

Brassica Bioactives May Ameliorate Endometriosis

Subjects: Pathology | Biotechnology & Applied Microbiology | Food Science & Technology

Contributor: Diego Moreno-Fernandez, Antonio J. Ruiz-Alcaraz, María Martínez-Esparza

Endometriosis is a chronic, inflammatory, hormone-dependent disease characterized by histological lesions produced by the presence of endometrial tissue outside the uterine cavity. Despite the fact that an estimated 176 million women are affected worldwide by this gynecological disorder, risk factors that cause endometriosis have not been properly defined and current treatments are not efficient. Although the interaction between diet and human health has been the focus of many studies, little information about the correlation of foods and their bioactive derivatives with endometriosis is available. In this framework, *Brassica* crops have emerged as potential candidates for ameliorating the chronic inflammatory condition of endometriosis, due to their abundant content of health-promoting compounds such as glucosinolates and their hydrolysis products, isothiocyanates.

Keywords: aquaporins ; Brassica ; isothiocyanates ; indoles ; endometriosis ; inflammation

1. Introduction

Endometriosis is a frequently diagnosed, incurable, hormone-dependent gynecological disorder characterized by a chronic inflammatory profile and histological lesions generated by the abnormal growth of endometrial-like tissue outside the uterine cavity. These lesions mainly occur within the peritoneal cavity and are engrafted in different locations such as the peritoneum wall, ovaries, colon, and bladder, although it can also develop in distant organs, such as the liver, lung, and brain, among others [1][2]. Endometriosis may appear in diverse forms and locations, and thus it can be classified as peritoneal endometriosis; ovarian endometriosis; deep infiltrating endometriosis (DIE); or extragenital endometriosis [1][2].

It is estimated that approximately 176 million women worldwide are affected by endometriosis, which represents about 10% of women of reproductive age [2]. However, it is suspected that these data could underestimate the real number of affected women, since many of them are accidentally diagnosed after going into surgery for the treatment of other pathologies [1][2]. Women affected by endometriosis may present severe symptoms, which include, among others, chronic pelvic pain, dysmenorrhea, and dyspareunia [1][2][3][4]. In this context, 2–4% of women of reproductive age may suffer from sexual dysfunction due to the painful symptoms produced during intercourse [5]. Furthermore, pelvic pain can be incapacitating, especially in cases of DIE [6][7]. Endometriosis causes infertility in approximately 30% of affected women, likely because of the endometriotic lesion scars present in the reproductive organs [8]. Furthermore, it has been demonstrated that endometriosis is a disease with a high impact on health care costs, being comparable with other chronic diseases such as diabetes [9].

On the other hand, knowledge of the direct influence of diet on human health has grown quickly in recent years [10][11]. However, to date, no conclusive results relating a diet rich in vegetables with endometriosis development or an improvement in its symptoms have been found [12]. In this regard, Parazzini *et al.* [13] performed a case-control study with 504 women suffering from endometriosis. Food frequency questionnaires revealed that women with a higher intake of vegetables showed a lower endometriosis risk. Nevertheless, another case-control study based on dietary questionnaires found no association between a higher intake of vegetables and a decrease in endometriosis risk [14]. A more recent study showed that women who consumed more than one serving of cruciferous vegetables per day had a 13% higher risk of developing endometriosis compared with those who ingested them once a week or less [15]. However, these data came from retrospective studies based on self-reported questionnaires, while there is little or no information on the effect of a diet rich in *Brassica* spp. for women affected with endometriosis and even less information about the effect of the bioactive compounds present in cruciferous foods.

2. Brassicaceae and Their Bioactive Compounds in Inflammation

Brassica crops are well known for their high content in health-promoting compounds [16][17]. Specifically, glucosinolates (GLSs) and their bioactive breakdown product, isothiocyanates (ITCs), have shown different types of activity, such as induction of detoxification Phase II enzymes [18] and anti-tumorigenic [19] or anti-inflammatory effects [20]. Glucosinolates

are secondary plant metabolites that are mainly present in vegetables from the Brassicaceae family. Their basic structure is a thiohydrozimate-O-sulfonate group linked with a glucose, whose side chain can vary depending on the amino acid from which they are derived [21]. Inside the plant cells, GLSs are stable, but when tissue disruption takes place, for example, chewing or wounding, these biomolecules are hydrolyzed by the enzyme myrosinase (EC 3.2.1.147), producing ITCs [22]. Cooking methods can also affect the content in glucosinolates and its degree of conversion to ITCs. For example, steaming, microwave cooking, and stir frying are processes that reduce the glucosinolate content the least and can even improve its extraction from the food matrix [23]. However, boiling or blanching usually inactivate myrosinase while leaching glucosinolates and breakdown products into the water [24]. A link between chronic inflammatory-related diseases, such as cancer or obesity, and these bioactive ITCs has been reported [25]. Furthermore, different ITCs are involved in diverse mechanisms and pathways of inflammatory processes; this analyzed here under the perspective of endometriosis.

2.1. Aliphatic ITCs and Related Metabolites

One of the most frequently studied aliphatic ITCs is sulforaphane (SFN), the resulting hydrolysis derivative of glucoraphanin, due to its effects on human health. SFN is widely considered to be a multi-faceted agent due to its role in several cellular pathways—attenuating, reversing, or even blocking different activities in the cellular metabolism [19]. For example, it has been reported that SFN specially induces the activation of Phase II detoxification enzymes, alters cellular signaling pathways, and takes part in the suppression of pro-inflammatory responses [26]. Specifically, SFN has been described as an inducer of the Nrf2 transcription factor (Figure 1A), which is responsible for the transcription of various genes involved in antioxidant activities or anti-inflammatory pathways [27]. The influence of SFN is based on its ability to bind the cysteine residues present in the Nrf2 repressor Keap1. Under normal conditions, Keap1 promotes the degradation of Nrf2 in the proteasome, but when SFN interacts with it, Nrf2 is released and translocated to the nucleus [28].

Figure 1. Effect of *Brassicaceae* bioactives compounds upon the transcription of genes related with inflammation. **(A)** In physiological conditions, Keap1 promotes the degradation of Nrf2 by the proteasome. However, when sulforaphane (SFN) interacts with Keap1 by binding its cysteine residues, Nrf2 is released and translocated to the nucleus. In this way, Nrf2 up regulates the transcription of genes related with antioxidant activities and anti-inflammatory pathways. **(B)** In an inflammatory process, Toll-like receptor 4 (TLR4) undergoes dimerization after recognizing pathogen-associated molecular patterns (PAMPs), activating the NFκB pathway and up-regulating genes related with inflammation. When SFN and iberin interact with TLR4, it cannot oligomerize, and the inflammatory response is inhibited.

Furthermore, SFN is involved in the NFκB pathway, decreasing its ability to bind to target genes related to the inflammatory response, such as pro-inflammatory interleukins, including tumor necrosis factor (TNF)-α [29]. In this signaling cascade, Toll-like receptor (TLR)-4 is the first element, and it is responsible for recognizing pathogen-associated molecular patterns (PAMPs) or intrinsic molecules (Figure 1B), such as fibronectin and heparan sulfate [29][30]. It has been reported that SFN can bind to the cysteine residues in this receptor, suppressing its oligomerization and inhibiting the inflammatory response [31].

Additionally, several pre-clinical studies have demonstrated the efficiency of this ITC in models of chronic inflammatory diseases [32]. For example, Zhao *et al.* [33] demonstrated that the administration of SFN to a nephropathy murine model reduced the level of reactive oxygen species (ROS) and activated Nrf2 transcription factor. However, little to no information about the effects in models of endometriosis has been found. Recently, Zhou *et al.* [34] revealed that SFN attenuated endometriosis symptoms in rats by diminishing the levels of TNF-α and IL-6 in peritoneal fluid and plasma. Moreover, SFN seems to decrease the expression of VEGF, affecting the neoangiogenesis of the endometriotic foci. In addition, SFN has been reported to inhibit the growth of ectopic endometrial tissue in sciatic endometriosis rat models, showing a decrease in both the size of lesions and the VEGF level [35]. However, more studies are needed in order to determine the effect of SFN in long-term dietary interventions.

SFN has a structural analog, sulforaphene (SFE), which differs only by having a double bond in the alkyl chain. It is derived from the aliphatic GLS glucoraphenin and can be found mainly in radishes [36]. Although research on SFE has been focused on its anti-carcinogenic and pro-apoptotic effects [37], little is known about its direct effect on inflammatory processes. However, it is known that SFE can lose its double bond, turning into SFN, whose properties are better known [38][39].

Another aliphatic ITC present in *Brassica* foods, including broccoli sprouts, is erucin [40]. Although an inter-conversion of SFN to erucin and viceversa has been reported in humans after its consumption, the effects of erucin in inflammation are also significant [41]. For example, it has been reported that it decreases the DNA-binding capacity of NFκB, thus reducing the transcription of target genes related to the inflammatory process, including TNF-α, IL-6, COX-2, and inducible nitric

oxide synthase (iNOS) [42]. Additionally, glucoiberin is a glucosinolate that is present in products such as cabbage and kale, and its degradation product is iberin [43][44]. As mentioned previously, TLRs play an important role in the induction of the innate immune response, and iberin can prevent the dimerization of TLRs, down-regulating NFκB signaling [45].

In oilseed rape and other herbaceous *Brassic*as, the aliphatic ITC allyl-ITC (AITC) derived from sinigrin does not have sulfur atoms in its side chain, unlike SFN and SFE. The action of AITC in the NFκB pathway when administered to the LPS-activated macrophage RAW 264.7 has been reported to decrease the production of TNF-α, IL-6, and nitric oxide [46]. In addition, *in vivo* studies performed in rat models of mammary carcinogenesis showed that AITC administration did not provoke a decrease in p65-NFκB expression and, thus, in pro-inflammatory cytokines such as IL-6 [47]. The same group also reported that AITC has a preventive effect on DMBA-induced mammary carcinogenesis by modulating the aryl hydrocarbon receptor (AhR)/Nrf2 signaling pathway [48]. This cytoplasmic receptor and transcription factor has a major role in environmental pollutant detoxification [49]. Nevertheless, recent studies have highlighted the role of AhR as a negative regulator of the immune response [50]. In this regard, it has been reported that AhR-null mice produce higher levels of pro-inflammatory cytokines, such as TNF-α and IL-12 [51].

2.2. Indoles and Related Compounds

The hydrolysis of glucobrassicin, a major indole glucosinolate present in *Brassic*as, including broccoli or cabbage [52], produces indole-3-carbinol (I3C). I3C can be converted into its dimeric condensation product 3,3-diindolylmethane (3,3-DIM) [53]. Both biomolecules are AhR agonists that decrease the level of IL-1β and increase the rate of the detoxification cascade [54]. In addition, when I3C was administered to murine models of doxorubicin-induced damage, positive stimulation of the Nrf2 response and down-regulation of p50-NFκB expression was observed [55]. However, when macrophages derived from monocytes of systemic lupus erythematosus patients were treated with I3C, a switch toward a M2 phenotype was described [56]. As mentioned before, M2 macrophages could help with the survival of endometrial cysts. Thus, in the absence of *in vivo* studies of I3C applied to endometriosis, the specific effect on this disease remains unknown.

On the other hand, 3,3-DIM alone has been reported to inhibit the inflammatory response by also down-regulating the NFκB pathway and decreasing the levels of prostaglandin E₂ (PGE₂), TNF-α, IL-6, and IL-1β in murine macrophage cell cultures [57], and the oral administration of 3,3-DIM to a model of acute colitis in mice provoked the down-regulation of various types of VEGF and the expression of the VEGF receptor-2 [58]. This function would be beneficial for endometriosis patients to reduce the neoangiogenesis of lesions. In addition, the effects of 3,3-DIM have been also tested in combination with dienogest, a common progestin that is commonly prescribed to treat pain associated with endometriosis. Women who were administered with this combination for 3 months reported less endometriosis-associated pelvic pain [59]. However, only eight patients finished the study; therefore, studies with a larger number of volunteers are needed to better understand its effects.

Ascorbigen is a biomolecule obtained from glucobrassicin degradation products generated after the enzymatic hydrolysis of I3C in the presence of ascorbic acid that has shown high antioxidant potential [60]. Fermented cabbage extracts enriched in ascorbigen were found to have improved antioxidant capacity and nitric oxide inhibition in RAW 264.7 murine cell cultures [61]. Nonetheless, information about the exact effects of this metabolite in inflammation is very limited since no recent *in vivo* studies are available.

3. Conclusion and perspectives

To summarize the results presented in this review, macrophages are key immune cells in endometriotic lesions, which are characterized by an anomalous inflammatory environment and cell growth in women with endometriosis. We have presented and discussed a new approach based on the potential role of glucosinolates/isothiocyanates from brassicas as putative natural anti-inflammatory compounds, which could reduce the symptoms and progression of endometriosis, improving the quality of life for affected women. The potential role of glucosinolates in the development chronic inflammatory diseases should be investigated since there is still little information in this respect.

References

1. Vercellini, P.; Viganò, P.; Somigliana, E.; Fedele, L. Endometriosis: Pathogenesis and treatment. *Nat. Rev. Endocrinol.* 2014, 10, 261–275.
2. Zondervan, K.T.; Becker, C.M.; Koga, K.; Missmer, S.A.; Taylor, R.N.; Viganò, P. Endometriosis. *Nat. Rev. Dis. Primers* 2018, 4, 9.

3. As-Sanie, S.; Black, R.; Giudice, L.C.; Gray Valbrun, T.; Gupta, J.; Jones, B.; Laufer, M.R.; Milspaw, A.T.; Missmer, S.A.; Norman, A.; et al. Assessing research gaps and unmet needs in endometriosis. *Am. J. Obstet. Gynecol.* 2019, 221, 86–94.
4. Denny, E.; Mann, C.H. A clinical overview of endometriosis: A misunderstood disease. *Br. J. Nurs.* 2007, 16, 1112–1116.
5. La Rosa, V.L.; de Franciscis, P.; Barra, F.; Schiattarella, A.; Tropea, A.; Tesarik, J.; Shah, M.; Kahramanoglu, I.; Marques Cerentini, T.; Ponta, M.; et al. Sexuality in women with endometriosis: A critical narrative review. *Minerva Med.* 2020, 111, 79–89.
6. Jones, G.; Jenkinson, C.; Kennedy, S. The impact of endometriosis upon quality of life: A qualitative analysis. *J. Psychosom. Obstet. Gynaecol.* 2004, 25, 123–133.
7. Aerts, L.; Grangier, L.; Streuli, I.; Dällenbach, P.; Marci, R.; Wenger, J.-M.; Pluchino, N. Psychosocial impact of endometriosis: From co-morbidity to intervention. *Best Pract. Res. Clin. Obstet. Gynaecol.* 2018, 50, 2–10.
8. Mahmood, T.A.; Templeton, A.A.; Thomson, L.; Fraser, C. Menstrual symptoms in women with pelvic endometriosis. *Br. J. Obstet. Gynaecol.* 1991, 98, 558–563.
9. La Rosa, V.L.; De Franciscis, P.; Barra, F.; Schiattarella, A.; Török, P.; Shah, M.; Karaman, E.; Marques Cerentini, T.; Di Guardo, F.; Gullo, G.; et al. Quality of life in women with endometriosis: A narrative overview. *Minerva Med.* 2020, 111, 68–78.
10. López-Chillón, M.T.; Carazo-Díaz, C.; Prieto-Merino, D.; Zafrilla, P.; Moreno, D.A.; Villaño, D. Effects of long-term consumption of broccoli sprouts on inflammatory markers in overweight subjects. *Clin. Nutr.* 2019, 38, 745–752.
11. Finicelli, M.; Squillaro, T.; Di Cristo, F.; Di Salle, A.; Melone, M.A.B.; Galderisi, U.; Peluso, G. Metabolic syndrome, Mediterranean diet, and polyphenols: Evidence and perspectives. *J. Cell. Physiol.* 2019, 234, 5807–5826.
12. Soave, I.; Occhiali, T.; Wenger, J.M.; Pluchino, N.; Caserta, D.; Marci, R. Endometriosis and food habits: Can diet make the difference? *J. Endometr. Pelvic Pain Disord.* 2018, 10, 59–71.
13. Parazzini, F.; Esposito, G.; Tozzi, L.; Noli, S.; Bianchi, S. Epidemiology of endometriosis and its comorbidities. *Eur. J. Obstet. Gynecol. Reprod. Biol.* 2017, 209, 3–7.
14. Trabert, B.; Peters, U.; De Roos, A.J.; Scholes, D.; Holt, V.L. Diet and risk of endometriosis in a population-based case-control study. *Br. J. Nutr.* 2011, 105, 459–467.
15. Harris, H.R.; Eke, A.C.; Chavarro, J.E.; Missmer, S.A. Fruit and vegetable consumption and risk of endometriosis. *Hum. Reprod.* 2018, 33, 715–727.
16. Hallmann, E.; Kazimierczak, R.; Marszałek, K.; Drela, N.; Kiernożek, E.; Toomik, P.; Matt, D.; Luik, A.; Rembiałkowska, E. The nutritive value of organic and conventional white cabbage (*Brassica oleracea* L. var. *capitata*) and anti-apoptotic activity in gastric adenocarcinoma cells of sauerkraut juice produced thereof. *J. Agric. Food Chem.* 2017, 65, 8171–8183.
17. Wiczorek, M.N.; Walczak, M.; Skrzypczak-Zielińska, M.; Jeleń, H.H. Bitter taste of Brassica vegetables: The role of genetic factors, receptors, isothiocyanates, glucosinolates, and flavor context. *Crit. Rev. Food Sci. Nutr.* 2018, 58, 3130–3140.
18. Russo, M.; Spagnuolo, C.; Russo, G.L.; Skalicka-Woźniak, K.; Daglia, M.; Sobarzo-Sánchez, E.; Nabavi, S.F.; Nabavi, S.M. Nrf2 targeting by sulforaphane: A potential therapy for cancer treatment. *Crit. Rev. Food Sci. Nutr.* 2018, 58, 1391–1405.
19. Bayat Mokhtari, R.; Baluch, N.; Homayouni, T.S.; Morgatskaya, E.; Kumar, S.; Kazemi, P.; Yeger, H. The role of Sulforaphane in cancer chemoprevention and health benefits: A mini-review. *J. Cell Commun. Signal.* 2018, 12, 91–101.
20. Burčul, F.; Generalić Mekinić, I.; Radan, M.; Rollin, P.; Blažević, I. Isothiocyanates: Cholinesterase inhibiting, antioxidant, and anti-inflammatory activity. *J. Enzyme Inhib. Med. Chem.* 2018, 33, 577–582.
21. Agerbirk, N.; Olsen, C.E. Glucosinolate structures in evolution. *Phytochemistry* 2012, 77, 16–45.
22. Bhat, R.; Vyas, D. Myrosinase: Insights on structural, catalytic, regulatory, and environmental interactions. *Crit. Rev. Biotechnol.* 2019, 39, 508–523.
23. Nugraedi, P.Y.; Verkerk, R.; Widianarko, B.; Dekker, M. A Mechanistic Perspective on Process-Induced Changes in Glucosinolate Content in Brassica Vegetables: A Review. *Crit. Rev. Food Sci. Nutr.* 2015, 55, 823–838.
24. Soares, A.; Carrascosa, C.; Raposo, A. Influence of Different Cooking Methods on the Concentration of Glucosinolates and Vitamin C in Broccoli. *Food Bioprocess Technol.* 2017, 10, 1387–1411.

25. Bessler, H.; Djaldetti, M. Broccoli and human health: Immunomodulatory effect of sulforaphane in a model of colon cancer. *Int. J. Food Sci. Nutr.* 2018, 69, 946–953.
26. Houghton, C.A.; Fassett, R.G.; Coombes, J.S. Sulforaphane and other nutrigenomic Nrf2 activators: Can the clinician's expectation be matched by the reality? *Oxid. Med. Cell. Longev.* 2016, 2016, 785186.
27. Eren, E.; Tufekci, K.U.; Isci, K.B.; Tastan, B.; Genc, K.; Genc, S. Sulforaphane inhibits lipopolysaccharide-induced inflammation, cytotoxicity, oxidative stress, and miR-155 expression and switches to mox phenotype through activating extracellular signal-regulated kinase 1/2-nuclear factor erythroid 2-related factor 2/An. *Front. Immunol.* 2018, 23, 9–36.
28. Greaney, A.J.; Maier, N.K.; Leppla, S.H.; Moayeri, M. Sulforaphane inhibits multiple inflammasomes through an Nrf2-independent mechanism. *J. Leukoc. Biol.* 2016, 99, 189–199.
29. Okamura, Y.; Watari, M.; Jerud, E.S.; Young, D.W.; Ishizaka, S.T.; Rose, J.; Chow, J.C.; Strauss, J.F. The extra domain A of fibronectin activates Toll-like Receptor 4. *J. Biol. Chem.* 2001, 276, 10229–10233.
30. O'Callaghan, P.; Zhang, X.; Li, J.P. Heparan sulfate proteoglycans as relays of neuroinflammation. *J. Histochem. Cytochem.* 2018, 66, 305–319.
31. Youn, H.S.; Kim, Y.S.; Park, Z.Y.; Kim, S.Y.; Choi, N.Y.; Joung, S.M.; Seo, J.A.; Lim, K.-M.; Kwak, M.-K.; Hwang, D.H.; et al. Sulforaphane Suppresses Oligomerization of TLR4 in a Thiol-Dependent Manner. *J. Immunol.* 2010, 184, 305–319.
32. Mazarakis, N.; Snibson, K.; Licciardi, P.V.; Karagiannis, T.C. The potential use of L-sulforaphane for the treatment of chronic inflammatory diseases: A review of the clinical evidence. *Clin. Nutr.* 2019, 39, 664–675.
33. Zhao, Z.; Liao, G.; Zhou, Q.; Lv, D.; Holthfer, H.; Zou, H. Sulforaphane attenuates contrast-induced nephropathy in rats via Nrf2/HO-1 pathway. *Oxid. Med. Cell. Longev.* 2016, 2016, 11–13.
34. Zhou, A.; Hong, Y.; Lv, Y. Sulforaphane attenuates endometriosis in rat models through inhibiting PI3K/Akt signaling pathway. *Dose-Response* 2019, 17, 1559325819855538.
35. Liu, Y.; Zhang, Z.; Lu, X.; Meng, J.; Qin, X.; Jiang, J. Anti-nociceptive and anti-inflammatory effects of sulforaphane on sciatic endometriosis in a rat model. *Neurosci. Lett.* 2020, 723, 134858.
36. Baenas, N.; Piegholdt, S.; Schloesser, A.; Moreno, D.A.; García-Viguera, C.; Rimbach, G.; Wagner, A.E. Metabolic activity of radish sprouts derived isothiocyanates in drosophila melanogaster. *Int. J. Mol. Sci.* 2016, 17, 251.
37. Pawlik, A.; Wała, M.; Hać, A.; Felczykowska, A.; Herman-Antosiewicz, A. Sulforaphane, an isothiocyanate present in radish plants, inhibits proliferation of human breast cancer cells. *Phytomedicine* 2017, 29, 1–10.
38. Pocasap, P.; Weerapreeyakul, N.; Barusrux, S. Cancer preventive effect of Thai rat-tailed radish (*Raphanus sativus* L. var. *caudatus* Alef). *J. Funct. Foods* 2013, 5, 1372–1381.
39. Baenas, N.; Suárez-Martínez, C.; García-Viguera, C.; Moreno, D.A. Bioavailability and new biomarkers of cruciferous sprouts consumption. *Food Res. Int.* 2017.
40. Westphal, A.; Riedl, K.M.; Cooperstone, J.L.; Kamat, S.; Balasubramaniam, V.M.; Schwartz, S.J.; Böhm, V. High-pressure processing of broccoli sprouts: Influence on bioactivation of glucosinolates to isothiocyanates. *J. Agric. Food Chem.* 2017, 65, 8578–8585.
41. Clarke, J.D.; Hsu, A.; Riedl, K.; Bella, D.; Schwartz, S.J.; Stevens, J.F.; Ho, E. Bioavailability and inter-conversion of sulforaphane and erucin in human subjects consuming broccoli sprouts or broccoli supplement in a cross-over study design. *Pharmacol. Res.* 2011, 64, 456–463.
42. Cho, H.J.; Lee, K.W.; Yoon Park, J.H. Erucin exerts anti-inflammatory properties in murine macrophages and mouse skin: Possible mediation through the inhibition of NFκB signaling. *Int. J. Mol. Sci.* 2013, 14, 20564–20577.
43. Luang-In, V.; Deeseenthum, S.; Udomwong, P.; Saengha, W.; Gregori, M. Formation of sulforaphane and iberin products from Thai cabbage fermented by myrosinase-positive bacteria. *Molecules* 2018, 23, 955.
44. Baenas; Marhuenda; García-Viguera; Zafrilla; Moreno Influence of cooking methods on glucosinolates and isothiocyanates content in novel cruciferous foods. *Foods* 2019, 8, 257.
45. Shibata, T.; Nakashima, F.; Honda, K.; Lu, Y.J.; Kondo, T.; Ushida, Y.; Aizawa, K.; Suganuma, H.; Oe, S.; Tanaka, H.; et al. Toll-like receptors as a target of food-derived anti-inflammatory compounds. *J. Biol. Chem.* 2014, 289, 32757–32772.
46. Chang, W.J.; Chen, B.H.; Inbaraj, B.S.; Chien, J.T. Preparation of allyl isothiocyanate nanoparticles, their anti-inflammatory activity towards RAW 264.7 macrophage cells and anti-proliferative effect on HT1376 bladder cancer cells. *J. Sci. Food Agric.* 2019, 99, 3106–3116.
47. Rajakumar, T.; Pugalendhi, P.; Jayaganesh, R.; Ananthkrishnan, D.; Gunasekaran, K. Effect of allyl isothiocyanate on NF-κB signaling in 7,12-dimethylbenz(a)anthracene and N-methyl-N-nitrosourea-induced mammary carcinogenesis.

48. Rajakumar, T.; Pugalendhi, P.; Thilagavathi, S.; Ananthakrishnan, D.; Gunasekaran, K. Allyl isothiocyanate, a potent chemopreventive agent targets AhR/Nrf2 signaling pathway in chemically induced mammary carcinogenesis. *Mol. Cell. Biochem.* 2018, 437, 1–12.
49. Bessede, A.; Gargaro, M.; Pallotta, M.T.; Matino, D.; Servillo, G.; Brunacci, C.; Bicciato, S.; Mazza, E.M.C.; Macchiarulo, A.; Vacca, C.; et al. Aryl hydrocarbon receptor control of a disease tolerance defence pathway. *Nature* 2014, 511, 184–190.
50. Murray, I.A.; Patterson, A.D.; Perdew, G.H. Aryl hydrocarbon receptor ligands in cancer: Friend and foe. *Nat. Rev. Cancer* 2014, 14, 801–814.
51. Sanchez, Y.; de Dios Rosado, J.; Vega, L.; Elizondo, G.; Estrada-Muñiz, E.; Saavedra, R.; Juárez, I.; Rodríguez-Sosa, M. The unexpected role for the aryl hydrocarbon receptor on susceptibility to experimental toxoplasmosis. *J. Biomed. Biotechnol.* 2010, 2010, 505694.
52. Hwang, I.M.; Park, B.; Dang, Y.M.; Kim, S.Y.; Seo, H.Y. Simultaneous direct determination of 15 glucosinolates in eight Brassica species by UHPLC-Q-Orbitrap-MS. *Food Chem.* 2019, 282, 127–133.
53. Grose, K.R.; Bjeldanes, L.F. Oligomerization of Indole-3-carbinol in aqueous acid. *Chem. Res. Toxicol.* 1992, 5, 188–193.
54. Wang, T.T.Y.; Pham, Q.; Kim, Y.S. Elucidating the role of CD84 and AHR in modulation of LPS-induced cytokines production by cruciferous vegetable-derived compounds indole-3-carbinol and 3,3'-diindolylmethane. *Int. J. Mol. Sci.* 2018, 19, 339.
55. Hajra, S.; Patra, A.R.; Basu, A.; Bhattacharya, S. Prevention of doxorubicin (DOX)-induced genotoxicity and cardiotoxicity: Effect of plant derived small molecule indole-3-carbinol (I3C) on oxidative stress and inflammation. *Biomed. Pharmacother.* 2018, 101, 228–243.
56. Mohammadi, S.; Memarian, A.; Sedighi, S.; Behnampour, N.; Yazdani, Y. Immunoregulatory effects of indole-3-carbinol on monocyte-derived macrophages in systemic lupus erythematosus: A crucial role for aryl hydrocarbon receptor. *Autoimmunity* 2018, 51, 199–209.
57. Cho, H.J.; Seon, M.R.; Lee, Y.M.; Kim, J.; Kim, J.-K.; Kim, S.G.; Park, J.H.Y. 3,3'-Diindolylmethane suppresses the inflammatory response to lipopolysaccharide in murine macrophages. *J. Nutr.* 2008, 138, 17–23.
58. Jeon, E.J.; Davaatseren, M.; Hwang, J.T.; Park, J.H.; Hur, H.J.; Lee, A.S.; Sung, M.J. Effect of oral administration of 3,3'-Diindolylmethane on dextran sodium sulfate-induced acute colitis in mice. *J. Agric. Food Chem.* 2016, 64, 7702–7709.
59. Morales-Prieto, D.M.; Herrmann, J.; Osterwald, H.; Kochhar, P.S.; Schleussner, E.; Markert, U.R.; Oettel, M. Comparison of dienogest effects upon 3,3'-diindolylmethane supplementation in models of endometriosis and clinical cases. *Reprod. Biol.* 2018, 18, 252–258.
60. Tai, A.; Fukunaga, K.; Ohno, A.; Ito, H. Antioxidative properties of ascorbigen in using multiple antioxidant assays. *Biosci. Biotechnol. Biochem.* 2014, 78, 1723–1730.
61. Martinez-Villaluenga, C.; Peñas, E.; Sidro, B.; Ullate, M.; Frias, J.; Vidal-Valverde, C. White cabbage fermentation improves ascorbigen content, antioxidant and nitric oxide production inhibitory activity in LPS-induced macrophages. *LWT Food Sci. Technol.* 2012, 46, 77–83.