# Hydrogen Peroxide

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Hydrogen peroxide (H2O2) is an important oxidation molecule regulating aerobic metabolism. Redox signals include physiological oxidative stress (EU stress), and excessive oxidative stress can damage molecules. The main enzyme sources of H2O2 are nicotinamide adenine dinucleotide phosphate oxidase or NADPH oxidase (NOx), mitochondrial respiratory chain and various kinds of oxidase. NOx family consists of seven enzyme subtypes, which produce a superoxide anion (O2 -) which can be converted to H2O2 by superoxide dismutase or spontaneously. H2O2 passes through the membrane through some aquaporin (AQP), which is called porphyrin hydroperoxide. It diffuses in cells and tissues, triggering cellular effects such as proliferation, immune cell recruitment and morphological changes.

Keywords: Hydrogen Peroxide ; NADPH Oxidases ; oxidative stress

## 1. Introducción

Living organisms that develop under aerobic conditions generate reactive oxygen species (ROS) by cellular metabolism <sup>[1]</sup> <sup>[2][3]</sup>. ROS are formed during cellular processes, such as cellular respiration, NADPH oxidase (NOX) activity, and processes related to peroxisomes, nitric oxide synthetase, lipoxygenases, xanthine oxidase, cyclooxygenases, and the cytochrome P450 system, as well as inflammatory processes, mainly in hepatocytes and macrophages <sup>[1][2][3][4][5][6][7]</sup>.

For a long time, ROS were considered as molecules or entities that could generate damage at different levels, be they cellular, tissue or even at the organ level, or that their production was linked to enzymatic reactions which involve oxygen intermediaries. Its production causes an imbalance in the oxide-reduction (redox) state of the cells. In phagocytes, ROS are produced by the activation of NOX to destroy pathogens during phagocytosis <sup>[8]</sup>. However, in the last 20 years, it has been shown that the generation of ROS is linked to a variety of essential biological processes, where they participate in the regulation of some metabolic pathways, either negatively or positively. These processes include cell development and differentiation, generation or crosslinking of the extracellular matrix, hormone biosynthesis, cell aging, apoptosis, the responses of oxygen changes (oxygen detection), growth factors, hormones, and cytokines.

Hydrogen peroxide ( $H_2O_2$ ) is an important oxidizing molecule that regulates the metabolisms of aerobic organisms. Redox signaling comprises physiological oxidative stress (eustress), while excessive oxidative stress causes damage to molecules. The main enzymatic generators of  $H_2O_2$  are nicotinamide adenine dinucleotide phosphate oxidases or NADPH oxidases (NOXs) and mitochondrial respiratory chains, as well as various oxidases. The NOX family is constituted of seven enzyme isoforms that produce a superoxide anion ( $O_2^-$ ), which can be converted to  $H_2O_2$  by superoxide dismutase or spontaneously.  $H_2O_2$  passes through the membranes by some aquaporins (AQPs), known as peroxyporins. It diffuses through cells and tissues to initiate cellular effects, such as proliferation, the recruitment of immune cells, and cell shape changes. Therefore, it has been proposed that  $H_2O_2$  has the same importance as Ca<sup>2+</sup> or adenosine triphosphate (ATP) to act as modulators in signaling and the metabolism.

# 2. NOX Family

NOXs represent a family of enzyme complexes present in the membrane which generate ROS. This complex is widely distributed in eukaryotic organisms <sup>[9]</sup>, and recent results indicate that they are also present in bacteria <sup>[10]</sup>. The first NOX was described in neutrophils and produces a large amount of ROS as a defense system <sup>[11]</sup>. The catalytic subunit of this enzyme was named  $gp91^{phox}$  and is now called NOX2. The development of more of sensitive assays allowed the detection of low amounts of ROS in other cells <sup>[12]</sup>. NOX1, also known as Mox1, was the first NOX2 homolog to be identified <sup>[13]</sup>. In mammals, the NOX family is formed by seven members: NOX1, 2, and 3, activated by cytoplasmic subunits; NOX4, which is constitutively active; and NOX5 and DUOX1 and 2, which bind calcium. The product of NOX isoforms is  $O_2^-$ , which rapidly changes to  $H_2O_2$  <sup>[14]</sup>.

#### 2.1. Hydrogen Peroxide Production

NOX family members have different regulation processes, but they share a catalytic core with two domains: the dehydrogenase and the transmembrane domains <sup>[15]</sup>. Electrons are transferred from cytosolic NADPH to FAD in the dehydrogenase domain, then to the inner and outer hemes in the transmembrane domain, and finally to the  $O_2$  outside the cell, producing  $O_2^-$ , as shown in the following reaction:

$$NADPH + 2O_2 \leftrightarrow NADP^+ + 2O_2^- + H^+$$
(1)

A superoxide is highly reactive, short-lived, and its charge prevents its diffusion through membranes  $\frac{[16]}{1}$ . However, it can dismutase to H<sub>2</sub>O<sub>2</sub> spontaneously (rate constant = 8 × 10<sup>4</sup> M<sup>-1</sup>s<sup>-1</sup>) or by superoxide dismutase (SOD) (rate constant = 2 × 10<sup>9</sup> M<sup>-1</sup>s<sup>-1</sup>)  $\frac{[17]}{1}$ . H<sub>2</sub>O<sub>2</sub> is diffusible, transverses membranes by aquaporins (AQPs),  $\frac{[18]}{18}$  and initiates cell signaling  $\frac{[19]}{19}$ .

#### 2.2. Tissue Distribution of NOX Proteins

NOXs localize in different tissues and membrane compartments, allowing the spatiotemporal organization of ROS signaling. The expression of NOX enzymes in various tissues supports the idea that ROS production is not exclusive to phagocytic cells. The fact that some of these enzymes produce low levels of ROS suggests that they may be participating in other cellular functions than the inflammatory response <sup>[20]</sup>. NOX2 is referred as phagocyte NADPH oxidase. However, it is also found in other cells such as hepatocytes, endothelial cells, smooth and skeletal muscle, neurons, cardiomyocytes, and hematopoietic stem cells. NOX1 and NOX2 share a 60% amino acid identity. NOX1 is expressed more in the colon epithelium, while NOX3 is almost exclusively expressed in the inner ear. NOX4 expression is strongly observed in the kidney and at moderate levels in vascular endothelial cells, vascular smooth muscle, murine osteoclasts, murine adipocytes, and the pancreas. NOX5 is expressed in the human spleen, testis, and vascular smooth muscle. The dual oxidases DUOX1 and DUOX2 are the  $H_2O_2$ -producing isoforms of the NADPH oxidases (DUOXs) are highly expressed in the thyroid, but they can also be expressed in other cells.

### 2.3. Structure and Regulation of NOX

NOX isoform structures were deduced from studies on NOX2 [14]. There are structural properties of NOX2 that are similar to all members of the family. NOX complexes consist of one catalytic subunit and five regulatory subunits (p22<sup>phox</sup>, p47<sup>phox</sup>, p67<sup>phox</sup>, p40<sup>phox</sup>, and Rac). The catalytic subunit has six transmembrane domains (seven for DUOX1-2) and four hemes in the third and fifth transmembrane domains, as well as an NADPH-binding domain and an FAD-binding domain in the C-terminal in the cytosolic region. NOX5 has additional EF-hands and Ca<sup>2+</sup>-binding domains at the N-terminus, and DUOX enzymes have EF-hands and a peroxidase-like domain at the N-terminus. NOX2 forms a complex membrane with a stabilizing factor, p22<sup>phox [21]</sup>. In unstimulated cells, NOX2 and p22<sup>phox</sup> are found in the plasma membrane, with p67<sup>phox</sup> associating with the proline-rich region (PRR) of the p47<sup>phox</sup> in the cytosol. The p47<sup>phox</sup> has two Src homology 3 (SH3) domains for its association with the PRR of the p22<sup>phox [22]</sup>. In the resting state, the auto inhibitory region AIR in the p47<sup>phox</sup> combines with the bis-SH3 domain and the domain phagocyte oxidase (PX) of the p47<sup>phox</sup>, preventing its binding to p22<sup>phox</sup> and membrane lipids <sup>[23]</sup>. The cytoplasmic component p40<sup>phox</sup> is associated with p67<sup>phox</sup>. Rac attached to guanosine 5'-diphosphate (GDP) is associated with RhoGDP dissociation inhibitor (RhoGDI) in the cytosol. Rac is a member of the Rho family, and there are three expressed isoforms of Rac: Rac1, Rac2, and Rac3. Rac1 is expressed in non-hematopoietic cells and activates many NOX isoforms. In contrast, Rac2 is expressed in hematopoietic cells, and Rac3 is predominantly found in the central nervous system <sup>[24][25]</sup>. The phagocytosis of bacteria induces phosphorylation in p47<sup>phox</sup> and conformational change, allowing its interaction with p22<sup>phox</sup> [14][22]. Subunit p47<sup>phox</sup>, attached to the membrane, transports the activator subunits p67<sup>phox</sup> and p40<sup>phox</sup> to NOX2 <sup>[26]</sup>. Phagocytosis also causes the generation of Rac2-GTP, which is released from the RhoGDI. Rac-GTP goes toward the membrane and interacts with NOX2 and p67<sup>phox</sup> [27][28]. The assembled complex generates superoxide in the extracellular space (Figure 1).

**Figure 1.** NADPH oxidase (NOX) types and activation of the NOX2 macrophage for the formation of reactive oxygen species. (a) Structure of the isoforms that are part of the NOX family. (b) Activation of the NOX2 system, which occurs by the assembly of cytosolic regulatory proteins ( $p40^{phox}$ ,  $p47^{phox}$ ,  $p67^{phox}$ , and Rac2) with the catalytic subunit  $gp91^{phox}$  and  $p22^{phox}$  in the membrane, to activate the enzyme complex and generate superoxide anions ( $O_2^-$ ) and hydrogen peroxide ( $H_2O_2$ ).

The other isoforms share some characteristics of NOX2 activation. The NOX1 isoform exhibits 60% of its identity with NOX2. Its activity depends on cytosolic subunits. NOX1 requires p22<sup>phox</sup>, Rac GTPase, the first organizer (NOXO1), which is a homolog of p47<sup>phox</sup>, and the first activator (NOXA1), a homolog of p67<sup>phox</sup> <sup>[29]</sup>. NOX3 shares 56% of its amino acid identity with NOX2 [30]. This NOX requires NOXO1 and NOXA1 for activation, but p47<sup>phox</sup>, p67<sup>phox</sup>, and Rac have not been studied [31]. NOX4 shares approximately 39% of it sequence homology with NOX2. This isoform is constitutively active, and its activity depends on p22<sup>phox [32]</sup>. Recently, it has been described that polymerase  $\delta$ -interacting protein 2 (POLDIP2) regulates NOX4 [33]. NOX5 comprises five isoforms: a, b, c, d, and e or S [34][35]. NOX5 a-d isoforms have an intracellular domain that contains a Ca<sup>+2</sup> -binding EF-hand region <sup>[36]</sup>. NOX5 a-d isoforms do not require p22<sup>phox</sup> or cytosolic subunits for activation, but they need cytosolic calcium. Heat shock protein 90 (HSP90) has been described as a regulatory protein for NOX1–3 and NOX5 [32]. The NOX5 e isoform lacks a Ca<sup>2+</sup>-binding domain and depends on the cAMP response element-binding protein for activity [38]. DUOX1 and DUOX2 were first identified in the thyroid gland. These enzymes share 50% of their amino acid identity with NOX2 <sup>[39]</sup>. They have a NOX domain similar to that of NOX1-4, an EF-hand region similar to NOX5, and a seventh transmembrane peroxidase-like domain. Both dual oxidases have been identified in the plasma membrane and endoplasmic reticulum <sup>[40]</sup>. The permanence of DUOX proteins in the endoplasmic reticulum depends on the maturation factors of DUOXA1 and DUOXA2. The DUOX2 immature form generates  $O_2^-$ , whereas the mature form generates  $H_2O_2^{[41]}$ . Las proteínas DUOX no necesitan un activador citosólico o subunidades organizadoras para su activación, pero sí requieren calcio [42].

## References

- 1. He, L.; He, T.; Farrar, S.; Ji, L.; Liu, T.; Ma, X. Antioxidants maintain cellular redox homeostasis by elimination of reactive oxygen species. Cell Physiol. Biochem. 2017, 44, 532–553.
- Snezhkina, A.V.; Kudryavtseva, A.V.; Kardymon, O.L.; Savvateeva, M.V.; Melnikova, N.V.; Krasnov, G.S.; Dmitriev, A.A. ROS generation and antioxidant defense systems in normal and malignant cells. Oxid. Med. Cell Longev. 2019, 2019, 6175804.
- 3. Ighodaro, O.M.; Akinloye, O.A. First line defence antioxidants-superoxide dismutase (SOD), catalase (CAT) and glutathione peroxidase (GPX): Their fundamental role in the entire antioxidant defence grid. Alexandria J. Med. 2018, 54, 287–293.
- 4. Yang, S.; Lian, G. ROS and diseases: Role in metabolism and energy supply. Mol. Cell Biochem. 2020, 467, 1–12.
- Osorio Alves, J.; Matta Pereira, L.; Cabral Coutinho do Rego Monteiro, I.; Pontes Dos Santos, L.H.; Soares Marreiros Ferraz, A.; Carneiro Loureiro, A.C.; Calado Lima, C.; Leal-Cardoso, J.H.; Pires Carvalho, D.; Soares Fortunato, R.; et al. Strenuous acute exercise induces slow and fast twitch-dependent NADPH oxidase expression in rat skeletal muscle. Antioxidants (Basel) 2020, 9, 57.

- Sakellariou, G.K.; Vasilaki, A.; Palomero, J.; Kayani, A.; Zibrik, L.; McArdle, A.; Jackson, M.J. Studies of mitochondrial and nonmitochondrial sources implicate nicotinamide adenine dinucleotide phosphate oxidase(s) in the increased skeletal muscle superoxide generation that occurs during contractile activity. Antioxid. Redox Signal. 2013, 18, 603– 621.
- Breton-Romero, R.; Lamas, S. Hydrogen peroxide signaling in vascular endothelial cells. Redox Biol. 2014, 2, 529– 534.
- 8. Sundaresan, M.; Yu, Z.X.; Ferrans, V.J.; Sulciner, D.J.; Gutkind, J.S.; Irani, K.; Goldschmidt-Clermont, P.J.; Finkel, T. Regulation of reactive-oxygen-species generation in fibroblasts by Rac1. Biochem. J. 1996, 318 Pt 2, 379–382.
- 9. Bedard, K.; Krause, K.H. The NOX family of ROS-generating NADPH oxidases: Physiology and pathophysiology. Physiol. Rev. 2007, 87, 245–313.
- 10. Hajjar, C.; Cherrier, M.V.; Dias Mirandela, G.; Petit-Hartlein, I.; Stasia, M.J.; Fontecilla-Camps, J.C.; Fieschi, F.; Dupuy, J. The NOX family of proteins is also present in bacteria. mBio 2017, 8.
- 11. Babior, B.M.; Lambeth, J.D.; Nauseef, W. The neutrophil NADPH oxidase. Arch. Biochem. Biophys. 2002, 397, 342–344.
- 12. Burdon, R.H. Superoxide and hydrogen peroxide in relation to mammalian cell proliferation. Free Radic. Biol. Med. 1995, 18, 775–794.
- 13. Lambeth, J.D.; Cheng, G.; Arnold, R.S.; Edens, W.A. Novel homologs of gp91phox. Trends Biochem. Sci. 2000, 25, 459–461.
- 14. Sumimoto, H. Structure, regulation and evolution of Nox-family NADPH oxidases that produce reactive oxygen species. FEBS J. 2008, 275, 3249–3277.
- 15. Magnani, F.; Nenci, S.; Millana Fananas, E.; Ceccon, M.; Romero, E.; Fraaije, M.W.; Mattevi, A. Crystal structures and atomic model of NADPH oxidase. Proc. Natl. Acad. Sci. USA 2017, 114, 6764–6769.
- 16. Pryor, W.A. Oxy-radicals and related species: Their formation, lifetimes, and reactions. Annu. Rev. Physiol. 1986, 48, 657–667.
- 17. Ferrer-Sueta, G.; Radi, R. Chemical biology of peroxynitrite: Kinetics, diffusion, and radicals. ACS Chem. Biol. 2009, 4, 161–177.
- 18. Bienert, G.P.; Chaumont, F. Aquaporin-facilitated transmembrane diffusion of hydrogen peroxide. Biochim. Biophys. Acta 2014, 1840, 1596–1604.
- 19. Rhee, S.G.; Bae, Y.S.; Lee, S.R.; Kwon, J. Hydrogen peroxide: A key messenger that modulates protein phosphorylation through cysteine oxidation. Sci. STKE 2000, 2000, pe1.
- 20. Krause, K.H. Tissue distribution and putative physiological function of NOX family NADPH oxidases. Jpn. J. Infect. Dis. 2004, 57, S28–S29.
- Ambasta, R.K.; Kumar, P.; Griendling, K.K.; Schmidt, H.H.; Busse, R.; Brandes, R.P. Direct interaction of the novel Nox proteins with p22phox is required for the formation of a functionally active NADPH oxidase. J. Biol. Chem. 2004, 279, 45935–45941.
- 22. Groemping, Y.; Lapouge, K.; Smerdon, S.J.; Rittinger, K. Molecular basis of phosphorylation-induced activation of the NADPH oxidase. Cell 2003, 113, 343–355.
- 23. Hiroaki, H.; Ago, T.; Ito, T.; Sumimoto, H.; Kohda, D. Solution structure of the PX domain, a target of the SH3 domain. Nat. Struct. Biol. 2001, 8, 526–530.
- 24. Bokoch, G.M. Regulation of innate immunity by Rho GTPases. Trends Cell Biol. 2005, 15, 163–171.
- 25. Mizuno, T.; Kaibuchi, K.; Ando, S.; Musha, T.; Hiraoka, K.; Takaishi, K.; Asada, M.; Nunoi, H.; Matsuda, I.; Takai, Y. Regulation of the superoxide-generating NADPH oxidase by a small GTP-binding protein and its stimulatory and inhibitory GDP/GTP exchange proteins. J. Biol. Chem. 1992, 267, 10215–10218.
- 26. Han, C.H.; Freeman, J.L.; Lee, T.; Motalebi, S.A.; Lambeth, J.D. Regulation of the neutrophil respiratory burst oxidase. Identification of an activation domain in p67(phox). J. Biol. Chem. 1998, 273, 16663–16668.
- 27. Diebold, B.A.; Bokoch, G.M. Molecular basis for Rac2 regulation of phagocyte NADPH oxidase. Nat. Immunol. 2001, 2, 211–215.
- Lapouge, K.; Smith, S.J.; Walker, P.A.; Gamblin, S.J.; Smerdon, S.J.; Rittinger, K. Structure of the TPR domain of p67phox in complex with Rac.GTP. Mol. Cell 2000, 6, 899–907.
- 29. Geiszt, M.; Lekstrom, K.; Witta, J.; Leto, T.L. Proteins homologous to p47phox and p67phox support superoxide production by NAD(P)H oxidase 1 in colon epithelial cells. J. Biol. Chem. 2003, 278, 20006–20012.

- Ueno, N.; Takeya, R.; Miyano, K.; Kikuchi, H.; Sumimoto, H. The NADPH oxidase Nox3 constitutively produces superoxide in a p22phox-dependent manner: Its regulation by oxidase organizers and activators. J. Biol. Chem. 2005, 280, 23328–23339.
- 31. Ueyama, T.; Geiszt, M.; Leto, T.L. Involvement of Rac1 in activation of multicomponent Nox1- and Nox3-based NADPH oxidases. Mol. Cell Biol. 2006, 26, 2160–2174.
- 32. Martyn, K.D.; Frederick, L.M.; von Loehneysen, K.; Dinauer, M.C.; Knaus, U.G. Functional analysis of Nox4 reveals unique characteristics compared to other NADPH oxidases. Cell Signal. 2006, 18, 69–82.
- Lyle, A.N.; Deshpande, N.N.; Taniyama, Y.; Seidel-Rogol, B.; Pounkova, L.; Du, P.; Papaharalambus, C.; Lassegue, B.; Griendling, K.K. Poldip2, a novel regulator of Nox4 and cytoskeletal integrity in vascular smooth muscle cells. Circ. Res. 2009, 105, 249–259.
- 34. Banfi, B.; Molnar, G.; Maturana, A.; Steger, K.; Hegedus, B.; Demaurex, N.; Krause, K.H. A Ca(2+)-activated NADPH oxidase in testis, spleen, and lymph nodes. J. Biol. Chem. 2001, 276, 37594–37601.
- 35. Cheng, G.; Cao, Z.; Xu, X.; van Meir, E.G.; Lambeth, J.D. Homologs of gp91phox: Cloning and tissue expression of Nox3, Nox4, and Nox5. Gene 2001, 269, 131–140.
- 36. Banfi, B.; Tirone, F.; Durussel, I.; Knisz, J.; Moskwa, P.; Molnar, G.Z.; Krause, K.H.; Cox, J.A. Mechanism of Ca2+ activation of the NADPH oxidase 5 (NOX5). J. Biol. Chem. 2004, 279, 18583–18591.
- 37. Chen, F.; Pandey, D.; Chadli, A.; Catravas, J.D.; Chen, T.; Fulton, D.J. Hsp90 regulates NADPH oxidase activity and is necessary for superoxide but not hydrogen peroxide production. Antioxid. Redox Signal 2011, 14, 2107–2119.
- Fu, X.; Beer, D.G.; Behar, J.; Wands, J.; Lambeth, D.; Cao, W. cAMP-response element-binding protein mediates acidinduced NADPH oxidase NOX5-S expression in Barrett esophageal adenocarcinoma cells. J. Biol. Chem. 2006, 281, 20368–20382.
- 39. Dupuy, C.; Ohayon, R.; Valent, A.; Noel-Hudson, M.S.; Deme, D.; Virion, A. Purification of a novel flavoprotein involved in the thyroid NADPH oxidase. Cloning of the porcine and human cdnas. J. Biol. Chem. 1999, 274, 37265–37269.
- 40. De Deken, X.; Wang, D.; Dumont, J.E.; Miot, F. Characterization of ThOX proteins as components of the thyroid H(2)O(2)-generating system. Exp. Cell Res. 2002, 273, 187–196.
- 41. Sugawara, M.; Sugawara, Y.; Wen, K.; Giulivi, C. Generation of oxygen free radicals in thyroid cells and inhibition of thyroid peroxidase. Exp. Biol Med. (Maywood) 2002, 227, 141–146.
- 42. Ameziane-El-Hassani, R.; Morand, S.; Boucher, J.L.; Frapart, Y.M.; Apostolou, D.; Agnandji, D.; Gnidehou, S.; Ohayon, R.; Noel-Hudson, M.S.; Francon, J.; et al. Dual oxidase-2 has an intrinsic Ca2+-dependent H2O2-generating activity. J. Biol. Chem. 2005, 280, 30046–30054.

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