

GTS's Candidate Genes and Pathways

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Gilles de la Tourette syndrome (GTS) is a childhood-onset neurodevelopmental and -psychiatric tic-disorder of complex etiology which is often comorbid with obsessive-compulsive disorder (OCD) and/or attention deficit hyperactivity disorder (ADHD). Twin and family studies of GTS individuals have shown a high level of heritability suggesting, that genetic risk factors play an important role in disease etiology. However, the identification of major GTS susceptibility genes has been challenging, presumably due to the complex interplay between several genetic factors and environmental influences, low penetrance of each individual factor, genetic diversity in populations, and the presence of comorbid disorders. Even though several strong candidate genes have hitherto been identified, none of these have turned out to be major susceptibility genes yet.

Keywords: Gilles de la Tourette syndrome ; GTS ; tics ; human genetics ; neurotransmission ; neurodevelopmental disorders ; dopamine ; serotonin ; movement disorders

1. Introduction

Gilles de la Tourette syndrome (GTS) is a complex early-onset neurodevelopmental/-psychiatric disorder with a prevalence between 0.52% and 0.77% in children ^{[1][2]}. GTS is one of the four tic disorders included in the fifth edition of the American Diagnostic and Statistical Manual of Mental Disorders (DSM-5): GTS, persistent (chronic) tic disorder (CTD), provisional tic disorder, and other specified and unspecified tic disorders. Diagnostic criteria for GTS are the presence of multiple motor tics and at least one vocal tic persisting for at least one year with onset of the first tic before age 18 ^[3]. Diagnostic criteria for CTD includes all of these criteria that exception that an individual has either motor or vocal tics, but not both. Provisional tic disorder meets all of the criteria for GTS except for the fact that tics do not last longer than 12 months. Finally, it is categorized as other specified and unspecified tic disorder when none of the above criteria are met ^[3]. Recently, the term "tic spectrum disorders" was suggested to replace the GTS and CTD diagnoses ^[4]. In addition to tics, most GTS individuals present a variety of symptoms due to comorbidities, the most common of which are obsessive-compulsive disorder (OCD) and/or attention deficit hyperactivity disorder (ADHD), and to a lesser extent autism spectrum disorder (ASD), migraine, anxiety, depression, sleep disorders, and rage attacks, implying an overlapping etiology ^{[5][6][7][8][9][10]}. Several neurotransmitter systems have been implicated in disease pathogenesis and amongst these, the dopaminergic and the serotonergic pathways are the most widely studied.

GTS is a complex disorder, where several environmental factors (such as streptococcus infections, birth complications and maternal smoking) are thought to interact with multiple genes in yet undiscovered ways ^[11]. Altered immune regulation is also suggested to predispose to inflammation and infection, thereby triggering GTS ^[11], and there is growing evidence for dysregulation of the brain's resident immune cells, the microglia ^[12]. Several lines of evidence suggest that GTS has a strong genetic component ^[13], and it has been suggested as one of the non-Mendelian neuropsychiatric disorders with the highest heritability ^[9]. However, the identification of susceptibility genes has been challenging, which is likely due to the complex and heterogeneous genetic architecture of GTS, wherein common and rare variants in several different genes and biological pathways are involved. Even though neurophysiology and neuroimaging studies suggest that GTS is associated with altered synaptic neurotransmission, the pathophysiology is far from understood, which also hampers the identification of possible associated genetic factors. The overall results of the hitherto genetic studies suggest that in lieu of one or more major susceptibility loci, several rare and low-penetrance variants account for the genetic risk factors in GTS etiology.

2. Candidate Gene and Pathway

Most of the studies utilizing a candidate gene/pathway approach have focused on the genes encoding the proteins of the neurotransmitter pathways of the cortico-basal ganglia-thalamo-cortical (CBGTC) loops. The CBGTC loops are the primary neural circuits associated with GTS pathogenesis, and they are altered in both structure and activity in GTS

individuals [14][15][16]. The CBGTC loops connect specific regions of the cerebral cortex, the thalamus, and the basal ganglia, which is associated with several movement disorders [17]; and notably, GTS is also categorized as a movement disorder. Multiple neurotransmission systems present within the CBGTC loops, including dopaminergic, serotonergic, glutamatergic, and γ -aminobutyric acid (GABA)ergic systems, have been the target for GTS candidate gene studies.

2.1. The Dopaminergic Pathway

The most extensively studied neurotransmission system in GTS is the dopaminergic pathway (**Figure 1**). Upon the excitation of the dopaminergic neurons, dopamine is released from the synaptic vesicles of the presynaptic neurons into the synaptic cleft, where it is either transported back to the cytosol facilitated by the dopamine transporter DAT1 (encoded by *SLC6A3*) or interacts with post- or presynaptic dopamine receptors [18]. Dopamine receptors are categorized in two classes; D1-like receptors (D1 and D5 are encoded by *DRD1* and *DRD5*, respectively) and D2-like receptors (D2, D3, and D4 are encoded by *DRD2*, *DRD3*, and *DRD4*, respectively) coupled to excitatory and inhibitory G proteins, respectively [19]. Other players of the dopaminergic pathway include monoamine oxidase A (encoded by *MAOA*) and catechol O-methyltransferase (encoded by *COMT*), which metabolize unbound cytosolic dopamine, and dopamine β -hydroxylase (encoded by *DBH*), which catalyzes the conversion of dopamine to norepinephrine [18].

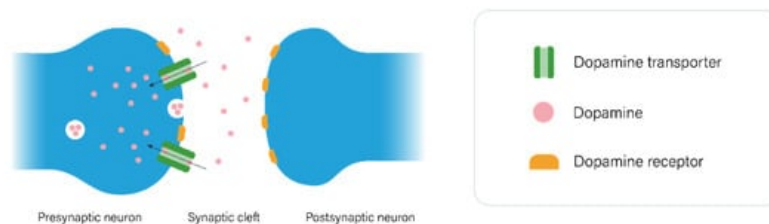


Figure 1. Visualization of dopaminergic neurotransmission in the synaptic cleft. Dopamine is released from the vesicles in the presynaptic neuron into the synaptic cleft, where it binds to either presynaptic or postsynaptic dopamine receptors or is transported back to the cytosol of the presynaptic neuron by dopamine transporters.

Most of the studies investigating the genes of the dopaminergic pathway in GTS have been carried out in small cohorts with limited statistical power and an unequivocal conclusion regarding the involvement of the dopaminergic pathway genes in GTS etiology cannot be drawn. Similarly, none of the later GTS genome-wide association studies (GWASs) identified any significant association, altogether suggesting that genes of the dopaminergic pathway do not markedly contribute to GTS pathogenesis.

2.2. The Serotonergic Pathway

The serotonergic pathway, which is closely linked to the dopaminergic pathway, has also been studied in GTS, especially in OCD etiology [20]. Similar to dopamine, serotonin (5-hydroxytryptamine, 5-HT) is released from a presynaptic neuron into the synaptic cleft and is transported back to the cytosol by the serotonin transporter SERT (encoded by *SLC6A4*) or is bound to serotonin receptors. There are at least 14 different classes of serotonin receptors on the post- or presynaptic membranes [21]. Serotonin metabolism involves tryptophan 5-hydroxylase 2 (encoded by *TPH2*) which is responsible for the biosynthesis of serotonin from L-tryptophan; as for dopamine, MAOA is responsible for the degradation of serotonin to 5-hydroxyindole acetic acid (5-HIAA).

Several serotonin receptors, including 5-HT1A, 5-HT1B, 5-HT2A, 5-HT2C, 5-HT3, and 5-HT4, are capable of facilitating and inhibiting dopamine activity in addition to that of serotonin [22][23][24], just as SERT can act both as a serotonin and dopamine transporter [25]. This has collectively led to the speculation that these two neurotransmitter pathways might be involved in various neuropsychiatric disorders in a combined manner. However, while the genes encoding the proteins of the dopaminergic pathway have been extensively studied in GTS, similar studies for the serotonergic pathway are scarce.

SLC6A4 encoding SERT has repeatedly been investigated in GTS cohorts focusing particularly on the SERT-linked polymorphic region (5-HTTLPR) in the promoter region of *SLC6A4*. The 5-HTTLPR is a 43bp insertion/deletion giving rise to a long (L) allele and a short (S) allele (**Figure 2**). The L_{AC} haplotype, which is a combination of the L allele and the major alleles of the rs25531 (A/G) and rs25532 (C/T) SNPs and which is associated with the highest rate of *SLC6A4* mRNA expression (**Figure 2**), was found to be more prevalent in 151 GTS individuals compared to 858 controls [26]. Recently, *SLC6A4* promoter variants and mRNA levels were investigated in a Danish cohort comprising 72 GTS individuals and 87 controls. *SLC6A4* mRNA expression levels were found to be higher in GTS individuals, particularly when only considering the individuals with the L_{AC}/L_{AC} genotype [27]. In earlier studies, an association between GTS and

the 5-HTTLPR variants was not detected [28][29][30]. These contradictory results may be due to the previously unnoticed effect of the rs25531 (A/G) and rs25532 (C/T) SNPs on *SLC6A4* mRNA expression, as it is likely that higher expression of *SLC6A4* mRNA is associated with the 5-HTTLPR/rs25531 L_A allele but not with the L_G and S alleles [31] (**Figure 2**). Most recently, the exome sequencing of 13 multiplex GTS families suggested *SLC6A4* as a high confidence GTS risk gene [32].

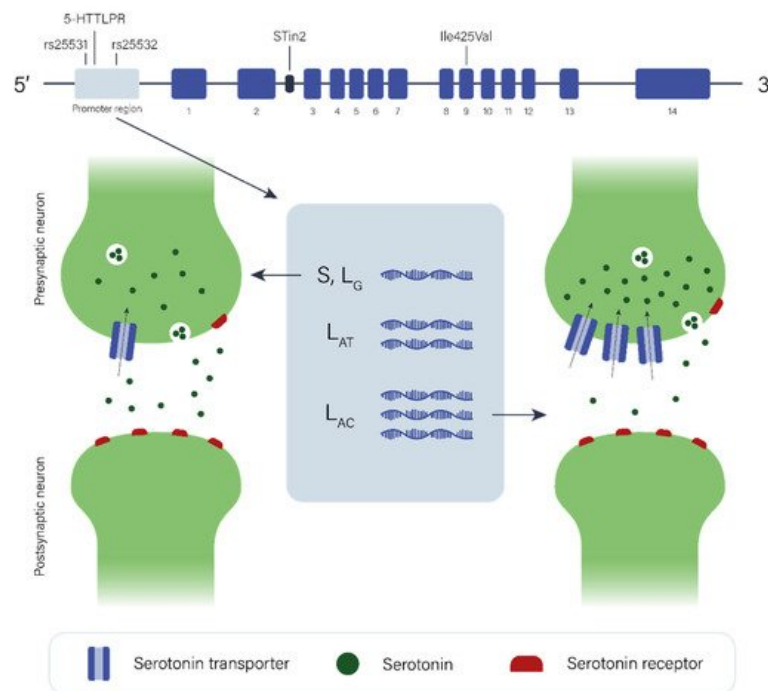


Figure 2. *SLC6A4* and serotonergic neurotransmission. *SLC6A4* gene, exon numbers, and the polymorphisms 5-HTTLPR, rs25531, rs25532, STin2, and Ile425Val in the top part of the figure. The different 5-HTTLPR/rs25531/rs25532 three-locus haplotypes (**bottom middle**) are likely to affect *SLC6A4* mRNA expression (**bottom middle**). It is hypothesized that S and L_G haplotypes result in low *SLC6A4* mRNA expression (**bottom left**), while the L_{AC} haplotype results in *SLC6A4* overexpression, leading to increased SERT in the presynaptic neuron followed by increased serotonin clearance from the synaptic cleft (**bottom right**). 5-HTTLPR: SERT-linked polymorphic region; S: short allele of 5-HTTLPR; L_G: long allele of 5-HTTLPR and minor allele of rs25531; L_{AT} and L_{AC}: 5-HTTLPR/rs25531/rs25532 three-locus haplotypes, differing in rs25532 alleles.

The data regarding the role of the serotonergic pathway in GTS etiology are limited and again, individual studies have been of limited statistical power with low sample size. However, when studies of the dopaminergic system are taken into consideration, it is plausible to hypothesize that the serotonergic system may play a complex role in GTS etiology, directly and/or indirectly in combination with or through modulating dopaminergic neurotransmission. Additional neurotransmitter pathways (e.g., glutamatergic and GABAergic) may also be involved in the pathology of GTS, as the dysfunction of one pathway may affect others due to interaction or self-regulation among them.

3. Linkage Analyses, GWAS & Other Studies

3.1. *SLITRK1*

SLITRK1 (Slit and Trk-like 1) is the most investigated GTS candidate gene, since one of the breakpoints of a *de novo* inversion of chromosome 13 in a child with GTS was mapped within its proximity [33]. Functional studies showed that neurite outgrowth was affected by a truncating *SLITRK1* variant (varCDF) identified in a GTS individual, and a missense variant (var321) in the 3' untranslated region (3'-UTR) of the gene altered a microRNA binding site leading to decreased mRNA expression [33].

SLITRK1 promotes neurite outgrowth and *SLITRK1* is predominantly expressed in the brain, including the CBGTC circuits [34]. In addition, the six proteins belonging to the *SLITRK* family play important roles in the development of the central nervous system and neuronal processes and have been implicated in several other neurodevelopmental and -psychiatric disorders [35]. The *SLITRK*-genes have therefore been attractive candidates in the search for the genes involved in GTS etiology. However, it is likely that these genes, especially *SLITRK1*, do not play a major role in GTS etiology.

3.2. *IMMPL2*

Similar to *SLITRK1*, *IMMPL2* (inner mitochondrial membrane peptidase, subunit 2) was suggested as a candidate GTS gene by the finding of a chromosome aberration in a GTS individual, where the gene was disrupted by a 7q31 breakpoint [36], a region previously linked to GTS [37]. The role of *IMMPL2* and the 7q31 region in GTS etiology was further supported by the finding of another Caucasian individual with tics and a disruption of *IMMPL2* [38] and a family based association study (86 French Canadian GTS trios) suggesting an association between GTS and several 7q31 markers, including a tendency of association with an intragenic *IMMPL2* marker [39]. A recent meta-analysis of previously reported GTS candidate genes in European GTS individuals and controls detected an association between GTS and *IMMPL2* [40], further implicating *IMMPL2* in GTS etiology. However, how a defective *IMMPL2* function may be involved in GTS pathogenesis is still unknown.

3.3. *HDC*

HDC encodes the histidine decarboxylase enzyme essential for histamine synthesis and hence for histaminergic neurotransmission, and the striatum has a high density of histamine receptors which modulate both dopaminergic and serotonergic neurotransmission [41]. *HDC* was first implicated as a candidate GTS susceptibility gene by the identification of a nonsense truncating variant (Trp317Ter) segregating with the disorder in a Caucasian, nonconsanguineous two-generation family (eight siblings and their parents) with an extremely high prevalence of GTS, as only a single family member was unaffected [42]. This variant was not present in 3,000 Caucasian controls, in a replication cohort of 720 GTS individuals, nor in 360 controls not screened for psychiatric disorders. These results suggested the Trp317Ter variant is a very rare genetic GTS risk factor with high penetrance. This study implicated disrupted histamine production in GTS etiology and raised the histaminergic hypothesis. Since then, several studies have found an association between GTS and *HDC* variants in populations of European ancestry [43][44][45], and *HDC* knockout mice exhibited tic-like stereotypies [46]. A genome-wide analysis of *de novo* copy number variations (CNVs) in 460 Caucasian GTS individuals (including 148 trios) and 1131 ancestry-matched controls showed enrichment of genes within the histamine receptor (H1R and H2R) signaling pathways in GTS individuals, further supporting the histaminergic hypothesis [47]. However, two studies among Han Chinese GTS individuals, one with a case-control approach (120 cases and 240 controls) [48] and the other with a family-based approach (241 nuclear family trios) [49], did not detect an association between *HDC* and GTS. Taken together, this could indicate that a causal role for *HDC* in GTS etiology primarily applies to individuals of European ancestry, in which case it has the potential to be an important candidate gene with high penetrance in very rare cases, an uncommon feature in complex disorders such as GTS.

3.4. *CELSR3*, *WWC1*, *FN1*, and *NIPBL*

In 2017, exome sequencing of 511 GTS trios with various ancestries suggested that *de novo* damaging variants in approximately 400 genes contributed to the genetic risk load in 12% of the individuals [50]. In addition, four likely susceptibility genes with multiple *de novo* damaging variants were identified in unrelated GTS individuals: *CELSR3* (cadherin EGF LAG seven-pass G-type receptor 3), *WWC1* (WW and C2 domain containing 1), *FN1* (Fibronectin 1), and *NIPBL* (Nipped-B-like) [50]. In a follow-up study, additional variants of these four genes were identified [51]. All of these variants were either absent or present in very low frequency in the gnomAD database, which comprised about 250,000 individuals [52]. The two former genes were suggested as high confidence GTS risk genes and the two latter were suggested as probable GTS risk genes.

3.5. *ASH1L*

One of the most recent and quite notable additions to the list of potential GTS candidate genes is *ASH1L* (ASH1-like histone lysine methyltransferase), identified through the exome sequencing of 100 GTS trios [53]. *ASH1L* was suggested to be associated with GTS based on a transmission disequilibrium test (TDT) as well as a finding of *de novo* variants. Of the nineteen reported damaging *ASH1L* variants, five were present in very low frequency in the gnomAD database. The association was replicated through targeted sequencing of 524 GTS samples (and 2,822 East Asian ExAC controls). Some of the variants altered the methyltransferase activity of the protein, and *Ash1l*^{+/-} transgenic mice manifested compulsive and tic-like behaviors that could be rescued by a tic-relieving drug. The disruption of *Ash1l* also affected dopaminergic modulation in the dorsal striatum in the basal ganglia [53], altogether indicating a role of *ASH1L* in GTS physiopathology.

3.6. *FLT3*

Recently, in a GWAS with a cohort comprising 4,819 GTS individuals and 9,488 controls, Yu et al. reported a single genome-wide significant SNP (rs2504235) within *FLT3*, encoding FMS-like tyrosine kinase 3 [54]. rs2504235 is in strong linkage disequilibrium with the common *FLT3* missense variant rs1933437 (p.Thr227Met), which in the same study had a p-value of 8.2×10^{-8} [54]. This latter variant drove the association between GTS and a lymphocytic gene set in a recent large pathway analysis based on genome-wide genotypic data from 3,581 GTS individuals and 7,682 controls [55]. The association between lymphocytic genes, *FLT3*, and GTS not only provides further support for a role of *FLT3* in GTS, but also suggests an involvement of a neuroinflammatory element in disease pathogenesis.

3.7. *NRXN1* and *CNTN6*

Rare CNVs are known to be associated with a number of neuropsychiatric disorders [56], including GTS [47]. The investigation of rare CNVs in a large cohort (2,434 GTS individuals and 4,093 controls) of European ancestry showed an increased global burden of CNVs in the GTS sample [57]. Notably, *NRXN1* deletions and *CNTN6* duplications were present in 1% of the GTS individuals and survived genome-wide correction for multiple testing and were suggested as definitive GTS susceptibility loci. *CNTN6* encodes a cell adhesion molecule, contactin-6, and has been implicated in ASD and intellectual disability [58][59]. *NRXN1* encodes the presynaptic cell adhesion molecule neurexin 1 which is involved in glutamatergic and GABAergic neurotransmission and synaptogenesis [60]. *NRXN1* deletions have previously been associated with GTS, although the sample sizes have been considerably smaller (111 Caucasian GTS individuals with 73 ancestry-matched controls and 263 Latin American GTS individuals with 285 ancestry-matched controls, respectively) [61][62]. The two susceptibility loci are important additions to the list of candidate GTS susceptibility loci and demonstrate the importance of rare structural variations in GTS etiology.

4. Conclusions

The collaborative research conducted over the last few years and the investigation of large cohorts to identify the genetic susceptibility factors involved in GTS etiology are beginning to bear fruit. The employment of uniform methodology and clinically homogeneous cohorts taking comorbidities and ancestry into consideration are crucial in achieving more consistent and comparable results in future studies. Furthermore, functional analyses and cross-disorder studies might offer novel insights into the etiology and pathophysiology of GTS and overlapping neurodevelopmental, -psychiatric, and movement disorders. A better understanding of the complex etiology of GTS and new insights into the pathophysiology of GTS, which is far from being fully understood, are crucial in new treatment strategies for this disorder.

References

1. Knight, T.; Steeves, T.; Day, L.; Lowerison, M.; Jette, N.; Pringsheim, T. Prevalence of Tic Disorders: A Systematic Review and Meta-Analysis. *Pediatr. Neurol.* 2012, 47, 77–90.
2. Scharf, J.M.; Miller, L.L.; Bs, C.A.G.; Ma, J.A.; Mathews, C.A.; Ben-Shlomo, Y. Population prevalence of Tourette syndrome: A systematic review and meta-analysis. *Mov. Disord.* 2014, 30, 221–228.
3. American Psychiatric Association. *Diagnostic and Statistical Manual of Mental Disorders*, 5th ed.; American Psychiatric Association: Arlington, VA, USA, 2013.
4. Müller-Vahl, K.R.; Sambrani, T.; Jakubovski, E. Tic disorders revisited: Introduction of the term “tic spectrum disorders”. *Eur. Child Adolesc. Psychiatry* 2019, 28, 1129–1135.
5. Conte, G.; Valente, F.; Fioriello, F.; Cardona, F. Rage attacks in Tourette Syndrome and Chronic Tic Disorder: A systematic review. *Neurosci. Biobehav. Rev.* 2020, 119, 21–36.
6. Darrow, S.M.; Grados, M.; Sandor, P.; Hirschtritt, M.E.; Illmann, C.; Osiecki, L.; Dion, Y.; King, R.; Pauls, D.; Budman, C.L.; et al. Autism Spectrum Symptoms in a Tourette's Disorder Sample. *J. Am. Acad. Child Adolesc. Psychiatry* 2017, 56, 610–617.e1.
7. Huisman-van Dijk, H.M.; van de Schoot, R.; Rijkeboer, M.M.; Mathews, C.A.; Cath, D.C. The relationship between tics, OC, ADHD and autism symptoms: A cross-disorder symptom analysis in Gilles de la Tourette syndrome patients and family-members. *Psychiatry Res.* 2016, 237, 138–146.
8. Jiménez-Jiménez, F.J.; Alonso-Navarro, H.; García-Martín, E.; Agundez, J. Sleep disorders in tourette syndrome. *Sleep Med. Rev.* 2020, 53, 101335.

9. Robertson, M.M.; Eapen, V.; Singer, H.S.; Martino, D.; Scharf, J.M.; Paschou, P.; Roessner, V.; Woods, D.W.; Hariz, M.; Mathews, C.A.; et al. Gilles de la Tourette syndrome. *Nat. Rev. Dis. Prim.* 2017, 3, 16097.
10. Vermilion, J.; Pedraza, C.; Augustine, E.F.; Adams, H.R.; Vierhile, A.; Lewin, A.B.; Collins, A.T.; McDermott, M.P.; O'Connor, T.; Kurlan, R.; et al. Anxiety Symptoms Differ in Youth With and Without Tic Disorders. *Child Psychiatry Hum. Dev.* 2021, 52, 301–310.
11. Hoekstra, P.J.; Dietrich, A.; Edwards, M.J.; Elamin, I.; Martino, D. Environmental factors in Tourette syndrome. *Neurosci. Biobehav. Rev.* 2013, 37, 1040–1049.
12. Frick, L.; Pittenger, C. Microglial Dysregulation in OCD, Tourette Syndrome, and PANDAS. *J. Immunol. Res.* 2016, 2016, 1–8.
13. Paschou, P. The genetic basis of Gilles de la Tourette Syndrome. *Neurosci. Biobehav. Rev.* 2013, 37, 1026–1039.
14. Draganski, B.; Martino, D.; Cavanna, A.E.; Hutton, C.; Orth, M.; Robertson, M.M.; Critchley, H.D.; Frackowiak, R.S. Multispectral brain morphometry in Tourette syndrome persisting into adulthood. *Brain* 2010, 133, 3661–3675.
15. Peterson, B.S.; Thomas, P.; Kane, M.J.; Scahill, L.; Zhang, H.; Bronen, R.; King, R.A.; Leckman, J.F.; Staib, L. Basal Ganglia Volumes in Patients With Gilles de la Tourette Syndrome. *Arch. Gen. Psychiatry* 2003, 60, 415–424.
16. Wang, Z.; Maia, T.; Marsh, R.; Colibazzi, T.; Gerber, A.; Peterson, B.S. The Neural Circuits That Generate Tics in Tourette's Syndrome. *Am. J. Psychiatry* 2011, 168, 1326–1337.
17. Neuner, I.; Schneider, F.; Shah, N.J. Functional Neuroanatomy of Tics. In *International Review of Neurobiology*; Harris, R.A., Jenner, P., Eds.; Elsevier Inc.: Amsterdam, The Netherlands, 2013; pp. 35–71.
18. Cumming, P. The life history of dopamine. In *Imaging Dopamine*; Cambridge University Press: Cambridge, UK, 2009; pp. 5–18.
19. Beaulieu, J.-M.; Gainetdinov, R. The Physiology, Signaling, and Pharmacology of Dopamine Receptors. *Pharmacol. Rev.* 2011, 63, 182–217.
20. Sinopoli, V.M.; Burton, C.; Kronenberg, S.; Arnold, P.D. A review of the role of serotonin system genes in obsessive-compulsive disorder. *Neurosci. Biobehav. Rev.* 2017, 80, 372–381.
21. Olivier, B. Serotonin: A never-ending story. *Eur. J. Pharmacol.* 2015, 753, 2–18.
22. Bortolozzi, A.; Diaz-Mataix, L.; Scorza, M.C.; Celada, P.; Artigas, F. The activation of 5-HT_{2A} receptors in prefrontal cortex enhances dopaminergic activity. *J. Neurochem.* 2005, 95, 1597–1607.
23. De Deurwaerdère, P.; Navailles, S.; Berg, K.A.; Clarke, W.P.; Spampinato, U. Constitutive Activity of the Serotonin_{2C} Receptor Inhibits In Vivo Dopamine Release in the Rat Striatum and Nucleus Accumbens. *J. Neurosci.* 2004, 24, 3235–3241.
24. Esposito, E.; Di Matteo, V.; Di Giovanni, G. Serotonin–dopamine interaction: An overview. *Prog. Brain Res.* 2008, 172, 3–6.
25. Larsen, M.B.; Sonders, M.S.; Mortensen, O.V.; Larson, G.A.; Zahniser, N.R.; Amara, S.G. Dopamine Transport by the Serotonin Transporter: A Mechanistically Distinct Mode of Substrate Translocation. *J. Neurosci.* 2011, 31, 6605–6615.
26. Moya, P.; Wendland, J.R.; Rubenstein, L.M.; Timpano, K.; Heiman, G.; Tischfield, J.; King, R.A.; Andrews, A.; Ramamoorthy, S.; McMahon, F.; et al. Common and rare alleles of the serotonin transporter gene, SLC6A4, associated with Tourette's disorder. *Mov. Disord.* 2013, 28, 1263–1270.
27. Hildonen, M.; Levy, A.M.; Dahl, C.; Bjerregaard, V.A.; Møller, L.B.; Guldberg, P.; Debes, N.M.; Tümer, Z. Elevated Expression of SLC6A4 Encoding the Serotonin Transporter (SERT) in Gilles de la Tourette Syndrome. *Genes* 2021, 12, 86.
28. Dehning, S.; Müller, N.; Matz, J.; Bender, A.; Kerle, I.; Benninghoff, J.; Musil, R.; Spellmann, I.; Bondy, B.; Möller, H.-J.; et al. A genetic variant of HTR2C may play a role in the manifestation of Tourette syndrome. *Psychiatr. Genet.* 2010, 20, 35–38.
29. Cavallini, M.C.; Di Bella, D.; Catalano, M.; Bellodi, L. An association study between 5-HTTLPR polymorphism, COMT polymorphism, and Tourette's syndrome. *Psychiatry Res.* 2000, 97, 93–100.
30. Liu, S.; Zhang, X.; Yin, Y.; Wang, M.; Che, F.; Ma, X. An Association Analysis between 5-HTTLPR Polymorphism and Obsessive-Compulsive Disorder, Tourette Syndrome in a Chinese Han Population. *CNS Neurosci. Ther.* 2011, 17, 793–795.
31. Hu, X.-Z.; Lipsky, R.; Zhu, G.; Akhtar, L.A.; Taubman, J.; Greenberg, B.D.; Xu, K.; Arnold, P.D.; Richter, M.A.; Kennedy, J.L.; et al. Serotonin Transporter Promoter Gain-of-Function Genotypes Are Linked to Obsessive-Compulsive Disorder. *Am. J. Hum. Genet.* 2006, 78, 815–826.

32. Cao, X.; Zhang, Y.; Abdulkadir, M.; Deng, L.; Fernandez, T.V.; Julie, B.G.; Pieter, H.; Robert, J.H.; Justin, A.K.; Kuperman, S.; et al. Whole-exome sequencing identifies genes associated with Tourette's disorder in multiplex families. *Mol. Psychiatry* 2021, 1–15.
33. Abelson, J.F.; Kwan, K.Y.; O'Roak, B.J.; Baek, D.Y.; Stillman, A.A.; Morgan, T.M.; Mathews, C.A.; Pauls, D.L.; Rašin, M.-R.; Gunel, M.; et al. Sequence Variants in SLITRK1 Are Associated with Tourette's Syndrome. *Science* 2005, 310, 317–320.
34. Stillman, A.A.; Krsnik, Ž.; Sun, J.; Rašin, M.-R.; State, M.W.; Šestan, N.; Louvi, A. Developmentally regulated and evolutionarily conserved expression of SLITRK1 in brain circuits implicated in Tourette syndrome. *J. Comp. Neurol.* 2009, 513, 21–37.
35. Proenca, C.C.; Gao, K.P.; Shmelkov, S.V.; Rafii, S.; Lee, F.S. Slitrks as emerging candidate genes involved in neuropsychiatric disorders. *Trends Neurosci.* 2011, 34, 143–153.
36. Petek, E.; Windpassinger, C.; Vincent, J.B.; Cheung, J.; Boright, A.P.; Scherer, S.; Kroisel, P.M.; Wagner, K. Disruption of a Novel Gene (IMMP2L) by a Breakpoint in 7q31 Associated with Tourette Syndrome. *Am. J. Hum. Genet.* 2001, 68, 848–858.
37. Boghosian-Sell, L.; Comings, D.E.; Overhauser, J. Tourette syndrome in a pedigree with a 7;18 translocation: Identification of a YAC spanning the translocation breakpoint at 18q22.3. *Am. J. Hum. Genet.* 1996, 59, 999–1005.
38. Patel, C.J.; Cooper-Charles, L.; McMullan, D.J.; Walker, J.M.; Davison, V.; Morton, J.E. Translocation breakpoint at 7q31 associated with tics: Further evidence for IMMP2L as a candidate gene for Tourette syndrome. *Eur. J. Hum. Genet.* 2011, 19, 634–639.
39. Díaz-Anzaldúa, A.; Joobor, R.; Rivière, J.-B.; Dion, Y.; Lespérance, P.; Chouinard, S.; Richer, F.; Rouleau, G.A. Association between 7q31 markers and tourette syndrome. *Am. J. Med. Genet. Part A* 2004, 127A, 17–20.
40. Pagliaroli, L.; Vereczkei, A.; Padmanabhuni, S.S.; Tarnok, Z.; Farkas, L.; Nagy, P.; Rizzo, R.; Wolanczyk, T.; Szymanska, U.; Kapisyzi, M.; et al. Association of Genetic Variation in the 3'UTR of LHX6, IMMP2L, and AADAC With Tourette Syndrome. *Front. Neurol.* 2020, 11, 803.
41. Haas, H.L.; Sergeeva, O.A.; Selbach, O. Histamine in the Nervous System. *Physiol. Rev.* 2008, 88, 1183–1241.
42. Ercan-Sencicek, A.G.; Stillman, A.A.; Ghosh, A.K.; Bilguvar, K.; O'Roak, B.J.; Mason, C.E.; Abbott, T.; Gupta, A.; King, R.A.; Pauls, D.L.; et al. L-Histidine Decarboxylase and Tourette's Syndrome. *N. Engl. J. Med.* 2010, 362, 1901–1908.
43. Alexander, J.; Potamianou, H.; Xing, J.; Deng, L.; Karagiannidis, I.; Tsetsos, F.; Drineas, P.; Tarnok, Z.; Rizzo, R.; Wolanczyk, T.; et al. Targeted Re-Sequencing Approach of Candidate Genes Implicates Rare Potentially Functional Variants in Tourette Syndrome Etiology. *Front. Neurosci.* 2016, 10, 428.
44. Depienne, C.; Ciura, S.; Trouillard, O.; Bouteiller, D.; Leitão, E.; Nava, C.; Keren, B.; Marie, Y.; Guegan, J.; Forlani, S.; et al. Association of Rare Genetic Variants in Opioid Receptors with Tourette Syndrome. *Tremor Other Hyperkinet. Mov. (N. Y.)* 2019, 9.
45. Karagiannidis, I.; Dehning, S.; Sandor, P.; Tarnok, Z.; Rizzo, R.; Wolanczyk, T.; Madruga-Garrido, M.; Hebebrand, J.; Nöthen, M.; Lehmkuhl, G.; et al. Support of the histaminergic hypothesis in Tourette Syndrome: Association of the histamine decarboxylase gene in a large sample of families. *J. Med. Genet.* 2013, 50, 760–764.
46. Baldan, L.C.; Williams, K.A.; Gallezot, J.-D.; Pogorelov, V.; Rapanelli, M.; Crowley, M.; Anderson, G.M.; Loring, E.; Gorczyca, R.; Billingslea, E.; et al. Histidine Decarboxylase Deficiency Causes Tourette Syndrome: Parallel Findings in Humans and Mice. *Neuron* 2014, 81, 77–90.
47. Fernandez, T.V.; Sanders, S.; Yurkiewicz, I.R.; Ercan-Sencicek, A.G.; Kim, Y.-S.; Fishman, D.O.; Raubeson, M.J.; Song, Y.; Yasuno, K.; Ho, W.S.; et al. Rare Copy Number Variants in Tourette Syndrome Disrupt Genes in Histaminergic Pathways and Overlap with Autism. *Biol. Psychiatry* 2012, 71, 392–402.
48. Lei, J.; Deng, X.; Zhang, J.; Su, L.; Xu, H.; Liang, H.; Huang, X.; Song, Z.; Deng, H. Mutation screening of the HDC gene in Chinese Han patients with Tourette syndrome. *Am. J. Med. Genet. Part B Neuropsychiatr. Genet.* 2011, 159B, 72–76.
49. Dong, H.; Liu, W.; Liu, M.; Xu, L.; Li, Q.; Zhang, R.; Zhang, X.; Liu, S. Investigation of a Possible Role for the Histidine Decarboxylase Gene in Tourette Syndrome in the Chinese Han Population: A Family-Based Study. *PLoS ONE* 2016, 11, e0160265.
50. Willsey, A.J.; Fernandez, T.V.; Yu, D.; King, R.A.; Dietrich, A.; Xing, J.; Sanders, S.J.; Mandell, J.D.; Huang, A.Y.; Richer, P.; et al. De Novo Coding Variants Are Strongly Associated with Tourette Disorder. *Neuron* 2017, 94, 486–499.e9.
51. Wang, S.; Mandell, J.D.; Kumar, Y.; Sun, N.; Morris, M.T.; Arbelaez, J.; Nasello, C.; Dong, S.; Duhn, C.; Zhao, X.; et al. De Novo Sequence and Copy Number Variants Are Strongly Associated with Tourette Disorder and Implicate Cell

52. gnomAD. Genome Aggregation Database. Available online: <https://gnomad.broadinstitute.org/> (accessed on 19 February 2021).
53. Liu, S.; Tian, M.; He, F.; Li, J.; Xie, H.; Liu, W.; Zhang, Y.; Zhang, R.; Yi, M.; Che, F.; et al. Mutations in ASH1L confer susceptibility to Tourette syndrome. *Mol. Psychiatry* 2020, 25, 476–490.
54. Yu, D.; Sul, J.H.; Tsetsos, F.; Nawaz, M.S.; Huang, A.Y.; Zelaya, I.; Illmann, C.; Osiecki, L.; Darrow, S.M.; Hirschtritt, M.E.; et al. Interrogating the Genetic Determinants of Tourette's Syndrome and Other Tic Disorders Through Genome-Wide Association Studies. *Am. J. Psychiatry* 2019, 176, 217–227.
55. Tsetsos, F.; Yu, D.; Sul, J.H.; Huang, A.Y.; Illmann, C.; Osiecki, L.; Darrow, S.M.; Hirschtritt, M.E.; Greenberg, E.; Muller-Vahl, K.R.; et al. Synaptic processes and immune-related pathways implicated in Tourette syndrome. *Transl. Psychiatry* 2021, 11, 1–12.
56. Malhotra, D.; Sebat, J. CNVs: Harbingers of a Rare Variant Revolution in Psychiatric Genetics. *Cell* 2012, 148, 1223–1241.
57. Huang, A.Y.; Yu, D.; Davis, L.K.; Sul, J.H.; Tsetsos, F.; Ramensky, V.; Zelaya, I.; Ramos, E.M.; Osiecki, L.; Chen, J.A.; et al. Rare Copy Number Variants in NRXN1 and CNTN6 Increase Risk for Tourette Syndrome. *Neuron* 2017, 94, 1101–1111.e7.
58. Mercati, O.; Huguet, G.; Danckaert, A.; André-Leroux, G.; Maruani, A.; Bellinzoni, M.; Rolland, T.; Gouder, L.; Mathieu, A.; Buratti, J.; et al. CNTN6 mutations are risk factors for abnormal auditory sensory perception in autism spectrum disorders. *Mol. Psychiatry* 2017, 22, 625–633.
59. Kashevarova, A.A.; Nazarenko, L.P.; Schultz-Pedersen, S.; Skryabin, N.A.; Salyukova, O.A.; Chechetkina, N.N.; Tolmacheva, E.N.; Rudko, A.A.; Magini, P.; Graziano, C.; et al. Single gene microdeletions and microduplication of 3p26.3 in three unrelated families: CNTN6 as a new candidate gene for intellectual disability. *Mol. Cytogenet.* 2014, 7, 97.
60. Pak, C.; Danko, T.; Zhang, Y.; Aoto, J.; Anderson, G.; Maxeiner, S.; Yi, F.; Wernig, M.; Südhof, T.C. Human Neuropsychiatric Disease Modeling using Conditional Deletion Reveals Synaptic Transmission Defects Caused by Heterozygous Mutations in NRXN1. *Cell Stem Cell* 2015, 17, 316–328.
61. Sundaram, S.K.; Huq, A.M.; Wilson, B.J.; Chugani, H.T. Tourette syndrome is associated with recurrent exonic copy number variants. *Neurology* 2010, 74, 1583–1590.
62. Nag, A.; Bochukova, E.; Kremeyer, B.; Campbell, D.; Muller, H.; Valencia-Duarte, A.V.; Cardona, J.; Rivas, I.C.; Mesa, S.C.; Cuartas, M.; et al. CNV Analysis in Tourette Syndrome Implicates Large Genomic Rearrangements in COL8A1 and NRXN1. *PLoS ONE* 2013, 8, e59061.