PPAR α and Ocular Dieseases

Subjects: Others Contributor: Toshihide Kurihara, Yohei Tomita, Deokho Lee

Mounting evidence suggests that peroxisome proliferator-activator receptor alpha (PPAR α) activation can be a therapeutic target for various ocular diseases including diabetic retinopathy (DR). Here we describe functions of PPAR α in the eye contributing to ocular homeostasis.

Keywords: diabetic retinopathy ; PPARa ; SPPARMa ; HIF-1a

1. Introduction

Diabetic retinopathy (DR) is a complication of diabetes that affects the eyes in subjects with type 1 or type 2 diabetes mellitus ^[1]. DR develops because of a chronic abnormality of glycemic control ^[2]. In detail, DR progresses from an initial stage where high blood glucose levels damage the microvasculature ^[3]. Then, microvascular irregularities including hemorrhage, ischemia, and microaneurysms bring retinal neovascularization ^[3]. Abnormal vasculatures by retinal neovascularization lead to severe hypoxic conditions in the retina of the eye ^[3]. At the final stage, fibrovascular proliferation resulting in tractional retinal detachment by chronic severe hypoxic conditions causes vision loss ^[3]. However, recent evidence indicates that pathogenesis of DR contains far more complex mechanisms that are the involvement of multiple interlinked alterations via impairment of crosstalk between retinal neurons, glial cells, and vasculatures ^{[4][5][6][7]}. Emerging studies have demonstrated that neurons and glial cells in the central nervous system act as oxygen sensors and vascular homeostasis ^{[8][9]}. Although exact mechanisms have not been completely defined, possible major factors such as mitochondrial oxidative stress, inflammation, production of advanced glycation end products (AGEs), and activation of the protein kinase C (PKC) pathway have been proposed (along with the traditional concept of retinal neovascularization by hyperglycemia-induced microvascular irregularities) for the comprehensive pathogenesis of DR ^[10].

PPARα is from a nuclear receptor PPAR family (PPARα, PPARδ, and PPARγ), which regulates expressions of several genes affecting lipid and carbohydrate metabolism ^[12]. PPARα is named based on its ability to be activated by peroxisome proliferator chemicals, and it is the first member to be cloned among the PPAR isotypes ^[13]. PPARα is expressed in various types of cells in the skeletal muscle, heart, liver, brown adipose, kidney, intestinal mucosa, adrenal gland, eye, and most cell types present in the vasculature including endothelial cells, smooth muscle cells, monocytes, and macrophages ^[14][15][16][17][18][19][20][21]</sup>. PPARα activation was found to increase circulating levels of high-density lipoprotein cholesterol and decrease serum levels of triglycerides, free fatty acids and apolipoprotein, which improves an overall serum lipid profile and finally exerts positive effects on inflammation and insulin resistance ^{[22][23][24]}. Increasing evidence suggests that PPARα activation can be a strong therapeutic target for various types of diseases such as cardiovascular diseases ^[25], dyslipidemia ^[26], and diabetes and its complications including DR ^[22]. However, its molecular mechanisms are far from being elucidated. In this paper, we review the therapeutic effects of PPARα agonists as a promising approach for the treatment of DR.

2. Functions of PPARα in the Eye

Experimental evidence indicates that PPAR α is expressed in various tissues in diabetic microvascular diseases—the retina as well as kidney and nerve ^{[20][21]}. PPAR α has been spotlighted as its expression levels decreased in the retinas with diabetes ^{[27][28]}. Decreased PPAR α expression has been found to contribute to retinal inflammation and neovascularization, and pharmacological activation of PPAR α has been shown to exert therapeutic effects against various ocular degenerative disorders ^{[28][29][30]}. A previous study showed that more severe retinal acellular capillary formation and pericyte dropout were observed in PPAR $\alpha^{-/-}$ mice with diabetes, compared with those in diabetic wild-type mice ^[29]. Another study demonstrated that retinal neurodegeneration was exacerbated in PPAR $\alpha^{-/-}$ mice with diabetes in comparison with that in diabetic wild-type mice ^[31]. Multiple proteomics data indicated that several oxidative stress

markers such as *Gstm1* (glutathione-s-transferase m1), *Prdx6* (peroxidase 6) and *Txnrd1* (thioredoxin reductase 1) increased in diabetic retinas and that there were further increases in diabetic PPAR $\alpha^{-/-}$ retinas ^[31]. This implies that oxidative stress may become worsened by PPAR α ablation in DR. PPAR α activation increased retinal NADH (nicotinamide adenine dinucleotide + hydrogen) oxidation in diabetic mice, and treatment of fenofibric acid, an active metabolite of fenofibrate, reduced mitochondrial oxidative stress and cell death in retinal neuronal cell lines under 4-hydroxynonenal (4-HNE)-induced oxidative stress conditions ^[31]. This implies mitochondrial dysfunction by oxidative stress could be restored by PPAR α activation.

In terms of an ischemic model other than the diabetic model, PPAR $\alpha^{-/-}$ mice with a laser-induced choroidal neovascularization (CNV) developed more severe CNV compared with wild-type CNV mice ^[32]. PPAR $\alpha^{-/-}$ mice with oxygen-induced retinopathy (OIR) also showed deleterious effects (such as increased retinal cell death and glial activation) in comparison with wild-type OIR mice ^[33]. Overexpression of PPAR α using an adenovirus system attenuated increased endothelial progenitor cell circulation in OIR mice through inhibition of the hypoxia-inducible factor (HIF)-1 α pathway, and mouse brain endothelial cells from PPAR $\alpha^{-/-}$ mice showed prominent activation of HIF-1 α induced by hypoxia, compared with wild-type mouse brain endothelial cells ^[34]. This suggests anti-angiogenic effects of PPAR α via HIF-1 α inhibition, a novel protective molecular mechanism against retinal ischemic conditions.

Pharmacological PPAR α activation by palmitoylethanolamide (PEA) reduced retinal neovascularization and fibrotic changes and suppressed glial activation in proliferative retinopathy and neovascular age-related macular degeneration mouse models ^[35]. In cardiovascular studies, PEA exerted direct vaso-relaxation of the bovine ophthalmic artery by using PPAR α transcription factors, which suggests a role of PPAR α on physiological vascular regulation ^[36]. This vaso-relaxing effect could increase supply of oxygen to the retina and prevent ischemic lesions, as observed in patients with ocular hypertension ^[37]. Another study demonstrated that administration of PEA showed enhancement of aqueous humor outflow facility and this effect appeared to be mediated partially by the involvement of PPAR α ^[38]. Those studies imply PPAR α modulation could also be a promising therapeutic target for glaucoma ^[39].

Even though there are not many reports available on roles of PPAR α in the ocular surface, recent evidence suggests that PPAR α may play a critical role in regulation of inflammatory processes in the ocular surface ^[40]. Fenofibrate ameliorated a severity of ocular surface squamous metaplasia, commonly seen in patients with long-term deficiency of tear film ^[41] and suppressed formation of tear film instability via inhibition of macrophages and downregulation of pro-inflammatory factors ^[40]. In the corneal epithelium of mice with dry eyes by sleep deprivation, downregulation in PPAR α expression was detected ^[42], and fenofibrate increased PPAR α expression in cultured corneal epithelium sheets and restored a microvilli morphology ^[42]. This implies PPAR α activation could have potential for use as a preventive agent in patients with a high risk of the dry eye. Other cornea studies indicated therapeutic roles of PPAR α agonists against corneal inflammation and neovascularization ^{[43][44]}. Corneal neovascularization is closely related to a reduction in corneal transparency which is important for a visual acuity ^{[45][46]}. In rat corneal alkali burn models, corneal neovascularization was seen and a topical injection of fenofibrate suppressed its neovascularization through upregulation of PPAR α mRNA expression and suppression of *II-6*, *II-1β*, *Vegf* and *Ang-2* mRNA expressions ^{[43][44]}.

The above, taken together as accumulating evidence, supports the concept that it may be important to restore or boost PPAR α expression for prevention of various ocular diseases. However, studies on downstream signaling effector molecules regarding PPAR α activation need to be further unraveled.

3. Selective PPARα modulatorα (SPPARMα)

Pemafibrate is a novel selective PPAR α modulator (SPPARM α) and has higher potency and selectivity for the activation of PPAR α than other PPAR α agonists, especially fenofibrate ^{[43][47][48][49]}. Several studies have shown that pemafibrate has therapeutic effects on retinal diseases ^{[50][51][52]}. One report showed that oral administration of pemafibrate increased plasma fibroblast growth factor 21 (FGF21) levels in OIR and inhibited retinal neovascularization through inhibition of HIF-1 α and *Vegf* expressions ^[51]. In addition, long-acting FGF21 inhibited HIF activity in a photoreceptor cell line under hypoxic conditions ^[51]. Another report demonstrated oral administration of pemafibrate increased serum FGF21 levels in a streptozotocin-induced diabetic mouse model and preserved retinal function through maintaining synaptophysin expression, which regulates synaptic vesicle endocytosis ^[50]. Additionally, long-acting FGF21 directly upregulated synaptophysin expression in differentiated neurons in vitro ^[50]. The other study reported that pemafibrate directly inhibited diabetes-induced vascular leukostasis and leakage in the rat retina through upregulation of *THBD* expression in human umbilical vein endothelial cells and human retinal microvascular endothelial cells in vitro ^[52]. Even though further studies are required to elucidate various mechanisms, pemafibrate could offer retinal protection through upregulation of blood FGF21 levels to work on the damaged retina with suppression of pathological neovascularization, maintenance of

retinal function, modulation of systemic metabolisms such as triglyceride or blood glucose levels, or a direct increase in thrombomodulin expression in retinal endothelial cells to work on the damaged retina with suppression of vascular leakage, leukostasis and inflammation.

4. Conclusions

Diabetes mellitus is a complex metabolic disorder which is associated with insulin resistance, insulin signaling impairment, β -cell dysfunction, abnormal glucose and lipid metabolisms, inflammation and mitochondrial oxidative stress ^[53]. Likewise, development of DR has multiple interlinked alterations via dysfunctions of various cell types with enormous complex pathological mechanisms. Its development in turn cannot be prevented or protected by one therapeutic molecular target. Fortunately, PPAR α targeting could be an alternative therapy to other present therapeutic agents in that it covers systemic enhancement of glucose and lipid metabolisms, anti-inflammation and anti-oxidative stress. Moreover, PPAR α activation could ameliorate development of DR at an early stage through prevention of retinal vascular leakage^{[54][50][51]}. This summary enables comprehensive understandings of protective roles of PPAR α agonists against DR development. It can be useful for future studies on the protective effects of PPAR α agonists against DR.

References

- 1. Duh, E.J.; Sun, J.K.; Stitt, A.W. Diabetic retinopathy: Current understanding, mechanisms, and treatment strategies. JC I Insight 2017, 2, e93751.
- 2. Chatziralli, I.P. The Role of Glycemic Control and Variability in Diabetic Retinopathy. Diabetes 2018, 9, 431–434.
- 3. Wang, W.; Lo, A.C.Y. Diabetic Retinopathy: Pathophysiology and Treatments. Int. J. Mol. Sci. 2018, 19, 1816.
- 4. Frank, R.N. Diabetic retinopathy. N. Engl. J. Med. 2004, 350, 48–58.
- 5. Curtis, T.M.; Gardiner, T.A.; Stitt, A.W. Microvascular lesions of diabetic retinopathy: Clues towards understanding path ogenesis? Eye 2009, 23, 1496–1508.
- Barber, A.J.; Gardner, T.W.; Abcouwer, S.F. The significance of vascular and neural apoptosis to the pathology of diabet ic retinopathy. Invest. Ophthalmol. Vis. Sci. 2011, 52, 1156–1163.
- Antonetti, D.A.; Barber, A.J.; Bronson, S.K.; Freeman, W.M.; Gardner, T.W.; Jefferson, L.S.; Kester, M.; Kimball, S.R.; K rady, J.K.; LaNoue, K.F.; et al. Diabetic retinopathy: Seeing beyond glucose-induced microvascular disease. Diabetes 2 006, 55, 2401–2411.
- 8. Hawkins, B.T.; Davis, T.P. The Blood-Brain Barrier/Neurovascular Unit in Health and Disease. Pharmacol. Rev. 2005, 5 7, 173.
- 9. Yu, X.; Ji, C.; Shao, A. Neurovascular Unit Dysfunction and Neurodegenerative Disorders. Front. Neurosci. 2020, 14, 3 34.
- 10. Lechner, J.; O'Leary, O.E.; Stitt, A.W. The pathology associated with diabetic retinopathy. Vis. Res. 2017, 139, 7–14.
- 11. Robinson, R.; Barathi, V.A.; Chaurasia, S.S.; Wong, T.Y.; Kern, T.S. Update on animal models of diabetic retinopathy: F rom molecular approaches to mice and higher mammals. Dis. Models Mech. 2012, 5, 444–456.
- Tajoaek, P.; Petrovj, D.; Petrovi, M.G.N.; Kunej, T.J.P.R. Association of Peroxisome Proliferator-Activated Receptors (P PARs) with Diabetic Retinopathy in Human and Animal Models: Analysis of the Literature and Genome Browsers. PPA R Res. 2020, 2020, 1783564.
- 13. Tyagi, S.; Gupta, P.; Saini, A.S.; Kaushal, C.; Sharma, S. The peroxisome proliferator-activated receptor: A family of nu clear receptors role in various diseases. J. Adv. Pharm. Technol. Res. 2011, 2, 236–240.
- 14. Yu, X.H.; Zheng, X.L.; Tang, C.K. Peroxisome Proliferator-Activated Receptor α in Lipid Metabolism and Atherosclerosi s. Adv. Clin. Chem. 2015, 71, 171–203.
- Braissant, O.; Foufelle, F.; Scotto, C.; Dauça, M.; Wahli, W. Differential expression of peroxisome proliferator-activated r eceptors (PPARs): Tissue distribution of PPAR-alpha, -beta, and -gamma in the adult rat. Endocrinology 1996, 137, 35 4–366.
- Abbott, B.D. Review of the expression of peroxisome proliferator-activated receptors alpha (PPARα), beta (PPARβ), an d gamma (PPARγ) in rodent and human development. Reprod. Toxicol. 2009, 27, 246–257.
- 17. Auboeuf, D.; Rieusset, J.; Fajas, L.; Vallier, P.; Frering, V.; Riou, J.P.; Staels, B.; Auwerx, J.; Laville, M.; Vidal, H. Tissue distribution and quantification of the expression of mRNAs of peroxisome proliferator-activated receptors and liver X rec eptor-alpha in humans: No alteration in adipose tissue of obese and NIDDM patients. Diabetes 1997, 46, 1319–1327.

- Kliewer, S.A.; Forman, B.M.; Blumberg, B.; Ong, E.S.; Borgmeyer, U.; Mangelsdorf, D.J.; Umesono, K.; Evans, R.M. Dif ferential expression and activation of a family of murine peroxisome proliferator-activated receptors. Proc. Natl. Acad. S ci. USA 1994, 91, 7355–7359.
- 19. Touyz, R.M.; Schiffrin, E.L. Peroxisome proliferator-activated receptors in vascular biology-molecular mechanisms and clinical implications. Vasc. Pharmacol. 2006, 45, 19–28.
- 20. Lefebvre, P.; Chinetti, G.; Fruchart, J.C.; Staels, B. Sorting out the roles of PPAR alpha in energy metabolism and vasc ular homeostasis. J. Clin. Investig. 2006, 116, 571–580.
- 21. Desouza, C.V.; Rentschler, L.; Fonseca, V. Peroxisome proliferator-activated receptors as stimulants of angiogenesis in cardiovascular disease and diabetes. Diabetes Metab. Syndr. Obes. 2009, 2, 165–172.
- 22. Krauss, R.M. Lipids and Lipoproteins in Patients with Type 2 Diabetes. Diabetes Care 2004, 27, 1496.
- 23. Katsiki, N.; Nikolic, D.; Montalto, G.; Banach, M.; Mikhailidis, D.P.; Rizzo, M. The role of fibrate treatment in dyslipidemi a: An overview. Curr. Pharm. Des. 2013, 19, 3124–3131.
- 24. Staels, B.; Dallongeville, J.; Auwerx, J.; Schoonjans, K.; Leitersdorf, E.; Fruchart, J.-C. Mechanism of Action of Fibrates on Lipid and Lipoprotein Metabolism. Circulation 1998, 98, 2088–2093.
- 25. Han, L.; Shen, W.J.; Bittner, S.; Kraemer, F.B.; Azhar, S. PPARs: Regulators of metabolism and as therapeutic targets i n cardiovascular disease. Part I: PPAR-α. Future Cardiol. 2017, 13, 259–278.
- 26. Tenenbaum, A.; Fisman, E.Z. Fibrates are an essential part of modern anti-dyslipidemic arsenal: Spotlight on atherogen ic dyslipidemia and residual risk reduction. Cardiovasc. Diabetol. 2012, 11, 125.
- 27. Wang, F.; Gao, L.; Gong, B.; Hu, J.; Li, M.; Guan, Q.; Zhao, J. Tissue-specific expression of PPAR mRNAs in diabetic r ats and divergent effects of cilostazol. Can. J. Physiol. Pharmacol. 2008, 86, 465–471.
- 28. Hu, Y.; Chen, Y.; Ding, L.; He, X.; Takahashi, Y.; Gao, Y.; Shen, W.; Cheng, R.; Chen, Q.; Qi, X.; et al. Pathogenic role o f diabetes-induced PPAR-α down-regulation in microvascular dysfunction. Proc. Natl. Acad. Sci. USA 2013, 110, 15401 –15406.
- Ding, L.; Cheng, R.; Hu, Y.; Takahashi, Y.; Jenkins, A.J.; Keech, A.C.; Humphries, K.M.; Gu, X.; Elliott, M.H.; Xia, X.; et al. Peroxisome proliferator-activated receptor α protects capillary pericytes in the retina. Am. J. Pathol. 2014, 184, 2709 –2720.
- 30. Chen, Y.; Hu, Y.; Lin, M.; Jenkins, A.J.; Keech, A.C.; Mott, R.; Lyons, T.J.; Ma, J.-X. Therapeutic Effects of PPARα Agoni sts on Diabetic Retinopathy in Type 1 Diabetes Models. Diabetes 2013, 62, 261.
- 31. Pearsall, E.A.; Cheng, R.; Matsuzaki, S.; Zhou, K.; Ding, L.; Ahn, B.; Kinter, M.; Humphries, K.M.; Quiambao, A.B.; Farj o, R.A.; et al. Neuroprotective effects of PPARα in retinopathy of type 1 diabetes. PLoS ONE 2019, 14, e0208399.
- 32. Qiu, F.; Matlock, G.; Chen, Q.; Zhou, K.; Du, Y.; Wang, X.; Ma, J.X. Therapeutic Effects of PPARα Agonist on Ocular Ne ovascularization in Models Recapitulating Neovascular Age-Related Macular Degeneration. Invest. Ophthalmol. Vis. Sc i. 2017, 58, 5065–5075.
- 33. Moran, E.; Ding, L.; Wang, Z.; Cheng, R.; Chen, Q.; Moore, R.; Takahashi, Y.; Ma, J.X. Protective and antioxidant effect s of PPARα in the ischemic retina. Invest. Ophthalmol. Vis. Sci. 2014, 55, 4568–4576.
- 34. Wang, Z.; Moran, E.; Ding, L.; Cheng, R.; Xu, X.; Ma, J.X. PPARα regulates mobilization and homing of endothelial pro genitor cells through the HIF-1α/SDF-1 pathway. Invest. Ophthalmol. Vis. Sci. 2014, 55, 3820–3832.
- 35. Ye, S.; Chen, Q.; Jiang, N.; Liang, X.; Li, J.; Zong, R.; Huang, C.; Qiu, Y.; Ma, J.-X.; Liu, Z. PPARα-Dependent Effects o f Palmitoylethanolamide Against Retinal Neovascularization and Fibrosis. Investig. Ophthalmol. Vis. Sci. 2020, 61, 15.
- 36. Romano, M.R.; Lograno, M.D. Involvement of the peroxisome proliferator-activated receptor (PPAR) alpha in vascular r esponse of endocannabinoids in the bovine ophthalmic artery. Eur. J. Pharmacol. 2012, 683, 197–203.
- 37. Strobbe, E.; Cellini, M.; Campos, E.C. Effectiveness of palmitoylethanolamide on endothelial dysfunction in ocular hype rtensive patients: A randomized, placebo-controlled cross-over study. Invest. Ophthalmol. Vis. Sci. 2013, 54, 968–973.
- Kumar, A.; Qiao, Z.; Kumar, P.; Song, Z.H. Effects of palmitoylethanolamide on aqueous humor outflow. Invest. Ophthal mol. Vis. Sci. 2012, 53, 4416–4425.
- 39. Keppel Hesselink, J.M.; Costagliola, C.; Fakhry, J.; Kopsky, D.J. Palmitoylethanolamide, a Natural Retinoprotectant: Its Putative Relevance for the Treatment of Glaucoma and Diabetic Retinopathy. J. Ophthalmol. 2015, 2015, 430596.
- 40. He, H.; Liang, M.; Li, L.; Luo, S.; Fang, X.; He, H.; Xiao, X.; Wu, H.; Lin, Z. PPAR-α Agonist Fenofibrate Suppressed the Formation of Ocular Surface Squamous Metaplasia Induced by Topical Benzalkonium Chloride. Invest. Ophthalmol. Vi s. Sci. 2020, 61, 54.

- 41. Li, W.; Hayashida, Y.; Chen, Y.T.; He, H.; Tseng, D.Y.; Alonso, M.; Chen, S.Y.; Xi, X.; Tseng, S.C. Air exposure induced s quamous metaplasia of human limbal epithelium. Invest. Ophthalmol. Vis. Sci. 2008, 49, 154–162.
- 42. Tang, L.; Wang, X.; Wu, J.; Li, S.M.; Zhang, Z.; Wu, S.; Su, T.; Lin, Z.; Chen, X.; Liao, X.; et al. Sleep Deprivation Induc es Dry Eye Through Inhibition of PPARα Expression in Corneal Epithelium. Investig. Ophthalmol. Vis. Sci. 2018, 59, 54 94–5508.
- 43. Sana Raza-Iqbal; Toshiya Tanaka; Motonobu Anai; Takeshi Inagaki; Yoshihiro Matsumura; Kaori Ikeda; Akashi Taguchi; Frank J. Gonzalez; Juro Sakai; Tatsuhiko Kodama; et al. Transcriptome Analysis of K-877 (a Novel Selective PPARα M odulator (SPPARMα))-Regulated Genes in Primary Human Hepatocytes and the Mouse Liver. *Journal of Atherosclerosi* s and Thrombosis **2015**, *22*, 754-772, <u>10.5551/jat.28720</u>.
- 44. Arima, T.; Uchiyama, M.; Nakano, Y.; Nagasaka, S.; Kang, D.; Shimizu, A.; Takahashi, H. Peroxisome proliferator-activa ted receptor alpha agonist suppresses neovascularization by reducing both vascular endothelial growth factor and angi opoietin-2 in corneal alkali burn. Sci. Rep. 2017, 7, 17763.
- 45. Nakano, Y.; Arima, T.; Tobita, Y.; Uchiyama, M.; Shimizu, A.; Takahashi, H. Combination of Peroxisome Proliferator-Acti vated Receptor (PPAR) Alpha and Gamma Agonists Prevents Corneal Inflammation and Neovascularization in a Rat Al kali Burn Model. Int. J. Mol. Sci. 2020, 21, 5093.
- 46. Sharma, N.; Kaur, M.; Agarwal, T.; Sangwan, V.S.; Vajpayee, R.B. Treatment of acute ocular chemical burns. Surv. Oph thalmol. 2018, 63, 214–235.
- 47. Jean-Charles Fruchart; Pemafibrate (K-877), a novel selective peroxisome proliferator-activated receptor alpha modula tor for management of atherogenic dyslipidaemia. *Cardiovascular Diabetology* **2017**, *16*, 1-12, <u>10.1186/s12933-017-06</u> <u>02-y</u>.
- 48. Jean-Charles Fruchart; Selective peroxisome proliferator-activated receptorα modulators (SPPARMα): The next genera tion of peroxisome proliferator-activated receptor α-agonists. *Cardiovascular Diabetology* **2013**, *12*, 82-82, <u>10.1186/147</u> <u>5-2840-12-82</u>.
- 49. Yusuke Sasaki; Sana Raza-Iqbal; Toshiya Tanaka; Kentaro Murakami; Motonobu Anai; Tsuyoshi Osawa; Yoshihiro Mat sumura; Juro Sakai; Tatsuhiko Kodama; Gene Expression Profiles Induced by a Novel Selective Peroxisome Proliferat or-Activated Receptor α Modulator (SPPARMα) Pemafibrate. *International Journal of Molecular Sciences* **2019**, *20*, 568 2, <u>10.3390/ijms20225682</u>.
- 50. Yohei Tomita; Deokho Lee; Yukihiro Miwa; Xiaoyan Jiang; Kazuo Tsubota; Toshihide Kurihara; Toshihide Kurihara; Pem afibrate Protects Against Retinal Dysfunction in a Murine Model of Diabetic Retinopathy. *International Journal of Molecu lar Sciences* **2020**, *21*, 6243, <u>10.3390/ijms21176243</u>.
- 51. Yohei Tomita; Nobuhiro Ozawa; Yukihiro Miwa; Ayako Ishida; Masayuki Ohta; Kazuo Tsubota; Toshihide Kurihara; Pem afibrate Prevents Retinal Pathological Neovascularization by Increasing FGF21 Level in a Murine Oxygen-Induced Reti nopathy Model. *International Journal of Molecular Sciences* **2019**, *20*, 5878, <u>10.3390/ijms20235878</u>.
- 52. Akira Shiono; Hiroki Sasaki; Reio Sekine; Yohei Abe; Yoshihiro Matsumura; Takeshi Inagaki; Toshiya Tanaka; Tatsuhiko Kodama; Hiroyuki Aburatani; Juro Sakai; et al. PPARα activation directly upregulates thrombomodulin in the diabetic ret ina. *Scientific Reports* **2020**, *10*, 10837, <u>10.1038/s41598-020-67579-1</u>.
- 53. Wagoner, M.D. Chemical injuries of the eye: Current concepts in pathophysiology and therapy. Surv. Ophthalmol. 1997, 41, 275–313.
- 54. Wu, T.; Qiao, S.; Shi, C.; Wang, S.; Ji, G. Metabolomics window into diabetic complications. J. Diabetes Investig. 2018, 9, 244–255.
- 55. Akira Shiono; Hiroki Sasaki; Reio Sekine; Yohei Abe; Yoshihiro Matsumura; Takeshi Inagaki; Toshiya Tanaka; Tatsuhiko Kodama; Hiroyuki Aburatani; Juro Sakai; et al. PPARα activation directly upregulates thrombomodulin in the diabetic ret ina. *Scientific Reports* **2020**, *10*, 10837, <u>10.1038/s41598-020-67579-1</u>.

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