berger, W. Identification and functional characterization of a novel rhodopsin mutation associated with autosomal dominant CSNB. Investig. Ophthalmol. Vis. Sci. 2008, 49, 4105–4114.
- 94. Gross, A.K.; Rao, V.R.; Oprian, D.D. Characterization of rhodopsin congenital night blindness mutant T94I. Biochemistry 2003, 42, 2009–2015.
- 95. Singhal, A.; Guo, Y.; Matkovic, M.; Schertler, G.; Deupi, X.; Yan, E.C.; Standfuss, J. Structural role of the T941 rhodopsin mutation in congenital stationary night blindness. EMBO Rep. 2016, 17, 1431–1440.
- 96. Chen, J.; Shi, G.; Concepcion, F.A.; Xie, G.; Oprian, D.; Chen, J. Stable rhodopsin/arrestin complex leads to retinal degeneration in a transgenic mouse model of autosomal dominant retinitis pigmentosa. J. Neurosci. 2006, 26, 11929–11937.
- 97. Tam, B.M.; Moritz, O.L. Characterization of rhodopsin P23H-induced retinal degeneration in a Xenopus laevis model of retinitis pigmentosa. Investig. Ophthalmol. Vis. Sci. 2006, 47, 3234–3241.
- 98. Ramon, E.; del Valle, L.J.; Garriga, P. Unusual thermal and conformational properties of the rhodopsin congenital night blindness mutant Thr-94 --> Ile. J. Biol. Chem. 2003, 278, 6427–6432.
- 99. Azam, M.; Khan, M.I.; Gal, A.; Hussain, A.; Shah, S.T.; Khan, M.S.; Sadeque, A.; Bokhari, H.; Collin, R.W.J.; Orth, U.; et al. A homozygous p.Glu150Lys mutation in the opsin gene of two Pakistani families with autosomal recessive retinitis pigmentosa. Mol. Vis. 2009, 15, 2526–2534.
- 100. Saqib, M.A.; Nikopoulos, K.; Ullah, E.; Sher Khan, F.; Iqbal, J.; Bibi, R.; Jarral, A.; Sajid, S.; Nishiguchi, K.M.; Venturini, G.; et al. Homozygosity mapping reveals novel and known mutations in Pakistani families with inherited retinal dystrophies. Sci. Rep. 2015, 5, 9965.
- 101. Van Schil, K.; Karlstetter, M.; Aslanidis, A.; Dannhausen, K.; Azam, M.; Qamar, R.; Leroy, B.P.; Depasse, F.; Langmann, T.; De Baere, E. Autosomal recessive retinitis pigmentosa with homozygous rhodopsin mutation E150K and non-coding cis-regulatory variants in CRX-binding regions of SAMD7. Sci. Rep. 2016, 6, 21307.
- 102. Collin, R.W.; van den Born, L.I.; Klevering, B.J.; de Castro-Miro, M.; Littink, K.W.; Arimadyo, K.; Azam, M.; Yazar, V.; Zonneveld, M.N.; Paun, C.C.; et al. High-resolution homozygosity mapping is a powerful tool to detect novel mutations causative of autosomal recessive RP in the Dutch population. Investig. Ophthalmol. Vis. Sci. 2011, 52, 2227–2239.

- 103. Kartasasmita, A.; Fujiki, K.; Iskandar, E.; Sovani, I.; Fujimaki, T.; Murakami, A. A novel nonsense mutation in rhodopsin gene in two Indonesian families with autosomal recessive retinitis pigmentosa. Ophthalmic Genet. 2011, 32, 57–63.
- 104. Rosenfeld, P.J.; Cowley, G.S.; McGee, T.L.; Sandberg, M.A.; Berson, E.L.; Dryja, T.P. A null mutation in the rhodopsin gene causes rod photoreceptor dysfunction and autosomal recessive retinitis pigmentosa. Nat. Genet. 1992, 1, 209–213.
- 105. Sullivan, L.S.; Bowne, S.J.; Birch, D.G.; Hughbanks-Wheaton, D.; Heckenlively, J.R.; Lewis, R.A.; Garcia, C.A.; Ruiz, R.S.; Blanton, S.H.; Northrup, H.; et al. Prevalence of disease-causing mutations in families with autosomal dominant retinitis pigmentosa: A screen of known genes in 200 families. Investig. Ophthalmol. Vis. Sci. 2006, 47, 3052–3064.
- 106. Jacobson, S.G.; Kemp, C.M.; Sung, C.H.; Nathans, J. Retinal function and rhodopsin levels in autosomal dominant retinitis pigmentosa with rhodopsin mutations. Am. J. Ophthalmol. 1991, 112, 256–271.
- 107. Dryja, T.P.; McEvoy, J.A.; McGee, T.L.; Berson, E.L. Novel rhodopsin mutations Gly114Val and Gln184Pro in dominant retinitis pigmentosa. Investig. Ophthalmol. Vis. Sci. 2000, 41, 3124–3127.
- 108. Shi, W.; Sports, C.D.; Raman, D.; Shirakawa, S.; Osawa, S.; Weiss, E.R. Rhodopsin arginine-135 mutants are phosphorylated by rhodopsin kinase and bind arrestin in the absence of 11-cis retinal. Biochemistry 1998, 37, 4869–4874.
- 109. Del Porto, G.; Vingolo, E.M.; David, D.; Steindl, K.; Wedemann, H.; Forte, R.; Iannccone, A.; Gal, A.; Pannarale, M.R. Clinical features of autosomal dominant retinitis pigmentosa associated with the GLY-188-ARG mutation of the rhodopsin gene. In Retinal Degeneration; Hollyfield, J.G., Anderson, R.E., LaVail, M.M., Eds.; Springer: Boston, MA, USA, 1993; pp. 91–101.
- 110. Van Woerkom, C.; Ferrucci, S. Sector retinitis pigmentosa. Optometry 2005, 76, 309-317.
- 111. Ramon, E.; Cordomi, A.; Aguila, M.; Srinivasan, S.; Dong, X.; Moore, A.T.; Webster, A.R.; Cheetham, M.E.; Garriga, P. Differential light-induced responses in sectorial inherited retinal degeneration. J. Biol. Chem. 2014, 289, 35918–35928.
- 112. Sanchez-Reyes, O.B.; Cooke, A.L.G.; Tranter, D.B.; Rashid, D.; Eilers, M.; Reeves, P.J.; Smith, S.O. G Protein-Coupled Receptors Contain Two Conserved Packing Clusters. Biophys. J. 2017, 112, 2315–2326.
- 113. Jastrzebska, B.; Chen, Y.; Orban, T.; Jin, H.; Hofmann, L.; Palczewski, K. Disruption of rhodopsin dimerization with synthetic peptides targeting an interaction interface. J. Biol. Chem. 2015, 290, 25728–25744.
- 114. Kota, P.; Reeves, P.J.; Rajbhandary, U.L.; Khorana, H.G. Opsin is present as dimers in COS1 cells: Identification of amino acids at the dimeric interface. Proc. Natl. Acad. Sci. USA 2006, 103, 3054–3059.
- 115. Gunkel, M.; Schoneberg, J.; Alkhaldi, W.; Irsen, S.; Noe, F.; Kaupp, U.B.; Al-Amoudi, A. Higher-order architecture of rhodopsin in intact photoreceptors and its implication for phototransduction kinetics. Structure 2015, 23, 628–638.
- 116. Ploier, B.; Caro, L.N.; Morizumi, T.; Pandey, K.; Pearring, J.N.; Goren, M.A.; Finnemann, S.C.; Graumann, J.; Arshavsky, V.Y.; Dittman, J.S.; et al. Dimerization deficiency of enigmatic retinitis pigmentosa-linked rhodopsin mutants. Nat. Commun. 2016, 7, 12832.
- 117. Davies, W.I.; Downes, S.M.; Fu, J.K.; Shanks, M.E.; Copley, R.R.; Lise, S.; Ramsden, S.C.; Black, G.C.M.; Gibson, K.; Foster, R.G.; et al. Next-generation sequencing in health-care delivery: Lessons from the functional analysis of rhodopsin. Genet. Med. 2012, 14, 891–899.
- 118. Lim, K.P.; Yip, S.P.; Cheung, S.C.; Leung, K.W.; Lam, S.T.; To, C.H. Novel PRPF31 and PRPH2 mutations and co-occurrence of PRPF31 and RHO mutations in Chinese patients with retinitis pigmentosa. Arch. Ophthalmol. 2009, 127, 784–790.
- 119. Cideciyan, A.V.; Hood, D.C.; Huang, Y.; Banin, E.; Li, Z.Y.; Stone, E.M.; Milam, A.H.; Jacobson, S.G. Disease sequence from mutant rhodopsin allele to rod and cone photoreceptor degeneration in man. Proc. Natl. Acad. Sci. USA 1998, 95, 7103–7108.
- 120. Li, J.; Edwards, P.C.; Burghammer, M.; Villa, C.; Schertler, G.F. Structure of bovine rhodopsin in a trigonal crystal form. J. Mol. Biol. 2004, 343, 1409–1438.
- 121. Iannaccone, A.; Man, D.; Waseem, N.; Jennings, B.J.; Ganapathiraju, M.; Gallaher, K.; Reese, E.; Bhattacharya, S.S.; Klein-Seetharaman, J. Retinitis pigmentosa associated with rhodopsin mutations: Correlation between phenotypic variability and molecular effects. Vision Res. 2006, 46, 4556–4567.
- 122. Tuan-Phat Huynh; Shivani N. Mann; Nawajes A. Mandal; Botanical Compounds: Effects on Major Eye Diseases. *Evidence-Based Complementary and Alternative Medicine* **2013**, *2013*, 549174, <u>10.1155/2013/549174</u>.
- 123. Constance Saw; Yue Guo; Yuqing Yang; Ximena Paredes-Gonzalez; Christina Ramirez; Douglas Pung; Ah-Ng Tony Kong; The berry constituents quercetin, kaempferol, and pterostilbene synergistically attenuate reactive oxygen

- species: Involvement of the Nrf2-ARE signaling pathway. *Food and Chemical Toxicology* **2014**, *72*, 303-311, <u>10.1016/j.f</u> ct.2014.07.038.
- 124. Virginia Miraldi Utz; Wanda Pfeifer; Susannah Q. Longmuir; Richard Olson; Kai Wang; Arlene Drack; Presentation of TRPM1-Associated Congenital Stationary Night Blindness in Children. *JAMA Ophthalmology* **2018**, *136*, 389-398, <u>10.1</u> 001/jamaophthalmol.2018.0185.
- 125. Mandeep S. Singh; Robert E. MacLaren; Stem Cell Treatment for Age-Related Macular Degeneration: the Challenges. *Investigative Opthalmology & Visual Science* **2018**, 59, AMD78-AMD82, <u>10.1167/iovs.18-24426</u>.
- 126. Joseph T. Ortega; Tanu Parmar; Beata Jastrzebska; Flavonoids enhance rod opsin stability, folding, and self-association by directly binding to ligand-free opsin and modulating its conformation. *Journal of Biological Chemistry* **2019**, *294*, 8101-8122, <u>10.1074/jbc.ra119.007808</u>.
- 127. Joseph T. Ortega; Tanu Parmar; Marcin Golczak; Beata Jastrzebska; Protective Effects of Flavonoids in Acute Models of Light-Induced Retinal Degeneration. *Molecular Pharmacology* **2020**, *99*, 60-77, <u>10.1124/molpharm.120.000072</u>.
- 128. Joseph T. Ortega; Beata Jastrzebska; The Retinoid and Non-Retinoid Ligands of the Rod Visual G Protein-Coupled Receptor. *International Journal of Molecular Sciences* **2019**, *20*, 6218, <u>10.3390/ijms20246218</u>.
- 129. Xiaoguang Cao; Melissa Liu; Jingsheng Tuo; Defen Shen; Chi-Chao Chan; The effects of quercetin in cultured human RPE cells under oxidative stress and in Ccl2/Cx3cr1 double deficient mice. *Experimental Eye Research* **2010**, *91*, 15-25, 10.1016/j.exer.2010.03.016.
- 130. Minsup Lee; Seohyeon Yun; Hyesook Lee; Jaewook Yang; Quercetin Mitigates Inflammatory Responses Induced by Vascular Endothelial Growth Factor in Mouse Retinal Photoreceptor Cells through Suppression of Nuclear Factor Kappa B. *International Journal of Molecular Sciences* **2017**, *18*, 2497, <u>10.3390/ijms18112497</u>.
- 131. Sisi Weng; Lei Mao; Yuanyuan Gong; Tao Sun; Qing Gu; Role of quercetin in protecting ARPE-19 cells against H2O2-induced injury via nuclear factor erythroid 2 like 2 pathway activation and endoplasmic reticulum stress inhibition. *Molecular Medicine Reports* **2017**, *16*, 3461-3468, <u>10.3892/mmr.2017.6964</u>.
- 132. Daniel Kook; Armin H. Wolf; Alice L. Yu; Aljoscha S. Neubauer; Siegfried G. Priglinger; Anselm Kampik; Ulrich C. Welge-Lu"ssen; The Protective Effect of Quercetin against Oxidative Stress in the Human RPE In Vitro. *Investigative Opthalmology & Visual Science* **2008**, *49*, 1712-1720, 10.1167/iovs.07-0477.
- 133. Jun Kim; Hong Lan Jin; Dae Sik Jang; Kwang Won Jeong; Se-Young Choung; Quercetin-3-O-α-l-arabinopyranoside protects against retinal cell death via blue light-induced damage in human RPE cells and Balb-c mice. *Food & Function* **2018**, 9, 2171-2183, <u>10.1039/c7fo01958k</u>.
- 134. Abolfazl Barzegar; Antioxidant activity of polyphenolic myricetin in vitro cell- free and cell-based systems. *Molecular biology research communications* **1970**, *5*, 87-95, .
- 135. Minjuan Bian; Yong Zhang; Xiaoye Du; Jing Xu; Jingang Cui; Jiangping Gu; Weiliang Zhu; Teng Zhang; Yu Chen; Apigenin-7-diglucuronide protects retinas against bright light-induced photoreceptor degeneration through the inhibition of retinal oxidative stress and inflammation. *Brain Research* **2017**, *1663*, 141-150, <u>10.1016/j.brainres.2017.03.019</u>.
- 136. Wen-Wen Chou; Yung-Song Wang; Ku-Chung Chen; Jing-Mei Wu; Chung-Ling Liang; Suh-Hang Hank Juo; Tannic acid suppresses ultraviolet B-induced inflammatory signaling and complement factor B on human retinal pigment epithelial cells. *Cellular Immunology* **2012**, *273*, 79-84, <u>10.1016/j.cellimm.2011.11.003</u>.
- 137. Maria Hytti; Dora Szabó; Niina Piippo; Eveliina Korhonen; Paavo Honkakoski; Kai Kaarniranta; Goran Petrovski; Anu Kauppinen; Two dietary polyphenols, fisetin and luteolin, reduce inflammation but augment DNA damage-induced toxicity in human RPE cells. *The Journal of Nutritional Biochemistry* **2017**, *42*, 37-42, <u>10.1016/j.jnutbio.2016.12.014</u>.
- 138. Pascal Escher; Daniel F. Schorderet; Sandra Cottet; Altered Expression of the Transcription Factor Mef2c during Retinal Degeneration inRpe65–/–Mice. *Investigative Opthalmology & Visual Science* **2011**, *52*, 5933-5940, <u>10.1167/iov s.10-6978</u>.
- 139. Anne Wolf; Alexander Aslanidis; Thomas Langmann; Retinal expression and localization of Mef2c support its important role in photoreceptor gene expression. *Biochemical and Biophysical Research Communications* **2017**, *4*83, 346-351, <u>1</u> 0.1016/j.bbrc.2016.12.141.
- 140. Daniela Calzia; Paolo Degan; Federico Caicci; Maurizio Bruschi; Lucia Manni; Luca A. Ramenghi; Giovanni Candiano; Carlo Enrico Traverso; Isabella Panfoli; Modulation of the rod outer segment aerobic metabolism diminishes the production of radicals due to light absorption. *Free Radical Biology and Medicine* **2018**, *117*, 110-118, <u>10.1016/j.freerad biomed.2018.01.029</u>.
- 141. Lucio G. Costa; Jacqueline M. Garrick; Pamela J. Roquè; Claudia Pellacani; Mechanisms of Neuroprotection by Quercetin: Counteracting Oxidative Stress and More. *Oxidative Medicine and Cellular Longevity* **2016**, *2016*, 2986796, 10.1155/2016/2986796.

- 142. Joe G Hollyfield; Vera L Bonilha; Mary E Rayborn; Xiaoping Yang; Karen G Shadrach; Liang Lu; Rafael L Ufret; Robert G Salomon; Victor L Perez; Oxidative damage—induced inflammation initiates age-related macular degeneration. *Nature Medicine* **2008**, *14*, 194-198, <u>10.1038/nm1709</u>.
- 143. Moritz Veltmann; Margrit Hollborn; Andreas Reichenbach; Peter Wiedemann; Leon Kohen; Andreas Bringmann; Osmotic Induction of Angiogenic Growth Factor Expression in Human Retinal Pigment Epithelial Cells. *PLOS ONE* 2016, 11, e0147312-e0147312, 10.1371/journal.pone.0147312.
- 144. Sun-Myung Yoon; Bom-Lee Lee; Yuan-Ri Guo; Se-Young Choung; Preventive effect of Vaccinium uliginosum L. extract and its fractions on age-related macular degeneration and its action mechanisms. *Archives of Pharmacal Research* **2016**, *39*, 21-32, <u>10.1007/s12272-015-0683-7</u>.
- 145. Yong Wang; Hye Jin Kim; Janet R. Sparrow; Quercetin and cyanidin-3-glucoside protect against photooxidation and photodegradation of A2E in retinal pigment epithelial cells. *Experimental Eye Research* **2017**, *160*, 45-55, <u>10.1016/j.exer.2017.04.010</u>.
- 146. Zhao Zhao; Tao Sun; Yun Jiang; Lijiang Wu; Xiangzhong Cai; Xiaodong Sun; Xiangjun Sun; Photooxidative damage in retinal pigment epithelial cells via GRP78 and the protective role of grape skin polyphenols. *Food and Chemical Toxicology* **2014**, *74*, 216-224, <u>10.1016/j.fct.2014.10.001</u>.
- 147. Ludmila F.M.F. Cardozo; Liliana M. Pedruzzi; Peter Stenvinkel; Milena Barcza Stockler-Pinto; Julio Daleprane; Maurilo Leite; Denise Mafra; Nutritional strategies to modulate inflammation and oxidative stress pathways via activation of the master antioxidant switch Nrf2. *Biochimie* **2013**, 95, 1525-1533, 10.1016/j.biochi.2013.04.012.
- 148. Amita Mishra; Amit Kumar Sharma; Shashank Kumar; Ajit K. Saxena; Abhay K. Pandey; Bauhinia variegataLeaf Extracts Exhibit Considerable Antibacterial, Antioxidant, and Anticancer Activities. *BioMed Research International* **2013**, *2013*, 915436, <u>10.1155/2013/915436</u>.
- 149. S Kumar; U K Sharma; A K Sharma; Abhay K Pandey; Protective efficacy of Solanum xanthocarpum root extracts against free radical damage: phytochemical analysis and antioxidant effect.. *Cellular and molecular biology* **2012**, *58*, 174-181, .
- 150. Swapna Upadhyay; Madhulika Dixit; Role of Polyphenols and Other Phytochemicals on Molecular Signaling. *Oxidative Medicine and Cellular Longevity* **2015**, *2015*, 504253, <u>10.1155/2015/504253</u>.
- 151. Dyonne T Hartong; Eliot L Berson; Thaddeus P Dryja; Retinitis pigmentosa. *The Lancet* **2006**, *368*, 1795-1809, <u>10.101</u> <u>6/s0140-6736(06)69740-7</u>.
- 152. Hilda Petrs-Silva; Rafael Linden; Advances in gene therapy technologies to treat retinitis pigmentosa. *Clinical Ophthalmology* **2013**, *8*, 127-136, <u>10.2147/OPTH.S38041</u>.
- 153. Viviana Guadagni; Elena Novelli; Ilaria Piano; Maria Claudia Gargini; Enrica Strettoi; Pharmacological approaches to retinitis pigmentosa: A laboratory perspective. *Progress in Retinal and Eye Research* **2015**, *48*, 62-81, <u>10.1016/j.pretey</u> eres.2015.06.005.
- 154. Virginie Bernier; Daniel G Bichet; Michel Bouvier; Pharmacological chaperone action on G-protein-coupled receptors. *Current Opinion in Pharmacology* **2004**, *4*, 528-533, <u>10.1016/j.coph.2004.08.001</u>.
- 155. Gabriele D'Andrea; Quercetin: A flavonol with multifaceted therapeutic applications?. *Fitoterapia* **2015**, *106*, 256-271, <u>1</u> 0.1016/j.fitote.2015.09.018.
- 156. Tadao Maeda; Zhiqian Dong; Hui Jin; Osamu Sawada; Songqi Gao; Deepank Utkhede; Wendy Monk; Grazyna Palczewska; Krzysztof Palczewski; QLT091001, a 9-cis-Retinal Analog, Is Well-Tolerated by Retinas of Mice with Impaired Visual Cycles. *Investigative Opthalmology & Visual Science* 2013, 54, 455-466, 10.1167/iovs.12-11152.
- 157. Tadao Maeda; Akiko Maeda; Gemma Casadesus; Krzysztof Palczewski; Philippe Margaron; Evaluation of 9-cis-Retinyl Acetate Therapy inRpe65–/–Mice. *Investigative Opthalmology & Visual Science* **2009**, *50*, 4368-4378, <u>10.1167/iovs.09-</u>3700.
- 158. J. Preston Van Hooser; Yan Liang; Tadao Maeda; Vladimir Kuksa; Geeng-Fu Jang; Yu-Guang He; Fred Rieke; Henry K. W. Fong; Peter Detwiler; Krzysztof Palczewski; et al. Recovery of Visual Functions in a Mouse Model of Leber Congenital Amaurosis. *Journal of Biological Chemistry* **2002**, *277*, 19173-19182, 10.1074/jbc.m112384200.