### Inflammaging, an Imbalanced Immune Response

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Nowadays, new advances in society and health have brought an increased life expectancy. However, at the same time, aging comes with complications that impact the development of autoimmunity, neurodegenerative diseases and cancer. These complications affect the quality of life and impact the public health system. Specifically, with aging, a low-grade chronic sterile systemic inflammation with self-reactivity in the absence of acute infection occurs termed inflammaging. Inflammaging is related to an imbalanced immune response that can be either naturally acquired with aging or accelerated due to external triggers. Different molecules, metabolites and inflammatory forms of cell death are highly involved in these processes. Importantly, adoptive cellular immunotherapy is a modality of treatment for cancer patients that administers ex vivo expanded immune cells in the patient. The manipulation of these cells confers them enhanced proinflammatory properties. A general consequence of proinflammatory events is the development of autoimmune diseases and cancer.

Keywords: inflammaging ; immunosenescence ; SASP ; immunotherapy ; T cells ; NK cells

## **1.** Introduction: Immunosenescence and Inflammation during Aging, and its Consequences in Cancer and Other Age-Related Diseases

Nowadays, the elderly population (>65-year-old) in Europe represents 19.7% of the population. This number is predicted to continue increasing and reach 28.5% in 2050 <sup>[1]</sup>. With that in mind, those numbers will impact social life and public healthcare. Thus, a new discipline termed "Geroscience" has emerged to decipher the link between mechanisms of aging and susceptibility to age-related diseases <sup>[2][3]</sup>.

Biologically, aging is associated with a physiological process of tissue degeneration related to chronic inflammation <sup>[4]</sup>. This age-related chronic inflammation is highly associated with inflammaging, which was initially defined as a progressive increase of proinflammation in aged organisms <sup>[5]</sup>, leading to increased morbidity and mortality <sup>[6]</sup>. Currently, inflammaging is defined as the elevated low-grade chronic sterile systemic inflammation with self-reactivity in the elderly in the absence of acute infection <sup>[I]</sup>.

"Immunosenescence", a process associated with aging that impairs the immune function, is highly responsible for inflammaging. Different age-associated events cause immunosenescence. Specifically, during aging, occurs a thymic involution that reduces the pool of naïve T cells and amplifies the oligo-clonal expansion of memory T cells. These events will cause a reduced immune repertoire diversity <sup>[B]</sup>, leading to reduced ability to fight infections and increased cancer incidence <sup>[9]</sup>. Thymic involution also leads to an amplified release of self-reactive T cells and reduced capacity of T-regulatory (reg) cells to suppress these self-reactive T cells and preserve immune homeostasis. Consequently, these events will enhance tissue damage with autoimmunity and chronic inflammation, being essential contributors to inflammaging <sup>[Z][10]</sup>.

Immunosenescence also occurs in the BM, which constitutes the primary site of hematopoiesis <sup>[11]</sup>. Thus, aging causes both a gradual replacement of the different cellular components of the BM by adipocytes and a skew towards the generation of myeloid cells <sup>[12]</sup>. These changes negatively impact the repertoire and activity of T and B lymphocytes <sup>[13]</sup>. Moreover, this cellular degeneration in the BM will increase the production of proinflammatory cytokines <sup>[12][14][15]</sup>, impacting the activity of immune cells.

The innate immune system is also impacted by immunosenescence. Thus, neutrophil and macrophage capacity for phagocytosis and subsequent elimination of dead cells is reduced with aging <sup>[16][17]</sup>. Macrophages also acquire an increased polarization towards M2 cells <sup>[18]</sup>, and natural killer (NK) cells present a reduced capacity to secrete cytotoxic molecules <sup>[19]</sup>.

Overall, immunosenescent cells will not be able to remove senescent somatic cells that also accumulate with aging  $^{[20][21]}$   $^{[22][23][24]}$  and are characterized by secretion of proinflammatory molecules known as senescence-associated secretory phenotype (SASP)  $^{[25]}$ . The SASP is another crucial contributor to inflammaging  $^{[I][25]}$ . This accumulation of senescent cells will enhance the SASP promoting further inflammaging and accelerated aging  $^{[22]}$  and will contribute to cancer development  $^{[I][25][26]}$ . Furthermore, the SASP transmits cellular senescence to neighboring non-senescent cells  $^{[27][28]}$ , leading to enhanced senescence and inflammaging. The SASP is also increased with anti-cancer therapies that induce senescence in both immune and tumor cells, leading to enhanced inflammation and treatment resistance  $^{[29][30]}$ .

Moreover, microbes debris of exogenous origin and cell debris of endogenous origin are recognized through the pathogen-associated molecular pattern (PAMPs) and damage-associated molecular patterns (DAMPs), respectively <sup>[31]</sup>, the latter being part of the SASP <sup>[32]</sup>. PAMPs and DAMPs become more abundant during aging, and PAMP stimulation induces DAMP secretion by immune cells <sup>[33]</sup>, leading to enhanced inflammaging.

This feedback occurring between immunosenescence and inflammaging explains the involvement of both processes in age-related diseases, including cancer, neurodegenerative diseases, metabolic diseases and cardiovascular diseases <sup>[Z]</sup> (see **Table 1**). For instance, Alzheimer's disease is a chronic neurodegenerative disease with pathological accumulation of amyloid-beta (A $\beta$ ) peptides and neurofibrillary tangles containing tau protein. A $\beta$  and tau deposition cause an age-dependent deterioration of the blood-brain barrier that leads to the infiltration of immune cells into the central nervous system exacerbating the neurodegenerative process and promoting inflammatory responses <sup>[33]</sup>. Type-2 diabetes is a multifactorial metabolic disease with chronic hyperglycemia and dyslipidemia as main pathological features. A chronic low-grade inflammation resembling inflammaging induces insulin resistance and dysfunction of  $\beta$ -cells, emerging as a relevant factor contributing to the development of diabetes <sup>[34]</sup>.

| Age-Related Diseases                         | Mediators  | References                  |
|--|--|-----------------------------|
| Atherosclerosis                              | Secretion of IL1 $\beta$ , IL18 and IL6 among others | [7][35]                     |
| Cardiovascular diseases                      | CRP and IL6 in blood                                 | [Z]                         |
| Frailty, Sarcopenia                          | Inflammatory markers in blood, IL6                   | [36]                        |
| Decline of innate and adaptive immune system | Immunosenescence                                     | [8][9]                      |
| Type 2 diabetes                              | Secretion of IL1β among others                       | [34][37][38]                |
| Cancer                                       | CRP, IL6, immunosenescence                           | [7][25][26][39][40][41][42] |
| Osteoporosis, bone remodeling                | IL1, IL6, TNFα                                       | [43]                        |
| Neurodegenerative disease                    | Immune cells infiltration                            | [33]                        |

Table 1. Side effects of inflammaging.

In cancer, aging and chronic inflammation are highly involved in its development <sup>[39][40]</sup>. However, the intricate relationship between aging and cancer is not clear. In detail, half of the cancers occur in individuals older than 70. Yet, whereas aging and cancer share disease mechanisms, such as genomic instability, they also present opposite features, such as hypoactive cells in aging vs. hyperactive cells in cancer <sup>[40]</sup>. The role of chronic inflammation in cancer is also controversial. Thus, inflammation is required initially for immune surveillance; however, failure to resolve inflammation will promote tumor growth <sup>[44][45]</sup>. The relevant impact of chronic inflammation in cancer is suggested by different studies that estimate that 15–20% of cancers are inflammation-related <sup>[46]</sup>.

For instance, autoimmune diseases such asinflammatory bowel disease (IBD) increase the risk of developing colorectal cancer <sup>[47]</sup>. Moreover, numerous studies have revealed associations of high levels of inflammatory markers, such as C-reactive protein (CRP) <sup>[41]</sup> and IL6 <sup>[42]</sup>, with an increased risk of developing different types of cancer.

The relevant role of chronic inflammation in cancer and of the immune response in the development of inflammaging should be considered incancer patients treated with adoptive cellular immunotherapy. These treatments administer various immune cells in patients, such as chimeric antigen receptor (CAR)-T cells <sup>[48]</sup>, tumor-infiltrating lymphocyte (TIL) <sup>[49]</sup> or NK cells <sup>[50]</sup> which previously have been modified and expanded in vitro. The in vitro expansion changes the phenotype of immune cells and their cytotoxic mechanisms that activate inflammatory forms of cell death <sup>[50][51][52][53]</sup>. For instance, after encountering tumor cells, CAR-T cells <sup>[53]</sup> and NK cells <sup>[41]</sup> initiate pyroptosis, an inflammatory form of cell death. Pyroptosis was initially described as a type of cell death triggered by the innate immune system after recognition of

intracellular pathogens by intracellular receptors. Nucleotide-binding oligomerization domain (NOD)-like receptors (NLRs), and among them NLRP3, belong to these receptors. They initiate the assembly of inflammasomes that will activate caspase-1 leading to release of IL1β and IL18 and the pore-forming protein gasdermin-D (GSDMD) the latter inducing pyroptosis <sup>[54]</sup>. Of interest, in CAR-T cellimmunotherapy, NLRP3 activates pyroptosis with release of DAMPs, IL1β and IL6 <sup>[53]</sup>. In addition, immune cells after encountering tumor cells release different types of Granzymes (Gzm). Besides the classic GzmB, other inflammatory Gzm, such as GzmA and GzmK, are involved in the anti-tumor activity of immune cells <sup>[55]</sup>.

These relevant associations of inflammaging with an inadequate immune response and the development of inflammatory diseases and cancer, added to the fact that cancer associates with aging suggest their relevance in the field of cellular immunotherapy. Here, we will review the contribution to inflammaging of different subsets of T cells and NK cells, as they are administered in cancer patients, either unmodified or modified with a CAR <sup>[51][57][58]</sup>. Moreover, the role of NLRP3 and inflammatory granzymes, activated during the innate and adaptive immune response, will be presented, focusing on their impact on inflammation. Other intrinsic and external inflammation triggers related to cancer will be mentioned, and some preclinical models that associate inflammation with cancer development will be cited.

# 2. Variation of T-reg Cells and Th17 Cells during Aging and Their Impact on the Development of Inflammaging

Human centenarians represent a model with low inflammaging to study healthy aging. Of interest, although those human centenarians present a systemic inflammatory state (e.g., high levels of IL6 and IL8 in plasma), they also count on efficient anti-inflammatory networks termed anti-inflammaging that compensate for inflammaging <sup>[59]</sup>. Analyses of the immune cell populations in centenarians have concluded that longer survivors present higher leucocytes with a higher number of naïve, activated/memory and effector/memory CD4 and CD8 T cells <sup>[60]</sup>. Proteomic studies in centenarians also show a pattern with less inflammaging and autoimmunity, increased B cell-mediated immune response, higher expression of proteins involved in angiogenesis and enhanced intercellular junctions <sup>[61]</sup>.On the other side, elderly cancer patients, such as multiple myeloma (MM), present immunosenescent T cells with deficient immune responses <sup>[62]</sup> that will increase inflammaging.

Among the different subsets of T lymphocytes, we will focus on T-reg and Th17 cells that share a common precursor and present opposing roles in developing inflammaging. Thus, Th17 cells cause autoimmunity and inflammation, and T-reg cells inhibit their activity <sup>[63]</sup>. Specifically, during aging, there is an increased production of Th17 cells that will contribute to inflammaging <sup>[64]</sup> and a decrease in the functionality of T-reg cells that will increase chronic inflammation <sup>[65]</sup>. Even though Th17 cells are very well-known for their role in inflammation and autoimmunity, their role in cancer is less understood. Notably, an intricate balance between T-regs and Th17 cells must be maintained to avoid developing these pathologies <sup>[64]</sup>[66][67].

#### 2.1. Changes in the T-reg Cell Compartment during Aging and Impact in Inflammation and Cancer

The impact of T-reg cells during aging should be analyzed considering the variation in numbers and their functionality. As previously mentioned, thymic involution with aging reduces the capacity of T-reg cells to suppress self-reactive T cells and preserve immune homeostasis <sup>[7][10]</sup>. Two different origins have been described for T-reg cells. The first one is the thymus, at the early stages of life, which gives rise to naturally occurring T-reg (nT-reg) cells after escaping from the negative selection in the thymus, followed by appropriate TCR stimulation. The second one is in the peripheral blood (PB) and secondary lymphoid organs, where different triggers induce the expression of Foxp3 in naïve T cells, originating inducible T-reg (iT-reg) cells. iT-reg cells have a similar phenotype and suppressive function to nT-regs <sup>[68]</sup>. Data suggest that aging induces a decline in iT-regs and an increase in the number of nT-reg cells <sup>[69]</sup>.

Regarding the functionality, it remains controversial whether aging induces a loss of T-reg functionality or just an effect of the variation in the number of T-reg cells <sup>[69]</sup>. Studies in aged mice have observed an increased proportion of functional CD4 T-regs in PB and lymphoid tissues, decreasing the effector T cell responses against *Leishmania* infection <sup>[70]</sup>. In humans, there is also an increase in the number of CD4 T-regs in PB with immunosuppressive properties <sup>[71]</sup>. In addition, the increased number of T-reg cells with aging can be explained by the polarization of CD4 conventional T cells to cells with T-reg cell properties, an event observed in aged mice <sup>[72]</sup>. Moreover, CD8 T-reg cells are a relevant immunosuppressive cell population <sup>[73]</sup>, increasing with aging in absolute numbers in PB, the spleen and lymph nodes presenting functionality <sup>[74][75]</sup>.

Various studies have associated the functionality of T-regs with the progression of different tumors due to their immunosuppressive activity <sup>[76][77]</sup>. Thus, in melanoma and colon carcinoma models, intratumoral T-reg cells inhibit the anti-tumor activity of TILs <sup>[78]</sup>. In these models, T-reg cells can adapt to the lactic acid-enriched TME through CD36 up-regulation that enhances their mitochondrial fitness <sup>[78]</sup>. In MM, where most patients represent an elderly cancer population, elevated frequencies of functional T-reg cells are present in newly diagnosed and relapsed patients compared to healthy volunteers <sup>[79]</sup>.

On the other side, autoimmunity and chronic low-grade inflammation, both hallmarks of inflammaging <sup>[80]</sup>, are also recognized as drivers of cancer <sup>[39][46]</sup>. In this scenario, murine models of autoimmunity have shown the beneficial impact of T-reg cells ameliorating inflammation. For instance, in models of multiple sclerosis, T-reg cells produce CCL1 that upregulates its receptor, CCR8, and induces the expression of CD39, granzyme B and IL10, which suppress the disease <sup>[81]</sup>. In autoimmune colitis, aged T-reg cells present equal suppressive in vitro activity than young T-regs to mitigate the disease. In these models, aged T-reg cells were able to restrain IFN- $\gamma$  T cell responses. Even though, they controlled Th17 cells only in cases of acute inflammation and not in cases of chronic inflammation, leading to autoimmunity and promoting colitis <sup>[82]</sup>. T-reg cells also present contradictory roles in inflammation. Thus, IBD is another autoimmune disease that increases the risk of developing colorectal cancer <sup>[47]</sup>. In this scenario, different murine models have demonstrated the protective role of T-reg cells in IBD development through the suppression of T effector cells. In detail, IL35 secretion by T-reg cells suppresses the proliferation of effector T cells. However, on the other side, IL35 overexpression associates with the induction of gastrointestinal cancer <sup>[83][84]</sup>.

Another relevant model that contradicts the relationship of chronic inflammation mediated by T lymphocytes and cancer and where T-reg cells are involved is the graft versus host disease (GVHD). Chronic GVHD (cGVHD) is a relevant complication after allogeneic stem cell transplantation (allo-SCT) mediated by donor's T lymphocytes that enhances mortality due to a chronic inflammatory response and at the same time reduces the risk of cancer relapse <sup>[85]</sup>. T-reg cells associate with reduced development of GVHD <sup>[86]</sup>. Of interest, pediatric allo-SCT recipients have a lower incidence of cGvHD than adults <sup>[87]</sup>, which might reflect in this context the beneficial impact of lower immunosenescence levels in pediatric patients compared to adult patients. Indeed, it has been observed that cGVHD-derived T-cells present high expression of genes that positively regulate cellular senescence (*CDKN2A*, *SERPINB9*, *LYPLA1* and *CKTM1A/B*) <sup>[88]</sup>.

To summarize, two opposite scenarios,"enhanced immunosuppression and chronic inflammation", associate with cancer, and T-reg cells play either a detrimental or beneficial role in both systems. These findings bring the question of the exact contribution of T-reg cells in the regulation of inflammation and cancer development, specifically in the elderly.

#### 2.2. Th17 Compartment and Its Delicate Balance with T-reg Cells

Th17 cells are critical players in maintaining mucosal immune homeostasis and protection against pathogens. They are also very well-known for their role in inflammation and autoimmunity. An intricate balance between T-regs and Th17 cells is maintained to avoid developing these pathologies <sup>[66][67]</sup>. A common precursor for T-reg cells and Th17 cells will differentiate into one cell subtype depending on the cytokine environment <sup>[67]</sup>. In detail, TGF $\beta$  is required for differentiation from naïve CD4 T cells to both Th17 and iTreg. Thus, TGF $\beta$  upregulates the retinoic acid-related orphan receptors-yt (RORyt) and Foxp3, which give rise to a common precursor of T-regs and Th17 cells. In the presence of TGF- $\beta$ , both IL6 and IL21 induce differentiation to Th17 cells. Otherwise, T cells will differentiate to T-reg cells. Moreover, Foxp3 inhibits Th17 development through binding to RORyt. Without IL6, TGF $\beta$  reinforces this inhibition and favors the formation of T-reg cells. In addition, Th17 and T-reg cells can also polarize to each other <sup>[67]</sup>.

The role of T-reg cells in maintaining the number of Th17 cells has been observed in different contexts. For instance, intestinal T-reg cells constrain microbiota-dependent IL-17-production by Th17 cells. This activity is dependent on the transcription factor c-Maf that controls IL10 production by T-reg cells <sup>[89]</sup>. In a murine model of neuroinflammation, imaging of T-reg and Th17 cells in the spinal cord demonstrated that T-regs suppress Th17 cells by inhibiting Ca<sup>2+</sup> signaling and limiting the access of Th17 cells to APCs, avoiding neuroinflammation <sup>[90]</sup>. On the contrary, in hepatic carcinoma, increased Th17 levels are detected in the PB, correlating positively with metastasis progression and T-reg cells in the TME <sup>[91]</sup>.

Altogether, T-reg and Th17 cells present opposite roles with an intricate regulation between them. Monitoring their changes in elderly cancer patients and patients receiving adoptive cellular immunotherapy will provide relevant information in this field.

#### 2.3. Changes in the Th17 Compartment during Aging and Implications for Autoimmunity and Cancer

Aging causes an increased Th17/T-reg ratio that contributes to inflammaging <sup>[92]</sup>. Indeed, older subjects present higher Th17 cytokine production than younger subjects. One of the causes described is defective autophagy in CD4 T cells occurring with aging, leading to reduced mitophagy with an accumulation of malfunctioning mitochondria. These events result in the upregulation of Th17 cytokines contributing to inflammaging <sup>[64][93]</sup>. The detrimental impact of this higher Th17/T-reg ratio in cancer is observed at specific stages of tumors. Thus, oral squamous cell carcinoma patients increase the Th17/T-reg ratio at early stages and decrease it at late stages <sup>[94]</sup>. In colorectal tumor specimens, patients with increased expression of Th17 genes presented a poor prognosis <sup>[95]</sup>. Others have found that the increased IL1 $\beta$  and IL2 reduction in aged mice contributed to an elevated Th17 differentiation <sup>[96]</sup>.

In MM, a variety of studies confirm the detrimental role of Th17 cells. Thus, Th17 cells promote MM growth and inhibit immune functions <sup>[97]</sup>; and in MM patients with lytic bone disease, numbers of Th17 cells were the highest <sup>[98]</sup>. Of interest, IL6, which is over-expressed in MM, creates a proinflammatory TME, a crucial factor mediating the conversion of T-regs into Th17 cells <sup>[99]</sup>. Th17 cells also cause osteoclast-dependent bone damage in vitro and in vivo, where miR-21 activates differentiation of naïve T cells in Th17 cells, promoting these detrimental effects in MM <sup>[100]</sup>. IL17, produced by Th17, cells induces osteoblasts pyroptosis in vitro, through activation of the NLRP3 inflammasome complex with Caspase-1 execution and release of IL1 $\beta$  <sup>[101]</sup>. In newly diagnosed MM patients, Th17 cell levels fluctuate considerably. Of interest, Th17 increased further when the disease reached partial remission, decreased to normal levels when complete remission was achieved and increased again when the disease recurred <sup>[102]</sup>.

Moreover, in MM, dendritic cells (DCs) infiltrate the BM as efficient inducers of Th17 cells and promote higher levels of Th17 in BM than PB. Of interest, in monoclonal gammopathy of undetermined significance (MGUS) patients, an initial stage of the MM disease, this increase in Th17 cells was not observed. Another study analyzing the microbiota in MM observed that *Prevotella heparinolytica* promotes the differentiation of Th17 cells that colonize the gut and migrate to the BM, to favor the progression of MM. Similarly, in smoldering MM patients, higher BM IL17 levels predicted faster disease progression<sup>[103]</sup>.

Moreover, the imbalance of the Th17/T-reg ratio in MM is reinforced by studies, where MM and MGUS patients show a reduction in the number of T-reg cells compared to healthy donors, being these T-reg cells dysfunctional <sup>[104]</sup>. Another study observed fewer T-regs in the BM of MM patients compared to healthy individuals, where Th17 cells are responsible for osteoclast activation mediating lytic bone disease <sup>[105]</sup>.

To summarize, Th17 cells are highly involved in this connection between chronic inflammation and cancer development. Moreover, they are related to different types of cancer and to the pathogenic events of MM patients, who represent elderly cancer patients. Novel studies are required to decipher their role in the progression of these diseases.

#### References

- 1. Ageing Europe. Available online: https://ec.europa.eu/eurostat/web/products-statistical-books/-/ks-02-19-681 (accessed on 23 September 2021).
- 2. Kennedy, B.K.; Berger, S.L.; Brunet, A.; Campisi, J.; Cuervo, A.M.; Epel, E.S.; Franceschi, C.; Lithgow, G.J.; Morimoto, R.I.; Pessin, J.E.; et al. Geroscience: Linking Aging to Chronic Disease. Cell 2014, 159, 709–713.
- 3. Sierra, F. The Emergence of Geroscience as an Interdisciplinary Approach to the Enhancement of Health Span and Life Span. Cold Spring Harb. Perspect. Med. 2016, 6, a025163.
- 4. López-Otín, C.; Blasco, M.A.; Partridge, L.; Serrano, M.; Kroemer, G. The Hallmarks of Aging. Cell 2013, 153, 1194– 1217.
- 5. Franceschi, C.; Bonafè, M.; Valensin, S.; Olivieri, F.; De Luca, M.; Ottaviani, E.; De Benedictis, G. Inflamm-Aging. An Evolutionary Perspective on Immunosenescence. Ann. N. Y. Acad. Sci. 2000, 908, 244–254.
- 6. Franceschi, C.; Campisi, J. Chronic Inflammation (Inflammaging) and Its Potential Contribution to Age-Associated Diseases. J. Gerontol. A Biol. Sci. Med. Sci. 2014, 69 (Suppl. 1), S4–S9.
- 7. Thomas, R.; Wang, W.; Su, D.-M. Contributions of Age-Related Thymic Involution to Immunosenescence and Inflammaging. Immun. Ageing 2020, 17, 2.
- Drabkin, M.J.; Meyer, J.I.; Kanth, N.; Lobel, S.; Fogel, J.; Grossman, J.; Krumenacker, J.H. Age-Stratified Patterns of Thymic Involution on Multidetector CT. J. Thorac. Imaging 2018, 33, 409–416.
- 9. Zinger, A.; Cho, W.C.; Ben-Yehuda, A. Cancer and Aging—The Inflammatory Connection. Aging Dis. 2017, 8, 611–627.

- Coder, B.D.; Wang, H.; Ruan, L.; Su, D.-M. Thymic Involution Perturbs Negative Selection Leading to Autoreactive T Cells That Induce Chronic Inflammation. J. Immunol. 2015, 194, 5825–5837.
- 11. Fröbel, J.; Landspersky, T.; Percin, G.; Schreck, C.; Rahmig, S.; Ori, A.; Nowak, D.; Essers, M.; Waskow, C.; Oostendorp, R.A.J. The Hematopoietic Bone Marrow Niche Ecosystem. Front. Cell Dev. Biol. 2021, 9, 705410.
- 12. Rodrigues, L.P.; Teixeira, V.R.; Alencar-Silva, T.; Simonassi-Paiva, B.; Pereira, R.W.; Pogue, R.; Carvalho, J.L. Hallmarks of Aging and Immunosenescence: Connecting the Dots. Cytokine Growth Factor Rev. 2021, 59, 9–21.
- 13. Beerman, I.; Maloney, W.J.; Weissmann, I.L.; Rossi, D.J. Stem Cells and the Aging Hematopoietic System. Curr. Opin. Immunol. 2010, 22, 500–506.
- 14. Zheng, X.; Wang, Q.; Xie, Z.; Li, J. The Elevated Level of IL-1α in the Bone Marrow of Aged Mice Leads to MSC Senescence Partly by down-Regulating Bmi-1. Exp. Gerontol. 2021, 148, 111313.
- 15. Wang, J.; Chen, J.; Zhang, B.; Jia, X. IL-6 Regulates the Bone Metabolism and Inflammatory Microenvironment in Aging Mice by Inhibiting Setd7. Acta Histochem. 2021, 123, 151718.
- 16. Aprahamian, T.; Takemura, Y.; Goukassian, D.; Walsh, K. Ageing Is Associated with Diminished Apoptotic Cell Clearance in Vivo. Clin. Exp. Immunol. 2008, 152, 448–455.
- 17. Gasparoto, T.H.; Dalboni, T.M.; Amôr, N.G.; Abe, A.E.; Perri, G.; Lara, V.S.; Vieira, N.A.; Gasparoto, C.T.; Campanelli, A.P. Fcy Receptors on Aging Neutrophils. J. Appl. Oral. Sci. 2021, 29, e20200770.
- 18. Van Beek, A.A.; Van den Bossche, J.; Mastroberardino, P.G.; de Winther, M.P.J.; Leenen, P.J.M. Metabolic Alterations in Aging Macrophages: Ingredients for Inflammaging? Trends Immunol. 2019, 40, 113–127.
- 19. Solana, R.; Campos, C.; Pera, A.; Tarazona, R. Shaping of NK Cell Subsets by Aging. Curr. Opin. Immunol. 2014, 29, 56–61.
- 20. Muñoz-Espín, D.; Serrano, M. Cellular Senescence: From Physiology to Pathology. Nat. Rev. Mol. Cell Biol. 2014, 15, 482–496.
- 21. Jeyapalan, J.C.; Ferreira, M.; Sedivy, J.M.; Herbig, U. Accumulation of Senescent Cells in Mitotic Tissue of Aging Primates. Mech. Ageing Dev. 2007, 128, 36–44.
- 22. Ovadya, Y.; Landsberger, T.; Leins, H.; Vadai, E.; Gal, H.; Biran, A.; Yosef, R.; Sagiv, A.; Agrawal, A.; Shapira, A.; et al. Impaired Immune Surveillance Accelerates Accumulation of Senescent Cells and Aging. Nat. Commun. 2018, 9, 5435.
- 23. Xue, W.; Zender, L.; Miething, C.; Dickins, R.A.; Hernando, E.; Krizhanovsky, V.; Cordon-Cardo, C.; Lowe, S.W. Senescence and Tumour Clearance Is Triggered by P53 Restoration in Murine Liver Carcinomas. Nature 2007, 445, 656–660.
- 24. Sagiv, A.; Biran, A.; Yon, M.; Simon, J.; Lowe, S.W.; Krizhanovsky, V. Granule Exocytosis Mediates Immune Surveillance of Senescent Cells. Oncogene 2013, 32, 1971–1977.
- 25. Battram, A.M.; Bachiller, M.; Martín-Antonio, B. Senescence in the Development and Response to Cancer with Immunotherapy: A Double-Edged Sword. Int. J. Mol. Sci. 2020, 21, 4346.
- 26. Tsukishiro, T.; Donnenberg, A.D.; Whiteside, T.L. Rapid Turnover of the CD8(+)CD28(-) T-Cell Subset of Effector Cells in the Circulation of Patients with Head and Neck Cancer. Cancer Immunol. Immunother. 2003, 52, 599–607.
- Acosta, J.C.; Banito, A.; Wuestefeld, T.; Georgilis, A.; Janich, P.; Morton, J.P.; Athineos, D.; Kang, T.-W.; Lasitschka, F.; Andrulis, M.; et al. A Complex Secretory Program Orchestrated by the Inflammasome Controls Paracrine Senescence. Nat. Cell Biol. 2013, 15, 978–990.
- 28. Hoare, M.; Narita, M. Transmitting Senescence to the Cell Neighbourhood. Nat. Cell Biol 2013, 15, 887–889.
- Onyema, O.O.; Decoster, L.; Njemini, R.; Forti, L.N.; Bautmans, I.; De Waele, M.; Mets, T. Chemotherapy-Induced Changes and Immunosenescence of CD8+ T-Cells in Patients with Breast Cancer. Anticancer Res. 2015, 35, 1481– 1489.
- 30. Bruni, E.; Cazzetta, V.; Donadon, M.; Cimino, M.; Torzilli, G.; Spata, G.; Leonardi, G.; Dieli, F.; Mikulak, J.; Mavilio, D. Chemotherapy Accelerates Immune-Senescence and Functional Impairments of Vδ2pos T Cells in Elderly Patients Affected by Liver Metastatic Colorectal Cancer. J. Immunother. Cancer 2019, 7, 347.
- 31. Martín-Antonio, B.; Granell, M.; Urbano-Ispizua, A. Genomic Polymorphisms of the Innate Immune System and Allogeneic Stem Cell Transplantation. Expert Rev. Hematol. 2010, 3, 411–427.
- Basisty, N.; Kale, A.; Jeon, O.H.; Kuehnemann, C.; Payne, T.; Rao, C.; Holtz, A.; Shah, S.; Sharma, V.; Ferrucci, L.; et al. A Proteomic Atlas of Senescence-Associated Secretomes for Aging Biomarker Development. PLoS Biol. 2020, 18, e3000599.

- 33. Zenaro, E.; Piacentino, G.; Constantin, G. The Blood-Brain Barrier in Alzheimer's Disease. Neurobiol. Dis. 2017, 107, 41–56.
- Prattichizzo, F.; De Nigris, V.; Spiga, R.; Mancuso, E.; La Sala, L.; Antonicelli, R.; Testa, R.; Procopio, A.D.; Olivieri, F.; Ceriello, A. Inflammageing and Metaflammation: The Yin and Yang of Type 2 Diabetes. Ageing Res. Rev. 2018, 41, 1– 17.
- Dinarello, C.A. Interleukin-1 in the Pathogenesis and Treatment of Inflammatory Diseases. Blood 2011, 117, 3720– 3732.
- Fabbri, E.; An, Y.; Zoli, M.; Simonsick, E.M.; Guralnik, J.M.; Bandinelli, S.; Boyd, C.M.; Ferrucci, L. Aging and the Burden of Multimorbidity: Associations with Inflammatory and Anabolic Hormonal Biomarkers. J. Gerontol. A Biol. Sci. Med. Sci. 2015, 70, 63–70.
- 37. Masters, S.L.; Dunne, A.; Subramanian, S.L.; Hull, R.L.; Tannahill, G.M.; Sharp, F.A.; Becker, C.; Franchi, L.; Yoshihara, E.; Chen, Z.; et al. Activation of the NLRP3 Inflammasome by Islet Amyloid Polypeptide Provides a Mechanism for Enhanced IL-1β in Type 2 Diabetes. Nat. Immunol. 2010, 11, 897–904.
- Stienstra, R.; Joosten, L.A.B.; Koenen, T.; van Tits, B.; van Diepen, J.A.; van den Berg, S.A.A.; Rensen, P.C.N.; Voshol, P.J.; Fantuzzi, G.; Hijmans, A.; et al. The Inflammasome-Mediated Caspase-1 Activation Controls Adipocyte Differentiation and Insulin Sensitivity. Cell Metab. 2010, 12, 593–605.
- 39. Grivennikov, S.I.; Greten, F.R.; Karin, M. Immunity, Inflammation, and Cancer. Cell 2010, 140, 883–899.
- 40. Aunan, J.R.; Cho, W.C.; Søreide, K. The Biology of Aging and Cancer: A Brief Overview of Shared and Divergent Molecular Hallmarks. Aging Dis. 2017, 8, 628–642.
- 41. Van't Klooster, C.C.; Ridker, P.M.; Hjortnaes, J.; van der Graaf, Y.; Asselbergs, F.W.; Westerink, J.; Aerts, J.G.J.V.; Visseren, F.L.J. On behalf of the UCC-SMART study group The Relation between Systemic Inflammation and Incident Cancer in Patients with Stable Cardiovascular Disease: A Cohort Study. Eur. Heart J. 2019, 40, 3901–3909.
- 42. Heikkilä, K.; Harris, R.; Lowe, G.; Rumley, A.; Yarnell, J.; Gallacher, J.; Ben-Shlomo, Y.; Ebrahim, S.; Lawlor, D.A. Associations of Circulating C-Reactive Protein and Interleukin-6 with Cancer Risk: Findings from Two Prospective Cohorts and a Meta-Analysis. Cancer Causes Control 2009, 20, 15–26.
- Pietschmann, P.; Mechtcheriakova, D.; Meshcheryakova, A.; Föger-Samwald, U.; Ellinger, I. Immunology of Osteoporosis: A Mini-Review. Gerontology 2016, 62, 128–137.
- 44. Zhang, Q.; Zhu, B.; Li, Y. Resolution of Cancer-Promoting Inflammation: A New Approach for Anticancer Therapy. Front. Immunol. 2017, 8, 71.
- 45. Hanahan, D.; Weinberg, R.A. Hallmarks of Cancer: The Next Generation. Cell 2011, 144, 646–674.
- 46. Mantovani, A.; Allavena, P.; Sica, A.; Balkwill, F. Cancer-Related Inflammation. Nature 2008, 454, 436–444.
- 47. Waldner, M.J.; Neurath, M.F. Colitis-Associated Cancer: The Role of T Cells in Tumor Development. Semin. Immunopathol. 2009, 31, 249–256.
- Maude, S.L.; Laetsch, T.W.; Buechner, J.; Rives, S.; Boyer, M.; Bittencourt, H.; Bader, P.; Verneris, M.R.; Stefanski, H.E.; Myers, G.D.; et al. Tisagenlecleucel in Children and Young Adults with B-Cell Lymphoblastic Leukemia. N. Engl. J. Med. 2018, 378, 439–448.
- 49. Wang, S.; Sun, J.; Chen, K.; Ma, P.; Lei, Q.; Xing, S.; Cao, Z.; Sun, S.; Yu, Z.; Liu, Y.; et al. Perspectives of Tumor-Infiltrating Lymphocyte Treatment in Solid Tumors. BMC Med. 2021, 19, 140.
- Shah, N.; Martin-Antonio, B.; Yang, H.; Ku, S.; Lee, D.A.; Cooper, L.J.N.; Decker, W.K.; Li, S.; Robinson, S.N.; Sekine, T.; et al. Antigen Presenting Cell-Mediated Expansion of Human Umbilical Cord Blood Yields Log-Scale Expansion of Natural Killer Cells with Anti-Myeloma Activity. PLoS One 2013, 8, e76781.
- 51. Bachiller, M.; Battram, A.M.; Perez-Amill, L.; Martín-Antonio, B. Natural Killer Cells in Immunotherapy: Are We Nearly There? Cancers 2020, 12, 3139.
- 52. Martín-Antonio, B.; Suñe, G.; Najjar, A.; Perez-Amill, L.; Antoñana-Vildosola, A.; Castella, M.; León, S.; Velasco-de Andrés, M.; Lozano, F.; Lozano, E.; et al. Extracellular NK Histones Promote Immune Cell Anti-Tumor Activity by Inducing Cell Clusters through Binding to CD138 Receptor. J. Immunother. Cancer 2019, 7, 259.
- 53. Liu, Y.; Fang, Y.; Chen, X.; Wang, Z.; Liang, X.; Zhang, T.; Liu, M.; Zhou, N.; Lv, J.; Tang, K.; et al. Gasdermin E-Mediated Target Cell Pyroptosis by CAR T Cells Triggers Cytokine Release Syndrome. Sci. Immunol. 2020, 5.
- 54. Wen, J.; Xuan, B.; Liu, Y.; Wang, L.; He, L.; Meng, X.; Zhou, T.; Wang, Y. Updating the NLRC4 Inflammasome: From Bacterial Infections to Autoimmunity and Cancer. Front. Immunol. 2021, 12, 702527.
- 55. Zhou, Z.; He, H.; Wang, K.; Shi, X.; Wang, Y.; Su, Y.; Wang, Y.; Li, D.; Liu, W.; Zhang, Y.; et al. Granzyme A from Cytotoxic Lymphocytes Cleaves GSDMB to Trigger Pyroptosis in Target Cells. Science 2020, 368, eaaz7548.

- Martín-Antonio, B.; Suñe, G.; Perez-Amill, L.; Castella, M.; Urbano-Ispizua, A. Natural Killer Cells: Angels and Devils for Immunotherapy. Int. J. Mol. Sci. 2017, 18, 1868.
- 57. Perez-Amill, L.; Marzal, B.; Urbano-Ispizua, A.; Juan, M.; Martín-Antonio, B. CAR-T Cell Therapy: A Door Is Open to Find Innumerable Possibilities of Treatments for Cancer Patients. Turk. J. Haematol. 2018, 35, 217–228.
- Liu, E.; Marin, D.; Banerjee, P.; Macapinlac, H.A.; Thompson, P.; Basar, R.; Nassif Kerbauy, L.; Overman, B.; Thall, P.; Kaplan, M.; et al. Use of CAR-Transduced Natural Killer Cells in CD19-Positive Lymphoid Tumors. N. Engl. J. Med. 2020, 382, 545–553.
- 59. Franceschi, C.; Capri, M.; Monti, D.; Giunta, S.; Olivieri, F.; Sevini, F.; Panourgia, M.P.; Invidia, L.; Celani, L.; Scurti, M.; et al. Inflammaging and Anti-Inflammaging: A Systemic Perspective on Aging and Longevity Emerged from Studies in Humans. Mech. Ageing Dev. 2007, 128, 92–105.
- 60. Bucci, L.; Ostan, R.; Giampieri, E.; Cevenini, E.; Pini, E.; Scurti, M.; Vescovini, R.; Sansoni, P.; Caruso, C.; Mari, D.; et al. Immune Parameters Identify Italian Centenarians with a Longer Five-Year Survival Independent of Their Health and Functional Status. Exp. Gerontol. 2014, 54, 14–20.
- Santos-Lozano, A.; Valenzuela, P.L.; Llavero, F.; Lista, S.; Carrera-Bastos, P.; Hampel, H.; Pareja-Galeano, H.; Gálvez, B.G.; López, J.A.; Vázquez, J.; et al. Successful Aging: Insights from Proteome Analyses of Healthy Centenarians. Aging 2020, 12, 3502–3515.
- Zelle-Rieser, C.; Thangavadivel, S.; Biedermann, R.; Brunner, A.; Stoitzner, P.; Willenbacher, E.; Greil, R.; Jöhrer, K. T Cells in Multiple Myeloma Display Features of Exhaustion and Senescence at the Tumor Site. J. Hematol. Oncol. 2016, 9, 116.
- 63. Lee, G.R. The Balance of Th17 versus Treg Cells in Autoimmunity. Int. J. Mol. Sci. 2018, 19, 730.
- 64. Kroemer, G.; Zitvogel, L. CD4+ T Cells at the Center of Inflammaging. Cell Metab. 2020, 32, 4–5.
- 65. Thomas, R.; Oh, J.; Wang, W.; Su, D.-M. Thymic Atrophy Creates Holes in Treg-Mediated Immuno-Regulation via Impairment of an Antigen-Specific Clone. Immunology 2021, 163, 478–492.
- 66. Knochelmann, H.M.; Dwyer, C.J.; Bailey, S.R.; Amaya, S.M.; Elston, D.M.; Mazza-McCrann, J.M.; Paulos, C.M. When Worlds Collide: Th17 and Treg Cells in Cancer and Autoimmunity. Cell Mol. Immunol. 2018, 15, 458–469.
- 67. Sun, L.; Fu, J.; Zhou, Y. Metabolism Controls the Balance of Th17/T-Regulatory Cells. Front. Immunol. 2017, 0.
- Feuerer, M.; Hill, J.A.; Mathis, D.; Benoist, C. Foxp3 + Regulatory T Cells: Differentiation, Specification, Subphenotypes. Nat. Immunol. 2009, 10, 689–695.
- Jagger, A.; Shimojima, Y.; Goronzy, J.J.; Weyand, C.M. Regulatory T Cells and the Immune Aging Process: A Mini-Review. Gerontology 2014, 60, 130–137.
- Lages, C.S.; Suffia, I.; Velilla, P.A.; Huang, B.; Warshaw, G.; Hildeman, D.A.; Belkaid, Y.; Chougnet, C. Functional Regulatory T Cells Accumulate in Aged Hosts and Promote Chronic Infectious Disease Reactivation. J. Immunol. 2008, 181, 1835–1848.
- Gregg, R.; Smith, C.M.; Clark, F.J.; Dunnion, D.; Khan, N.; Chakraverty, R.; Nayak, L.; Moss, P.A. The Number of Human Peripheral Blood CD4+ CD25high Regulatory T Cells Increases with Age. Clin. Exp. Immunol. 2005, 140, 540– 546.
- 72. Shimizu, J.; Moriizumi, E. CD4+CD25- T Cells in Aged Mice Are Hyporesponsive and Exhibit Suppressive Activity. J. Immunol. 2003, 170, 1675–1682.
- 73. Peterson, R.A. Regulatory T-Cells: Diverse Phenotypes Integral to Immune Homeostasis and Suppression. Toxicol. Pathol. 2012, 40, 186–204.
- Sharma, S.; Dominguez, A.L.; Lustgarten, J. High Accumulation of T Regulatory Cells Prevents the Activation of Immune Responses in Aged Animals. J. Immunol. 2006, 177, 8348–8355.
- 75. Simone, R.; Zicca, A.; Saverino, D. The Frequency of Regulatory CD3+CD8+CD28- CD25+ T Lymphocytes in Human Peripheral Blood Increases with Age. J. Leukoc. Biol. 2008, 84, 1454–1461.
- Togashi, Y.; Shitara, K.; Nishikawa, H. Regulatory T Cells in Cancer Immunosuppression Implications for Anticancer Therapy. Nat. Rev. Clin. Oncol. 2019, 16, 356–371.
- 77. Dong, Y.; Wan, Z.; Gao, X.; Yang, G.; Liu, L. Reprogramming Immune Cells for Enhanced Cancer Immunotherapy: Targets and Strategies. Front. Immunol. 2021, 12, 609762.
- Wang, H.; Franco, F.; Tsui, Y.-C.; Xie, X.; Trefny, M.P.; Zappasodi, R.; Mohmood, S.R.; Fernández-García, J.; Tsai, C.-H.; Schulze, I.; et al. CD36-Mediated Metabolic Adaptation Supports Regulatory T Cell Survival and Function in Tumors. Nat. Immunol. 2020, 21, 298–308.

- 79. Raja, K.R.M.; Rihova, L.; Zahradova, L.; Klincova, M.; Penka, M.; Hajek, R. Increased T Regulatory Cells Are Associated with Adverse Clinical Features and Predict Progression in Multiple Myeloma. PLoS ONE 2012, 7, e47077.
- 80. Watad, A.; Bragazzi, N.L.; Adawi, M.; Amital, H.; Toubi, E.; Porat, B.-S.; Shoenfeld, Y. Autoimmunity in the Elderly: Insights from Basic Science and Clinics - A Mini-Review. Gerontology 2017, 63, 515–523.
- Barsheshet, Y.; Wildbaum, G.; Levy, E.; Vitenshtein, A.; Akinseye, C.; Griggs, J.; Lira, S.A.; Karin, N. CCR8+FOXp3+ Treg Cells as Master Drivers of Immune Regulation. Proc. Natl. Acad. Sci. USA 2017, 114, 6086–6091.
- 82. Sun, L.; Hurez, V.J.; Thibodeaux, S.R.; Kious, M.J.; Liu, A.; Lin, P.; Murthy, K.; Pandeswara, S.; Shin, T.; Curiel, T.J. Aged Regulatory T Cells Protect from Autoimmune Inflammation despite Reduced STAT3 Activation and Decreased Constraint of IL-17 Producing T Cells. Aging Cell 2012, 11, 509–519.
- 83. Fan, Y.-G.; Zhai, J.-M.; Wang, W.; Feng, B.; Yao, G.-L.; An, Y.-H.; Zeng, C. IL-35 over-Expression Is Associated with Genesis of Gastric Cancer. Asian Pac. J. Cancer Prev. 2015, 16, 2845–2849.
- Van Herk, E.H.; Velde, A.A. Treg Subsets in Inflammatory Bowel Disease and Colorectal Carcinoma: Characteristics, Role, and Therapeutic Targets. J. Gastroenterol. Hepatol. 2016, 31, 1393–1404.
- 85. Cutler, C.S.; Koreth, J.; Ritz, J. Mechanistic Approaches for the Prevention and Treatment of Chronic GVHD. Blood 2017, 129, 22–29.
- 86. Whangbo, J.S.; Antin, J.H.; Koreth, J. The Role of Regulatory T Cells in Graft-versus-Host Disease Management. Expert Rev. Hematol. 2020, 13, 141–154.
- Qayed, M.; Wang, T.; Hemmer, M.T.; Spellman, S.; Arora, M.; Couriel, D.; Alousi, A.; Pidala, J.; Abdel-Azim, H.; Aljurf, M.; et al. Influence of Age on Acute and Chronic GVHD in Children Undergoing HLA-Identical Sibling Bone Marrow Transplantation for Acute Leukemia: Implications for Prophylaxis. Biol. Blood Marrow Transplant. 2018, 24, 521–528.
- Serrano-López, J.; Fernández, J.L.; Lumbreras, E.; Serrano, J.; Martínez-Losada, C.; Martín, C.; Hernández-Rivas, J.M.; Sánchez-García, J. Machine Learning Applied to Gene Expression Analysis of T-Lymphocytes in Patients with CGVHD. Bone Marrow Transplant. 2020, 55, 1668–1670.
- Neumann, C.; Blume, J.; Roy, U.; Teh, P.P.; Vasanthakumar, A.; Beller, A.; Liao, Y.; Heinrich, F.; Arenzana, T.L.; Hackney, J.A.; et al. C-Maf-Dependent Treg Cell Control of Intestinal TH17 Cells and IgA Establishes Host-Microbiota Homeostasis. Nat. Immunol. 2019, 20, 471–481.
- 90. Othy, S.; Jairaman, A.; Dynes, J.L.; Dong, T.X.; Tune, C.; Yeromin, A.V.; Zavala, A.; Akunwafo, C.; Chen, F.; Parker, I.; et al. Regulatory T Cells Suppress Th17 Cell Ca2+ Signaling in the Spinal Cord during Murine Autoimmune Neuroinflammation. PNAS 2020, 117, 20088–20099.
- 91. Wang, W.; Wang, Z.; Qin, Y.; Tang, G.; Cai, G.; Liu, Y.; Zhang, J.; Zhang, P.; Shen, Q.; Shen, L.; et al. Th17, Synchronically Increased with Tregs and Bregs, Promoted by Tumour Cells via Cell-Contact in Primary Hepatic Carcinoma. Clin. Exp. Immunol. 2018, 192, 181–192.
- 92. Schmitt, V.; Rink, L.; Uciechowski, P. The Th17/Treg Balance Is Disturbed during Aging. Exp. Gerontol. 2013, 48, 1379–1386.
- Bharath, L.P.; Agrawal, M.; McCambridge, G.; Nicholas, D.A.; Hasturk, H.; Liu, J.; Jiang, K.; Liu, R.; Guo, Z.; Deeney, J.; et al. Metformin Enhances Autophagy and Normalizes Mitochondrial Function to Alleviate Aging-Associated Inflammation. Cell Metab. 2020, 32, 44–55.e6.
- 94. Gaur, P.; Qadir, G.A.; Upadhyay, S.; Singh, A.K.; Shukla, N.K.; Das, S.N. Skewed Immunological Balance between Th17 (CD4(+)IL17A (+)) and Treg (CD4 (+)CD25 (+)FOXP3 (+)) Cells in Human Oral Squamous Cell Carcinoma. Cell Oncol. 2012, 35, 335–343.
- 95. Tosolini, M.; Kirilovsky, A.; Mlecnik, B.; Fredriksen, T.; Mauger, S.; Bindea, G.; Berger, A.; Bruneval, P.; Fridman, W.-H.; Pagès, F.; et al. Clinical Impact of Different Classes of Infiltrating T Cytotoxic and Helper Cells (Th1, Th2, Treg, Th17) in Patients with Colorectal Cancer. Cancer Res. 2011, 71, 1263–1271.
- 96. Lim, M.-A.; Lee, J.; Park, J.-S.; Jhun, J.-Y.; Moon, Y.-M.; Cho, M.-L.; Kim, H.-Y. Increased Th17 Differentiation in Aged Mice Is Significantly Associated with High IL-1β Level and Low IL-2 Expression. Exp. Gerontol. 2014, 49, 55–62.
- 97. Prabhala, R.H.; Pelluru, D.; Fulciniti, M.; Prabhala, H.K.; Nanjappa, P.; Song, W.; Pai, C.; Amin, S.; Tai, Y.-T.; Richardson, P.G.; et al. Elevated IL-17 Produced by TH17 Cells Promotes Myeloma Cell Growth and Inhibits Immune Function in Multiple Myeloma. Blood 2010, 115, 5385–5392.
- 98. Dhodapkar, K.M.; Barbuto, S.; Matthews, P.; Kukreja, A.; Mazumder, A.; Vesole, D.; Jagannath, S.; Dhodapkar, M.V. Dendritic Cells Mediate the Induction of Polyfunctional Human IL17-Producing Cells (Th17-1 Cells) Enriched in the Bone Marrow of Patients with Myeloma. Blood 2008, 112, 2878–2885.

- 99. Harmer, D.; Falank, C.; Reagan, M.R. Interleukin-6 Interweaves the Bone Marrow Microenvironment, Bone Loss, and Multiple Myeloma. Front. Endocrinol. 2019, 0.
- 100. Rossi, M.; Altomare, E.; Botta, C.; Gallo Cantafio, M.E.; Sarvide, S.; Caracciolo, D.; Riillo, C.; Gaspari, M.; Taverna, D.; Conforti, F.; et al. MiR-21 Antagonism Abrogates Th17 Tumor Promoting Functions in Multiple Myeloma. Leukemia 2021, 35, 823–834.
- 101. Lei, L.; Sun, J.; Han, J.; Jiang, X.; Wang, Z.; Chen, L. Interleukin-17 Induces Pyroptosis in Osteoblasts through the NLRP3 Inflammasome Pathway in Vitro. Int. Immunopharmacol. 2021, 96, 107781.
- 102. Ma, T.; Zhang, Y.; Zhou, X.; Xie, P.; Li, J. A Unique Role of T Helper 17 Cells in Different Treatment Stages of Multiple Myeloma. Clin. Lymphoma Myeloma Leuk. 2020, 20, 190–197.
- 103. Calcinotto, A.; Brevi, A.; Chesi, M.; Ferrarese, R.; Garcia Perez, L.; Grioni, M.; Kumar, S.; Garbitt, V.M.; Sharik, M.E.; Henderson, K.J.; et al. Microbiota-Driven Interleukin-17-Producing Cells and Eosinophils Synergize to Accelerate Multiple Myeloma Progression. Nat. Commun. 2018, 9, 4832.
- 104. Prabhala, R.H.; Neri, P.; Bae, J.E.; Tassone, P.; Shammas, M.A.; Allam, C.K.; Daley, J.F.; Chauhan, D.; Blanchard, E.; Thatte, H.S.; et al. Dysfunctional T Regulatory Cells in Multiple Myeloma. Blood 2006, 107, 301–304.
- 105. Noonan, K.; Marchionni, L.; Anderson, J.; Pardoll, D.; Roodman, G.D.; Borrello, I. A Novel Role of IL-17–Producing Lymphocytes in Mediating Lytic Bone Disease in Multiple Myeloma. Blood 2010, 116, 3554–3563.

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