

PKCtheta in Cancer

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Protein Kinase C theta (PKC θ) is a serine/threonine kinase that belongs to the novel PKC sub-family. PKC θ has been extensively studied for its role in the immune system where it plays a critical role in T cell activation. Beyond its physiological role in immune responses, increasing evidence implicates PKC θ in the pathology of various diseases, especially autoimmune disorders and cancers. Particularly, in various types of cancers, the high PKC θ expression leads to aberrant cell proliferation, migration and invasion, thereby promoting cancer aggressiveness. The recent development and application of PKC θ inhibitors in the context of auto-immune diseases could benefit the emergence of treatment for cancers in which PKC θ has been implicated.

PKCtheta

cancer

tumoral function

mechanisms of action

1. Introduction

The Protein Kinase C (PKC) family is a family of serine/threonine kinases that are involved in various cellular processes for different cell types. The PKC family is classified into three subfamilies: classical (α , β I, β II, γ), novel (δ , ϵ , η , θ) and atypical (ζ , ι/λ) PKC isoforms. This classification is based on their structure and ability to respond to calcium and/or diacylglycerol (DAG) [1]. Among this family, the novel PKC θ isoform is different from other PKC isoforms since its physiological expression is limited to a few types of cells, such as T cells, platelets and skeletal muscle cells. This specific expression confers to this isoform a central role in the immune system where PKC θ controls T cell activation, survival and differentiation [2]. In skeletal muscle cells, PKC θ regulates muscle cell development, homeostasis and remodeling [3]. Beyond its physiological functions, PKC θ is also involved in the pathology of various diseases. In the context of the immune system and skeletal muscle tissue, the dysregulation of PKC θ activity leads to both autoimmune and inflammatory diseases and to insulin resistance and Type 2 diabetes, respectively [3][4]. In the last decade, growing evidence implicated the PKC θ signaling in the biology of cancer where it controls cancer cell proliferation, migration and invasion at the cytoplasmic or nuclear levels. Here, we discuss this emerging function of PKC θ in cancer by analyzing its diverse modes of action and their consequence on critical biological processes involved in tumorigenesis and cancer progression.

2. PKC θ Structure and Physiological Function

In this section, we provide a brief overview of the PKC θ structure and the PKC θ physiological function mainly in the immune system. For extensive details, the readers can refer to several excellent reviews written by the experts in the field of T cell biology (reviewed in [2][4][5][6][7]).

2.1. PKCθ Structure

The novel PKCθ isoform is a protein kinase encoded by the *PRKCQ* gene and composed of 706 amino acids with a molecular weight of approximately 82 kDa [8]. PKCθ is a DAG-dependent but Ca^{2+} -independent, protein kinase whose structure consists of several functional domains that are conserved among the novel PKC subfamily (Figure 1) [1]. The N-terminal regulatory domain contains the C2-like domain, the pseudosubstrate region and the DAG-binding domain (C1A/B) while the C-terminal catalytic domain contains the ATP-binding domain (C3) and the substrate-binding domain (C4). The regulatory and catalytic domains are separated by a hinge region, called the V3 motif, which is unique and highly specific to each PKC isoforms.

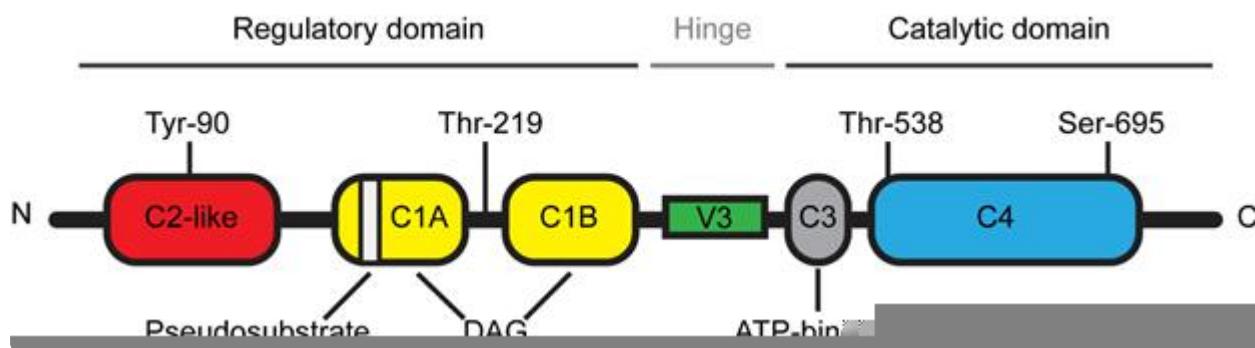


Figure 1. Schematic representation of Protein Kinase C theta (PKCθ) structure.

2.2. PKCθ Function in the Immune System

Due to the high expression levels of PKCθ in T lymphocytes, extensive research has studied the biological function of this novel PKC isoform in the immune system. The generation and analysis of PKCθ-deficient mice have unraveled the selective role of PKCθ in the T cell immune response [9][10]. While PKCθ is critical for the T helper (Th)2- and Th17-mediated responses, the Th1- and cytotoxic T cell-driven responses remain relatively intact in the absence of PKCθ [4][7]. However, a few studies reported that some specific Th1 responses were altered in PKCθ deficient mice [11][12]. T lymphocyte activation is a central step of the T cell immune response during which T cell interacts with an antigen-presenting cell (APC) [4]. This cell-cell junction forms a well-organized and dynamic structure called the immunological synapse [13]. Following this T cell-APC interaction, cytoplasmic PKCθ is translocated to the membrane at the immunological synapse [6] and this specific and critical relocalization is highly dependent on the unique V3 motif of PKCθ [14]. In addition, other events are also required for the proper localization and activation of PKCθ at the immunological synapse. Concerning the PKCθ localization, several studies indicated that the Ick-mediated phosphorylation of PKCθ at tyr-90 participated in the PKCθ recruitment to the immunological synapse [14][15] and a report from Thuille et al. suggested that the PKCθ autophosphorylation at thr-219 was required for the cell membrane localization of PKCθ [16]. Moreover, the data from Cartwright et al. suggested that PKCθ required its active kinase domain in order to be maintained at the immunological synapse [17]. More recently, Wang et al. reported that the sumoylation of PKCθ upon T cell activation was involved in the specific localization of PKCθ and in the organization of the immunological synapse [18]. Concerning the PKCθ activation, the phosphorylation at Thr-538 in the activation loop regulates the PKCθ activity by maintaining PKCθ in an active

conformation and thus this phosphorylation has been used as a marker reflecting the PKC θ activation [19]. GCK-like kinase (GLK, MAP4K3) has been identified as one kinase capable of directly phosphorylating this Thr-538 residue during the T cell activation [20]. Moreover, the auto-phosphorylation at Ser-695 induced during T cell activation is also required for the PKC θ kinase activity [19][21].

Once translocated to the immunological synapse, PKC θ integrates various signaling cascades that conduct to the activation of important transcription factors, including Nuclear Factor κ B (NF- κ B), Activating Protein 1 (AP-1) and, to a lesser extent, Nuclear Factor of Activated T-cells (NFAT) [5]. This transcriptional machinery then induces the production of interleukin-2, a cytokine essential for the T cell proliferation [5]. Moreover, the PKC θ function is not only limited to the activation of signaling pathways that leads to the transcriptional regulation of gene expression. For example, PKC θ has been involved in the actin cytoskeletal reorganization that occurs during the formation of the immunological synapse and the related polarization of activated T cells [18][22][23]. PKC θ can also enter the nucleus of activated T cells to directly bind to the chromatin in order to regulate the expression of immune response genes and microRNAs involved in the cytokine regulation [24].

2.3. Implication of PKC θ in Immunological Disorders

As a selective regulator of the Th2 and Th17 immune responses, the perturbation of PKC θ expression and activity leads to the development of Th2-driven inflammatory diseases and Th17-mediated autoimmune diseases. Indeed, PKC θ is highly expressed and activated in these immunological disorders [4]. Studies from the PKC θ -deficient mice showed that the PKC θ suppression decreased the T cell inflammatory response in autoimmunity, allergy and allograft rejection [4]. Therefore, the therapeutic use of specific PKC θ inhibitors could provide an interesting approach for these PKC θ -dependent pathologies [25]. Clinical studies using sotрастaurин (AEB071) as the PKC θ inhibitor showed some encouraging results in the context of immunosuppressive therapy for autoimmune diseases such as psoriasis and organ transplantation [4][26]. However, sotрастaurин is not specific to PKC θ and it also shows strong and specific inhibitory activity against PKC α and PKC β and to a lesser extend against PKC δ , PKC ϵ and PKC η . It thus suggests that sotрастaurин would inhibit not only the PKC θ -mediated functions but also the functions from other PKCs [27]. Therefore, current research works aim to develop more selective PKC θ inhibitors [28][29]. These inhibitors are currently tested in mouse models and further studies are needed to validate them in the clinical trials.

References

1. Steinberg, S.F. Structural basis of protein kinase C isoform function. *Physiol. Rev.* 2008, 88, 1341–1378.
2. Hayashi; K.; Altman, A. Protein kinase C theta (PKCtheta): A key player in T cell life and death. *Pharmacol. Res.* 2007, 55, 537–544.

3. Marrocco, V.; Fiore, P.; Madaro, L.; Crupi, A.; Lozanoska-Ochser, B.; Bouché, M. Targeting PKCtheta in skeletal muscle and muscle diseases: Good or bad? *Biochem. Soc. Trans.* 2014, 42, 1550–1555.
4. Zhang; Y.E.; Kong, K.F.; Altman, A. The yin and yang of protein kinase C-theta (PKCtheta): A novel drug target for selective immunosuppression. *Adv. Pharmacol.* 2013, 66, 267–312.
5. Isakov; N.; Altman, A. Protein kinase C(theta) in T cell activation. *Annu. Rev. Immunol.* 2002, 20, 761–794.
6. Kong; F.K.; Altman, A. In and out of the bull's eye: Protein kinase Cs in the immunological synapse. *Trends Immunol.* 2013, 34, 234–342.
7. Marsland, J.B.; Kopf, M. T-cell fate and function: PKC-theta and beyond. *Trends Immunol.* 2008, 29, 179–185.
8. Baier, G.; Telford, D.; Giampa, L.; Coggeshall, K.M.; Bitterlich, G.B.; Isakov, N.; Altman, A. Molecular cloning and characterization of PKC theta, a novel member of the protein kinase C (PKC) gene family expressed predominantly in hematopoietic cells. *J. Biol. Chem.* 1993, 268, 4997–5004.
9. Pfeifhofer, C.; Kofler, K.; Gruber, T.; Tabrizi, N.G.; Lutz, C.; Maly, K.; Leitges, M.; Baier, G. Protein kinase C theta affects Ca²⁺ mobilization and NFAT cell activation in primary mouse T cells. *J. Exp. Med.* 2003, 197, 1525–1535.
10. Sun, Z.; Arendt, C.W.; Ellmeier, W.; Schaeffer, E.M.; Sunshine, M.J.; Gandhi, L.; Annes, J.; Petrzilka, D.; Kupfer, A.; Schwartzberg, P.L.; et al., PKC-theta is required for TCR-induced NF-kappaB activation in mature but not immature T lymphocytes. *Nature* 2000, 404, 402–407.
11. Nishanth, G.; Burkiewicz, M.S.; Händel, U.; Kliche, S.; Wang, X.; Naumann, M.; Deckert, M.; Schlüter, D. Protective Toxoplasma gondii-specific T-cell responses require T-cell-specific expression of protein kinase C-theta. *Infect. Immun.* 2010, 78, 3454–3464.
12. Ohayon, A.; Golenser, J.; Sinay, R.; Tamir, A.; Altman, A.; Pollack, Y.; Isakov, N. Protein kinase C theta deficiency increases resistance of C57BL/6J mice to Plasmodium berghei infection-induced cerebral malaria. *Infect. Immun.* 2010, 78, 4195–4205.
13. Grakoui, A.; Bromley, S.K.; Sumen, C.; Davis, M.M.; Shaw, A.S.; Allen, P.M.; Dustin, M.L. The immunological synapse: A molecular machine controlling T cell activation. *Science* 1999, 285, 221–227.
14. Kong, K.O.; Yokosuka, T.; Balancio, A.J.C.; Isakov, N.; Saito, T.; Altman, A. A motif in the V3 domain of the kinase PKC-theta determines its localization in the immunological synapse and functions in T cells via association with CD28. *Nat. Immunol.* 2011, 12, 1105–1112.

15. Liu, Y.; Witte, S.; Liu, Y.C.; Doyle, M.; Elly, C.; Altman, A. Regulation of protein kinase C θ function during T cell activation by Lck-mediated tyrosine phosphorylation. *J. Biol. Chem.* 2000, 275, 3603–3609.

16. Thuille, N.; Heit, I.; Fresser, F.; Krumböck, N.; Bauer, B.; Leuthaeusser, S.; Dammeier, S.; Graham, C.; Copeland, T.D.; Shaw, S.; et al., Critical role of novel Thr-219 autophosphorylation for the cellular function of PKC θ in T lymphocytes. *EMBO J.* 2005, 24, 3869–3880.

17. Cartwright, G.N.; Kashyap, A.K.; Schaefer, B.C. An active kinase domain is required for retention of PKC θ at the T cell immunological synapse. *Mol. Biol. Cell* 2011, 22, 3491–3497.

18. Wang, X.D.; Gong, Y.; Chen, Z.L.; Gong, B.N.; Xie, J.J.; Zhong, C.Q.; Wang, Q.L.; Diao, L.H.; Xu, A.; Han, J.; et al., TCR-induced sumoylation of the kinase PKC-theta controls T cell synapse organization and T cell activation. *Nat. Immunol.* 2015, 16, 1195–1203.

19. Liu, Y.; Graham, C.; Li, A.; Fisher, R.J.; Shaw, S. Phosphorylation of the protein kinase C-theta activation loop and hydrophobic motif regulates its kinase activity, but only activation loop phosphorylation is critical to in vivo nuclear-factor-kappaB induction. *Biochem. J.* 2002, 361, 255–265.

20. Chuang, H.C.; Lan, J.L.; Chen, d.; Yang, C.Y.; Chen, Y.M.; Li, J.P.; Huang, C.Y.; Liu, P.E.; Wang, X.; Tan, T.H. The kinase GLK controls autoimmunity and NF-kappaB signaling by activating the kinase PKC-theta in T cells. *Nat. Immunol.* 2011, 12, 1113–1118.

21. Czerwinski, R.; Aulabaugh, A.; Greco, R.M.; Olland, S.; Malakian, K.; Wolfrom, S.; Lin, L.; Kriz, R.; Stahl, M.; Huang, Y.; et al. Characterization of protein kinase C theta activation loop autophosphorylation and the kinase domain catalytic mechanism. *Biochemistry* 2005, 44, 9563–9573.

22. Britton, G.J.; Ambler, R.; Clark, D.J.; Hill, E.V.; Tunbridge, H.M.; McNally, K.E.; Burton, B.R.; Butterweck, P.; Peyton, C.S.; O’Neil, L.A.H.; et al., PKC θ links proximal T cell and Notch signaling through localized regulation of the actin cytoskeleton. *Elife* 2017, 6, e20003.

23. Quann, E.J.; Liu, X.; Bonnet, G.A.; Huse, M. A cascade of protein kinase C isozymes promotes cytoskeletal polarization in T cells. *Nat. Immunol.* 2011, 12, 647–654.

24. Sutcliffe, E.L.; Bunting, K.L.; He, Y.Q.; Li, J.; Phetsouphanh, C.; Seddiki, N.; Zafar, A.; Hindmarsh, E.J.; Parish, C.R.; Kelleher, A.D.; et al., Chromatin-associated protein kinase C-theta regulates an inducible gene expression program and microRNAs in human T lymphocytes. *Mol. Cell* 2011, 41, 704–719.

25. Kwon, M.Y.; Wang, R.; Ma, J.; Sun, Z. PKC-theta is a drug target for prevention of T cell-mediated autoimmunity and allograft rejection. *Endocr. Metab. Immune Disord. Drug Targets* 2010, 10, 367–372.

26. Sleiman, R.H.; Hamze, A.B.; Reslan, L.; Kobeissy, H.; Dbaibo, G. The Novel PKCtheta from Benchtop to Clinic. *J. Immunol. Res.* 2015, 2015, 348798.
27. Evenou, J.P.; Wagner, J.; Zenke, G.; Brinkmann, V.; Wagner, K.; Kovarik, J.; Welzenbach, K.A.; Schmidt, G.W.; Guntermann, C.; Towbin, H.; et al., The potent protein kinase C-selective inhibitor AEB071 (sotrasaurin) represents a new class of immunosuppressive agents affecting early T-cell activation. *J. Pharmacol. Exp. Ther.* 2009, 330, 792–801.
28. Kunikawa, S.; Tanaka, A.; Takasuna, Y.; Tasaki, M.; Chida, N. Discovery of 2,4-diamino-5-cyanopyrimidine derivatives as protein kinase C theta inhibitors with mitigated time-dependent drug-drug interactions. *Bioorg. Med. Chem.* 2019, 27, 790–799.
29. Wang, J.; Jin, W.; Zhou, X.; Li, J.; Xu, C.; Ma, Z.; Wang, J.; Qin, L.; Zhou, B.; Ding, W.; et al., Identification, Structure-Activity Relationships of Marine-Derived Indolocarbazoles, and a Dual PKCtheta/delta Inhibitor with Potent Antipancreatic Cancer Efficacy. *J. Med. Chem.* 2020, 63, 12978–12991.

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