

EV miRNAs Modulate Inflammation

Subjects: [Immunology](#)

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In rheumatoid arthritis (RA), extracellular vesicles (EVs) are associated with both the propagation and attenuation of joint inflammation and destruction. However, the specific EV content responsible for these processes is largely unknown. Investigations into identifying EV content are confounded by the challenges in obtaining high-quality EV preparations from synovial fluid. Implementing a size exclusion chromatography-based method of EV isolation, coupled with small RNA sequencing, we accurately characterised EV miRNAs in synovial fluid obtained from RA patients and investigated the differences between joints with high- and low-grade inflammation.

rheumatoid arthritis

extracellular vesicles

miRNA

synovial fluid

1. Introduction

Rheumatoid arthritis (RA) is a chronic systemic autoimmune disease that targets synovial joints. RA causes chronic, progressive joint inflammation with exacerbations and remissions. Unless it is adequately treated, disease persistence leads to irreversible joint destruction and subsequent joint deformity, resulting in progressive disability. In RA, joint damage occurs due to an immune response that drives inflammatory cell infiltration. The pathophysiology of RA is incompletely understood. To improve patient outcomes and guide the development of more effective ways to control this disease, a better understanding of the disease processes is required.

Extracellular vesicles (EVs)—including exosomes, microvesicles and apoptotic bodies—are phospholipid bilayer-bound vesicles that are released from host cells and which contain a bioactive cargo of RNAs, proteins and lipids [\[1\]](#). In RA, synovial fluid (SF) EVs have immunomodulatory functionality and are described to both contribute to and protect against joint destruction [\[2\]\[3\]](#).

MicroRNAs (miRNAs) are ~18–25 nucleotide RNAs that can regulate protein translation through complementary binding to mRNA transcripts [\[4\]](#). In RA, fluctuations in the expression of miRNAs may contribute to the severity of joint inflammation and destruction. For example, elevated miR-146a levels are reported in RA SF [\[5\]](#) and in CD4+ T cells isolated from RA SF [\[6\]](#), but not in osteoarthritis SF, whereas miR-146a expression in SF CD4+ T cells is positively correlated with SF TNF levels [\[6\]](#). A direct role for miR-146a in promoting inflammation through upregulating T cell activity is supported by miR-146a overexpression, suppressing apoptosis in Jurkat T-like cells [\[6\]](#).

An increasing number of studies have also demonstrated that miRNAs encapsulated within EVs can promote inflammatory disease. For instance, miRNAs in Treg-derived EVs suppressed T helper 1 cell proliferation and pro-

inflammatory cytokine release [7]. Conversely, GU-rich motifs present in some EV miRNAs have been described to act as endolysosomal TLR8 (TLR7 in mice) agonists [8]. Consistent with this, EVs in the sera of HIV patients were found to contain GU-rich viral miRNAs that act as ligands for TLR8 and stimulate TNF release in recipient macrophages [9].

2. Characterisation of EV Isolation

EVs were first prepared by means of size exclusion chromatography (SEC) from SF obtained from a cohort of RA patients with either high- or low-grade inflammation (Table 1). To confirm that EV preparations were of satisfactory quality, EV enrichments were assessed by Western blotting for the canonical EV markers: HSP70, syntenin, FLOT1, Rab 27b, TSG101 and annexin 1 (Figure 1A). Transmission electron microscopy confirmed the presence of EVs with minimal apparent non-EV contaminating material (Figure 1B). These observations are consistent with our previous report detailing SEC as a method for obtaining high-quality EV enrichments from SF [10]. Previously, we demonstrated that RA patients with high-grade synovial inflammation have increased concentrations of SF EVs [11]. Consistent with this, we detected a 3.5-fold increase in the EV miRNA concentration per mL of SF in joints with high-grade inflammation (p -value = 0.0017; Figure 1C), suggesting the potential involvement of SF EV miRNAs in inflammatory processes in RA.

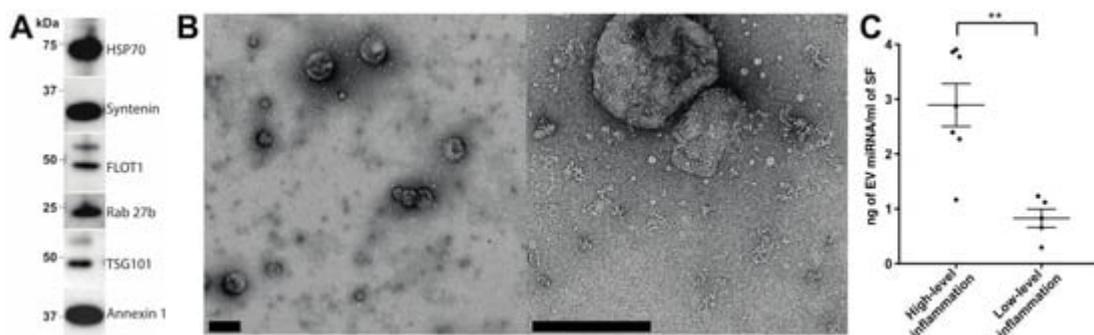


Figure 1. Assessment of synovial fluid EV enrichments prepared using size-exclusion chromatography. (A) Synovial fluid EV enrichments prepared by SEC were assessed for the presence of the canonical EV markers (HSP70, syntenin, FLOT1, Rab 27b, TSG101 and annexin 1) via Western blotting. (B) The quality of EV enrichments was further assessed by transmission electron microscopy (scale bars = 200 nm). (C) miRNA concentration per mL of rheumatoid arthritis synovial fluid in joints with high- or low-grade joint inflammation. Group means are indicated. Data analysed with Student's *t*-test. Error bars = SEM. ** denotes p -value < 0.01.

Table 1. Summary of patient details and clinical parameters.

	RA (High-Grade Inflammation)	RA (Low-Grade Inflammation)	<i>p</i> -Value
<i>n</i>	7	5	-
Age—mean (s.d.)	64.4 (11.8)	67.6 (10.0)	0.64

	RA (High-Grade Inflammation)	RA (Low-Grade Inflammation)	p-Value
Sex—number of females/males	3/4	2/3	>0.99
White cell count—mean (s.d.) cells μL^{-1}	8940 *	171.8	<0.001
Anti-citrullinated protein antibody (%) positive)	71%	20%	0.24
Rheumatoid factor (% positive)	86%	60%	0.52
Disease Activity Score 28—median (range)	4.7 (3.31–5.41)	3.5 (2.74–5.0)	0.15
C-reactive protein—median (range) mg L^{-1}	20 (6–164)	2 (1.4–2)	0.16

3. Highly Ranked SF EV miRNAs Target Immunomodulatory SF EV Proteins

To characterise the miRNA present in the SF of RA patients, small RNA sequencing was performed. A total of 1415 miRNA species were identified and, after filtering out lowly abundant species, 318 miRNAs remained. The 10 highest-ranked miRNAs across the entire patient cohort are specified in [Table 2](#). To investigate biological processes associated with these prevalent miRNAs, experimentally validated gene targets were identified using miRTarBase and biological processes associated with the gene targets were characterised ([Figure 2](#)). A robust enrichment for genes involved in regulating gene expression was observed, including AKT1 (miR-100-5p, miR-10b-5p and miR-99a-5p), FOXO1 (let-7a-5p and miR-21-5p), KRAS (let-7a-5p), IKBKB (miR-148a-3p), STAT3 (let-7a-5p, miR-148a-3p, miR-21-5p and miR-92a-3p), TGFBR1 (let-7b-5p) and TP53 (miR-10b-5p). In addition, genes associated with cytokine-mediated signalling were targets of highly prevalent miRNA, including the inflammatory mediators CCR1 (miR-21-5p), IL-1 β (miR-21-5p), IL-6 (let-7a-5p and miR-26a-5p), MCL1 (miR-26a-5p), NOS2 (miR-26a-5p), PIK3CA (miR-10b-5p) and SOCS5 (miR-92a-3p).

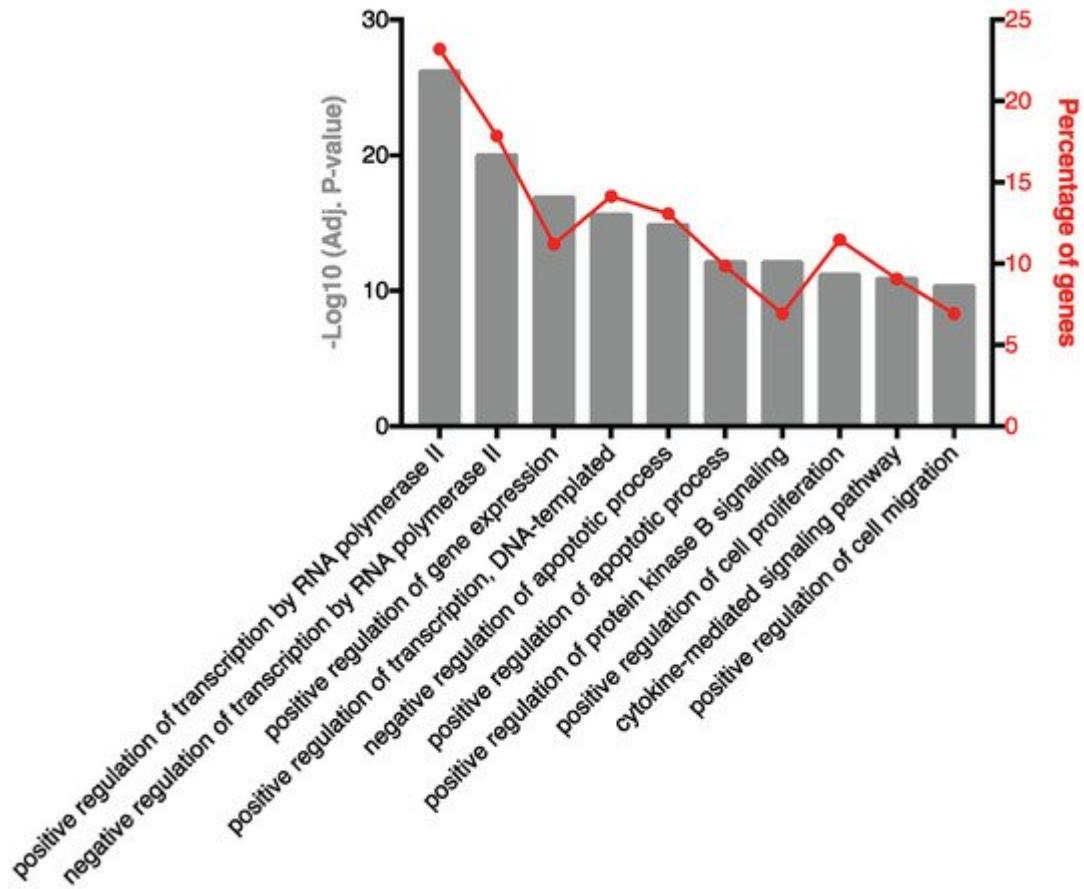


Figure 2. Biological processes associated with experimentally validated targets of highly ranked SF EV miRNA. Pathway analysis was performed on experimentally validated gene targets of the 10 highest-ranked miRNAs. The percentage of target genes associated with each biological process are indicated.

Table 2. Highest-ranked SF EV miRNAs across the entire patient cohort.

Rank	miRNA	Average Expression (CPM)
1	hsa-miR-100-5p	132,441
2	hsa-miR-21-5p	179,929
3	hsa-miR-148a-3p	86,059
4	hsa-let-7a-5p	43,627
5	hsa-miR-92a-3p	45,881
6	hsa-let-7b-5p	31,280
7	hsa-miR-10b-5p	35,499
8	hsa-miR-99b-5p	31,569
9	hsa-miR-26a-5p	34,975

Rank	miRNA	Average Expression (CPM)
10	hsa-miR-99a-5p	45,879

To investigate if these 10 highly-ranked miRNAs might synergise to repress the expression of individual genes, common gene targets were identified. Genes targeted by four or more miRNA are listed in [Table 3](#). Interestingly, the Akt activator insulin-like growth factor 1 receptor (IGF1R) is a target of five of the 10 highest-ranked miRNAs, and the pro-inflammatory transcription factor STAT3 is targeted by four of the 10 highest-ranked miRNAs.

Table 3. Common targets of the 10 highest-ranked EV miRNAs.

Target Gene ID	Target Gene Name	miRNA Regulators
IGF1R	Insulin like growth factor 1 receptor	hsa-let-7b-5p hsa-miR-100-5p hsa-miR-21-5p hsa-miR-99a-5p hsa-miR-99b-5p
CCND2	Cyclin D2	hsa-let-7a-3p hsa-let-7a-5p hsa-let-7b-5p hsa-miR-26a-5p
E2F2	E2F transcription factor 2	hsa-let-7a-3p hsa-let-7a-5p hsa-let-7b-5p hsa-miR-26a-5p
PTEN	Phosphatase and tensin homolog	hsa-miR-10b-5p hsa-miR-21-5p hsa-miR-26a-5p hsa-miR-92a-3p
STAT3	Signal transducer and activator of transcription 3	hsa-let-7a-5p hsa-miR-148a-3p hsa-miR-21-5p hsa-miR-92a-3p

4. Seventy-Eight SF EV miRNAs Are Differentially Expressed between RA Patients with High- and Low-Grade Inflammation

To investigate how SF EV miRNAs might more specifically contribute to RA pathophysiology, differences in miRNA expression in RA joints with either high- or low-grade inflammation were characterised. Overall, 78 differentially expressed miRNAs were defined. Of these, 49 were elevated in high-grade inflammation, whereas 29 were elevated in RA joints with low-grade inflammation ([Figure 3A](#) and [Table 4](#)).

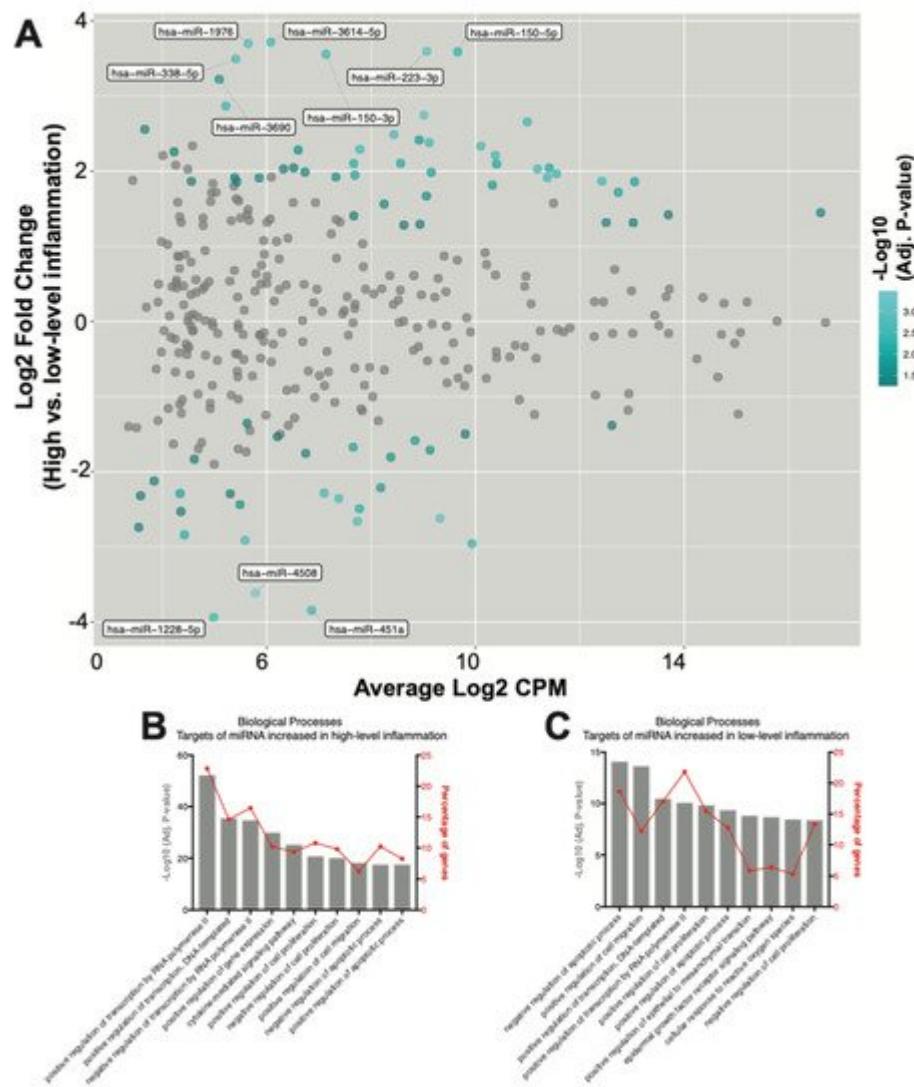


Figure 3. SF EV miRNAs have diverse immunoregulatory capacities. **(A)** MA plot of miRNA counts per million reads (CPM) vs. fold change. Differentially expressed miRNAs with an adjusted *p*-value < 0.05 are highlighted in blue. miRNAs with an adjusted *p*-value < 0.05 and a log2 fold change >3 are labelled. **(B,C)** Biological processes associated with target genes of the miRNAs significantly increased in joints with **(B)** high-, and **(C)** low-grade inflammation. The percentage of target genes associated with each biological process is indicated.

Table 4. Differentially expressed miRNAs.

miRNA	Average Read Count	Log2 Fold Change(High- vs. Low-Grade Inflammation)	Adjusted <i>p</i> -Value
	(Log2 CPM)		
hsa-miR-4508	5.78		-3.61 3.94×10^{-4}
hsa-miR-223-3p	9.07		3.59 6.42×10^{-4}
hsa-miR-3529-3p	9.01		2.74 6.42×10^{-4}

miRNA	Average Read Count (Log2 CPM)	Log2 Fold Change(High- vs. Low-Grade Inflammation)	Adjusted p-Value
hsa-miR-615-3p	9.32	-2.62	6.42×10^{-4}
hsa-miR-1976	5.65	3.70	1.15×10^{-3}
hsa-miR-543	7.74	-2.66	1.15×10^{-3}
hsa-miR-338-5p	5.41	3.49	1.36×10^{-3}
hsa-miR-146b-3p	8.43	2.49	1.36×10^{-3}
hsa-miR-433-3p	7.38	-2.36	1.36×10^{-3}
hsa-miR-485-3p	5.59	-2.91	1.36×10^{-3}
hsa-miR-101-3p	10.10	2.33	1.52×10^{-3}
hsa-miR-27a-5p	11.19	2.03	1.53×10^{-3}
hsa-miR-361-3p	11.38	1.91	1.76×10^{-3}
hsa-miR-3614-5p	6.08	3.72	1.86×10^{-3}
hsa-miR-150-3p	7.14	3.55	1.89×10^{-3}
hsa-miR-223-5p	9.11	2.38	1.89×10^{-3}
hsa-miR-142-5p	7.78	2.29	1.89×10^{-3}
hsa-miR-106b-3p	10.38	2.21	1.89×10^{-3}
hsa-miR-28-3p	12.42	1.87	1.89×10^{-3}
hsa-miR-455-5p	7.10	-2.28	1.89×10^{-3}
hsa-miR-451a	6.86	-3.84	1.89×10^{-3}

miRNA	Average Read Count (Log2 CPM)	Log2 Fold Change(High- vs. Low-Grade Inflammation)	Adjusted p-Value
hsa-miR-143-3p	10.99	2.66	2.04×10^{-3}
hsa-miR-1228-5p	4.99	-3.94	2.11×10^{-3}
hsa-miR-30e-3p	11.55	1.96	2.25×10^{-3}
hsa-miR-486-5p	9.93	-2.96	2.30×10^{-3}
hsa-miR-1273h-3p	5.22	2.87	2.56×10^{-3}
hsa-miR-150-5p	9.66	3.59	3.01×10^{-3}
hsa-miR-378c	7.67	2.10	3.61×10^{-3}
hsa-miR-92b-5p	7.78	-2.49	3.61×10^{-3}
hsa-miR-4448	4.42	-2.84	3.61×10^{-3}
hsa-miR-103b	8.57	2.10	4.76×10^{-3}
hsa-miR-941	11.41	2.04	4.76×10^{-3}
hsa-miR-103a-3p	7.69	1.95	5.24×10^{-3}
hsa-miR-1246	10.40	2.09	5.64×10^{-3}
hsa-miR-125b-1-3p	7.66	-1.67	5.64×10^{-3}
hsa-miR-769-5p	9.15	1.98	6.68×10^{-3}
hsa-miR-378a-3p	13.05	1.86	6.68×10^{-3}
hsa-miR-140-3p	12.73	1.72	6.68×10^{-3}
hsa-miR-214-5p	4.34	-2.29	6.68×10^{-3}

miRNA	Average Read Count (Log2 CPM)	Log2 Fold Change(High- vs. Low-Grade Inflammation)	Adjusted p-Value
hsa-miR-574-3p	8.84	-1.59	7.83×10^{-3}
hsa-miR-1180-3p	8.18	-2.21	8.45×10^{-3}
hsa-miR-155-5p	10.33	1.81	8.69×10^{-3}
hsa-miR-629-5p	8.92	2.41	9.41×10^{-3}
hsa-miR-328-3p	9.13	-1.71	1.05×10^{-2}
hsa-miR-3690	5.09	3.22	1.18×10^{-2}
hsa-miR-7704	8.38	-1.81	1.48×10^{-2}
hsa-miR-221-5p	9.06	1.67	1.51×10^{-2}
hsa-miR-486-3p	5.49	-2.44	1.51×10^{-2}
hsa-miR-589-5p	6.60	2.28	1.56×10^{-2}
hsa-miR-142-3p	7.33	1.92	1.56×10^{-2}
hsa-miR-192-5p	8.25	1.56	1.98×10^{-2}
hsa-miR-618	3.67	2.55	2.18×10^{-2}
hsa-miR-21-5p	16.61	1.45	2.22×10^{-2}
hsa-miR-21-3p	6.73	1.99	2.57×10^{-2}
hsa-miR-345-5p	5.86	1.91	2.59×10^{-2}
hsa-miR-214-3p	6.75	-1.76	2.59×10^{-2}
hsa-miR-532-5p	12.50	1.31	2.70×10^{-2}

miRNA	Average Read Count (Log2 CPM)	Log2 Fold Change(High- vs. Low-Grade Inflammation)	Adjusted p-Value
hsa-miR-378e	5.43	1.86	2.86×10^{-2}
hsa-miR-185-3p	6.32	2.03	3.05×10^{-2}
hsa-miR-6787-3p	3.55	-2.74	3.07×10^{-2}
hsa-miR-503-5p	4.23	2.26	3.11×10^{-2}
hsa-miR-25-3p	13.02	1.31	3.11×10^{-2}
hsa-miR-3120-5p	4.61	-1.83	3.24×10^{-2}
hsa-miR-203b-5p	3.84	-2.12	3.35×10^{-2}
hsa-miR-500a-3p	7.67	1.40	3.42×10^{-2}
hsa-miR-7-5p	4.56	1.86	3.56×10^{-2}
hsa-miR-3622a-5p	4.36	-2.53	3.56×10^{-2}
hsa-miR-365a-5p	5.30	-2.29	3.76×10^{-2}
hsa-miR-23b-3p	9.80	-1.50	3.85×10^{-2}
hsa-miR-424-3p	6.52	2.04	4.29×10^{-2}
hsa-miR-501-3p	8.94	1.29	4.30×10^{-2}
hsa-miR-92b-3p	12.61	-1.38	4.37×10^{-2}
hsa-miR-146b-5p	13.69	1.42	4.42×10^{-2}
hsa-miR-185-5p	8.63	1.28	4.48×10^{-2}
hsa-miR-23b-	6.21	-1.53	4.58×10^{-2}

targeting proteins typically associated with promoting inflammation, including IGF1R, PTEN, VEGFA and BCL2 ([Table 5](#)).

miRNA	Average Read Count (Log2 CPM)	Log2 Fold Change(High- vs. Low-Grade Inflammation)	Adjusted p-value with high-
5p			
Target Gene ID	Target Gene Name	miRNA Regulators	
IGF1R	Insulin like growth factor 1 receptor	hsa-miR-143-3p hsa-miR-150-3p hsa-miR-185-5p hsa-miR-21-5p hsa-miR-223-3p hsa-miR-223-5p hsa-miR-378a-3p hsa-miR-503-5p hsa-miR-7-5p	
PTEN	Phosphatase and tensin homolog	hsa-miR-103a-3p hsa-miR-106b-3p hsa-miR-142-5p hsa-miR-155-5p hsa-miR-21-3p hsa-miR-21-5p hsa-miR-25-3p	
VEGFA	Vascular endothelial growth factor A	hsa-miR-101-3p hsa-miR-150-5p hsa-miR-185-5p hsa-miR-21-5p hsa-miR-378a-3p hsa-miR-503-5p	
BCL2	BCL2, apoptosis regulator	hsa-miR-143-3p hsa-miR-192-5p hsa-miR-21-5p hsa-miR-503-5p hsa-miR-7-5p	
FBXW7	F-box and WD repeat domain containing 7	hsa-miR-155-5p hsa-miR-223-3p hsa-miR-223-5p hsa-miR-25-3p hsa-miR-503-5p	
MYB	MYB proto-oncogene, transcription factor	hsa-miR-103a-3p hsa-miR-150-3p hsa-miR-150-5p hsa-miR-155-5p hsa-miR-503-5p	
EGFR	Epidermal growth factor receptor	hsa-miR-146b-5p hsa-miR-21-5p	

Target Gene ID	Target Gene Name	miRNA Regulators
		hsa-miR-27a-5p hsa-miR-7-5p
RAC1	Rac family small GTPase 1	hsa-miR-101-3p hsa-miR-142-3p hsa-miR-142-5p hsa-miR-155-5p
TP53	Tumor protein p53	hsa-miR-150-3p hsa-miR-150-5p hsa-miR-25-3p hsa-miR-28-3p

ZEB1 Zinc finger E-box binding homeobox 1 hsa-miR-101-3p with low-
regulation of apoptotic processes was the highest-ranked biological process, with miRNA target genes including or genes
AKT1 (miR-451a), CD44 (miR-328-3p), IGF1R (miR-214-3p, miR-23b-3p, miR-486-5p and miR-92b-3p), IKBKB (miR-451a), IL6 (miR-451a), MIF (miR-1228-5p and miR-451a) and MYC (miR-451a). Interestingly, similarly to the 'Negative
above observations in joints with high-grade inflammation, multiple miRNAs found to be enriched in joints with low-
grade inflammation again target PTEN and TP53 (Table 6).

Table 6. Common targets of SF EV miRNAs found to be significantly enriched in the joints of RA patients with low-grade inflammation.

Target Gene ID	Target Gene Name	miRNA Regulators
PTEN	Phosphatase and tensin homolog	hsa-miR-214-3p hsa-miR-23b-3p hsa-miR-486-5p hsa-miR-92b-3p
TP53	Tumor protein p53	hsa-miR-125b-1-3p hsa-miR-214-3p hsa-miR-214-5p

References

1. Valadi, H.; Ekstrom, K.; Bossios, A.; Sjostrom, M.; Lee, J.J.; Lotvall, J.O. Exosome-mediated transfer of mRNAs and microRNAs is a novel mechanism of genetic exchange between cells. *Nat. Cell Biol.* 2007, 9, 654–659.
2. Foers, A.D.; Cheng, L.; Hill, A.F.; Wicks, I.P.; Pang, K.C. Review: Extracellular Vesicles in Joint Inflammation. *Arthritis Rheumatol.* 2017, 69, 1350–1362.

3. Malda, J.; Boere, J.; Van De Lest, C.H.A.; Van Weeren, P.R.; Wauben, M.H.M. Extracellular vesicles—New tool for joint repair and regeneration. *Nat. Rev. Rheumatol.* 2016, 12, 243–249.
4. O'Brien, J.; Hayder, H.; Zayed, Y.; Peng, C. Overview of MicroRNA Biogenesis, Mechanisms of Actions, and Circulation. *Front. Endocrinol.* 2018, 9, 402.
5. Murata, K.; Yoshitomi, H.; Tanida, S.; Ishikawa, M.; Nishitani, K.; Ito, H.; Nakamura, T. Plasma and synovial fluid microRNAs as potential biomarkers of rheumatoid arthritis and osteoarthritis. *Arthritis Res. Ther.* 2010, 12, 1–14.
6. Li, J.; Wan, Y.; Guo, Q.; Zou, L.; Zhang, J.; Fang, Y.; Zhang, J.; Zhang, J.; Fu, X.; Liu, H.; et al. Altered microRNA expression profile with miR-146a upregulation in CD4+ T cells from patients with rheumatoid arthritis. *Arthritis Res. Ther.* 2010, 12, R81.
7. Okoye, I.S.; Coomes, S.M.; Pelly, V.S.; Czieso, S.; Papayannopoulos, V.; Tolmachova, T.; Seabra, M.C.; Wilson, M.S. MicroRNA-containing T-regulatory-cell-derived exosomes suppress pathogenic T helper 1 cells. *Immunity* 2014, 41, 89–103.
8. Bosch, S.; Young, N.A.; Mignot, G.; Bach, J.-M. Epigenetic Mechanisms in Immune Disease: The Significance of Toll-Like Receptor-Binding Extracellular Vesicle-Encapsulated microRNA. *Front. Genet.* 2020, 11, 578335.
9. Bernard, M.A.; Zhao, H.; Yue, S.C.; Anandaiah, A.; Koziel, H.; Tachado, S.D. Novel HIV-1 MiRNAs Stimulate TNF α Release in Human Macrophages via TLR8 Signaling Pathway. *PLoS ONE* 2014, 9, e106006.
10. Foers, A.D.; Chatfield, S.; Dagley, L.F.; Scicluna, B.J.; Webb, A.I.; Cheng, L.; Hill, A.F.; Wicks, I.P.; Pang, K.C. Enrichment of extracellular vesicles from human synovial fluid using size exclusion chromatography. *J. Extracell. Vesicles* 2018, 7, 1490145.
11. Foers, A.D.; Dagley, L.F.; Chatfield, S.; Webb, A.I.; Cheng, L.; Hill, A.F.; Wicks, I.P.; Pang, K.C. Proteomic analysis of extracellular vesicles reveals an immunogenic cargo in rheumatoid arthritis synovial fluid. *Clin. Transl. Immunol.* 2020, 9, e1185.

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