

# Vitro Tissue Culture-Induced Variation

Subjects: [Agriculture, Dairy & Animal Science](#)

Contributor: Piotr Bednarek

Somaclonal variation includes genetic or epigenetic changes exhibited between clonal regenerants and their corresponding donor plants derived via in vitro tissue cultures (A. Leva, L.M.R. Rinaldi, in Encyclopedia of Applied Plant Sciences (Second Edition), 2017). It usually assumes that the changes are being transmitted during a generative cycle. However, in some cases, to stress the fact that not all changes are either not analyzed in the progeny or may not be sexually transmitted, the tissue culture-induced variation seems to reflect better the issue (Quantification of the tissue-culture induced variation in barley (*Hordeum vulgare* L.) Bednarek, PT., Orłowska, R., Koebner, RMD., Zimny, J. 2007 *BMC Plant Biology* 7 (1), 1-9).

[plant tissue cultures](#)

[signaling pathways](#)

## 1. Introduction

Tissue culture-induced variation (TCIV) is a well-established phenomenon of any changes affecting regenerants phenotype or genotype during in vitro plant regeneration [\[1\]](#)[\[2\]](#)[\[3\]](#)[\[4\]](#). If the changes are stably inherited via the generative cycle, it is usually called somaclonal variation (SV) [\[5\]](#)[\[6\]](#). However, the TCIV and SV terms are often used mutually [\[7\]](#). TCIV may be pronounced at the morphological [\[8\]](#), biochemical [\[9\]](#), genetic [\[10\]](#) and epigenetic [\[11\]](#) levels, which do not necessarily overlap [\[12\]](#). Possibly the most investigated level is based on DNA markers.

The studies of molecular aspects of TCIV started at the beginning of the 1980s [\[13\]](#)[\[14\]](#)[\[15\]](#). The development of the restriction fragment length polymorphism (RFLP) [\[16\]](#) and the randomly amplified polymorphic DNA (RAPD) [\[17\]](#) techniques (**Table 1**) resulted in a burst of experimental data suggesting that even using anther culture, plant regeneration was subjected to numerous mutations such as single nucleotide polymorphisms (SNPs) [\[18\]](#). Some studies suggested that variation was due to the so-called pre-existing variation [\[19\]](#)[\[20\]](#), the others addressed mutations to the activation of transposable elements [\[21\]](#) due to DNA methylation pattern alternations [\[22\]](#). For a long time, the origin of changes was not apparent. It was speculated that they might have come from the degradation of the cells during tissue cultures [\[23\]](#), the elevated production of reactive oxygen species (ROS) [\[24\]](#), due to stressful conditions [\[25\]](#). The degradation hypothesis claimed that cell death in culture under stress and programmed cell death leads to secondary metabolites (pigments, tannins). Their release into culture can induce somaclonal variation in an unspecified way [\[23\]](#). It was speculated that the callus phase and indirect embryogenesis could have contributed to the phenomenon [\[26\]](#). For about a decade, it was thought that studies on TCIV were deficient. However, the development of new molecular tools and progress in epigenetics led to the revision of previous data and pushed a deeper understanding of the phenomenon.

**Table 1.** The arrangement of molecular techniques used to study TCIV.

Abiotic Stress	Factor	Plant
Starvation/Heat	Sucrose-free medium, 25 °C, 33 °C	Wheat, tobacco [27]
Osmotic	Mannitol solutions	Barley [28]
Heat	41 °C	Rapeseed [29]
Cold	4 °C	Pepper [30]
Cold/Heat	4 °C/32 °C	Rye [31]

The introduction of the amplified fragment length polymorphism (AFLP) approach [32], followed by the methylation-sensitive amplified polymorphism (MSAP) one [33][34], allowed studies of DNA methylation changes addressed to in vitro tissue culture plant regeneration [35][36]. Although that changes are frequent but may vary in a wide spectrum, from 0.07% [37] to 52% [38]. While the MSAP approach was productive, it could identify only DNA methylation changes related to CG and CHG sequence contexts [39]. Moreover, the ways to quantify changes used distinct algorithms [40] that might have differed from study to study [41][42], making a comparison of results somewhat tricky. Thus, the semi-quantitative MSAP approach, allowing for quantifying DNA methylation changes, was suggested to overcome the limitation [43]. However, studies on sequence variation required the utilization of additional marker techniques. Consequently, another AFLP based technique, the so-called methylation-sensitive AFLP (metAFLP) approach, was developed [1][44], allowing quantification of sequence and DNA methylation changes during a single experiment. Further advances in the technique showed that one might study not only CG and CHG but also the CHH methylation context, an essential tool in studies on cereals [1][12].

The development of the new generation sequencing (NGS) opened innovative opportunities. The diversity arrays technology methylation analysis (DArTseqMet) techniques permitted for identifying a large number of markers that could be employed in quantification procedures evaluated for the semiquantitative MSAP approach [45]. MethylRAD [46] or MethylSeq [47] are the other alternatives. The MethylSeq method is an NGS variant of the bisulfite-based sequencing approach. It could be used to study different types of genomic DNA methylation, but its application requires at least 10X coverage. The MethylRAD approach uses Mrr-like enzymes to collect 32-bp methylated DNA fragments from the whole genome for high-throughput sequencing. It allows for de novo methylation analysis using low DNA input. The two approaches have incredible potential in DNA methylome studies; however, they were not yet used in tissue culture studies.

All the techniques mentioned above were exploited in studies on the whole genome. When it was of value to investigate DNA methylation pattern change of a specific sequence, the bisulfite approach was recommended [48][49]. It allowed the establishment of the extent to which the CG, CHG, and CHH sequences were methylated in the genome [50]. Thus, the tools to study molecular aspects of the TCIV were established. Moreover, by that time, the genetic model of the studies was also evaluated [51]. It became apparent that studies on TCIV should start from a

well-defined, preferentially homozygous double haploid plant that served as a donor of tissue for the regenerants subjected to further analysis. Utilizing specific plant materials and sophisticated molecular marker techniques, TCIV was studied in barley [52], triticale [53], *Polyscias filicifolia* [54], and *Gentiana pannonica* Scop. [4]. It was shown that the phenomenon is common to plants [55][56][57] and that at least some DNA methylation changes could be transmitted to the progeny [58][59]. Furthermore, DNA methylation changes affecting regenerants may require several generative cycles to be stabilized [53]. Using a modification of the metAFLP approach that utilized primers directed towards TEs, their putative role in sequence variation was demonstrated [52][60]. However, the mechanisms of the TCIV are still being discussed.

It is becoming understood that analyses of DNA methylation changes affecting tissue culture due to plant regeneration are insufficient to have an image of the phenomenon. Interdisciplinary studies are needed to have species-specific models describing consecutive steps required to change the cell fate towards somatic development. In this context, studies of the cell wall, cell membrane, subintinal layer, their components, and the way signals transmitting stresses are involved in the process are needed. An interesting approach to study the role of the cell wall or biochemical pathways affected by in vitro tissue culture is attenuated total reflectance Fourier transform infrared (ATR-FTIR) spectroscopy [61][62], which allows for the identification of putative cell wall components [63] or biochemical pathways [64] participating in sensing stresses. It was documented that  $\beta$ -glucans being built of glucose units [65], and probably present in the subintinal layer of some microspores [66], maybe sources of glucose for new anther-culture-derived regenerants [67]. Furthermore, stressful conditions might influence DNA methylation, probably disturbing the methionine cycle [67]. The role of copper and silver ions in sequence variation [68] and green plant development [45] was also documented in barley regenerants derived via anther culture. Similarly, in embryo-derived regenerants of barley copper ions participated in sequence variation, possibly via modification of methylated sequences [12]. The respective relationships could be evaluated using mediation and moderation analyses [69] applied to DNA methylation changes and the ATR-FTIR spectroscopy data. Such results could be used in structural equation modeling (SEM), resulting in a deeper insight into the phenomenon leading towards its practical applications. However, due to limited sample size, SEM analysis allowing construction of an in vitro plant regeneration either via andro- or embryogenesis was not evaluated. If not the most substantial, a further limitation is the understanding of the role of the way the cell senses and transmits stresses leading towards TCIV.

## 2. Stresses and Their Role in Plant Regeneration through Tissue Culture In Vitro

It is assumed that inductive stress treatment is necessary to initiate the cell reprogramming process required for the microspore to switch from gametophytic to embryogenic fate. The stress treatment needed to switch the developmental fate of microspores depends on the plant species and the species genotype. Osmotic, starvation, cold, heat, and many other stresses are employed to initiate andro- and embryogenesis [70][71] (Table 2).

**Table 2.** Summation of stresses that efficiently switch the developmental fate of microspores.

Abiotic Stress	Factor	Plant
Starvation/Heat	Sucrose-free medium, 25 °C, 33 °C	Wheat, tobacco [27]
Osmotic	Mannitol solutions	Barley [28]
Heat	41 °C	Rapeseed [29]
Cold	4 °C	Pepper [30]
Cold/Heat	4 °C/32 °C	Rye [31]

Three phases of embryogenic development due to androgenesis could be distinguished. The first one reflects the acquisition of embryogenic potential as the reaction to stress. At this stage, which lasts about three days, repression of gametophytic development and dedifferentiation of the cells occurs. The second stage follows the induction and culture initiation step. The microspores divide and produce multicellular structures persisting within the microspore wall (exine) [72], indicating the initiation of embryogenesis. This stage, at least in barley, may take two days. Next, the exine breaks down, and embryos follow a similar zygotic embryogenesis pathway through globular, heart, torpedo, and cotyledonary embryos. The formation of tissue culture structures reflects the third phase of embryogenic development that begins around the 21st day of in vitro tissue cultures [72]. Interestingly, the third phase of embryogenic development corresponds to the point when under increased copper concentration present in induction medium, regeneration of new barley plants via anther culture results in the highest output [45].

In the model system of *Hordeum vulgare* (barley), the inductive cold stress (4 °C) is applied, whereas, in *Brassica napus*, 32 °C is used to initiate a switch from gameto- to sporophytic path. Stress-related heat-shock proteins (HSP70 and HSP90) were detected in microspore embryogenesis cultures of *Brassica napus* and *Capsicum annuum* due to cold treatment [73]. Thus, a protective role for the chaperones was suggested; however, their role in microspore embryogenesis is not clear. Low temperatures may alter endogenous ethylene (ET) levels enhancing tolerance with higher ET concentrations [74].

During reprogramming stages, induced stresses result in increased cell death and oxidative stress. At this point, ROS arise in excess and are accumulated [75]. However, in the case of *Medicago sativa* leaf protoplasts, the application of oxidative stress-inducing agents resulted in acceleration of cell cycle re-entry that was accompanied by a lower level of ROS accumulation [76]. The equilibrium between ROS-scavenging and ROS-producing mechanisms administrates the cell's level of damage and oxidative stress. Several enzymes of the antioxidative machinery of the cell increase their activity in microspore cultures. These stress-related proteins may have a protective role [77].

Abiotic stress factors may impact auxin homeostasis [78] probably resulting in the induction of somatic embryogenesis [79][80][81]. While microspore culture does not require exogenous auxin (2,4-dichlorophenoxyacetic acid), endogenously, the phytohormone may participate in microspore reprogramming in *Brassica napus* and in vitro embryo formation [82]. It was shown that induction of microspore embryogenesis resulted in de novo

endogenous auxin biosynthesis and accumulation of indole-3-acetic acid (responsible for embryo patterning, polarization, and differentiation) in pre-embryo cells starting from the first embryogenic divisions [82]. Auxin activity, biosynthesis, and transport are essential for stress-induced microspore embryogenesis. There is a link between auxin biosynthesis, its perception, transport, gene expression, signaling, and non-transcriptional responses; however, the exact mechanisms remain elusive [83][84][85]. The other hormone vital for plant cells is cytokinin that acts antagonistically to auxin. Cytokinin participates in cell growth and may control the early stages of somatic embryogenesis [86]. Spatiotemporal localization of cytokinin and auxin responses during microspore embryogenesis was suggested [72]. However, data concerning the presence of endogenous cytokinin in microspore embryogenesis was not evidenced.

It is worth mentioning that stress conditions may induce or impact autophagy, a significant pathway for recycling cell materials [87][88]. Autophagy may promote plant cell survival under starvation and stress conditions [89]. Moreover, excretion of cytoplasmic material (occasionally containing whole organelles) via single membrane-bound autophagic bodies and those deposited in the cell wall (remnants from the digestion of cytoplasmic organelles) of embryogenic microspores in between the cell wall of embryogenic microspores and the plasma membrane was observed [90]. Some of the autophagosomes were transported out of the cell, creating extracytoplasmic fibrillar and membranous material deposits. It was shown that embryogenic microspores are associated with autophagy and excretion of the removed material [90]. Thus, autophagy is a kind of cytoplasmic cleaning, whereas excretion is a mechanism of avoiding unnecessary vacuolar system growth [90]. The excretion was essential for proper microspore embryogenesis; however, in some cases, the material was redirected to the cell [90], possibly implying 'recycling' of the materials for cellular processes. It should also be mentioned that autophagy can involve converting the lytic vacuole to a storage one [91][92].

The connection between ROS and autophagy in plants was suggested [93]. In barley, the activation of autophagy after inductive stress at 4 °C with upregulation of *HvATG5* and *HvATG6* genes, and a rise in the number of autophagosomes was reported [94]. Autophagy is activated and involved in cell death with the participation of cathepsins the proteases that degrade proteins during stress-induced microspore embryogenesis in barley [94]. It was also demonstrated that stress treatment of *Brassica napus* also resulted in induction of autophagy [90] in parallel to cysteine-dependent proteases (metacaspase) involved in programmed cell death, stress, and cell proliferation [95], and gene expression [96]. Interestingly, application of agents directed towards ROS (MnCl<sub>2</sub>), autophagy [3-methyladenine (3-MA), inhibitor], and protease activities, caspase 3-like, and metacaspase activities (E64, Ac-DEVD-CHO, and Ac-VRPR-FMK; inhibitors), reduces cell death levels, increasing embryogenesis in rapeseed and barley [94] opening up pathways reducing stress-induced cell death at the early stages of microspore embryogenesis.

Inductive stresses are responsible for cell reprogramming. The process involves DNA demethylation and de novo methylation [97]. A global DNA hypomethylation during the change of the gametophytic to sporophytic fate and first embryogenic divisions were observed in barley [98]. Among inductive stressful conditions, darkness is one of the most exploited in cereals [99]. When mannitol is applied, triggering microspores to dividing [31], the culture is subjected to osmotic, and carbon starvation discussed earlier [100]. Sometimes, instead of darkness, heat stress is

applied [101]. It was suggested that ethylene participates in the response of plants to heat stress [74]. Stresses induce ethylene accumulation. The higher the ethylene concentration, the higher the chance of plant survival under stressful conditions [102]. Most probably, ethylene modulates gene expression via ethylene signaling mechanisms [103].

The tissue culture medium itself may be treated as a stressful factor. The regeneration medium contains many ingredients such as 6-benzylaminopurine, 2,4-dichlorophenoxyacetic acid, alpha-naphthalene acetic acid, and indole-3-acetic acid not necessary are neutral to the cells may induce TCIV [99][104][105]. As such ingredients may be toxic, they may lead to genomic DNA degradation as indicated by comet analysis [99]. It was also demonstrated that copper and silver ions containing media might also affect DNA methylation patterns [12], resulting in DNA sequence changes [67]. Moreover, the two ions most probably change respiratory chain balance affecting the Yang cycle [67] and inducing sequence changes [68]. Interestingly, copper and silver may moderate relationships between de novo methylation/demethylation processes, leading to green plant regeneration [45]. For more details on that, see an excellent review published recently [72].

Assuming that different stresses may affect in vitro tissue culture, the mucilage layer, the cell wall, and the plasma membrane are reasonable candidates for sensing stresses [106][107] affecting the cell functioning and possibly promoting the TCIV. However, the information on how the mucilage layer, the cell wall, and plasma membrane perform due to in vitro tissue culture plant regeneration is not well addressed.

### 3. Cell Wall and Plasma Membrane as Sensors of Stresses

Even less, the pollen cell wall protects male sperm from abiotic stresses (i.e., abrasion, desiccation, and UV radiation). A durable wall consists of gametophyte-derived intine and sporophyte-derived exine layers covered with lipid-rich pollen coat or trephine [108][109]. The intine resembles the primary cellulosic plant cell wall [108][110][111] whereas the exine is formed of a heterogeneous polymer composed of polyhydroxylated aliphatic constituents with aromatic or conjugated side chains containing ether and ester bonds [112][113][114] called sporopollenin synthesized from precursors in tapetum [108][109] and anchored on to microspores [115].

Pollen wall formation starts after microspore meiosis generating tetrads covered with callose. At the late tetrad stage, the microspore surface is formed of primexine, a transient cell wall matrix-shaped of polysaccharides outside of microspore plasma membrane inside callose. Primexine is composed of sporopollenin. This layer may contain cellulase [116]. The callose wall degrades after the tetrad phase allowing free microspore release into the locule and tecta. At this stage, a thin microspore-derived intine wall forming between the developing exine and a microspore-derived intine wall is evidenced [117].

As the composition of the microspore cell wall is not apparent, available information on the plant cell wall in general with emphasis on cellulose and wall matrix polysaccharides [118] are discussed. All of them form a rigid structure. The essential cell wall function relies on providing mechanical strength to resist turgor pressure. It forms a barrier against biotic and abiotic stresses that may alter the cell wall components' deposition [119].

The cell wall cellulose may be present in amorphic, semi amorphic, or crystalline forms. Crystalline cellulose has significantly better stiffness than all other constituents. Amorphous cellulose is more penetrable and accessible to enzymes and has a higher enzyme binding capacity than its crystalline counterpart so that it has a higher hydrolysis rate. The ratio of these two regions characterizes crystallinity. The cellulose with high crystallinity usually has low enzymatic hydrolysis efficiency [120][121][122]. Cellulose synthase (CESA) complexes (CSCs) synthesize cellulose microfibrils assembled in the Golgi apparatus. CSCs actively synthesize cellulose when delivered to the plasma membrane (PM). CSCs move along cortical microtubule paths that define cellulose microfibrils synthesis. CSC traffic between the PM and various intracellular compartments plays a vital role in determining the level of cellulose synthesized [123]. Uridine diphosphate glucose (Glc) originates from the cytosolic invertase/uridine diphosphate Glc phosphorylase pathway [124]. It is the substrate for CESAs [125]. The CESA proteins are involved in the dimerization/oligomerization of CESA subunits [126] due to a cytosolic N-terminal region. Moreover, the proteins consist of two transmembrane domains, a large cytoplasmic central loop that contains the substrate binding and catalytic regions, six additional transmembrane domains [127][128], and an intracellular C-terminal domain. The CSC rosettes contain 18 CESA subunits synthetizing 18 glucans [129].

Phytohormones may influence cell wall composition. For example, the amount of cellulose increases in methyl jasmonate, whereas lignin content decreases after salicylic acid application. The profile of gene expression involved in cell wall biosynthesis is also modified [130]. Phytohormone signaling pathways regulate stress perception at the primary cell wall, followed by cellulose synthesis and microtubule arrangement [119]. Alternatively, ROS and peroxidases arising in response to stresses in the cell wall may cross-link phenolic compounds and glycoproteins, resulting in stiffening. If ROS levels remain high during continued stress, OH<sup>•</sup>-radicals lead to polymer cleavage [118]. Cellulosomes could accomplish the same, an extracellular supramolecular multienzyme complex that can efficiently degrade cellulose and hemicelluloses in plant cell walls [131].

The role of cellulose in sensing stresses was demonstrated on mutant plants. Cellulose-deficient mutants are typically more sensitive to abiotic stress than wild-type plants [119]. Thus, the cellulose synthesis machinery may be vital in abiotic stress responses [119]. The other option is that some cell wall components may sense stresses via cell wall integrity [132]. The cell wall proteins, i.e., the *Arabidopsis thaliana* leucine-rich repeat receptor kinase LRR-RK male discoverer 1-interacting receptor-like kinase 2 (MIK2) was suggested to play such a role [133]. The MIK2 regulates cell wall damage responses upon cellulose biosynthesis inhibition due to abiotic stress [132]. MIK2 has overlapping functions with receptor-like protein kinase THESEUS 1 (THE1), also proposed as a cell wall integrity sensor. Moreover, plants may coordinate stress responses by integrating phytohormones (auxins, cytokinins, gibberellins, abscisic acid, ethylene, brassinosteroids, salicylic acid, jasmonates, and strigolactones) and their pathways [119][134][135].

Recent studies have demonstrated that cellulose forms participate in a signaling pathway that links cellulose and mitochondria. Two *cell wall maintainer1* (*cwm1*) and *cwm2* pentatricopeptide repeat protein genes were shown to be involved in editing mitochondrial transcripts encoding subunits of respiratory complexes (i.e., complex III linked to the maintenance of cell wall integrity under stress) and activation of retrograde mitochondrial signaling via ANAC017, a transcription factor participating in retrograde signaling to the nucleus upon mitochondrial dysfunction

[136][137]. A complex hierarchy of transcription factors exists downstream of ANAC017. These involve ANAC and WRKY transcription factors associated with organellar signaling and senescence; moreover, the network includes ethylene- and gibberellic acid-related transcription factors involved in stress responses [138].

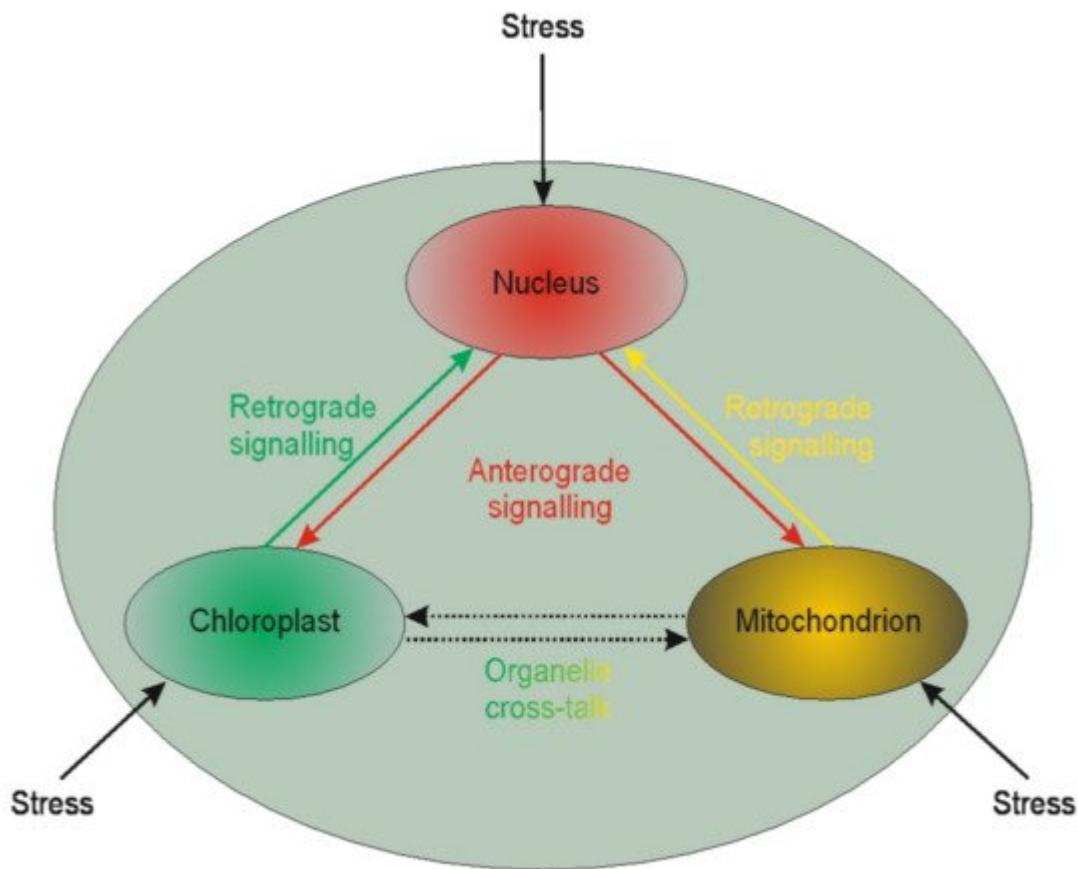
The development of plants requires the biosynthesis, deposition, and degradation of cell wall matrix polysaccharides. Biosynthesis of polysaccharides is performed in the endoplasmic reticulum and Golgi of the plant secretory system by polysaccharide glycosyltransferases (GTs) transmembrane proteins. The synthesis of the cell wall matrix glycans (pectins, xylans, xyloglucans, mannos, mixed-linkage glucans (MLGs), and arabinogalactan components of arabinogalactan proteins (AGPs) proteoglycans) was mapped to multigene families [139]. Different stresses can perturb targeted genes to promote functional adaptation [140]. The polysaccharide fraction of the cell wall may participate in stress sensing [141]. The environmental (mechanical) signals affect the cell wall determining its phenotypic features. However, the way stress is sensed and transmitted into the cell is still under investigation.

The plasma membrane was suggested to participate in signal transduction because of its position at the interface between the cell's interior and the cell wall. Thus, the crucial players that maintain cell status and respond to stresses are in the plasma membrane's same vicinity [142]. Interestingly, the layer between the plasma membrane and cell wall, the so-called subintinal layer present in microspores with embryogenic fate and composed of callose, is also considered essential in stress sensing. As callose is formed of glucans, it may serve as a source of glucose [143] required by glycolysis which impacts the Krebs cycle and the electron transport chain.

Cereal  $\beta$ -glucans have a specific combination of  $\beta$ -(1 $\rightarrow$ 4) and  $\beta$ -(1 $\rightarrow$ 3) linkages into linear long-chain polysaccharides of high molecular weight.  $\beta$ -glucans were identified in barley and oats [144]. Under elongated dark conditions,  $\beta$ -glucans deliver glucose for glycolysis, supporting biosynthesis of, i.e., lipids [145] that may act as signaling compounds [146][147], affecting gene expression [148][149]. Thus, the subintinal layer components are reasonable candidates for signals [150] of stresses in microspore culture. It was also demonstrated that  $\beta$ -glucans might impact DNA methylation of the CHG context in barley [67]. Whatever is the sensor of abiotic stresses at the cell wall level, the signal needs to be transmitted to the cell. It is being accepted that calcium is a second messenger that facilitates responses to stresses by activating calcium-binding proteins [151][152][153]. Another candidate that can sense stress and may act as a cell wall mechanism protecting plants from external stresses is  $\beta$ -1,3-linked D-glucose [119]. Furthermore, evidence shows that massively glycosylated hydroxyproline-rich proteins called AGP improve plant regeneration in barley anther culture [154]. The AGP content changes in *Brassica napus* are related to the developmental fate of microspores [155]. The proteins are present in the mucilage layer, the cell wall, and the cell membrane and participate in microspore fate change towards sporophytic pattern and may be involved in stress signaling [156].

## 4. Retrograde and Anterograde Signaling

It is becoming evident that nuclear genes' transcription regulation may be modulated by plastid gene expression [157][158][159]. Such regulation is called retrograde signaling. The reversed interaction when nucleus genes influence plastids' genome functioning is called anterograde [160] (Figure 1).



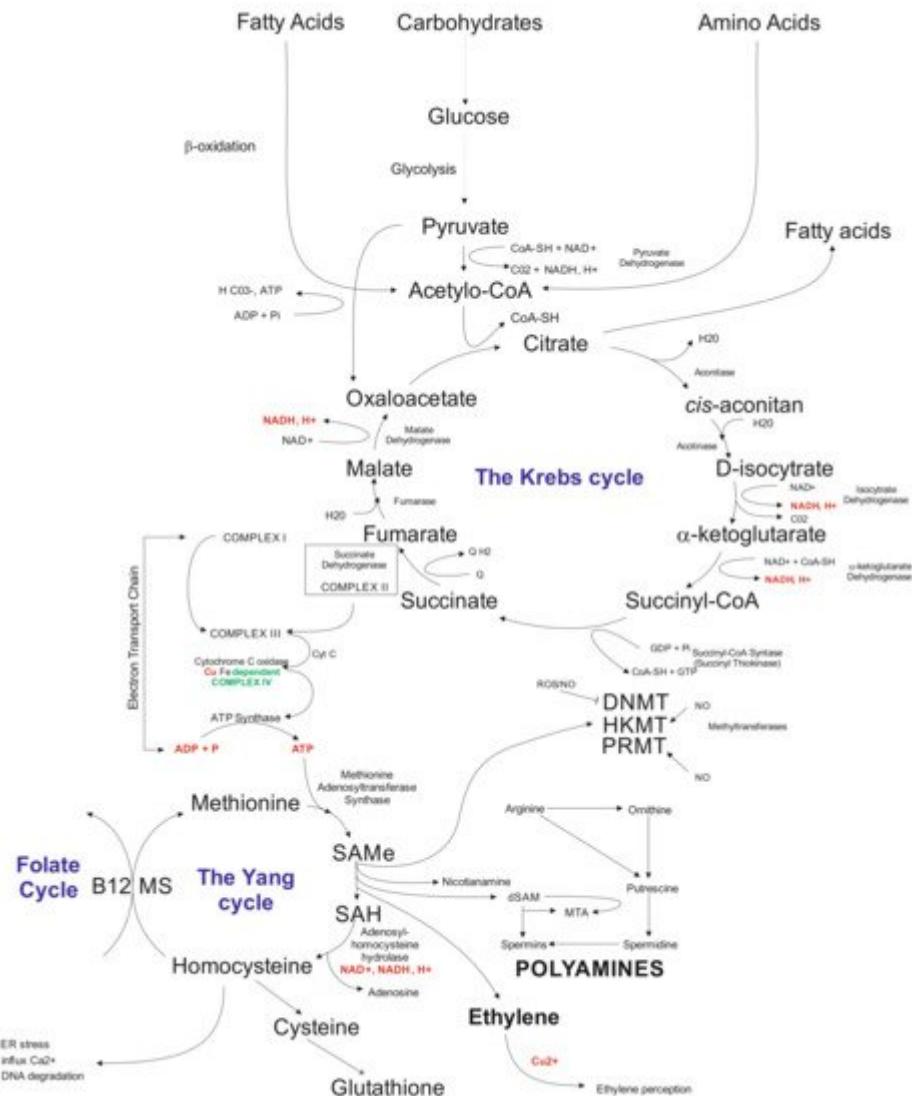
**Figure 1.** Schematic illustration of the interplay between cellular organelles. Anterograde is the signaling from the nucleus towards chloroplasts or mitochondria, whereas the reversed direction is called retrograde signaling. The communication between mitochondria and chloroplast (and other organelles except but nucleus) is called cross-talk.

Little is known of the two interactions in the context of microspores. The available literature is devoted chiefly to chloroplasts. Based on chloroplast biogenesis, retrograde signaling could be divided into “biogenic” and “operational” signals. The “biogenic signals” are the signals exhibited during early chloroplast development, whereas the “operational signals” are related to chloroplasts’ normal function in mature plants. Based on the signals’ origin, the retrograde signaling pathways in chloroplasts may include tetrapyrroles biosynthesis, redox state, chloroplast gene expression, reactive oxygen species, and protein import into plastids. The retrograde signaling pathway was described in barley mutant *albostrians*, which lacks plastid ribosomes and shows reduced amounts and/or activities of nuclear-encoded plastid proteins (the small subunit of ribulose-1,5-bisphosphate carboxylase/oxygenase (Rubisco), ferredoxin NADP<sup>+</sup> reductase, and enzymes of the Calvin cycle) [161]. Inhibition of tetrapyrrole biosynthesis and plastid gene expression activity resulted in the moderated expression of nuclear starch biosynthesis genes in tobacco [162]. It is well known that in vitro plant regeneration involves stressful conditions that induce chloroplast proteome remodeling. Nuclear factors regulate chloroplast gene expression, and this form of anterograde regulation has roles in plant adaptation to abiotic stress [163]. Furthermore, retrograde and anterograde signaling was demonstrated in barley in the case of albino plant regeneration via androgenesis [164] [165]. The other line of evidence suggests that retrograde mitochondrial signaling may also be essential for plants.

For instance, mitochondria have been proposed to play a vital role in programmed cell death (PCD) in anther tapetum cells. PCD is crucial during anther development providing lipids coating pollen exines [166]. The role of PCD in the tapetum was also demonstrated in sunflower [167]. However, it could not be excluded that such similar signaling may act for anther cultures. Thus, the retro and anterograde signaling pathways may affect organelle functioning's biochemical level, linked to epigenetic aspects of the in vitro tissue culture-induced variation.

## 5. Biochemical Aspects

Another layer that is being affected by in vitro tissue culture plant regeneration relates to biochemical pathways. The available data indicate that callose present in the subintinal layer [168] of microspores may affect the Krebs' cycle via complex II involved in the electron transport chain (ETC) [67]. Problems with ATP synthesis may disturb the Yang cycle [169], followed by DNA methylation problems [170] and induction of mutations [171], and possibly activation of transposable elements (TEs) [172]. Indirectly, the Yang cycle is responsible for spermine and spermidine synthesis [173]. The compounds may participate in gene expression regulation [174]. Furthermore, the Yang cycle is essential for the cell's glutathione synthesis as an antioxidant reagent preventing modification of, for example, methylated cytosines. It was documented that glutathione significantly improved plant regeneration via anther culture in the rye [100]. Possibly, that disturbances in the Krebs' cycle may change fatty acid synthesis (**Figure 2**). Fatty acids may influence, for example, gene transcription [175] and glycolysis. Furthermore, glycolysis could be disturbed under carbon starvation stress [176]. Thus, in vitro tissue culture plant regeneration is a complex, multidimensional process affecting all cell functioning levels. Understanding the relationships among all of the system's components is vital for the elaboration of knowledge-based approaches of in vitro plant regeneration and regulation of the levels of somaclonal variation.



**Figure 2.** Schematic illustration of biochemical cycles that may participate in the induction of in vitro tissue culture variation. Briefly, under cold treatment and darkness and maltose presence, carbon starvation stress results in dysfunction of glycolysis affecting the Krebs cycle. Disbalance in the Krebs cycle could be sensed at the electron transfer chain level via complex II and problems with ATP synthesis, which influence the Yang cycle. The latter is responsible for producing SAM involved in 80% of the cell compounds' methylation processes, including genomic cytosines. The Yang cycle's mal functioning may also influence glutathione production (antioxidant) used by the cell during oxidative stress. Oxidative stress may modify methylated cytosine, inducing point mutations. Furthermore, byproducts of the Yang cycle may regulate transcriptome as the result of abiotic stresses. Abbreviations: B12 (vitamin B12); CoA (Coenzyme A (acyl-CoA)); CoASH (coenzyme A not attached to acyl group); DNMT (DNA methyltransferase); dSAM (decarboxylated SAM); ER stress (endoplasmic reticulum stress); HKMT (histone lysine methyltransferase); PRMT; MS (methionine synthesis); MTA (5'-methyl thioadenosine); NAD (nicotinamide adenine dinucleotide); NADH (1,4-dihydro-nicotinamide adenine dinucleotide); PRMT (protein arginine N-methyltransferase); ROS (radical oxygen species); SAMe (S-adenosyl-L-methionine); SAH (S-adenosylhomocysteine).

## References

1. Orłowska, R.; Bednarek, P.T. Precise evaluation of tissue culture-induced variation during optimisation of in vitro regeneration regime in barley. *Plant Mol. Biol.* 2020, 103, 33–50.
2. Machczyńska, J.; Zimny, J.; Bednarek, P. Tissue culture-induced genetic and epigenetic variation in triticale ( $\times$  *Triticosecale* spp. Wittmack ex A. Camus 1927) regenerants. *Plant Mol. Biol.* 2015, 89, 279–292.
3. Mikuła, A.; Tomiczak, K.; Rybczynski, J.J. Cryopreservation enhances embryogenic capacity of *Gentiana cruciata* (L.) suspension culture and maintains (epi)genetic uniformity of regenerants. *Plant Cell Rep.* 2011, 30, 565–574.
4. Fiuk, A.; Bednarek, P.T.; Rybczyński, J.J. Flow Cytometry, HPLC-RP, and metAFLP Analyses to Assess Genetic Variability in Somatic Embryo-Derived Plantlets of *Gentiana pannonica* Scop. *Plant Mol. Biol. Rep.* 2010, 28, 413–420.
5. Agarwal, P.K.; Bhojwani, S.S. Genetic variability in the progeny of androgenic dihaploid plants and selection of high agronomic performing lines in *Brassica juncea*. *Biol. Plant.* 2004, 48, 503–508.
6. Zehr, B.E.; Williams, M.E.; Duncan, D.R.; Widholm, J.M. Somaclonal variation in the progeny of plants regenerated from callus cultures of seven inbred lines of maize. *Can. J. Bot.* 1987, 65, 491–499.
7. Ghosh, A.; Igamberdiev, A.U.; Debnath, S.C. Tissue culture-induced DNA methylation in crop plants: A review. *Mol. Biol. Rep.* 2021, 48, 823–841.
8. Cao, Z.; Sui, S.; Cai, X.; Yang, Q.; Deng, Z. Somaclonal variation in 'Red Flash' caladium: Morphological, cytological and molecular characterization. *Plant Cell Tissue Organ Cult.* 2016, 126, 269–279.
9. Deepthi, V.P. Somaclonal variation in micro propagated bananas. *Adv. Plants Agric. Res.* 2018, 8, 624–627.
10. Qin, Y.; Shin, K.-S.; Woo, H.-J.; Lim, M.-H. Genomic Variations of Rice Regenerants from Tissue Culture Revealed by Whole Genome Re-Sequencing. *Plant Breed. Biotechnol.* 2018, 6, 426–433.
11. Azizi, P.; Hanafi, M.M.; Sahebi, M.; Harikrishna, J.A.; Taheri, S.; Yassoralipour, A.; Nasehi, A. Epigenetic changes and their relationship to somaclonal variation: A need to monitor the micropropagation of plantation crops. *Funct. Plant Biol.* 2020, 47, 508–523.
12. Orłowska, R. Barley somatic embryogenesis-an attempt to modify variation induced in tissue culture. *J. Biol. Res. Thessalon.* 2021, 28, 9.
13. Skirvin, R.M.; Janick, J. Tissue culture-induced variation in scented *Pelargonium* spp. *J. Am. Soc. Hortic. Sci.* 1976, 101, 281–290.

14. Scowcroft, W.R. Somaclonal variation: The myth of clonal uniformity. In *Genetic Flux in Plants*; Hohn, B., Dennis, E.B., Eds.; Springer: Vienna, Austria, 1985; pp. 217–245.
15. Larkin, P.J.; Scowcroft, W.R. Somaclonal variation—A novel source of variability from cell cultures for plant improvement. *Theor. Appl. Genet.* 1981, **60**, 197–214.
16. Botstein, D.; White, R.L.; Skolnick, M.; Davis, R.W. Construction of a genetic linkage map in man using restriction fragment length polymorphisms. *Am. J. Hum. Genet.* 1980, **32**, 314–331.
17. Williams, J.G.; Kubelik, A.R.; Livak, K.J.; Rafalski, J.A.; Tingey, S.V. DNA polymorphisms amplified by arbitrary primers are useful as genetic markers. *Nucleic Acids Res.* 1990, **18**, 6531–6535.
18. Jeong, I.S.; Yoon, U.H.; Lee, G.S.; Ji, H.S.; Lee, H.J.; Han, C.D.; Hahn, J.H.; An, G.; Kim, T.H. SNP-based analysis of genetic diversity in anther-derived rice by whole genome sequencing. *Rice* 2013, **6**, 6.
19. Peschke, V.M.; Phillips, R.L.; Gengenbach, B.G. Discovery of transposable element activity among progeny of tissue-culture derived plants. *Science* 1987, **238**, 804–807.
20. Sato, M.; Hosokawa, M.; Doi, M. Somaclonal Variation Is Induced De Novo via the Tissue Culture Process: A Study Quantifying Mutated Cells in Saintpaulia. *PLoS ONE* 2011, **6**, e23541.
21. Barret, P.; Brinkman, M.; Beckert, M. A sequence related to rice Pong transposable element displays transcriptional activation by in vitro culture and reveals somaclonal variations in maize. *Genome* 2006, **49**, 1399–1407.
22. Tanurdzic, M.; Vaughn, M.W.; Jiang, H.; Lee, T.-J.; Slotkin, R.K.; Sosinski, B.; Thompson, W.F.; Doerge, R.W.; Martienssen, R.A. Epigenomic consequences of immortalized plant cell suspension culture. *PLoS Biol.* 2008, **6**, e302.
23. Kaepller, S.M.; Phillips, R.L.; Olhoft, P. Molecular basis of heritable tissue culture-induced variation in plants. In *Somaclonal Variation and Induced Mutation in Crop Improvement*; Jain, S.M., Brar, D.S., Ahloowalia, B.S., Eds.; Kluwer Academic: Dordrecht, The Netherlands, 1998; pp. 465–484.
24. Cassels, A.C.; Curry, R.F. Oxidative stress and physiological, epigenetic and genetic variability in plant tissue culture: Implications for micropropagators and genetic engineers. *Plant Cell Tissue Organ Cult.* 2001, **64**, 145–157.
25. Krishna, H.; Alizadeh, M.; Singh, D.; Singh, U.; Chauhan, N.; Eftekhari, M.; Sadh, R.K. Somaclonal variations and their applications in horticultural crops improvement. *3 Biotech* 2016, **6**, 54.
26. Dey, T.; Saha, S.; Ghosh, P.D. Somaclonal variation among somatic embryo derived plants—Evaluation of agronomically important somaclones and detection of genetic changes by RAPD in *Cymbopogon winterianus*. *S. Afr. J. Bot.* 2015, **96**, 112–121.

27. Touraev, A.; Indrianto, A.; Wratschko, I.; Vicente, O.; Heberle-Bors, E. Efficient microspore embryogenesis in wheat (*Triticum aestivum* L.) induced by starvation at high temperature. *Sex. Plant Reprod.* 1996, 9, 209–215.

28. Hoekstra, S.; Hoekstra, S.; Hoekstra, I.R.; Hoekstra, R.A.; Hoekstra, E. The Interaction of 2,4-D Application and Mannitol Pretreatment in Anther and Microspore Culture of *Hordeum vulgare* L. cv. Igri. *J. Plant Physiol.* 1996, 148, 696–700.

29. Binarova, P.; Hause, G.; Cenková, V.; Cordewener, J.H.; Campagne, M.L. A short severe heat shock is required to induce embryogenesis in late bicellular pollen of *Brassica napus* L. *Sex. Plant Reprod.* 1997, 10, 200–208.

30. Popova, T.; Grozeva, S.; Todorova, V.; Stankova, G.; Anachkov, N.; Rodeva, V. Effects of low temperature, genotype and culture media on in vitro androgenic answer of pepper (*Capsicum annuum* L.). *Acta Physiol. Plant.* 2016, 38, 273.

31. Tenhola-Roininen, T.; Tanhuanpää, P.; Immonen, S. The effect of cold and heat treatments on the anther culture response of diverse rye genotypes. *Euphytica* 2005, 145, 1–9.

32. Vos, P.; Hogers, R.; Bleeker, M.; Reijans, M.; van de Lee, T.; Hornes, M.; Frijters, A.; Pot, J.; Peleman, J.; Kuiper, M.; et al. AFLP: A new technique for DNA fingerprinting. *Nucleic Acids Res.* 1995, 23, 4407–4414.

33. Reyna-Lopez, G.E.; Simpson, J.; Ruiz-Herrera, J. Differences in DNA methylation patterns are detectable during the dimorphic transition of fungi by amplification of restriction polymorphisms. *Mol. Gen. Genet.* 1997, 253, 703–710.

34. Xiong, L.Z.; Xu, C.G.; Saghaf Maroof, M.A.; Zhang, Q. Patterns of cytosine methylation in an elite rice hybrid and its parental lines, detected by a methylation-sensitive amplification polymorphism technique. *Mol. Genet. Genom.* 1999, 261, 439–446.

35. Baranek, M.; Cechova, J.; Kovacs, T.; Eichmeier, A.; Wang, S.; Raddova, J.; Necas, T.; Ye, X. Use of Combined MSAP and NGS Techniques to Identify Differentially Methylated Regions in Somaclones: A Case Study of Two Stable Somatic Wheat Mutants. *PLoS ONE* 2016, 11, e0165749.

36. Francischini, J.H.M.B.; Kemper, E.L.; Costa, J.B.; Manechini, J.R.V.; Pinto, L.R. DNA methylation in sugarcane somaclonal variants assessed through methylation-sensitive amplified polymorphism. *Genet. Mol. Res.* 2017, 16.

37. Bobadilla Landey, R.; Cenci, A.; Georget, F.; Bertrand, B.; Camayo, G.; Dechamp, E.; Herrera, J.C.; Santoni, S.; Lashermes, P.; Simpson, J.; et al. High Genetic and Epigenetic Stability in *Coffea arabica* Plants Derived from Embryogenic Suspensions and Secondary Embryogenesis as Revealed by AFLP, MSAP and the Phenotypic Variation Rate. *PLoS ONE* 2013, 8, e56372.

38. Dann, A.L.; Wilson, C.R. Comparative assessment of genetic and epigenetic variation among regenerants of potato (*Solanum tuberosum*) derived from long-term nodal tissue-culture and cell selection. *Plant Cell Rep.* 2011, 30, 631–639.

39. Guevara, M.; de María, N.; Sáez-Laguna, E.; Vélez, M.D.; Cervera, M.T.; Cabezas, J.A. Analysis of DNA Cytosine Methylation Patterns Using Methylation-Sensitive Amplification Polymorphism (MSAP). *Methods Mol. Biol.* 2017, 1456, 99–112.

40. Fulneček, J.; Kovařík, A. How to interpret Methylation Sensitive Amplified Polymorphism (MSAP) profiles? *BMC Genet.* 2014, 15, 2.

41. Alonso, C.; Balao, F.; Bazaga, P.; Perez, R. Epigenetic contribution to successful polyploidizations: Variation in global cytosine methylation along an extensive ploidy series in *Dianthus broteri* (Caryophyllaceae). *New Phytol.* 2016, 212, 571–576.

42. Schulz, B.; Eckstein, R.L.; Durka, W. Scoring and analysis of methylation-sensitive amplification polymorphisms for epigenetic population studies. *Mol. Ecol. Resour.* 2013, 13, 642–653.

43. Bednarek, P.T.; Orłowska, R.; Niedziela, A. A relative quantitative Methylation-Sensitive Amplified Polymorphism (MSAP) method for the analysis of abiotic stress. *BMC Plant Biol.* 2017, 17, 79.

44. Machczyńska, J.; Orłowska, R.; Zimny, J.; Bednarek, P.T. Extended metAFLP approach in studies of the tissue culture induced variation (TCIV) in case of triticale. *Mol. Breed.* 2014, 34, 845–854.

45. Bednarek, P.T.; Orłowska, R. Time of In Vitro Anther Culture May Moderate Action of Copper and Silver Ions that Affect the Relationship between DNA Methylation Change and the Yield of Barley Green Regenerants. *Plants* 2020, 9, 1064.

46. Wang, S.; Lv, J.; Zhang, L.; Dou, J.; Sun, Y.; Li, X.; Fu, X.; Dou, H.; Mao, J.; Hu, X.; et al. MethylRAD: A simple and scalable method for genome-wide DNA methylation profiling using methylation-dependent restriction enzymes. *Open Biol.* 2015, 5, 150130.

47. Brunner, A.L.; Johnson, D.S.; Kim, S.W.; Valouev, A.; Reddy, T.E.; Neff, N.F.; Anton, E.; Medina, C.; Nguyen, L.; Chiao, E.; et al. Distinct DNA methylation patterns characterize differentiated human embryonic stem cells and developing human fetal liver. *Genome Res.* 2009, 19, 1044–1056.

48. Haque, N.; Nishiguchi, M. Bisulfite sequencing for cytosine-methylation analysis in plants. *Methods Mol. Biol.* 2011, 744, 187–197.

49. Zhang, Y.; Rohde, C.; Tierling, S.; Stamerjohanns, H.; Reinhardt, R.; Walter, J.; Jeltsch, A. DNA Methylation Analysis by Bisulfite Conversion, Cloning, and Sequencing of Individual Clones. In *DNA Methylation: Methods and Protocols*; Tost, J., Ed.; Humana Press: Totowa, NY, USA, 2009; pp. 177–187.

50. Sun, Q.; Qiao, J.; Zhang, S.; He, S.; Shi, Y.; Yuan, Y.; Zhang, X.; Cai, Y. Changes in DNA methylation assessed by genomic bisulfite sequencing suggest a role for DNA methylation in cotton fruiting branch development. *PeerJ* 2018, 6, e4945.

51. Bednarek, P.T.; Orłowska, R.; Koebner, R.M.D.; Zimny, J. Quantification of the tissue-culture induced variation in barley (*Hordeum vulgare* L.). *BMC Plant Biol.* 2007, 7, 10.

52. Orłowska, R.; Machczyńska, J.; Oleszczuk, S.; Zimny, J.; Bednarek, P.T. DNA methylation changes and TE activity induced in tissue cultures of barley (*Hordeum vulgare* L.). *J. Biol. Res.* 2016, 23, 19.

53. Machczyńska, J.; Orłowska, R.; Mańkowski, D.R.; Zimny, J.; Bednarek, P.T. DNA methylation changes in triticale due to in vitro culture plant regeneration and consecutive reproduction. *Plant Cell Tissue Organ Cult.* 2014, 119, 289–299.

54. Śliwińska, A.A.; Białek, A.; Orłowska, R.; Mańkowski, D.; Sykłowska-Baranek, K.; Pietrosiuk, A. Comparative Study of the Genetic and Biochemical Variability of *Polyscias filicifolia* (Araliaceae) Regenerants Obtained by Indirect and Direct Somatic Embryogenesis as a Source of Triterpenes. *Int. J. Mol. Sci.* 2021, 22, 5752.

55. Mikuła, A.; Tomiczak, K.; Wójcik, A.; Rybczynski, J.J. Encapsulation-dehydration method elevates embryogenic abilities of *Gentiana kurroo* cell suspension and carrying on genetic stability of its regenerants after cryopreservation. In *Acta Horticulturae*; International Society for Horticultural Science: Leuven, Belgium, 2011; Volume 908, pp. 143–154.

56. Oleszczuk, S.; Zimny, J.; Bednarek, P.T. The application of the AFLP method to determine the purity of homozygous lines of barley (*Hordeum vulgare* L.). *Cell. Mol. Biol. Lett.* 2002, 7, 777–783.

57. Coronel, C.J.; González, A.I.; Ruiz, M.L.; Polanco, C. Analysis of somaclonal variation in transgenic and regenerated plants of *Arabidopsis thaliana* using methylation related metAFLP and TMD markers. *Plant Cell Rep.* 2018, 37, 137–152.

58. Lukens, L.N.; Zhan, S. The plant genome's methylation status and response to stress: Implications for plant improvement. *Curr. Opin. Plant Biol.* 2007, 10, 317–322.

59. Jiang, C.; Mithani, A.; Gan, X.; Belfield, E.J.; Klingler, J.P.; Zhu, J.K.; Ragoussis, J.; Mott, R.; Harberd, N.P. Regenerant *Arabidopsis* lineages display a distinct genome-wide spectrum of mutations conferring variant phenotypes. *Curr. Biol.* 2011, 21, 1385–1390.

60. Orłowska, R.; Pachota, K.A.; Dynkowska, W.M.; Niedziela, A.; Bednarek, P.T. Androgenic-Induced Transposable Elements Dependent Sequence Variation in Barley. *Int. J. Mol. Sci.* 2021, 22, 6783.

61. Baker, M.J.; Trevisan, J.; Bassan, P.; Bhargava, R.; Butler, H.J.; Dorling, K.M.; Fielden, P.R.; Fogarty, S.W.; Fullwood, N.J.; Heys, K.A.; et al. Using Fourier transform IR spectroscopy to analyze biological materials. *Nat. Protoc.* 2014, 9, 1771–1791.

62. Kumar, S.; Lahlali, R.; Liu, X.; Karunakaran, C. Infrared spectroscopy combined with imaging: A new developing analytical tool in health and plant science. *Appl. Spectrosc. Rev.* **2016**, *51*, 466–483.

63. Kačuráková, M.; Čapek, P.; Sasinková, V.; Wellner, N.; Ebringerová, A. FT-IR study of plant cell wall model compounds: Pectic polysaccharides and hemicelluloses. *Carbohydr. Polym.* **2000**, *43*, 195–203.

64. Kazarian, S.G.; Chan, K.L.A. ATR-FTIR spectroscopic imaging: Recent advances and applications to biological systems. *Analyst* **2013**, *138*, 1940–1951.

65. Synytsya, A.; Novak, M. Structural analysis of glucans. *Ann. Transl. Med.* **2014**, *2*, 17.

66. Lou, Y.; Zhu, J.; Yang, Z. Molecular Cell Biology of Pollen Walls. In *Applied Plant Cell Biology: Cellular Tools and Approaches for Plant Biotechnology*; Nick, P., Opatrny, Z., Eds.; Springer: Berlin/Heidelberg, Germany, 2014; pp. 179–205.

67. Bednarek, P.T.; Zebrowski, J.; Orłowska, R. Exploring the Biochemical Origin of DNA Sequence Variation in Barley Plants Regenerated via in Vitro Anther Culture. *Int. J. Mol. Sci.* **2020**, *21*, 5770.

68. Bednarek, P.T.; Orłowska, R. CG Demethylation Leads to Sequence Mutations in an Anther Culture of Barley Due to the Presence of Cu, Ag Ions in the Medium and Culture Time. *Int. J. Mol. Sci.* **2020**, *21*, 4401.

69. Hayes, A.F. *Introduction to Mediation, Moderation, and Conditional Process Analysis. A Regression Bases Approach*; A Division of Guilford Publications, Inc.: New York, NY, USA, 2018; p. 507.

70. Maraschin, S.F.; de Priester, W.; Spalink, H.P.; Wang, M. Androgenic switch: An example of plant embryogenesis from the male gametophyte perspective. *J. Exp. Bot.* **2005**, *56*, 1711–1726.

71. Galán-Ávila, A.; García-Forteá, E.; Prohens, J.; Herraiz, F.J. Microgametophyte Development in *Cannabis sativa* L. and First Androgenesis Induction Through Microspore Embryogenesis. *Front. Plant Sci.* **2021**, *12*, 669424.

72. Testillano, P.S. Microspore embryogenesis: Targeting the determinant factors of stress-induced cell reprogramming for crop improvement. *J. Exp. Bot.* **2019**, *70*, 2965–2978.

73. Seguí-Simarro, J.M.; Bárány, I.; Suárez, R.; Fadón, B.; Testillano, P.S.; Risueño, M.C. Nuclear bodies domain changes with microspore reprogramming to embryogenesis. *Eur. J. Histochem.* **2006**, *50*, 35–44.

74. Wani, S.H.; Kumar, V.; Shriram, V.; Sah, S.K. Phytohormones and their metabolic engineering for abiotic stress tolerance in crop plants. *Crop J.* **2016**, *4*, 162–176.

75. Rodríguez-Serrano, M.; Bárány, I.; Prem, D.; Coronado, M.-J.; Risueño, M.C.; Testillano, P.S. NO, ROS, and cell death associated with caspase-like activity increase in stress-induced microspore

embryogenesis of barley. *J. Exp. Bot.* 2012, 63, 2007–2024.

76. Pasternak, T. Oxidative stress inducing agents' copper and alloxan accelerate cell cycle re-entering of somatic plant cells in the presence of suboptimal exogenous auxin. *bioRxiv* 2020. preprint.

77. Uváčková, L.; Takáč, T.; Boehm, N.; Obert, B.; Samaj, J. Proteomic and biochemical analysis of maize anthers after cold pretreatment and induction of androgenesis reveals an important role of anti-oxidative enzymes. *J. Proteom.* 2012, 75, 1886–1894.

78. Da Costa, C.; De Almeida, M.; Ruedell, C.; Schwambach, J.; Maraschin, F.; Fett-Neto, A. When stress and development go hand in hand: Main hormonal controls of adventitious rooting in cuttings. *Front. Plant Sci.* 2013, 4, 133.

79. Fehér, A. Somatic embryogenesis—Stress-induced remodeling of plant cell fate. *Biochim. Biophys. Acta* 2015, 1849, 385–402.

80. Pasternak, T.P.; Prinsen, E.; Ayaydin, F.; Miskolczi, P.I.; Potters, G.; Asard, H.; Van Onckelen, H.A.; Dudits, D.n.; Fehér, A. The Role of Auxin, pH, and Stress in the Activation of Embryogenic Cell Division in Leaf Protoplast-Derived Cells of Alfalfa. *Plant Physiol.* 2002, 129, 1807–1819.

81. Pasternak, T.P.; Ötvös, K.; Domoki, M.; Fehér, A. Linked activation of cell division and oxidative stress defense in alfalfa leaf protoplast-derived cells is dependent on exogenous auxin. *Plant Growth Regul.* 2007, 51, 109–117.

82. Rodríguez-Sanz, H.; Solís, M.T.; López, M.F.; Gómez-Cadenas, A.; Risueño, M.C.; Testillano, P.S. Auxin Biosynthesis, Accumulation, Action and Transport are Involved in Stress-Induced Microspore Embryogenesis Initiation and Progression in *Brassica napus*. *Plant Cell Physiol.* 2015, 56, 1401–1417.

83. Weijers, D.; Nemhauser, J.; Yang, Z. Auxin: Small molecule, big impact. *J. Exp. Bot.* 2018, 69, 133–136.

84. Li, S.-B.; Xie, Z.-Z.; Hu, C.-G.; Zhang, J.-Z. A Review of Auxin Response Factors (ARFs) in Plants. *Front. Plant Sci.* 2016, 7, 47.

85. Mironova, V.; Teale, W.; Shahriari, M.; Dawson, J.; Palme, K. The Systems Biology of Auxin in Developing Embryos. *Trends Plant Sci.* 2017, 22, 225–235.

86. Su, Y.H.; Liu, Y.B.; Bai, B.; Zhang, X.S. Establishment of embryonic shoot–root axis is involved in auxin and cytokinin response during *Arabidopsis* somatic embryogenesis. *Front. Plant Sci.* 2015, 5, 792.

87. Hofius, D.; Li, L.; Hafrén, A.; Coll, N.S. Autophagy as an emerging arena for plant-pathogen interactions. *Curr. Opin. Plant Biol.* 2017, 38, 117–123.

88. Masclaux-Daubresse, C.; Chen, Q.; Havé, M. Regulation of nutrient recycling via autophagy. *Curr. Opin. Plant Biol.* 2017, 39, 8–17.

89. Avin-Wittenberg, T.; Baluška, F.; Bozhkov, P.V.; Elander, P.H.; Fernie, A.R.; Galili, G.; Hassan, A.; Hofius, D.; Isono, E.; Le Bars, R.; et al. Autophagy-related approaches for improving nutrient use efficiency and crop yield protection. *J. Experimrntal Bot.* 2018, 69, 1335–1353.

90. Corral-Martínez, P.; Parra-Vega, V.; Seguí-Simarro, J.M. Novel features of *Brassica napus* embryogenic microspores revealed by high pressure freezing and freeze substitution: Evidence for massive autophagy and excretion-based cytoplasmic cleaning. *J. Exp. Bot.* 2013, 64, 3061–3075.

91. Michaeli, S.; Avin-Wittenberg, T.; Galili, G. Involvement of autophagy in the direct ER to vacuole protein trafficking route in plants. *Front. Plant Sci.* 2014, 5, 134.

92. Tan, X.; Li, K.; Wang, Z.; Zhu, K.; Tan, X.; Cao, J. A Review of Plant Vacuoles: Formation, Located Proteins, and Functions. *Plants* 2019, 8, 327.

93. Pérez-Pérez, M.E.; Lemaire, S.D.; Crespo, J.L. Reactive Oxygen Species and Autophagy in Plants and Algae. *Plant Physiol.* 2012, 160, 156–164.

94. Bárány, I.; Berenguer, E.; Solís, M.T.; Pérez-Pérez, Y.; Santamaría, M.E.; Crespo, J.L.; Risueño, M.C.; Díaz, I.; Testillano, P.S. Autophagy is activated and involved in cell death with participation of cathepsins during stress-induced microspore embryogenesis in barley. *J. Exp. Bot.* 2018, 69, 1387–1402.

95. Tsatsiani, L.; Van Breusegem, F.; Gallois, P.; Zavialov, A.; Lam, E.; Bozhkov, P.V. Metacaspases. *Cell Death Differ.* 2011, 18, 1279–1288.

96. Berenguer, E.; Bárány, I.; Solís, M.-T.; Pérez-Pérez, Y.; Risueño, M.C.; Testillano, P.S. Inhibition of Histone H3K9 Methylation by BIX-01294 Promotes Stress-Induced Microspore Totipotency and Enhances Embryogenesis Initiation. *Front. Plant Sci.* 2017, 8, 161.

97. Solís, M.-T.; El-Tantawy, A.-A.; Cano, V.; Risueño, M.C.; Testillano, P.S. 5-azacytidine promotes microspore embryogenesis initiation by decreasing global DNA methylation, but prevents subsequent embryo development in rapeseed and barley. *Front. Plant Sci.* 2015, 6, 472.

98. El-Tantawy, A.A.; Solís, M.T.; Risueño, M.C.; Testillano, P.S. Changes in DNA Methylation Levels and Nuclear Distribution Patterns after Microspore Reprogramming to Embryogenesis in Barley. *Cytogenet. Genome Res.* 2014, 143, 200–208.

99. Çakmak, E.; Uncuoğlu, A.A.; Aydın, Y. Evaluation of in vitro genotoxic effects induced by in vitro anther culture conditions in sunflower. *Plant Signal. Behav.* 2019, 14, 1633885.

100. Zieliński, K.; Krzewska, M.; Żur, I.; Juzoń, K.; Kopeć, P.; Nowicka, A.; Moravčíková, J.; Skrzypek, E.; Dubas, E. The effect of glutathione and mannitol on androgenesis in anther and isolated

microspore cultures of rye (*Secale cereale* L.). *Plant Cell Tissue Organ Cult.* 2020, 140, 577–592.

101. Castander-Olarieta, A.; Montalbán, I.A.; De Medeiros Oliveira, E.; Dell'Aversana, E.; D'Amelia, L.; Carillo, P.; Steiner, N.; Fraga, H.P.D.F.; Guerra, M.P.; Goicoa, T.; et al. Effect of Thermal Stress on Tissue Ultrastructure and Metabolite Profiles During Initiation of Radiata Pine Somatic Embryogenesis. *Front. Plant Sci.* 2019, 9, 4.

102. Gamalero, E.; Glick, B.R. Ethylene and Abiotic Stress Tolerance in Plants. In *Environmental Adaptations and Stress Tolerance of Plants in the Era of Climate Change*; Ahmad, P., Prasad, M.N.V., Eds.; Springer New York: New York, NY, USA, 2012; pp. 395–412.

103. Klay, I.; Pirrello, J.; Riahi, L.; Bernadac, A.; Cherif, A.; Bouzayen, M.; Bouzid, S. Ethylene response factor SI-ERF B. 3 is responsive to abiotic stresses and mediates salt and cold stress response regulation in tomato. *Sci. World J.* 2014, 2014, 167681.

104. Mu, C.; Wang, S.; Zhang, S.; Pan, J.; Chen, N.; Li, X.; Wang, Z.; Liu, H. Small heat shock protein LimHSP16.45 protects pollen mother cells and tapetal cells against extreme temperatures during late zygotene to pachytene stages of meiotic prophase I in David Lily. *Plant Cell Rep.* 2011, 30, 1981.

105. Orłowska, R.; Pachota, K.A.; Machczyńska, J.; Niedziela, A.; Makowska, K.; Zimny, J.; Bednarek, P.T. Improvement of anther cultures conditions using the Taguchi method in three cereal crops. *Electron. J. Biotechnol.* 2020, 43, 8–15.

106. Vaahtera, L.; Schulz, J.; Hamann, T. Cell wall integrity maintenance during plant development and interaction with the environment. *Nat. Plants* 2019, 5, 924–932.

107. De Lorenzo, G.; Ferrari, S.; Giovannoni, M.; Mattei, B.; Cervone, F. Cell wall traits that influence plant development, immunity, and bioconversion. *Plant J.* 2019, 97, 134–147.

108. Ariizumi, T.; Toriyama, K. Genetic regulation of sporopollenin synthesis and pollen exine development. *Annu. Rev. Plant Biol.* 2011, 62, 437–460.

109. Quilichini, T.D.; Grienberger, E.; Douglas, C.J. The biosynthesis, composition and assembly of the outer pollen wall: A tough case to crack. *Phytochemistry* 2015, 113, 170–182.

110. Heslop-Harrison, J. Pollen wall development. *Science* 1968, 161, 230–237.

111. Blackmore, S.; Wortley, A.H.; Skvarla, J.J.; Rowley, J.R. Pollen wall development in flowering plants. *New Phytol.* 2007, 174, 483–498.

112. Ahlers, F.; Lambert, J.; Wiermann, R. Acetylation and silylation of piperidine solubilized sporopollenin from pollen of *Typha angustifolia* L. *Z. Für Nat. C* 2003, 58, 807–811.

113. Blokker, P.; Boelen, P.; Broekman, R.; Rozema, J. The occurrence of p-coumaric acid and ferulic acid in fossil plant materials and their use as UV-proxy. In *Plants and Climate Change*; Springer: Berlin/Heidelberg, Germany, 2006; pp. 197–208.

114. Bubert, H.; Lambert, J.; Steuernagel, S.; Ahlers, F.; Wiermann, R. Continuous decomposition of sporopollenin from pollen of *Typha angustifolia* L. by acidic methanolysis. *Verl. Der Z. Für Nat.* 2002, 57, 1035–1041.

115. Li, W.L.; Liu, Y.; Douglas, C.J. Role of Glycosyltransferases in Pollen Wall Primexine Formation and Exine Patterning. *Plant Physiol.* 2017, 173, 167–182.

116. Heslop-Harrison, J. Wall development within the microspore tetrad of *Lilium longiflorum*. *Can. J. Bot.* 1968, 46, 1185–1192.

117. Quilichini, T.D.; Douglas, C.J.; Samuels, A.L. New views of tapetum ultrastructure and pollen exine development in *Arabidopsis thaliana*. *Ann. Bot.* 2014, 114, 1189–1201.

118. Tenhaken, R. Cell wall remodeling under abiotic stress. *Front. Plant Sci.* 2015, 5, 771.

119. Kesten, C.; Menna, A.; Sánchez-Rodríguez, C. Regulation of cellulose synthesis in response to stress. *Curr. Opin. Plant Biol.* 2017, 40, 106–113.

120. Lampugnani, E.R.; Khan, G.A.; Somssich, M.; Persson, S. Building a plant cell wall at a glance. *J. Cell Sci.* 2018, 131.

121. Majda, M.; Robert, S. The role of auxin in cell wall expansion. *Int. J. Mol. Sci.* 2018, 19, 951.

122. Saffer, A.M. Expanding roles for pectins in plant development. *J. Integr. Plant Biol.* 2018, 60, 910–923.

123. Polko, J.K.; Kieber, J.J. The Regulation of Cellulose Biosynthesis in Plants. *Plant Cell* 2019, 31, 282–296.

124. Barnes, W.J.; Anderson, C.T. Cytosolic invertases contribute to cellulose biosynthesis and influence carbon partitioning in seedlings of *Arabidopsis thaliana*. *Plant J.* 2018, 94, 956–974.

125. Verbančić, J.; Lunn, J.E.; Stitt, M.; Persson, S. Carbon supply and the regulation of cell wall synthesis. *Mol. Plant* 2018, 11, 75–94.

126. Kurek, I.; Kawagoe, Y.; Jacob-Wilk, D.; Doblin, M.; Delmer, D. Dimerization of cotton fiber cellulose synthase catalytic subunits occurs via oxidation of the zinc-binding domains. *Proc. Natl. Acad. Sci. USA* 2002, 99, 11109–11114.

127. Sethaphong, L.; Haigler, C.H.; Kubicki, J.D.; Zimmer, J.; Bonetta, D.; DeBolt, S.; Yingling, Y.G. Tertiary model of a plant cellulose synthase. *Proc. Natl. Acad. Sci. USA* 2013, 110, 7512–7517.

128. Slabaugh, E.; Davis, J.K.; Haigler, C.H.; Yingling, Y.G.; Zimmer, J. Cellulose synthases: New insights from crystallography and modeling. *Trends Plant Sci.* 2014, 19, 99–106.

129. Hill, J.L.; Hammudi, M.B.; Tien, M. The *Arabidopsis* cellulose synthase complex: A proposed hexamer of CESA trimers in an equimolar stoichiometry. *Plant Cell* 2014, 26, 4834–4842.

130. Napoleão, T.A.; Soares, G.; Vital, C.E.; Bastos, C.; Castro, R.; Loureiro, M.E.; Giordano, A. Methyl jasmonate and salicylic acid are able to modify cell wall but only salicylic acid alters biomass digestibility in the model grass *Brachypodium distachyon*. *Plant Sci.* 2017, 263, 46–54.

131. Wang, Y.; Leng, L.; Islam, M.K.; Liu, F.; Lin, C.S.K.; Leu, S.-Y. Substrate-Related Factors Affecting Cellulosome-Induced Hydrolysis for Lignocellulose Valorization. *Int. J. Mol. Sci.* 2019, 20, 3354.

132. Van der Does, D.; Boutrot, F.; Engelsdorf, T.; Rhodes, J.; McKenna, J.F.; Vernhettes, S.; Koevoets, I.; Tintor, N.; Veerabagu, M.; Miedes, E.; et al. The *Arabidopsis* leucine-rich repeat receptor kinase MIK2/LRR-KISS connects cell wall integrity sensing, root growth and response to abiotic and biotic stresses. *PLoS Genet.* 2017, 13, e1006832.

133. Humphrey, T.V.; Bonetta, D.T.; Goring, D.R. Sentinels at the wall: Cell wall receptors and sensors. *New Phytol.* 2007, 176, 7–21.

134. Kafkaletoú, M.; Fasseas, C.; Tsantili, E. Increased firmness and modified cell wall composition by ethylene were reversed by the ethylene inhibitor 1-methylcyclopropene (1-MCP) in the non-climacteric olives harvested at dark green stage—Possible implementation of ethylene for olive quality. *J. Plant Physiol.* 2019, 238, 63–71.

135. Salazar, R.; Pollmann, S.; Morales-Quintana, L.; Herrera, R.; Caparrós-Ruiz, D.; Ramos, P. In seedlings of *Pinus radiata*, jasmonic acid and auxin are differentially distributed on opposite sides of tilted stems affecting lignin monomer biosynthesis and composition. *Plant Physiol. Biochem.* 2019, 135, 215–223.

136. Hu, Z.; Vanderhaeghen, R.; Cools, T.; Wang, Y.; De Clercq, I.; Leroux, O.; Nguyen, L.; Belt, K.; Millar, A.H.; Audenaert, D.; et al. Mitochondrial Defects Confer Tolerance against Cellulose Deficiency. *Plant Cell* 2016, 28, 2276–2290.

137. Hofmann, N.R. A Functional Link between Mitochondria and the Cell Wall in Stress Responses. *Plant Cell* 2016, 28, 1996.

138. Meng, X.; Li, L.; De Clercq, I.; Narsai, R.; Xu, Y.; Hartmann, A.; Claros, D.L.; Custovic, E.; Lewsey, M.G.; Whelan, J.; et al. ANAC017 Coordinates Organellar Functions and Stress Responses by Reprogramming Retrograde Signaling. *Plant Physiol.* 2019, 180, 634–653.

139. Amos, R.A.; Mohnen, D. Critical Review of Plant Cell Wall Matrix Polysaccharide Glycosyltransferase Activities Verified by Heterologous Protein Expression. *Front. Plant Sci.* 2019, 10, 15.

140. Taylor-Teeple, M.; Lin, L.; de Lucas, M.; Turco, G.; Toal, T.W.; Gaudinier, A.; Young, N.F.; Trabucco, G.M.; Veling, M.T.; Lamothe, R.; et al. An *Arabidopsis* gene regulatory network for secondary cell wall synthesis. *Nature* 2015, 517, 571–575.

141. Uluisik, S.; Seymour, G.B. Pectate lyases: Their role in plants and importance in fruit ripening. *Food Chem.* 2020, 309, 125559.

142. Ackermann, F.; Stanislas, T. The Plasma Membrane—An Integrating Compartment for Mechano-Signaling. *Plants* **2020**, *9*, 505.

143. Burton, R.A.; Fincher, G.B. (1,3;1,4)- $\beta$ -D-Glucans in Cell Walls of the Poaceae, Lower Plants, and Fungi: A Tale of Two Linkages. *Mol. Plant* **2009**, *2*, 873–882.

144. Marković, S.M.; Đukić, N.H.; Knežević, D.; Leković, S.V. Divergence of barley and oat varieties according to their content of  $\beta$ -glucan. *J. Serb. Chem. Soc.* **2017**, *82*, 379–388.

145. Zhai, Z.; Liu, H.; Xu, C.; Shanklin, J. Sugar Potentiation of Fatty Acid and Triacylglycerol Accumulation. *Plant Physiol.* **2017**, *175*, 696–707.

146. Funck, D.; Winter, G.; Baumgarten, L.; Forlani, G. Requirement of proline synthesis during *Arabidopsis* reproductive development. *BMC Plant Biol.* **2012**, *12*, 191.

147. Price, J.; Laxmi, A.; St Martin, S.K.; Jang, J.-C. Global transcription profiling reveals multiple sugar signal transduction mechanisms in *Arabidopsis*. *Plant Cell* **2004**, *16*, 2128–2150.

148. Jang, J.C.; Sheen, J. Sugar sensing in higher plants. *Plant Cell* **1994**, *6*, 1665–1679.

149. Pego, J.V.; Smeekens, S.C. Plant fructokinases: A sweet family get-together. *Trends Plant Sci.* **2000**, *5*, 531–536.

150. Díaz-Sala, C. Molecular Dissection of the Regenerative Capacity of Forest Tree Species: Special Focus on Conifers. *Front. Plant Sci.* **2019**, *9*, 1943.

151. Gravino, M.; Savatin, D.V.; Macone, A.; De Lorenzo, G. Ethylene production in *Botrytis cinerea*- and oligogalacturonide-induced immunity requires calcium-dependent protein kinases. *Plant J.* **2015**, *84*, 1073–1086.

152. Wang, J.; Tergel, T.; Chen, J.; Yang, J.; Kang, Y.; Qi, Z. *Arabidopsis* transcriptional response to extracellular Ca<sup>2+</sup> depletion involves a transient rise in cytosolic Ca<sup>2+</sup>. *J. Integr. Plant Biol.* **2015**, *57*, 138–150.

153. Knight, H. Calcium Signaling during Abiotic Stress in Plants. In *International Review of Cytology*; Jeon, K.W., Ed.; Academic Press: Cambridge, MA, USA, 1999; Volume 195, pp. 269–324.

154. Makowska, K.; Kałużniak, M.; Oleszczuk, S.; Zimny, J.; Czaplicki, A.; Konieczny, R. Arabinogalactan proteins improve plant regeneration in barley (*Hordeum vulgare* L.) anther culture. *Plant Cell Tissue Organ Cult.* **2017**, *131*, 247–257.

155. Corral-Martínez, P.; Driouich, A.; Seguí-Simarro, J.M. Dynamic Changes in Arabinogalactan-Protein, Pectin, Xyloglucan and Xylan Composition of the Cell Wall During Microspore Embryogenesis in *Brassica napus*. *Front. Plant Sci.* **2019**, *10*, 332.

156. El-Tantawy, A.-A.; Solís, M.-T.; Da Costa, M.L.; Coimbra, S.; Risueño, M.-C.; Testillano, P.S. Arabinogalactan protein profiles and distribution patterns during microspore embryogenesis and

pollen development in *Brassica napus*. *Plant Reprod.* 2013, 26, 231–243.

157. Hernández-Verdeja, T.; Strand, Å. Retrograde Signals Navigate the Path to Chloroplast Development. *Plant Physiol.* 2017, 176, 967–976.

158. Singh, R.; Singh, S.; Parihar, P.; Singh, V.P.; Prasad, S.M. Retrograde signaling between plastid and nucleus: A review. *J. Plant Physiol.* 2015, 181, 55–66.

159. Pfalz, J.; Oelmüller, R. Plastid Retrograde Signals: More to Discover. In *Sensory Biology of Plants*; Sopory, S., Ed.; Springer: Singapore, 2019; pp. 477–507.

160. Fujii, S.; Toriyama, K. Genome Barriers between Nuclei and Mitochondria Exemplified by Cytoplasmic Male Sterility. *Plant Cell Physiol.* 2008, 49, 1484–1494.

161. Börner, T. The discovery of plastid-to-nucleus retrograde signaling—a personal perspective. *Protoplasma* 2017, 254, 1845–1855.

162. Enami, K.; Ozawa, T.; Motohashi, N.; Nakamura, M.; Tanaka, K.; Hanaoka, M. Plastid-to-Nucleus Retrograde Signals Are Essential for the Expression of Nuclear Starch Biosynthesis Genes during Amyloplast Differentiation in Tobacco BY-2 Cultured Cells. *Plant Physiol.* 2011, 157, 518–530.

163. Watson, S.J.; Sowden, R.G.; Jarvis, P. Abiotic stress-induced chloroplast proteome remodelling: A mechanistic overview. *J. Exp. Bot.* 2018, 69, 2773–2781.

164. Bélanger, S.; Marchand, S.; Jacques, P.-É.; Meyers, B.; Belzile, F. Differential Expression Profiling of Microspores During the Early Stages of Isolated Microspore Culture Using the Responsive Barley Cultivar Gobernadora. *G3* 2018, 8, 1603–1614.

165. Gajecka, M.; Marzec, M.; Chmielewska, B.; Jelonek, J.; Zbieszczak, J.; Szarejko, I. Plastid differentiation during microgametogenesis determines green plant regeneration in barley microspore culture. *Plant Sci.* 2020, 291, 110321.

166. Kawanabe, T.; Ariizumi, T.; Kawai-Yamada, M.; Uchimiya, H.; Toriyama, K. Abolition of the tapetum suicide program ruins microsporogenesis. *Plant Cell Physiol.* 2006, 47, 784–787.

167. Balk, J.; Leaver, C.J. The PET1-CMS mitochondrial mutation in sunflower is associated with premature programmed cell death and cytochrome c release. *Plant Cell* 2001, 13, 1803–1818.

168. Rivas-Sendra, A.; Corral-Martínez, P.; Porcel, R.; Camacho-Fernández, C.; Calabuig-Serna, A.; Seguí-Simarro, J.M. Embryogenic competence of microspores is associated with their ability to form a callosic, osmoprotective subtintinal layer. *J. Exp. Bot.* 2019, 70, 1267–1281.

169. Sauter, M.; Moffatt, B.; Saechao, M.C.; Hell, R.; Wirtz, M. Methionine salvage and S-adenosylmethionine: Essential links between sulfur, ethylene and polyamine biosynthesis. *Biochem. J.* 2013, 451, 145–154.

170. Moffatt, B.A.; Weretilnyk, E.A. Sustaining S-adenosyl-L-methionine-dependent methyltransferase activity in plant cells. *Physiol. Plant.* 2001, 113, 435–442.

171. Yoshida, T.; Furihata, H.Y.; To, T.K.; Kakutani, T.; Kawabe, A. Genome defense against integrated organellar DNA fragments from plastids into plant nuclear genomes through DNA methylation. *Sci. Rep.* 2019, 9, 2060.

172. Ong-Abdullah, M.; Ordway, J.M.; Jiang, N.; Ooi, S.-E.; Kok, S.-Y.; Sarpan, N.; Azimi, N.; Hashim, A.T.; Ishak, Z.; Rosli, S.K.; et al. Loss of Karma transposon methylation underlies the mantled somaclonal variant of oil palm. *Nature* 2015, 525, 533.

173. Zierer, W.; Hajirezaei, M.R.; Eggert, K.; Sauer, N.; von Wirén, N.; Pommerrenig, B. Phloem-Specific Methionine Recycling Fuels Polyamine Biosynthesis in a Sulfur-Dependent Manner and Promotes Flower and Seed Development. *Plant Physiol.* 2016, 170, 790–806.

174. Chen, D.; Shao, Q.; Yin, L.; Younis, A.; Zheng, B. Polyamine Function in Plants: Metabolism, Regulation on Development, and Roles in Abiotic Stress Responses. *Front. Plant Sci.* 2019, 9, 1945.

175. Shang, B.; Xu, C.; Zhang, X.; Cao, H.; Xin, W.; Hu, Y. Very-long-chain fatty acids restrict regeneration capacity by confining pericycle competence for callus formation in *Arabidopsis*. *Proc. Natl. Acad. Sci. USA* 2016, 113, 5101–5106.

176. Contento, A.L.; Kim, S.-J.; Bassham, D.C. Transcriptome Profiling of the Response of *Arabidopsis* Suspension Culture Cells to Suc Starvation. *Plant Physiol.* 2004, 135, 2330–2347.

Retrieved from <https://encyclopedia.pub/entry/history/show/30203>