

Hallmarks of Aging in Macrophages

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The skin is our largest organ and the outermost protective barrier. Its aging reflects both intrinsic and extrinsic processes resulting from the constant insults it is exposed to. Aging in the skin is accompanied by specific epigenetic modifications, accumulation of senescent cells, reduced cellular proliferation/tissue renewal, altered extracellular matrix, and a proinflammatory environment favoring undesirable conditions, including disease onset. Macrophages (M ϕ) are the most abundant immune cell type in the skin and comprise a group of heterogeneous and plastic cells that are key for skin homeostasis and host defense. However, they have also been implicated in orchestrating chronic inflammation during aging. Since M ϕ are related to innate and adaptive immunity, it is possible that age-modified skin M ϕ promote adaptive immunity exacerbation and exhaustion, favoring the emergence of proinflammatory pathologies, such as skin cancer.

immunosenescence

age-associated diseases

aging

1. Introduction

Aging is a time-dependent progressive accumulation of significant cellular and tissue changes, including physiological, structural, and functional changes, leading to functional disorders and increased vulnerability to death ^[1]. This process is associated with molecular events such as genomic instability, telomere attrition, epigenetic alterations, loss of proteostasis, deregulated nutrient-sensing, mitochondrial dysfunction, cellular senescence, stem cell exhaustion, and altered intercellular communication, which can be termed “hallmarks of aging” ^{[2][3]}.

The skin is our largest organ and constitutes a protective barrier that prevents excessive water loss and the entry of harmful substances and pathogens from the environment. Its aging reflects both intrinsic (or chronological) and extrinsic (such as radiation and pollution exposure) aging processes at the molecular and phenotypic levels ^[4]. Skin aging is a process accompanied by changes that alter the local microenvironment, such as weakening of the skin barrier and the accumulation of stressed and senescent cells, both of which foster inflammation through the invasion/release of Pathogen- and Damage-Associated Molecular Patterns ^[5]. The consequences of such an altered microenvironment include the promotion of the senescence-associated secretory phenotype (SASP), compromising tissue renewal and function, altered cellular interactions ^[6], and chronic low-grade inflammation ^[7]. This sterile inflammatory state, termed inflammaging, develops in several organs with advanced age and is associated with persistent inflammation that ultimately exhausts the skin’s defense system ^{[5][8]}.

Macrophages (M ϕ), a group of heterogeneous and plastic cells, play a central role in tissue homeostasis and repair, as well as host defense [9]. In the skin, M ϕ can be found in different layers, being classified as recruited M ϕ originating from monocytes following a recruitment process started by tissue injury, or as tissue-resident macrophages (TRM), which are derived from both adult and embryonic progenitors [10]. In the interfollicular epidermis, there are the Langerhans cells (LC), which can migrate to the lymph nodes to present antigens, being related to antimicrobial immunity, immune surveillance, and contact hypersensitivity [11]. Due to the shared characteristics with dendritic cells (DCs), LC have long been classified as such [12][13]. Nevertheless, after further ontogeny studies have demonstrated that LC arise from embryonic precursors and are maintained within the epidermis by local self-renewal under steady-state conditions, LC are currently considered a specialized subset of TRM [14]. M ϕ located in the dermis, on the other hand, are called dermal M ϕ and are associated with tissue repair and clearance [15].

To exert such a variety of functions, M ϕ may acquire different phenotypes in response to various stimuli. In this sense, based on in vitro assays, M ϕ have been divided into two groups based on their polarization phenotypes: M1 and M2. Classically activated M ϕ are deemed as M1 and constitute catabolic, proinflammatory cells that are involved in antimicrobial host defense. M2, or alternatively activated M ϕ , are anabolic cells with anti-inflammatory and tissue repair properties [16]. However, mainly due to recent advances in single-cell RNA sequencing (scRNA-Seq), it is now clear that such a dichotomy does not accurately represent M ϕ in vivo but represents the extremes of a wide range of continuous phenotypes which have been reported [17][18].

The aging process has a great impact on M ϕ , including alterations in M ϕ metabolic and immune function, impacting the M ϕ capability of clearance and immunosurveillance, constituting an important aspect of immunosenescence [19]. In fact, old M ϕ in a mice model were characterized with a senescent, proinflammatory profile [20], associated with increased oxidative stress, compromised antioxidant defenses, and impaired function [21].

Interestingly, the number of LC in the skin and their capacity to migrate to the lymph node and stimulate T cells seems to be reduced in aged subjects compared to young ones [22]. In aged mice, the same process is observed, accompanied by a decline in LC maturation, but not in LC proliferation and survival levels, suggesting either a deficiency in bone marrow-derived LC progenitors or the generation of progenitors that are less responsive to chemokine and cytokine signals [22]. The same study has also described a higher level of phagocytosis in M ϕ from older mice [22], which is probably a result of an age-related M ϕ hyperfunction, since during the aging process the skin barrier weakens, favoring the pathogen's invasion and stressed and senescent cells that should normally be eliminated are accumulated [23].

M ϕ are considered as gatekeepers of tissue homeostasis and integrity, constituting primary inflammatory cytokine producers, as well as initiators and regulators of inflammation, and representing one of the main cellular players in adaptive immunity exacerbation and exhaustion during aging [24][25]. With that being said, it is possible to consider M ϕ as important players in the promotion of chronic proinflammatory-associated pathologies, such as psoriasis [26][27], rosacea [28][29], vitiligo [30][31], and skin cancer [32][33].

2. Hallmarks of Aging and Macrophages

Aging is a progressive and common process for all cells and tissues and can be caused by both intracellular and extracellular factors. It leads to organismal dysfunction on multiple levels, the main underlying processes being identified as the hallmarks of aging (Figure 1). Such hallmarks are interconnected and converge to tissue inflammation and dysfunction [5].

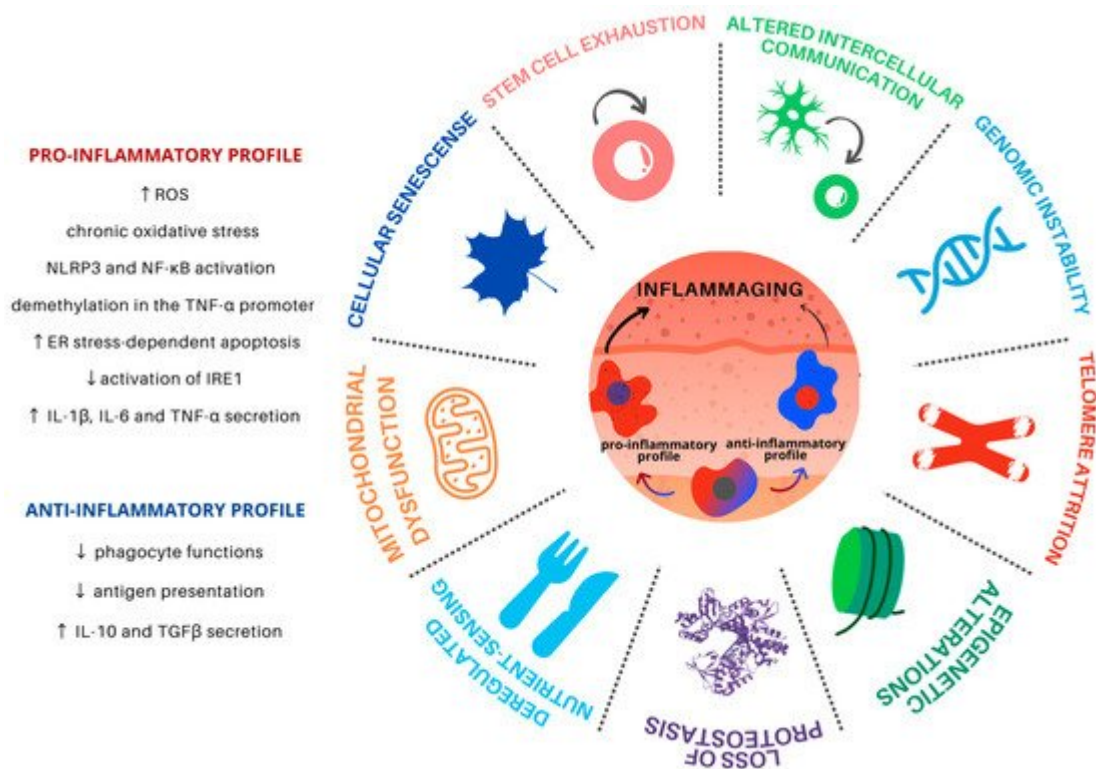


Figure 1. Hallmarks of aging in Mφ in the skin microenvironment. Skin inflammaging is fostered by different yet interconnected and synergistic aging hallmarks. Mφ are plastic cells that play a pivotal role in the immune system and have been associated with the persistent chronic inflammation levels found in aged skin. Skin inflammaging is characterized by a shift towards pro-inflammatory Mφ phenotypes, which promote further tissue inflammation in the skin microenvironment through the secretion of pro-inflammatory cytokines, activation of important inflammatory pathways, and increased oxidative stress. Chronic low-grade oxidative-inflammatory stress during the aging process is a key factor that stimulates a vicious cycle, contributing to age-associated disease onset. At the same time, the reduction of Mφ with an anti-inflammatory phenotype contributes to the decrease in antigen presentation and phagocytosis, contributing to tissue homeostasis disturbance.

2.1. Genomic Instability

DNA damage accumulation is expected to occur with aging and accumulates as a result of many endogenous and exogenous factors. Genomic instability in the aging process can be associated with somatic mutations, copy-number alterations, and chromosome abnormalities for nuclear as well as mitochondrial DNA (mtDNA) [2]. Such DNA alterations may affect essential genes and transcriptional pathways, resulting in dysfunctional cells.

In the literature, the causes of genomic instability in M ϕ are scarce and one work points out that it can be induced by pathogens such as *Mycobacterium tuberculosis* [34]. *M. tuberculosis* is the causative agent of tuberculosis with a pathological outcome associated with the formation of granulomas. The granulomas formed in the development of several chronic diseases (due to persistent inflammatory stimuli) can modulate molecular programs, which are involved in TRM differentiation and relate to clinical outcomes [35] and DNA damage [36].

Since M ϕ are the most abundant immune cell type in the skin, and this tissue is directly exposed to several environmental factors, such as UV radiation, genomic instability could be a central hallmark of aging in aged M ϕ ; after all, numerous DNA injuries can lead to accumulated damage and altered M ϕ function in this tissue [37]. This in turn can lead to altered gene expression of molecules such as cytokines, MHC class II, transcription factors, an exacerbated production of reactive oxygen species (ROS) [15], and NF- κ B signaling in response to DNA damage [38], rendering M ϕ more susceptible to apoptosis, impairing their phagocytosis function [39], and contributing to inflammaging [40].

2.2. Telomere Attrition

Telomeres protect the ends of chromosomes from degradation and abnormal recombination. Considered a primary hallmark of aging, telomere attrition causes the loss of chromosome protective structures as they gradually get shorter [2]. This shortening process has also been closely connected with inflammation [41][42].

Comparing young and old mice, Kang and colleagues (2018) observed that the shortening of telomeres in M ϕ leads to increased ROS production, the same phenotype observed for genomic instability (see [Section 2.1](#)). Furthermore, in their experiment, knockout mice for Telomerase RNA Component (*Terc*^{-/-}), a gene that encodes for the RNA that serves as a template for the telomere repeats, showed telomere dysfunction in M ϕ , which is associated with hyper inflammation and mitochondrial abnormality, followed by oxidative stress with hyperactivation of the Nod-like receptor protein 3 (NLRP3) inflammasome (see [Section 2.6](#)) [43].

The increase in oxidative stress can induce DNA breakdown, which can lead to mutations that may explain most of the changes described in the aged M ϕ . The main inflammatory signaling pathway, NF- κ B, regulates the maintenance of telomeres and telomerase activity [44], just as the latter regulate NF- κ B activity [45]. This relationship leads to a defective autophagic response and overexpression of inflammatory cytokines, such as TNF- α , IL-6, and IFN in circulating M ϕ [46].

2.3. Epigenetic Alterations

Epigenetic alterations involve changes in DNA methylation (DNAm) patterns, post-transcriptional modification of histones, and chromatin remodeling [2][47]. These aging-induced epigenetic changes in M ϕ are mainly responsible for controlling the inflammatory profile and cell differentiation [48][49].

DNAm undergoes predictable time-dependent modifications across CpG islands and is influenced by both intrinsic and extrinsic processes. Molecular clocks have been developed in order to calculate the “biological age” of

biological samples using methylome data [50][51], including a skin-specific Molecular Clock [51]. Age-associated epigenetic remodeling involves highly localized gain/loss of DNAm at the binding sites of transcription factors associated with the monocyte-macrophage differentiation process [52]. Despite a major lack of comprehension regarding the cause or effect role of epigenetic changes and aging phenotypes, recent studies have shed light on at least a few events that connect epigenetic changes in M ϕ and age-related phenotypes, such as inflammation and differentiation.

For instance, aging-associated changes in DNAm, particularly the demethylation in the tumor necrosis factor (TNF- α) promoter, a cytokine predominantly produced by M ϕ , revealed a possible link between inflammation, M ϕ , and chronic age-related diseases. The promoter demethylation has been described to occur in peripheral blood leukocytes and M ϕ of aging subjects and is accompanied by a reduction of TNF- α reporter gene activity [53], possibly associated with chronic inflammatory processes.

Other studies have also revealed that protein-3 containing the Jumonji domain (Jmjd-3), a H3K27 demethylase, promotes the induction of Irf-4, and SMYD-3, an H3K4 methyltransferase, of IL-4 and IL-12 [54][55]. These histone-modifying proteins play a central role in aging and their activities increase in the course of this process. In M ϕ , they contribute to the positive regulation of regenerative and anti-inflammatory profiles [55][56].

These epigenetic alterations in the skin microenvironment contribute to inflammaging and can be directly linked to clinical outcomes. For instance, the chronic exposure of M ϕ to inflammatory triggers and products of dead or senescent cells (see [Section 2.7](#)) can impose epigenetic changes that cause M ϕ ' altered response capacity during skin aging [52].

2.4. Loss of Proteostasis

Proteostasis is defined as the process of protein homeostasis maintenance and comprises a complex proteostasis network (PN), mainly composed by specialized proteins such as chaperones and cochaperones, translational machinery, the ubiquitin-proteasome system (UPS), and the autophagy machinery [57]. The PN has the role of controlling protein synthesis, modification, secretion, and degradation. It also reduces misfolded proteins by restoring, removing, or degrading them through the unfolded protein response (UPR) activity to prevent their accumulation in cellular compartments [58]. The chaperone-mediated autophagy (CMA) is another player in the proteostasis balance. HSPA8 is a central component of CMA and is an abundant protein in M ϕ and other immune cells. Together with a co-chaperone complex, HSPA8 recognizes "CMA-targeting recognition motifs" in the targeted protein sequence, unfolding the substrate and delivering it to a protein called lysosome-associated membrane protein 2A (LAMP2A), which internalizes the targeted proteins for subsequent degradation in the lysosomal lumen [59]. If those mechanisms fail to restore homeostasis, apoptotic pathways may be activated to ensure survival of the organism [58].

Autophagy is a paramount process in the maintenance of skin homeostasis throughout aging, the consequences of age-related autophagy decay affecting different skin types, including LC. An example describing the consequences of the loss of proteostasis on aged skin is the change of elastin, collagen, and melanin levels [60][61][62] found in

wrinkled and hypopigmented skin [60][63]. The balance in the composition of those proteins is essential for skin function and health and, interestingly, M ϕ play an important role in this context. For instance, M ϕ synthesize metalloelastases (e.g., metalloelastase 12) that participate in the elimination of nonfunctional elastin aggregates generated in the skin as a consequence of photoaging [64][65].

If on one side, healthy M ϕ are important contributors to the maintenance of proteostasis, aged M ϕ exhibit diminished inositol-requiring enzyme 1 α (IRE1 α) activation (a stress sensor that activates UPR) and increased susceptibility to endoplasmic reticulum (ER) stress-dependent apoptosis [66]. During high levels of ER stress, UPR activates IRE1 α , which in turn assists the alternative splicing of X-box binding protein 1 (XBP1) mRNA [67]. After its activation, the transcription factor XBP1 induces the expression of cytokines such as pro-IL-1 β [68]. However, it has been shown that Toll-like Receptors (TLR) in mice and human M ϕ can directly activate XBP1, without UPR mediation, or even in synergy with ER stress [67], leading to the splicing of XBP1 and activation of a sustained proinflammatory environment by IL-1 β , IL-6, and TNF [67][69].

In fact, it has been shown that the inflammasome is activated in the context of excessive misfolded protein accumulation, which is exacerbated in autophagy- or p62 (sequestosome 1)-deficient M ϕ [70][71]. In addition, ER stress can also be transferred from neighboring parenchymal cells to TRMs by upregulating the splicing of UPR components, such as *Grp78*, *Gadd34*, *Chop*, and *Xbp-1* [72][73]. This phenomenon of a “transmissible” ER stress state is mediated by the production of IL-4, IL-10, and by apoptotic bodies from stressed cells [72]. Therefore, the age-related loss of proteostasis in the skin affects M ϕ and seems to contribute to the local inflammaging phenotype.

2.5. Deregulated Nutrient Sensing

Aging directly affects the sensors and molecular targets of nutrients and fluid homeostatic regulation [74]. One of the underlying mechanisms by which aging promotes deregulated nutrient sensing is by promoting disturbed insulin sensitivity, which compromises the capacity of some tissues to uptake and metabolize nutrients [75].

Importantly, nutrient sensing components have been directly linked to longevity, including the mammalian targets of rapamycin (mTOR) and AMP-dependent protein kinase (AMPK), the two major components of nutrient sensing and metabolic regulation. AMPK has an inhibitory effect on mTOR signaling, which is activated during nutrient starvation, leading to a rise in the AMP:ATP ratio [76]. In association with nicotinamide adenine dinucleotide (NAD), high AMP levels activate sirtuins, responsible for insulin signaling pathway and longevity regulation. Consequently, the excess of nutrient availability can promote aging-associated diseases [77][78].

Accordingly, the modulation of nutrient sensing signaling influences several immune cell types including M ϕ . It has been said that the M ϕ immunometabolism influences M ϕ polarization and activation, processes which are tightly linked to skin homeostasis and inflammaging. M ϕ metabolic signatures have been closely connected with the M1-like and M2-like phenotypes; the M1-like M ϕ heavily relying on glycolysis, and the M2-like M ϕ being more dependent on oxidative phosphorylation [79]. During the aging process, M ϕ function and phenotypes are disturbed due to many factors, including nutrient sensing dysregulation and the installation of a chronic low-grade

inflammation environment in the tissue. In this sense, deregulated nutrient sensing can increase M ϕ glycolysis and suppress the oxidative phosphorylation (via attenuation of IL-4-induced anti-inflammatory responses), favoring the accumulation of M1-like M ϕ in the aged skin [80][81][82][83]. The presence of M2-like profile is therefore reduced, causing skin damage and promoting the progression of age-associated diseases [84][85][86].

Furthermore, FOXO and mTOR are targets of the insulin and insulin-like growth factor 1 (IGF-1) signaling (IIS) pathway and are influenced by nutrient status, altering tissue homeostasis, and inflammation [87][88]. In a very elegant experiment, the skin of mice lacking both the insulin and IGF-1 receptor in myeloid cells was enriched in noninflammatory M ϕ phenotype after the induction of dermatitis. When compared to controls, it showed evidence of a proinflammatory IR/IGF-1R-dependent pathway and a connection between cutaneous inflammatory responses and diseases such as insulin-resistant diabetes mellitus type 2 [89]. In addition, SASP has been highly associated with M ϕ inflammatory factors in conditions of hyperglycaemia, contributing to the fueling of low-grade inflammation in diabetes [90]. Under nutrient starvation, FOXO1 migrates to the nucleus after phosphorylation and seems to stimulate proinflammatory TLR4 signaling and IL-8 β production in M ϕ . FOXO1 migration also stimulates the expression of the anti-inflammatory cytokine IL-10 in M2-like cells [91], supporting the phenotypic development of aging M ϕ in distinct directions [19].

Bone marrow-derived M ϕ have also shown an increasing expression of growth hormone receptor (GH-R) and GH-R-dependent induction of inflammatory components in aged mice. Current evidence suggests that the downregulation of NLRP3 inflammasome in M ϕ by GH-R is capable of maintaining immune system homeostasis and extending health- and lifespan [92].

Since nutrient sensing signaling pathways can be pharmacologically modulated and are closely linked to inflammation, interesting observations could be made regarding the manipulation of M ϕ phenotypes in the skin. For instance, the modulation of AMPK/mTOR/NLRP3 inflammasome signaling using Metformin revealed that the drug treatment promoted reduced NLRP3 signaling and promoted the regenerative M2-like phenotype in skin M ϕ , paving a way to re-establish skin M ϕ equilibrium [93]. Several other anti-inflammatory drugs target immunometabolism and may also contribute in this sense, as revised by [94].

2.6. Mitochondrial Dysfunction

Mitochondrial production of ROS is crucial for the skin's defense against pathogens [95]. However, dysfunctional mitochondria in M ϕ can generate excessive production of ROS and cause damage to important intracellular structures, including the mtDNA [22], contributing to defective apoptosis and activation of inflammasomes [96]. Aged M ϕ present mitochondrial dysfunction associated with decreased ATP production, reduction of mitochondrial membrane potential ($\Delta\Psi_m$), and increased oxidative stress, as well as depreciated antioxidant defense response that can impair M ϕ functions and lead to senescence [97]. For instance, during lung *Streptococcus pneumoniae* infections, impaired mitochondrial function of aged M ϕ increases lung pathology and oxidative stress [98].

The activity of the oxidative phosphorylation system is also altered in aging. During aerobic respiration, oxygen can be reduced prematurely, generating a high amount of ROS, a process that is exacerbated in senescent cells [99].

Besides that, Minhas and colleagues (2019) demonstrated the importance of NAD⁺ levels to maintain mitochondrial respiration and regulate M ϕ phagocytosis in an anti-inflammatory homeostatic state, both in vitro and in vivo. NAD⁺ levels can be replenished by de novo synthesis and via the kynurenine pathway. Blockage of de novo NAD⁺ synthesis impaired phagocytosis and resolution of inflammation in aged M ϕ [100].

In addition to sensing and cleansing cellular debris, M ϕ also detect accumulation of mitochondrial garbage in the cellular microenvironment, leading to a continuous stimulation of these cells and thus their activation [101] and thus sustaining an environment of chronic low-grade inflammation with production of cytokines and ROS [97]. ROS accumulation in intracellular microenvironments (not only in the mitochondria) can cause DNA damage in aged tissues [28]. In the context of the skin, ultraviolet (UV) radiation-induced mtDNA injury also leads to more ROS production, accelerating photoaging [102]. In photodamaged skin, xanthine oxidase-induced ROS is reported to be the cause of alterations in collagen biosynthesis in cultured human dermal fibroblasts [103]. Moreover, other enzymatic and non-enzymatic sources of ROS are observed in the skin. Besides mitochondrial ROS production via electron transport chain and UV-induced ROS, there is production of ROS via peroxisomes, ER, and skin cell membranes [104]. In cultured M ϕ , Ives and colleagues (2015) demonstrated that xanthine oxidase (XO) is the major source of ROS [105]. XO expression and activity has also been shown to be increased in old mice, closely associating with oxidative stress and exacerbated ROS formation [21].

As extensively revised by Beek and colleagues (2019), there is a tight link between mitochondrial dysfunction and ER in aged M ϕ , that results in impaired calcium and redox homeostasis and leads to oxidative stress and activation of several pathways, including the inflammasome. Consequently, a proinflammatory environment is promoted by enhancing IL-1 β secretion and nuclear translocation of NF- κ B [19]. Furthermore, it has been proposed that this age-related oxidative-inflammatory stress (“oxi-inflammaging”) occurs in a vicious cycle during aging [106]. Taken together, it can be expected that age-related ER and oxidative stresses can contribute to an enhanced production of pro-inflammatory cytokines in M ϕ and thus to systemic inflammaging, favoring the onset of pathologies.

2.7. Cellular Senescence

Cellular senescence is a cellular state characterized by cell cycle arrest, even under growth-promoting conditions [2]. Other phenotypes of cellular senescence include apoptosis resistance and SASP [107]. In the human skin, senescent keratinocytes and fibroblasts accumulate with age and support a feedforward system mainly mediated by SASP to accelerate tissue function decay [108]. Senescent cells show increased production of proinflammatory cytokines, chemokines, growth factors and metalloproteinases [109], telomere attrition (see [Section 2.2](#)), epigenetic alterations (see [Section 2.3](#)), loss of proteostasis (see [Section 2.4](#)), and dysfunctional mitochondria (see [Section 2.6](#)), underscoring the many facets of the senescent cell phenotype.

The age-related alterations of immune system elements have been defined as immunosenescence and are characterized by changes in anatomical barriers, lymphoid organs and immune cell function, all of which synergize to result in a systemic organismal low-grade inflammation, deemed inflammaging [5].

Intrinsic and extrinsic factors of immunosenescence affect both recruited M ϕ and TRMs. In homeostatic conditions, activated M ϕ clear cell debris [110], but when senescent cell clearance is not effective, such cells accumulate and intensify SASP, causing several alterations in the local milieu, including M ϕ dysfunction [111]. Using a mouse model, Praticchizzo and colleagues (2018) demonstrated that in hyperglycemic conditions, both cellular senescence and SASP can be induced in M ϕ [90]. In aged tissues, the activated M ϕ produces molecules that drive inflammatory response, such as IL-6, matrix metalloproteinases, chemokines and other mediators [112][113]. Together, these indicate that dysfunctional M ϕ are both a result of dysfunctional niches and cellular senescence, in addition to contributing to the maintenance of low-grade tissue inflammation.

2.8. Stem Cell Exhaustion

Stem cell exhaustion is a consequence of the sum of several hallmarks of aging mentioned above and is likely one of the main culprits for the loss of tissue regenerative capacity, and consequently organismal aging. Examples of that loss have already been related to immunosenescence [114][115].

Skin homeostasis is mainly maintained by two stem cell types: dermal mesenchymal stem cells (dermal MSCs), present in the inner layer of dermis, and epidermal stem cells (ESCs), located in the basal epidermal layer. While ESCs are responsible for epidermal cell renewal, a consequence of the capacity to differentiate into different cell lineages of the skin, such as keratinocytes and melanocytes [116][117], dermal MSCs are capable of differentiating into subcutaneous adipocytes, osteoblasts, and chondrocytes [118][119]. With aging, a loss of ESC and dermal MSC production and differentiation is observed, with consequent deceleration of skin cell renewal and reduction of skin healing capacity [119][120].

Tissue-resident hematopoietic cells, progenitors of M ϕ , can self-maintain independently of hematopoietic stem cells (HSC). M ϕ derived from yolk-sac are replaced by HSC-derived ones only in a few organs, including epidermis skin layer (LC originating from erythro-myeloid progenitors) [121]. This statement suggests a more important role is played by the cell's origin than its tissue location in life span [14]. As a consequence, it is not surprising that the aged skin tends to have less LC in its composition, decreasing antigen-specific immunity [122].

Wound repair is a continuous process comprising four phases: hemostasis, inflammatory, proliferative, and remodeling (or resolution) phases [123]. With the advance in aging, there is a delayed lesion reepithelization and decreased tensile strength [124][125][126]. That may also indicate the importance of the presence of M ϕ subpopulations in key moments of wound skin repair [123]. The M1-like M ϕ profile contributes to the early stage of skin healing by promoting an inflammatory response with the production of high levels of proinflammatory cytokines. On the other hand, M2-like M ϕ are responsible for the tissue repair process itself, including the regulation of re-vascularisation processes, fibroblast proliferation, and myofibroblast conversion, in addition to collagen production via inhibition of the AMPK/mTOR/NLRP3 inflammasome signaling axis, as discussed in [Section 2.5](#). [93]. Immune cells such as M ϕ are capable of activating epidermal stem cells for re-epithelialization under the establishment of an inflammatory wound microenvironment [127].

Through single-cell transcriptomic data analysis, a study was capable of characterizing a dermal subpopulation of M ϕ that contributes to local nerve regeneration and axon sprouting after a mechanical injury [128]. Another study also observed that during skin repair, M ϕ are also capable of stimulating the proliferation of adipocyte precursors [129]. Aged M ϕ tend to lose their ability to migrate into wounds, with consequent retention of the M ϕ at the dermis and increased tissue damaging release of ROS and proinflammatory cytokines [130]. In the epidermis, such an excessive proinflammatory microenvironment depletes epidermal stem cells, further contributing to compromised skin healing capacity [131].

2.9. Altered Intercellular Communication

The age-related changes in intercellular communication have been characterized in the autocrine, paracrine, endocrine, and neuroendocrine levels. The neurohormonal signaling (e.g., renin-angiotensin, adrenergic, insulin-IGF1 signaling) tends to be deregulated in aging as inflammatory reactions increase, immunosurveillance against pathogens and premalignant cells declines, and the composition of the peri- and extracellular environment changes, thereby affecting the mechanical and functional properties of all tissues, including the skin [2][132].

Recently, it was observed that mast cells are important in the recruitment of M ϕ in aging through the change in the pattern of chemoattractant cytokines [133]. But there have still been few studies on how these intercellular communications involve M ϕ in the skin. Still, it is already known that the age-related changes in intercellular communication are associated with inflammation. The accumulation of tissue damage throughout life, the likelihood of cytokines being secreted by senescent cells, the enhanced activation of the NF- κ B transcription factor, and the occurrence of a defective autophagy response all seem to foster the immune system failure [96][134].

References

1. Chung, H.Y.; Kim, D.H.; Lee, E.K.; Chung, K.W.; Chung, S.; Lee, B.; Seo, A.Y.; Chung, J.H.; Jung, Y.S.; Im, E.; et al. Redefining Chronic Inflammation in Aging and Age-Related Diseases: Proposal of the Senoinflammation Concept. *Aging Dis.* 2019, 10, 367–382.
2. López-Otín, C.; Blasco, M.A.; Partridge, L.; Serrano, M.; Kroemer, G. The Hallmarks of Aging. *Cell* 2013, 153, 1194–1217.
3. Mc Auley, M.T.; Guimera, A.M.; Hodgson, D.; McDonald, N.; Mooney, K.M.; Morgan, A.E.; Proctor, C.J. Modelling the Molecular Mechanisms of Aging. *Biosci. Rep.* 2017, 37.
4. Tobin, D.J. Introduction to Skin Aging. *J. Tissue Viability* 2017, 26, 37–46.
5. Rodrigues, L.P.; Teixeira, V.R.; Alencar-Silva, T.; Simonassi-Paiva, B.; Pereira, R.W.; Pogue, R.; Carvalho, J.L. Hallmarks of Aging and Immunosenescence: Connecting the Dots. *Cytokine Growth Factor Rev.* 2021, 59, 9–21.

6. Solé-Boldo, L.; Raddatz, G.; Schütz, S.; Mallm, J.-P.; Rippe, K.; Lonsdorf, A.S.; Rodríguez-Paredes, M.; Lyko, F. Single-Cell Transcriptomes of the Human Skin Reveal Age-Related Loss of Fibroblast Priming. *Commun. Biol.* 2020, 3, 188.
7. Robert, L.; Labat-Robert, J.; Robert, A.-M. Physiology of Skin Aging. *Pathol. Biol.* 2009, 57, 336–341.
8. Franceschi, C.; Bonafè, M.; Valensin, S.; Olivieri, F.; De Luca, M.; Ottaviani, E.; De Benedictis, G. Inflamm-Aging. An Evolutionary Perspective on Immunosenescence. *Ann. N. Y. Acad. Sci.* 2000, 908, 244–254.
9. Yanez, D.A.; Lacher, R.K.; Vidyarthi, A.; Colegio, O.R. The Role of Macrophages in Skin Homeostasis. *Pflügers Arch. Eur. J. Physiol.* 2017, 469, 455–463.
10. Bian, Z.; Gong, Y.; Huang, T.; Lee, C.Z.W.; Bian, L.; Bai, Z.; Shi, H.; Zeng, Y.; Liu, C.; He, J.; et al. Deciphering Human Macrophage Development at Single-Cell Resolution. *Nature* 2020, 582, 571–576.
11. Doebel, T.; Voisin, B.; Nagao, K. Langerhans Cells—The Macrophage in Dendritic Cell Clothing. *Trends Immunol.* 2017, 38, 817–828.
12. Merad, M.; Ginhoux, F.; Collin, M. Origin, Homeostasis and Function of Langerhans Cells and Other Langerin-Expressing Dendritic Cells. *Nat. Rev. Immunol.* 2008, 8, 935–947.
13. Kaplan, D.H. Langerhans Cells: Not Your Average Dendritic Cell. *Trends Immunol.* 2010, 31, 437.
14. Gomez Perdiguero, E.; Klapproth, K.; Schulz, C.; Busch, K.; Azzoni, E.; Crozet, L.; Garner, H.; Trouillet, C.; de Bruijn, M.F.; Geissmann, F.; et al. Tissue-Resident Macrophages Originate from Yolk-Sac-Derived Erythro-Myeloid Progenitors. *Nature* 2015, 518, 547–551.
15. Malissen, B.; Tamoutounour, S.; Henri, S. The Origins and Functions of Dendritic Cells and Macrophages in the Skin. *Nat. Rev. Immunol.* 2014, 14, 417–428.
16. Shapouri-Moghaddam, A.; Mohammadian, S.; Vazini, H.; Taghadosi, M.; Esmaeili, S.-A.; Mardani, F.; Seifi, B.; Mohammadi, A.; Afshari, J.T.; Sahebkar, A. Macrophage Plasticity, Polarization, and Function in Health and Disease. *J. Cell. Physiol.* 2018, 233, 6425–6440.
17. Qian, J.; Olbrecht, S.; Boeckx, B.; Vos, H.; Laoui, D.; Etliglu, E.; Wauters, E.; Pomella, V.; Verbandt, S.; Busschaert, P.; et al. A Pan-Cancer Blueprint of the Heterogeneous Tumor Microenvironment Revealed by Single-Cell Profiling. *Cell Res.* 2020, 30, 745–762.
18. Zilionis, R.; Engblom, C.; Pfirschke, C.; Savova, V.; Zemmour, D.; Saatcioglu, H.D.; Krishnan, I.; Maroni, G.; Meyerovitz, C.V.; Kerwin, C.M.; et al. Single-Cell Transcriptomics of Human and Mouse Lung Cancers Reveals Conserved Myeloid Populations across Individuals and Species. *Immunity* 2019, 50, 1317–1334.e10.

19. Van Beek, A.A.; Van den Bossche, J.; Mastroberardino, P.G.; de Winther, M.P.J.; Leenen, P.J.M. Metabolic Alterations in Aging Macrophages: Ingredients for Inflammaging? *Trends Immunol.* 2019, 40, 113–127.
20. Tabula Muris Consortium A Single-Cell Transcriptomic Atlas Characterizes Ageing Tissues in the Mouse. *Nature* 2020, 583, 590–595.
21. Vida, C.; de Toda, I.M.; Cruces, J.; Garrido, A.; Gonzalez-Sanchez, M.; De la Fuente, M. Role of Macrophages in Age-Related Oxidative Stress and Lipofuscin Accumulation in Mice. *Redox Biol.* 2017, 12, 423–437.
22. Chambers, E.S.; Vukmanovic-Stejic, M. Skin Barrier Immunity and Ageing. *Immunology* 2020, 160, 116–125.
23. Xu, Y.-P.; Qi, R.-Q.; Chen, W.; Shi, Y.; Cui, Z.-Z.; Gao, X.-H.; Chen, H.-D.; Zhou, L.; Mi, Q.-S. Aging Affects Epidermal Langerhans Cell Development and Function and Alters Their miRNA Gene Expression Profile. *Aging* 2012, 4, 742–754.
24. Foley, K.G.; Pritchard, M.T.; Duncan, F.E. Macrophage-Derived Multinucleated Giant Cells: Hallmarks of the Aging Ovary. *Reproduction* 2021, 161, V5–V9.
25. Canan, C.H.; Gokhale, N.S.; Carruthers, B.; Lafuse, W.P.; Schlesinger, L.S.; Torrelles, J.B.; Turner, J. Characterization of Lung Inflammation and Its Impact on Macrophage Function in Aging. *J. Leukoc. Biol.* 2014, 96, 473–480.
26. Deng, G.; Chen, W.; Wang, P.; Zhan, T.; Zheng, W.; Gu, Z.; Wang, X.; Ji, X.; Sun, Y. Inhibition of NLRP3 Inflammasome-Mediated Pyroptosis in Macrophage by Cycloastragenol Contributes to Amelioration of Imiquimod-Induced Psoriasis-like Skin Inflammation in Mice. *Int. Immunopharmacol.* 2019, 74, 105682.
27. Lu, C.-H.; Lai, C.-Y.; Yeh, D.-W.; Liu, Y.-L.; Su, Y.-W.; Hsu, L.-C.; Chang, C.-H.; Catherine Jin, S.-L.; Chuang, T.-H. Involvement of M1 Macrophage Polarization in Endosomal Toll-Like Receptors Activated Psoriatic Inflammation. *Mediat. Inflamm.* 2018, 2018, 3523642.
28. Liu, T.; Deng, Z.; Xie, H.; Chen, M.; Xu, S.; Peng, Q.; Sha, K.; Xiao, W.; Zhao, Z.; Li, J. ADAMDEC1 Promotes Skin Inflammation in Rosacea via Modulating the Polarization of M1 Macrophages. *Biochem. Biophys. Res. Commun.* 2020, 521, 64–71.
29. Liu, Z.; Zhang, J.; Jiang, P.; Yin, Z.; Liu, Y.; Liu, Y.; Wang, X.; Hu, L.; Xu, Y.; Liu, W. Paeoniflorin Inhibits the Macrophage-Related Rosacea-like Inflammatory Reaction through the Suppressor of Cytokine Signaling 3-Apoptosis Signal-Regulating Kinase 1-p38 Pathway. *Medicine* 2021, 100, e23986.
30. Abdellatif, A.A.; Zaki, A.M.; Abdo, H.M.; Aly, D.G.; Emara, T.A.; El-Toukhy, S.; Emam, H.M.; Abdelwahab, M.S. Assessment of Serum Levels of Granulocyte-Macrophage Colony-Stimulating

- Factor (GM-CSF) among Non-Segmental Vitiligo Patients: A Pilot Study. *Acta Derm. Alp. Pannonica Adriat* 2015, 24, 43–45.
31. Farag, A.G.A.; Hammam, M.A.; Habib, M.S.; Elnaidany, N.F.; Kamh, M.E. Macrophage Migration Inhibitory Factor as an Incriminating Agent in Vitiligo. *An. Bras. Dermatol.* 2018, 93, 191–196.
 32. Ishida, Y.; Kuninaka, Y.; Yamamoto, Y.; Nosaka, M.; Kimura, A.; Furukawa, F.; Mukaida, N.; Kondo, T. Pivotal Involvement of the CX3CL1-CX3CR1 Axis for the Recruitment of M2 Tumor-Associated Macrophages in Skin Carcinogenesis. *J. Investig. Dermatol.* 2020, 140, 1951–1961.e6.
 33. Weber, C.; Telerman, S.B.; Reimer, A.S.; Sequeira, I.; Liakath-Ali, K.; Arwert, E.N.; Watt, F.M. Macrophage Infiltration and Alternative Activation during Wound Healing Promote MEK1-Induced Skin Carcinogenesis. *Cancer Res.* 2016, 76, 805–817.
 34. Castro-Garza, J.; Luévano-Martínez, M.L.; Villarreal-Treviño, L.; Gosálvez, J.; Fernández, J.L.; Dávila-Rodríguez, M.I.; García-Vielma, C.; González-Hernández, S.; Cortés-Gutiérrez, E.I. Mycobacterium Tuberculosis Promotes Genomic Instability in Macrophages. *Mem. Inst. Oswaldo Cruz* 2018, 113, 161–166.
 35. Herrtwich, L.; Nanda, I.; Evangelou, K.; Nikolova, T.; Horn, V.; Sagar, Erny, D.; Stefanowski, J.; Rogell, L.; Klein, C.; et al. DNA Damage Signaling Instructs Polyploid Macrophage Fate in Granulomas. *Cell* 2018, 174, 1325–1326.
 36. Horn, V.; Triantafyllopoulou, A. DNA Damage Signaling and Polyploid Macrophages in Chronic Inflammation. *Curr. Opin. Immunol.* 2018, 50, 55–63.
 37. Sidler, C. Chapter 29—Genomic Instability and Aging: Causes and Consequences. In *Genome Stability*; Kovalchuk, I., Kovalchuk, O., Eds.; Academic Press: Boston, MA, USA, 2016; pp. 511–525. ISBN 9780128033098.
 38. Janssens, S.; Tschopp, J. Signals from within: The DNA-Damage-Induced NF-κB Response. *Cell Death Differ.* 2006, 13, 773–784.
 39. Lloberas, J.; Tur, J.; Vico, T.; Celada, A. Molecular and Cellular Aspects of Macrophage Aging. In *Handbook of Immunosenescence*; Springer: Cham, Switzerland, 2018; pp. 1–32.
 40. Yue, Z.; Nie, L.; Zhang, P.; Chen, Q.; Lv, Q.; Wang, Q. Tissue-Resident Macrophage Inflammaging Aggravates Homeostasis Dysregulation in Age-Related Diseases. *Cell. Immunol.* 2021, 361, 104278.
 41. Stanley, S.E.; Armanios, M. The Short and Long Telomere Syndromes: Paired Paradigms for Molecular Medicine. *Curr. Opin. Genet. Dev.* 2015, 33, 1–9.
 42. De Maeyer, R.P.H.; Chambers, E.S. The Impact of Ageing on Monocytes and Macrophages. *Immunol. Lett.* 2021, 230, 1–10.

43. Kang, Y.; Zhang, H.; Zhao, Y.; Wang, Y.; Wang, W.; He, Y.; Zhang, W.; Zhang, W.; Zhu, X.; Zhou, Y.; et al. Telomere Dysfunction Disturbs Macrophage Mitochondrial Metabolism and the NLRP3 Inflammasome through the PGC-1 α /TNFAIP3 Axis. *Cell Rep.* 2018, 22, 3493–3506.
44. Jurk, D.; Wilson, C.; Passos, J.F.; Oakley, F.; Correia-Melo, C.; Greaves, L.; Saretzki, G.; Fox, C.; Lawless, C.; Anderson, R.; et al. Chronic Inflammation Induces Telomere Dysfunction and Accelerates Ageing in Mice. *Nat. Commun.* 2014, 2, 4172.
45. Jose, S.S.; Bendickova, K.; Kepak, T.; Krenova, Z.; Fric, J. Chronic Inflammation in Immune Aging: Role of Pattern Recognition Receptor Crosstalk with the Telomere Complex? *Front. Immunol.* 2017, 8, 1078.
46. Zhang, J.; Rane, G.; Dai, X.; Shanmugam, M.K.; Arfuso, F.; Samy, R.P.; Lai, M.K.P.; Kappei, D.; Kumar, A.P.; Sethi, G. Ageing and the Telomere Connection: An Intimate Relationship with Inflammation. *Ageing Res. Rev.* 2016, 25, 55–69.
47. Chen, S.; Yang, J.; Wei, Y.; Wei, X. Epigenetic Regulation of Macrophages: From Homeostasis Maintenance to Host Defense. *Cell. Mol. Immunol.* 2020, 17, 36–49.
48. Ivashkiv, L.B. Epigenetic Regulation of Macrophage Polarization and Function. *Trends Immunol.* 2013, 34, 216–223.
49. Daskalaki, M.G.; Tsatsanis, C.; Kampranis, S.C. Histone Methylation and Acetylation in Macrophages as a Mechanism for Regulation of Inflammatory Responses. *J. Cell. Physiol.* 2018, 233, 6495–6507.
50. Horvath, S. DNA Methylation Age of Human Tissues and Cell Types. *Genome Biol.* 2013, 14, R115.
51. Boroni, M.; Zonari, A.; de Oliveira, C.R.; Alkatib, K.; Cruz, E.A.O.; Brace, L.E.; de Carvalho, J.L. Highly Accurate Skin-Specific Methylome Analysis Algorithm as a Platform to Screen and Validate Therapeutics for Healthy Aging. *Clin. Epigenet.* 2020, 12, 105.
52. Dekkers, K.F.; Neele, A.E.; Jukema, J.W.; Heijmans, B.T.; de Winther, M.P.J. Human Monocyte-to-Macrophage Differentiation Involves Highly Localized Gain and Loss of DNA Methylation at Transcription Factor Binding Sites. *Epigenet. Chromatin.* 2019, 12, 34.
53. Gowers, I.R.; Walters, K.; Kiss-Toth, E.; Read, R.C.; Duff, G.W.; Wilson, A.G. Age-Related Loss of CpG Methylation in the Tumour Necrosis Factor Promoter. *Cytokine* 2011, 56, 792–797.
54. Satoh, T.; Takeuchi, O.; Vandenbon, A.; Yasuda, K.; Tanaka, Y.; Kumagai, Y.; Miyake, T.; Matsushita, K.; Okazaki, T.; Saitoh, T.; et al. The Jmjd3-Irf4 Axis Regulates M2 Macrophage Polarization and Host Responses against Helminth Infection. *Nat. Immunol.* 2010, 11, 936–944.
55. Kittan, N.A.; Allen, R.M.; Dhaliwal, A.; Cavassani, K.A.; Schaller, M.; Gallagher, K.A.; Carson, W.F., 4th; Mukherjee, S.; Grembecka, J.; Cierpicki, T.; et al. Cytokine Induced Phenotypic and

- Epigenetic Signatures Are Key to Establishing Specific Macrophage Phenotypes. *PLoS ONE* 2013, 8, e78045.
56. de Groot, A.E.; Pienta, K.J. Epigenetic Control of Macrophage Polarization: Implications for Targeting Tumor-Associated Macrophages. *Oncotarget* 2018, 9, 20908–20927.
57. Otoupalova, E.; Smith, S.; Cheng, G.; Thannickal, V.J. Oxidative Stress in Pulmonary Fibrosis. *Compr. Physiol.* 2011, 10, 509–547.
58. Williams, R.; Laskovs, M.; Williams, R.I.; Mahadevan, A.; Labbadia, J. A Mitochondrial Stress-Specific Form of HSF1 Protects against Age-Related Proteostasis Collapse. *Dev. Cell* 2020, 54, 758–772.e5.
59. Bonam, S.R.; Ruff, M.; Muller, S. HSPA8/HSC70 in Immune Disorders: A Molecular Rheostat That Adjusts Chaperone-Mediated Autophagy Substrates. *Cells* 2019, 8, 849.
60. Tsukahara, K.; Tamatsu, Y.; Sugawara, Y.; Shimada, K. Morphological Study of the Relationship between Solar Elastosis and the Development of Wrinkles on the Forehead and Lateral Canthus. *Arch. Dermatol.* 2012, 148, 913–917.
61. Rossetti, D.; Kielmanowicz, M.G.; Vigodman, S.; Hu, Y.P.; Chen, N.; Nkengne, A.; Oddos, T.; Fischer, D.; Seiberg, M.; Lin, C.B. A Novel Anti-Ageing Mechanism for Retinol: Induction of Dermal Elastin Synthesis and Elastin Fibre Formation. *Int. J. Cosmet. Sci.* 2011, 33, 62–69.
62. Genovese, L.; Corbo, A.; Sibilla, S. An Insight into the Changes in Skin Texture and Properties Following Dietary Intervention with a Nutricosmeceutical Containing a Blend of Collagen Bioactive Peptides and Antioxidants. *Skin Pharmacol. Physiol.* 2017, 30, 146–158.
63. Jiang, M.; Yan, F.; Avram, M.; Lu, Z. A Prospective Study of the Safety and Efficacy of a Combined Bipolar Radiofrequency, Intense Pulsed Light, and Infrared Diode Laser Treatment for Global Facial Photoaging. *Lasers Med. Sci.* 2017, 32, 1051–1061.
64. Charles-de-Sá, L.; Gontijo-de-Amorim, N.F.; Rigotti, G.; Sbarbati, A.; Bernardi, P.; Benati, D.; Bizon Vieira Carias, R.; Maeda Takiya, C.; Borojevic, R. Photoaged Skin Therapy with Adipose-Derived Stem Cells. *Plast. Reconstr. Surg.* 2020, 145, 1037e–1049e.
65. Pain, S.; Berthélémy, N.; Naudin, C.; Degrave, V.; André-Frei, V. Understanding Solar Skin Elastosis-Cause and Treatment. *J. Cosmet. Sci.* 2018, 69, 175–185.
66. Song, Y.; Shen, H.; Du, W.; Goldstein, D.R. Inhibition of X-Box Binding Protein 1 Reduces Tunicamycin-Induced Apoptosis in Aged Murine Macrophages. *Aging Cell* 2013, 12, 794–801.
67. Martinon, F.; Chen, X.; Lee, A.-H.; Glimcher, L.H. TLR Activation of the Transcription Factor XBP1 Regulates Innate Immune Responses in Macrophages. *Nat. Immunol.* 2010, 11, 411–418.
68. Shenderov, K.; Riteau, N.; Yip, R.; Mayer-Barber, K.D.; Oland, S.; Hieny, S.; Fitzgerald, P.; Oberst, A.; Dillon, C.P.; Green, D.R.; et al. Cutting Edge: Endoplasmic Reticulum Stress Licenses

- Macrophages To Produce Mature IL-1 β in Response to TLR4 Stimulation through a Caspase-8– and TRIF-Dependent Pathway. *J. Immunol.* 2014, 192, 2029–2033.
69. Fattah, E.A.; Bhattacharya, A.; Herron, A.; Safdar, Z.; Tony Eissa, N. Critical Role for IL-18 in Spontaneous Lung Inflammation Caused by Autophagy Deficiency. *J. Immunol.* 2015, 194, 5407–5416.
70. Shin, J.N.; Fattah, E.A.; Bhattacharya, A.; Ko, S.; Eissa, N.T. Inflammasome Activation by Altered Proteostasis. *J. Biol. Chem.* 2013, 288, 35886–35895.
71. Sun, W.; Pang, Y.; Liu, Z.; Sun, L.; Liu, B.; Xu, M.; Dong, Y.; Feng, J.; Jiang, C.; Kong, W.; et al. Macrophage Inflammasome Mediates Hyperhomocysteinemia-Aggravated Abdominal Aortic Aneurysm. *J. Mol. Cell. Cardiol.* 2015, 81, 96–106.
72. Xiu, F.; Catapano, M.; Diao, L.; Stanojcic, M.; Jeschke, M.G. Prolonged Endoplasmic Reticulum–Stressed Hepatocytes Drive an Alternative Macrophage Polarization. *Shock* 2015, 44, 44–51.
73. Mahadevan, N.R.; Rodvold, J.; Sepulveda, H.; Rossi, S.; Drew, A.F.; Zanetti, M. Transmission of Endoplasmic Reticulum Stress and pro-Inflammation from Tumor Cells to Myeloid Cells. *Proc. Natl. Acad. Sci. USA* 2011, 108, 6561–6566.
74. Aunan, J.R.; Watson, M.M.; Hagland, H.R.; Søreide, K. Molecular and Biological Hallmarks of Ageing. *Br. J. Surg.* 2016, 103, e29–e46.
75. Barzilai, N.; Ferrucci, L. Insulin Resistance and Aging: A Cause or a Protective Response? *J. Gerontol. A Biol. Sci. Med. Sci.* 2012, 67, 1329–1331.
76. Sanchez-Garrido, J.; Shenoy, A.R. Regulation and Repurposing of Nutrient Sensing and Autophagy in Innate Immunity. *Autophagy* 2020, 1–21.
77. Tabibzadeh, S. Signaling Pathways and Effectors of Aging. *Front. Biosci.* 2021, 26, 50–96.
78. Qin, K.; Han, C.; Zhang, H.; Li, T.; Li, N.; Cao, X. NAD⁺ Dependent Deacetylase Sirtuin 5 Rescues the Innate Inflammatory Response of Endotoxin Tolerant Macrophages by Promoting Acetylation of p65. *J. Autoimmun.* 2017, 81, 120–129.
79. Viola, A.; Munari, F.; Sánchez-Rodríguez, R.; Scolaro, T.; Castegna, A. The Metabolic Signature of Macrophage Responses. *Front. Immunol.* 2019, 10, 1462.
80. Ip, W.K.E.; Hoshi, N.; Shouval, D.S.; Snapper, S.; Medzhitov, R. Anti-Inflammatory Effect of IL-10 Mediated by Metabolic Reprogramming of Macrophages. *Science* 2017, 356, 513–519.
81. Wang, F.; Zhang, S.; Vuckovic, I.; Jeon, R.; Lerman, A.; Folmes, C.D.; Dzeja, P.P.; Herrmann, J. Glycolytic Stimulation Is Not a Requirement for M2 Macrophage Differentiation. *Cell Metab.* 2018, 28, 463–475.e4.

82. Binger, K.J.; Gebhardt, M.; Heinig, M.; Rintisch, C.; Schroeder, A.; Neuhofer, W.; Hilgers, K.; Manzel, A.; Schwartz, C.; Kleinewietfeld, M.; et al. High Salt Reduces the Activation of IL-4- and IL-13-Stimulated Macrophages. *J. Clin. Investig.* 2015, 125, 4223–4238.
83. Huang, S.C.-C.; Smith, A.M.; Everts, B.; Colonna, M.; Pearce, E.L.; Schilling, J.D.; Pearce, E.J. Metabolic Reprogramming Mediated by the mTORC2-IRF4 Signaling Axis Is Essential for Macrophage Alternative Activation. *Immunity* 2016, 45, 817–830.
84. Knipper, J.A.; Willenborg, S.; Brinckmann, J.; Bloch, W.; Maaß, T.; Wagener, R.; Krieg, T.; Sutherland, T.; Munitz, A.; Rothenberg, M.E.; et al. Interleukin-4 Receptor α Signaling in Myeloid Cells Controls Collagen Fibril Assembly in Skin Repair. *Immunity* 2015, 43, 803–816.
85. Minutti, C.M.; Jackson-Jones, L.H.; García-Fojeda, B.; Knipper, J.A.; Sutherland, T.E.; Logan, N.; Ringqvist, E.; Guillamat-Prats, R.; Ferenbach, D.A.; Artigas, A.; et al. Local Amplifiers of IL-4R α -mediated Macrophage Activation Promote Repair in Lung and Liver. *Science* 2017, 356, 1076–1080.
86. Weng, S.-Y.; Wang, X.; Vijayan, S.; Tang, Y.; Kim, Y.O.; Padberg, K.; Regen, T.; Molokanova, O.; Chen, T.; Bopp, T.; et al. IL-4 Receptor Alpha Signaling through Macrophages Differentially Regulates Liver Fibrosis Progression and Reversal. *EBioMedicine* 2018, 29, 92–103.
87. Gan, L.; Zheng, W.; Chabot, J.-G.; Unterman, T.G.; Quirion, R. Nuclear/cytoplasmic Shuttling of the Transcription Factor FoxO1 Is Regulated by Neurotrophic Factors. *J. Neurochem.* 2005, 93, 1209–1219.
88. Betz, C.; Hall, M.N. Where Is mTOR and What Is It Doing There? *J. Cell Biol.* 2013, 203, 563–574.
89. Knuever, J.; Willenborg, S.; Ding, X.; Akyüz, M.D.; Partridge, L.; Niessen, C.M.; Brüning, J.C.; Eming, S.A. Myeloid Cell-Restricted Insulin/IGF-1 Receptor Deficiency Protects against Skin Inflammation. *J. Immunol.* 2015, 195, 5296–5308.
90. Prattichizzo, F.; De Nigris, V.; Mancuso, E.; Spiga, R.; Giuliani, A.; Maccacchione, G.; Lazzarini, R.; Marcheselli, F.; Recchioni, R.; Testa, R.; et al. Short-Term Sustained Hyperglycaemia Fosters an Archetypal Senescence-Associated Secretory Phenotype in Endothelial Cells and Macrophages. *Redox Biol.* 2018, 15, 170–1811.
91. Chung, S.; Ranjan, R.; Lee, Y.G.; Park, G.Y.; Karpurapu, M.; Deng, J.; Xiao, L.; Kim, J.Y.; Unterman, T.G.; Christman, J.W. Distinct Role of FoxO1 in M-CSF- and GM-CSF-Differentiated Macrophages Contributes LPS-Mediated IL-10: Implication in Hyperglycemia. *J. Leukoc. Biol.* 2015, 97, 327–339.
92. Spadaro, O.; Goldberg, E.L.; Camell, C.D.; Youm, Y.-H.; Kopchick, J.J.; Nguyen, K.Y.; Bartke, A.; Sun, L.Y.; Dixit, V.D. Growth Hormone Receptor Deficiency Protects against Age-Related NLRP3 Inflammasome Activation and Immune Senescence. *Cell Rep.* 2016, 14, 1571–1580.

93. Qing, L.; Fu, J.; Wu, P.; Zhou, Z.; Yu, F.; Tang, J. Metformin Induces the M2 Macrophage Polarization to Accelerate the Wound Healing via Regulating AMPK/mTOR/NLRP3 Inflammasome Singling Pathway. *Am. J. Transl. Res.* 2019, 11, 655–668.
94. Koo, S.-J.; Garg, N.J. Metabolic Programming of Macrophage Functions and Pathogens Control. *Redox Biol.* 2019, 24, 101198.
95. Shekhova, E. Mitochondrial Reactive Oxygen Species as Major Effectors of Antimicrobial Immunity. *PLoS Pathog.* 2020, 16, e1008470.
96. Salminen, A.; Ojala, J.; Kaarniranta, K.; Kauppinen, A. Mitochondrial Dysfunction and Oxidative Stress Activate Inflammasomes: Impact on the Aging Process and Age-Related Diseases. *Cell. Mol. Life Sci.* 2012, 69, 2999–3013.
97. Yarbro, J.R.; Emmons, R.S.; Pence, B.D. Macrophage Immunometabolism and Inflammaging: Roles of Mitochondrial Dysfunction, Cellular Senescence, CD38, and NAD. *Immunometabolism* 2020, 2, e200026.
98. Plataki, M.; Cho, S.J.; Harris, R.M.; Huang, H.-R.; Yun, H.S.; Schiffer, K.T.; Stout-Delgado, H.W. Mitochondrial Dysfunction in Aged Macrophages and Lung during Primary Streptococcus Pneumoniae Infection Is Improved with Pirfenidone. *Sci. Rep.* 2019, 9, 971.
99. Giuliani, A.; Prattichizzo, F.; Micolucci, L.; Ceriello, A.; Procopio, A.D.; Rippo, M.R. Mitochondrial (Dys) Function in Inflammaging: Do MitomiRs Influence the Energetic, Oxidative, and Inflammatory Status of Senescent Cells? *Mediat. Inflamm.* 2017, 2017, 2309034.
100. Minhas, P.S.; Liu, L.; Moon, P.K.; Joshi, A.U.; Dove, C.; Mhatre, S.; Contrepois, K.; Wang, Q.; Lee, B.A.; Coronado, M.; et al. Macrophage de Novo NAD⁺ Synthesis Specifies Immune Function in Aging and Inflammation. *Nat. Immunol.* 2019, 20, 50–63.
101. Franceschi, C.; Garagnani, P.; Vitale, G.; Capri, M.; Salvioli, S. Inflammaging and “Garb-Aging”. *Trends Endocrinol. Metab.* 2017, 28, 199–212.
102. Sreedhar, A.; Aguilera-Aguirre, L.; Singh, K.K. Mitochondria in Skin Health, Aging, and Disease. *Cell Death Dis.* 2020, 11, 444.
103. Tanaka, H.; Okada, T.; Konishi, H.; Tsuji, T. The Effect of Reactive Oxygen Species on the Biosynthesis of Collagen and Glycosaminoglycans in Cultured Human Dermal Fibroblasts. *Arch. Dermatol. Res.* 1993, 285, 352–355.
104. Rinnerthaler, M.; Bischof, J.; Streubel, M.K.; Trost, A.; Richter, K. Oxidative Stress in Aging Human Skin. *Biomolecules* 2015, 5, 545–589.
105. Ives, A.; Nomura, J.; Martinon, F.; Roger, T.; LeRoy, D.; Miner, J.N.; Simon, G.; Busso, N.; So, A. Xanthine Oxidoreductase Regulates Macrophage IL1 β Secretion upon NLRP3 Inflammasome Activation. *Nat. Commun.* 2015, 6, 6555.

106. Fuente, M.; Miquel, J. An Update of the Oxidation-Inflammation Theory of Aging: The Involvement of the Immune System in Oxi-Inflamm-Aging. *Curr. Pharm. Des.* 2009, 15, 3003–3026.
107. Herranz, N.; Gil, J. Mechanisms and Functions of Cellular Senescence. *J. Clin. Investig.* 2018, 128, 1238–1246.
108. Gruber, F.; Kremslehner, C.; Eckhart, L.; Tschachler, E. Cell Aging and Cellular Senescence in Skin Aging—Recent Advances in Fibroblast and Keratinocyte Biology. *Exp. Gerontol.* 2020, 130, 110780.
109. Coppé, J.-P.; Desprez, P.-Y.; Krtolica, A.; Campisi, J. The Senescence-Associated Secretory Phenotype: The Dark Side of Tumor Suppression. *Annu. Rev. Pathol.* 2010, 5, 99–118.
110. Kale, A.; Sharma, A.; Stolzing, A.; Desprez, P.-Y.; Campisi, J. Role of Immune Cells in the Removal of Deleterious Senescent Cells. *Immun. Ageing* 2020, 17, 16.
111. Ogata, Y.; Yamada, T.; Hasegawa, S.; Sanada, A.; Iwata, Y.; Arima, M.; Nakata, S.; Sugiura, K.; Akamatsu, H. SASP-Induced Macrophage Dysfunction May Contribute to Accelerated Senescent Fibroblast Accumulation in the Dermis. *Exp. Dermatol.* 2021, 30, 84–91.
112. Gomez, C.R.; Karavitis, J.; Palmer, J.L.; Faunce, D.E.; Ramirez, L.; Nomellini, V.; Kovacs, E.J. Interleukin-6 Contributes to Age-Related Alteration of Cytokine Production by Macrophages. *Mediators Inflamm.* 2010, 2010, 475139.
113. Stout, R.D.; Suttles, J. Immunosenescence and Macrophage Functional Plasticity: Dysregulation of Macrophage Function by Age-Associated Microenvironmental Changes. *Immunol. Rev.* 2005, 205, 60–71.
114. Cao Dinh, H.; Njemini, R.; Onyema, O.O.; Beyer, I.; Liberman, K.; De Dobbeleer, L.; Renmans, W.; Vander Meeren, S.; Jochmans, K.; Delaere, A.; et al. Strength Endurance Training but Not Intensive Strength Training Reduces Senescence-Prone T Cells in Peripheral Blood in Community-Dwelling Elderly Women. *J. Gerontol. A Biol. Sci. Med. Sci.* 2019, 74, 1870–1878.
115. Díaz-Del Cerro, E.; Vida, C.; Martínez de Toda, I.; Félix, J.; De la Fuente, M. The Use of a Bed with an Insulating System of Electromagnetic Fields Improves Immune Function, Redox and Inflammatory States, and Decrease the Rate of Aging. *Environ. Health* 2020, 19, 118.
116. Sotiropoulou, P.A.; Blanpain, C. Development and Homeostasis of the Skin Epidermis. *Cold Spring Harb. Perspect. Biol.* 2012, 4, a008383.
117. Dunnwald, M.; Chinnathambi, S.; Alexandrunas, D.; Bickenbach, J.R. Mouse Epidermal Stem Cells Proceed through the Cell Cycle. *J. Cell. Physiol.* 2003, 195, 194–201.
118. Li, L.; Ngo, H.T.T.; Hwang, E.; Wei, X.; Liu, Y.; Liu, J.; Yi, T.-H. Conditioned Medium from Human Adipose-Derived Mesenchymal Stem Cell Culture Prevents UVB-Induced Skin Aging in Human Keratinocytes and Dermal Fibroblasts. *Int. J. Mol. Sci.* 2019, 21, 49.

119. Ma, T.; Fu, B.; Yang, X.; Xiao, Y.; Pan, M. Adipose Mesenchymal Stem Cell-Derived Exosomes Promote Cell Proliferation, Migration, and Inhibit Cell Apoptosis via Wnt/ β -Catenin Signaling in Cutaneous Wound Healing. *J. Cell. Biochem.* 2019, 120, 10847–10854.
120. Castilho, R.M.; Squarize, C.H.; Chodosh, L.A.; Williams, B.O.; Gutkind, J.S. mTOR Mediates Wnt-Induced Epidermal Stem Cell Exhaustion and Aging. *Cell Stem Cell* 2009, 5, 279–289.
121. Ide, S.; Yahara, Y.; Kobayashi, Y.; Strausser, S.A.; Ide, K.; Watwe, A.; Xu-Vanpala, S.; Privratsky, J.R.; Crowley, S.D.; Shinohara, M.L.; et al. Yolk-Sac-Derived Macrophages Progressively Expand in the Mouse Kidney with Age. *eLife* 2020, 9, e51756.
122. Ferrer, I.R.; West, H.C.; Henderson, S.; Ushakov, D.S.; e Sousa, P.S.; Strid, J.; Chakraverty, R.; Yates, A.J.; Bennett, C.L. A Wave of Monocytes Is Recruited to Replenish the Long-Term Langerhans Cell Network after Immune Injury. *Sci. Immunol.* 2019, 4, eaax8704.
123. Lucas, T.; Waisman, A.; Ranjan, R.; Roes, J.; Krieg, T.; Müller, W.; Roers, A.; Eming, S.A. Differential Roles of Macrophages in Diverse Phases of Skin Repair. *J. Immunol.* 2010, 184, 3964–3977.
124. Rodrigues, M.; Kosaric, N.; Bonham, C.A.; Gurtner, G.C. Wound Healing: A Cellular Perspective. *Physiol. Rev.* 2019, 99, 665–706.
125. Fisher, G.; Rittié, L. Restoration of the Basement Membrane after Wounding: A Hallmark of Young Human Skin Altered with Aging. *J. Cell Commun. Signal.* 2018, 12, 401–411.
126. Lee, S.; Szilagy, E.; Chen, L.; Premanand, K.; DiPietro, L.A.; Ennis, W.; Bartholomew, A.M. Activated Mesenchymal Stem Cells Increase Wound Tensile Strength in Aged Mouse Model via Macrophages. *J. Surg. Res.* 2013, 181, 20–24.
127. Morgun, E.I.; Vorotelyak, E.A. Epidermal Stem Cells in Hair Follicle Cycling and Skin Regeneration: A View From the Perspective of Inflammation. *Front. Cell Dev. Biol.* 2020, 8, 581697.
128. Kolter, J.; Feuerstein, R.; Zeis, P.; Hagemeyer, N.; Paterson, N.; d'Errico, P.; Baasch, S.; Amann, L.; Masuda, T.; Lösslein, A.; et al. A Subset of Skin Macrophages Contributes to the Surveillance and Regeneration of Local Nerves. *Immunity* 2019, 50, 1482–1497.e7.
129. Shook, B.A.; Wasko, R.R.; Rivera-Gonzalez, G.C.; Salazar-Gatzimas, E.; López-Giráldez, F.; Dash, B.C.; Muñoz-Rojas, A.R.; Aultman, K.D.; Zwick, R.K.; Lei, V.; et al. Myofibroblast Proliferation and Heterogeneity Are Supported by Macrophages during Skin Repair. *Science* 2018, 362, eaar2971.
130. Zhuang, Y.; Lyga, J. Inflammaging in Skin and Other Tissues—the Roles of Complement System and Macrophage. *Inflamm. Allergy Drug Targets* 2014, 13, 153–161.

131. Doles, J.; Storer, M.; Cozzuto, L.; Roma, G.; Keyes, W.M. Age-Associated Inflammation Inhibits Epidermal Stem Cell Function. *Genes Dev.* 2012, 26, 2144–2153.
132. Rando, T.A.; Chang, H.Y. Aging, Rejuvenation, and Epigenetic Reprogramming: Resetting the Aging Clock. *Cell* 2012, 148, 46–57.
133. Pilkington, S.M.; Barron, M.J.; Watson, R.E.B.; Griffiths, C.E.M.; Bulfone-Paus, S. Aged Human Skin Accumulates Mast Cells with Altered Functionality That Localize to Macrophages and Vasoactive Intestinal Peptide-Positive Nerve Fibres. *Br. J. Dermatol.* 2019, 180, 849–858.
134. Fulop, T.; Witkowski, J.M.; Olivieri, F.; Larbi, A. The Integration of Inflammaging in Age-Related Diseases. *Semin. Immunol.* 2018, 40, 17–35.

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