

βTCP-Poly(3hydroxybutyrate) for Bone Tissue Engineering

Subjects: **Others**

Contributor: Maciej Guzik

Tissue engineering is a technique that involves the in vitro seeding and attachment of cells onto a three-dimensional scaffold. In the case of bone tissue engineering, investigations have been focused mostly on synthetic bioceramic scaffolds based on calcium phosphates, such as hydroxyapatite or tricalcium phosphate. Due to their chemical similarity to an inorganic component of bone, hydroxyapatite, as well as β TCP- and α TCP-based materials, show excellent biocompatibility and osteoconductivity.

β tricalcium phosphate

silver-decorated scaffolds

P(3HB) coating

P(3HB) degradation

bacterial polymer

1. Overview

Implantations in orthopedics are associated with a high risk of bacterial infections in the surgery area. Therefore, biomaterials containing antibacterial agents, such as antibiotics, bactericidal ions or nanoparticles have been intensively investigated. In this work, silver decorated β tricalcium phosphate (β TCP)-based porous scaffolds were obtained and coated with a biopolymer—poly(3-hydroxybutyrate)-P(3HB). To the best of our knowledge, studies using silver-doped β TCP and P(3HB), as a component in ceramic-polymer scaffolds for bone tissue regeneration, have not yet been reported. Obtained materials were investigated by high-temperature X-ray diffraction, X-ray fluorescence, scanning electron microscopy with energy dispersive spectroscopy, hydrostatic weighing, compression tests and ultrahigh-pressure liquid chromatography with mass spectrometry (UHPLC-MS) measurements. The influence of sintering temperature (1150, 1200 °C) on the scaffolds' physicochemical properties (phase and chemical composition, microstructure, porosity, compressive strength) was evaluated. Materials covered with P(3HB) possessed higher compressive strength (3.8 ± 0.6 MPa) and surgical maneuverability, sufficient to withstand the implantation procedures. Furthermore, during the hydrolytic degradation of the composite material not only pure (R)-3-hydroxybutyric acid but also its oligomers were released which may nourish surrounding tissues. Thus, obtained scaffolds were found to be promising bone substitutes for use in non-load bearing applications

2. Calcium Phosphate

Tissue engineering is a technique that involves the in vitro seeding and attachment of cells onto a three-dimensional scaffold. In the case of bone tissue engineering, investigations have been focused mostly on synthetic

bioceramic scaffolds based on calcium phosphates, such as hydroxyapatite or tricalcium phosphate. Due to their chemical similarity to an inorganic component of bone, hydroxyapatite, as well as β TCP- and α TCP-based materials, show excellent biocompatibility and osteoconductivity. To further enhance their physicochemical and biological characteristics modifications with monovalent (Ag^+ , Na^+), divalent (Mg^{2+} , Zn^{2+}) and trivalent (Fe^{3+}) metal ions were done [1][2][3]. Due to the high risk of bacterial infections during implantation procedures, biomaterials containing antibacterial agents, such as antibiotics, bactericidal ions (e.g., Ag^+ , Au^+ , Cu^{2+}) and nanoparticles have been intensively investigated. Recently, silver-modified biomaterials have been prepared by various methods that, *inter alia*, involve the incorporation of silver ions into their structure or attachment of silver nanoparticles on their surface. Silver-containing tricalcium phosphate microspheres composed of α/β TCP phases were synthesized using an ultrasonic spray-pyrolysis technique [4]. Su et al. [5] studied silver containing calcium phosphate coatings on pure iron foam obtained via co-deposition and post-treatment method. Siek et al. [6] used wet chemical method to obtain bactericidal α TCP-based bone cements with silver-modified hydroxyapatite (Ag-HA) and CaCO_3 . It has been found that bone cement matrix did not impede the Ag^+ ions release from the Ag-HA agglomerates and antibacterial activity depended on the kind of bacterial strain. Hoover et al. [7] obtained silver-modified porous β -tricalcium phosphate (β TCP) scaffolds using liquid porogen based method. In their work silver added in the amount between 0.5 and 2 wt.% Ag_2O could be released over a long period of time without compromising the biocompatibility of the scaffolds. The bactericidal activity was achieved due to sustained release of Ag^+ ions through the continuous dissolution of Ag-modified β TCP. Materials for various biomedical applications, including chitosan [8], cellulose and its derivatives [9][10] as well as mesoporous carbons [11] decorated with silver nanoparticles were also examined. However, silver decorated calcium phosphate-based scaffolds seem to be an interesting and not yet fully explored area.

Calcium phosphate (CaP)-based bioceramic scaffolds are inherently brittle and often cannot match the mechanical properties of the bone. Thus, composites made of CaPs and bioresorbable polymers have also been investigated [12][13]. Recently, many researchers have been working on the fabrication of polymer-coated bone scaffolds including ceramic-biopolymer hybrid systems [14][15]. A new trend in the development of biomedical composites is a usage of polyhydroxyalkanoates (PHAs)—the bacterial derived polymers [16][17]. Under normal growth conditions, most bacteria produce only a small amount of PHA (1–15%). When special growth conditions and fermentation strategies are applied, the synthesized PHA can reach almost 90% [18]. Medical applications require a constant and reproducible quality of PHAs, which can be achieved via bacterial production in a rigorous culture [19]. Polyhydroxyalkanoates are a diverse group of materials with different applications and properties. It is known that more than 150 types of PHAs can be synthesized by microorganisms [20]. The material characteristics associated with PHAs are affected by many parameters including their chemical structure and type of monomer units with molecular mass. PHAs can be classified into short chain length (scl) PHAs with 4–5 carbons in pending monomers backbone or medium chain length (mcl) PHAs with (6–14 carbon atoms), when aliphatic monomers are present, and may further be grouped a homo-polymers (either scl-PHAs or mcl-PHAs) or copolymers (a mixture of different monomers of scl-PHA and/or mcl-PHA monomers). There were many attempts to use PHAs in medical applications due to their excellent biocompatibility and biodegradability [21][22][23][24]. PHAs have an advantage over other bioplastics such a poly(lactic acid) (PLA) or poly(D,L-lactide-co-glycolide) (PLGA) since their monomers (3-

hydroxylated acids) are quickly metabolized within the human body. Furthermore, they can be naturally detected in almost all parts of the body as a degradation product and were found not to cause carcinogenesis during long-term implantation [25][26]. Among the PHA family, the P(3HB) is the most common and well-characterized scl-PHA. The possibility of using P(3HB) as coating materials for composite type inorganic-organic scaffolds was demonstrated among others by Montazeri et al. [27]. It has been found that scaffold made of bioglass covered with polyhydroxybutyrate (PHB) possessed higher mechanical strength and bioactivity than nano-bioglass strut alone. In this study, to improve the mechanical properties and surgical maneuverability of brittle silver decorated β TCP scaffolds, they were covered with bioresorbable P(3HB) polymer. These kinds of highly-porous materials may be potentially applied for filling small bone defects in non-load or low-load bearing places. In the future the P(3HB) coating may serve as a drug delivery vehicle. Development of novel methodologies to fabricate bioceramic-PHAs composites may open new horizons for their applications in medicine. Therefore, silver decorated calcium phosphates, covered by a PHA layer seem to be interesting materials that have not yet been explored. This proof-of-concept study delivers some insights into the synthesis of such materials and their characterization, setting a benchmark for further developments of material for regenerative medicine based on biopolymers and bioceramics.

3. Conclusions

The macroporous β -tricalcium phosphate scaffolds modified with silver (Ag- β TCP) were prepared by a polyurethane foam replica method, followed by the coating of bacterially derived polymer-P(3HB). Materials with open porosity between $61.8 \pm 3.0\%$ – $68.1 \pm 4.6\%$ and pore sizes in the range of 100–700 μm , were obtained. Concerning the consolidation of bioceramic struts, an appropriate sintering regime was achieved at 1150 °C, which is reflected in the highly dense microstructure with a small amount of microcracks. In higher temperatures, exceeding the critical grain size resulted in regions with local intergranular cracks. The compressive strength of scaffolds was in the range from 2.1 ± 0.6 to 3.8 ± 0.6 MPa, which is comparable to the compressive strength of spongy bone and can be sufficient for implantation in the low-load-bearing places. The designed scaffolds cannot be applied for load-bearing applications. Scaffolds covered with polymer possessed higher compressive strength and surgical maneuverability, sufficient to withstand the implantation procedures. Notably, *in vivo* the mechanical properties of scaffolds should increase with time as the tissue grows and the scaffold degrades. Moreover, releasing not only pure (R)-3-hydroxybutyric acid but also its oligomers during hydrolytic degradation of the composite material was confirmed, which may be beneficial for the surrounding tissues as a nourishing agent. To further improve ductility and adhesion of P(3HB) film to the scaffold it would be reasonable to try blending P(3HB) with other polymers. This would be the subject of further investigations. Further biological studies considering biocompatibility and antibacterial properties of Ag- β TCP/poly(3hydroxybutyrate) scaffolds are necessary.

References

1. Yoshida, K.; Kondo, N.; Kita, H.; Mitamura, M.; Hashimoto, K.; Toda, Y. Effect of substitutional monovalent and divalent metal ions on mechanical properties of β -tricalcium phosphate. *J. Am.*

Ceram. Soc. 2005, 88, 2315–2318.

2. Matsumoto, N.; Sato, K.; Yoshida, K.; Hashimoto, K.; Toda, Y. Preparation and characterization of β -tricalcium phosphate co-doped with monovalent and divalent antibacterial metal ions. *Acta Biomater.* 2009, 5, 3157–3164.
3. Bohner, M.; Santoni, B.L.G.; Döbelin, N. β -tricalcium phosphate for bone substitution: Synthesis and properties. *Acta Biomater.* 2020, 113, 23–41.
4. Honda, M.; Kawanobe, Y.; Nagata, K.; Ishii, K.; Matsumoto, M.; Aizawa, M. Bactericidal and bioresorbable calcium phosphate cements fabricated by silver-containing tricalcium phosphate microspheres. *Int. J. Mol. Sci.* 2020, 21, 3745.
5. Su, Y.; Champagne, S.; Trenggono, A.; Tolouei, R.; Mantovani, D.; Hermawan, H. Development and characterization of silver containing calcium phosphate coatings on pure iron foam intended for bone scaffold applications. *Mater. Des.* 2018, 148, 124–134.
6. Siek, D.; Ślósarczyk, A.; Przekora, A.; Belcarz, A.; Zima, A.; Ginalska, G.; Czechowska, J. Evaluation of antibacterial activity and cytocompatibility of α -TCP based bone cements with silver-doped hydroxyapatite and CaCO₃. *Ceram. Int.* 2017, 43, 13997–14007.
7. Hoover, S.; Tarafder, S.; Bandyopadhyay, A.; Bose, S. Silver doped resorbable tricalcium phosphate scaffolds for bone graft applications. *Mater. Sci. Eng. C* 2017, 79, 763–769.
8. Dastidar, D.G.; Ghosh, D. Silver Nanoparticle Decorated Chitosan Scaffold for Wound Healing and Tissue Regeneration. *Macromolecules* 2018, 105, 1241–1249.
9. Hasan, A.; Waibhaw, G.; Saxena, V.; Pandey, L.M. Nano-biocomposite scaffolds of chitosan, carboxymethyl cellulose and silver nanoparticle modified cellulose nanowhiskers for bone tissue engineering applications. *Int. J. Biol. Macromol.* 2018, 111, 923–934.
10. Busuioc, C.; Nicoara, A.I. Bacterial cellulose hydroxyapatite composites decorated with silver nanoparticles for medical applications. *Eng. Biomater.* 2019, 22, 153.
11. Torre, E.; Giasafaki, D.; Steriotis, T.; Cassinelli, C.; Morra, M.; Fiorilli, S.; Vitale-Brovarone, C.; Charalambopoulou, G.; Ivgilia, G. Silver decorated mesoporous carbons for the treatment of acute and chronic wounds, in a tissue regeneration context. *Int. J. Nanomed.* 2019, 14, 10147.
12. Philippart, A.; Boccaccini, A.R.; Fleck, C.; Schubert, D.W.; Roether, J.A. Toughening and functionalization of bioactive ceramic and glass bone scaffolds by biopolymer coatings and infiltration: A review of the last 5 years. *Expert Rev. Med. Devices* 2015, 12, 93–111.
13. Dziadek, M.; Zima, A.; Cichoń, E.; Czechowska, J.; Ślósarczyk, A. Biomicroconcretes based on the hybrid HAp/CTS granules, α -TCP and pectins as a novel injectable bone substitutes. *Mater. Lett.* 2020, 265, 127457.

14. Cichoń, E.; Haraźna, K.; Skibiński, S.; Witko, T.; Zima, A.; Ślósarczyk, A.; Zimowska, M.; Witko, M.; Leszczynski, B.; Wrobel, A.; et al. Novel bioresorbable tricalcium phosphate/polyhydroxyoctanoate (TCP/PHO) composites as scaffolds for bone tissue engineering applications. *J. Mech. Behav. Biomed. Mater.* 2019, 98, 235–245.
15. Skibiński, S.; Cichoń, E.; Haraźna, K.; Marcello, E.; Roy, I.; Witko, M.; Słosarczyk, A.; Czechowska, J.; Guzik, M.; Zima, A. Functionalized tricalcium phosphate and poly (3-hydroxyoctanoate) derived composite scaffolds as platforms for the controlled release of diclofenac. *Ceram. Int.* 2021, 47, 3876–3883.
16. Ray, S.; Patel, S.K.; Singh, M.; Singh, G.P.; Kalia, V.C. Exploiting polyhydroxyalkanoates for tissue engineering. In *Biotechnological Applications of Polyhydroxyalkanoates*; Springer: Singapore, 2019; pp. 271–282.
17. Peptu, C.; Kowalcuk, M. Biomass-derived polyhydroxyalkanoates: Biomedical applications. In *Biomass as Renewable Raw Material to Obtain Bioproducts of High-Tech Value*; Elsevier: Amsterdam, The Netherlands, 2018; pp. 271–313.
18. Alves, M.I.; Macagnan, K.L.; Rodrigues, A.A.; de Assis, D.A.; Torres, M.M.; de Oliveira, P.D.; Furlan, L.; Vendruscolo, C.T.; Moreira, A.D.S. Poly (3-hydroxybutyrate)-P (3HB): Review of production process technology. *Ind. Biotechnol.* 2017, 13, 192–208.
19. Koller, M. A review on established and emerging fermentation schemes for microbial production of Polyhydroxyalkanoate (PHA) biopolymers. *Fermentation* 2018, 4, 30.
20. Steinbüchel, A.; Valentin, H.E. Diversity of bacterial polyhydroxyalkanoic acids. *FEMS Microbiol. Lett.* 1995, 128, 219–228.
21. Zinn, M.; Witholt, B.; Egli, T. Occurrence, synthesis and medical application of bacterial polyhydroxyalkanoate. *Adv. Drug Deliv. Rev.* 2001, 53, 5–21.
22. Singh, A.K.; Srivastava, J.K.; Chandel, A.K.; Sharma, L.; Mallick, N.; Singh, S.P. Biomedical applications of microbially engineered polyhydroxyalkanoates: An insight into recent advances, bottlenecks, and solutions. *Appl. Microbiol. Biotechnol.* 2019, 103, 2007–2032.
23. Ali, I.; Jamil, N. Polyhydroxyalkanoates: Current applications in the medical field. *Front. Biol.* 2016, 11, 19–27.
24. Sanhueza, C.; Acevedo, F.; Rocha, S.; Villegas, P.; Seeger, M.; Navia, R. Polyhydroxyalkanoates as biomaterial for electrospun scaffolds. *Int. J. Biol. Macromol.* 2019, 124, 102–110.
25. Philip, S.; Keshavarz, T.; Roy, I. Polyhydroxyalkanoates: Biodegradable polymers with a range of applications. *J. Chem. Technol. Biotechnol. Int. Res. Process Environ. Clean Technol.* 2007, 82, 233–247.

26. Peng, S.W.; Guo, X.Y.; Shang, G.G.; Li, J.; Xu, X.Y.; You, M.L.; Li, P.; Chen, G.Q. An assessment of the risks of carcinogenicity associated with polyhydroxyalkanoates through an analysis of DNA aneuploid and telomerase activity. *Biomaterials* 2011, 32, 2546–2555.
27. Montazeri, M.; Karbasi, S.; Foroughi, M.R.; Monshi, A.; Ebrahimi-Kahrizsangi, R. Evaluation of mechanical property and bioactivity of nano-bioglass 45S5 scaffold coated with poly-3-hydroxybutyrate. *J. Mater. Sci. Mater. Med.* 2015, 26, 62.

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