

HDL in Sepsis and SARS-CoV-2

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High-density lipoproteins (HDLs) are a class of blood particles, principally involved in mediating reverse cholesterol transport from peripheral tissue to liver. Omics approaches have identified crucial mediators in the HDL proteomic and lipidomic profile, which are involved in distinct pleiotropic functions. Besides their role as cholesterol transporter, HDLs display anti-inflammatory, anti-apoptotic, anti-thrombotic, and anti-infection properties. Experimental and clinical studies have unveiled significant changes in both HDL serum amount and composition that lead to dysregulated host immune response and endothelial dysfunction in the course of sepsis. Most SARS-Coronavirus-2-infected patients admitted to the intensive care unit showed common features of sepsis disease, such as the overwhelmed systemic inflammatory response and the alterations in serum lipid profile. Despite relevant advances, episodes of mild to moderate acute kidney injury (AKI), occurring during systemic inflammatory diseases, are associated with long-term complications, and high risk of mortality. The multi-faceted relationship of kidney dysfunction with dyslipidemia and inflammation encourages to deepen the clarification of the mechanisms connecting these elements.

Keywords: lipid profile changes ; dysfunctional HDL ; sepsis ; SARS-CoV-2 infection ; acute kidney injury (AKI)

1. Introduction

Infection and inflammation induce marked alteration of the lipid and lipoprotein profile, leading to impairment of host immune response and worsen outcome ^[1]. Accumulating evidence has shown that viruses, bacteria, and fungal pathogens are able to manipulate host lipid pathways, modifying energy storage and production in cells, immune signaling, and tissue repair ^[1].

High-density lipoprotein (HDL) is a key component of circulating blood and mainly contains phospholipids, free cholesterol, cholesteryl ester, triglycerides, apolipoproteins, and other proteins. Besides its role in reverse cholesterol transport, HDL displays pleiotropic functions during inflammation and endothelial dysfunction, decreasing inflammatory signaling in immune effector cells and inhibiting endothelial response ^{[2][3]}. It is increasingly recognized that HDL can bind and neutralize viruses and toxic bacterial substances such as lipopolysaccharide (LPS) ^{[4][5][6]} and lipoteichoic acid ^[7]. The binding of LPS by HDL has been shown to protect animals from the toxicity of endotoxin ^{[8][9]}. Moreover, HDL could block certain viruses to penetrate cells, reducing tissue invasion ^{[1][10]}.

Clinical studies have demonstrated that HDL levels drop by 40–70% during systemic inflammation, leading to a poor prognosis in septic subjects ^{[11][12][13][14][15]}. Moreover, low levels of HDL have been associated with increased risk of acute kidney injury (AKI) in the course of sepsis ^{[16][17]}. Renal function and plasma HDL are strongly related to each other as kidneys are implicated in the recycling of senescent HDL particles and their filtration function is associated with their levels and contents ^[18].

The new respiratory infectious disease, coronavirus disease 2019 (COVID-19), showed common features of sepsis pathogenesis, such as the dysregulated host immune response, alteration in serum lipids, endothelial dysfunction, and changes in the coagulation system ^{[19][20]}. In addition, some COVID-19 patients are admitted to the intensive care unit (ICU) developing acute respiratory distress syndrome (ARDS), acute cardiac injury, acute kidney injury (AKI), and shock ^{[21][22][23]}.

Since the major contributors to mortality in septic patients are the hyperinflammation, the endothelial dysfunction, and low HDL is prognostic of worsen outcome, considering the similarities between sepsis and COVID-19 infection, it is reasonable that HDL replacement therapy has been a well sought-after area of systemic inflammatory syndrome therapies.

2. HDLs Composition and Reverse Cholesterol Transport Function

HDLs are defined as a class of complex nanoparticles that consist of a broad variety of lipids and proteins [24]. Their structure includes a hydrophobic nucleus composed by triglycerides and esterified cholesterol and an external part with phospholipids and free cholesterol, associated with proteins named apoproteins [24].

The protein and lipid composition ratio depends on several physiological and pathological factors. The different protein and lipid cargo determines HDLs particles subdivision in five distinct classes: very large, large, medium, small, and very small HDL [24][25].

The principal pleiotropic function of HDL is Reverse Cholesterol Transport (RCT), leading the delivery of cholesterol from peripheral tissue to the liver where it is metabolized [26]. The cholesterol mobilization and transport are necessary to avoid its accumulation in cells, preventing cellular apoptosis [27][28]. This process starts with the biosynthesis of premature HDL, composed principally by ApoA-I, in the liver and small intestine [26]. Then, this premature HDL acquires phospholipids and free cholesterol through the interaction with ATP-binding cassette transporter A1 (ABCA1), principally expressed by macrophages and hepatocytes and becomes pre- β HDL [26]. The absence of a non-polar lipids core in pre- β HDL induces a disk-shaped structure, recognized as nascent HDL [26]. This nascent HDL quickly evolves toward mature HDL acquiring cholesterol, cholesterol ester, and triglycerides, and this remodeling process is catalyzed by a specific enzyme called lecithin-cholesterol acyltransferase (LCAT) [26]. Mature HDLs continue to acquire free cholesterol and apolipoproteins and exchange esterified cholesterol with triglycerides from other lipoproteins due to the action of lipid transfer proteins such as cholesteryl ester transfer protein (CETP) [26]. Then, these large HDLs return to the liver, bind the scavenger receptor class B type I (SR-BI), and facilitate cholesterol esters uptake, leading to their degradation by hepatocytes and their excretion into the bile [26]. This process is followed by the synthesis and release of a new premature HDL, enhancing a new cycle of RCT. There is a second mechanism for cholesterol degradation that is mediated by mature low-density lipoproteins (LDLs) that acquire cholesterol from HDLs by CEPT and are internalized by hepatocytes via the LDL receptor (LDL-R) [29].

The analysis of the HDL proteomic profile revealed a large protein content mainly characterized by ApoA-I, other major apolipoproteins including ApoA-II, Apo-CI/CII/CIII, and remodeling HDL enzymes and minor crucial proteins such as complement and coagulation regulatory factors, protease inhibitors, and acute-phase proteins that reflect HDL involvement in distinct pleiotropic functions [30]. The role of ApoA-II is still not clear in inflammatory diseases and some studies reported pro-inflammatory and anti-inflammatory role, respectively [31][32]. Indeed, Furlaneto et al. demonstrated that ApoA-II hampered neutrophil activation, reducing oxidative burst and IL-8 synthesis [32]. Another study showed that ApoA-II could increase monocyte response to LPS, thereby amplifying inflammatory reaction [31]. A minor apolipoprotein found in HDL is the apolipoprotein M (ApoM) that transports the Sphingosine 1-phosphate (S1P), a lysophospholipid mediator involved in several physiological functions such as cellular proliferation and survival, endothelial nitric oxide synthase (eNOS) activation, and inhibition of inflammation and endothelial dysfunction [33]. Recently, Kurano et al. demonstrated that ApoM was not only a carrier but it regulated and increased S1P content, reducing acute lung injury (ALI) and incidence of mortality in the endotoxemic mouse model [34].

Another functional protein, discovered by mapping HDL protein content by two-dimensional gel electrophoresis and mass spectrometry, is the alpha-1 antitrypsin (AAT), a natural inhibitor of serine proteases, chiefly neutrophil elastase but also chymotrypsin, cathepsin G (CathG), proteinase 3 (PR3), and thrombin [35]. AAT deficiency is strongly associated with an increasing risk to develop pulmonary emphysema due to lung damage caused by neutrophil elastase activation. To prevent pulmonary aggression by elastase, infusion of purified AAT proteins has been used as treatment not only in AAT deficient conditions but also in other inflammatory diseases [36].

The lipid composition of HDL includes different species among phospholipids, free and esterified cholesterol, sphingolipids, and triglycerides that could protect cells from infection. Indeed, Bricarello et al. found that gangliosides fixed in reconstituted HDLs (rHDLs) could bind cholera toxin, limiting its detrimental effects [37]. As previously indicated, S1P is an important player of inflammatory response and endothelial dysfunction [33]. In particular, S1P induces eNOS pathway activation and inhibition of monocyte chemotactic protein-1 (MCP-1) synthesis in endothelial cells by binding S1P-receptor-1 [38][39][40][41]. Several studies reported its involvement in regulating endothelial permeability, cytokines release, and inflammatory response in sepsis context [42][43]. In a single-center prospective-observational study, Winkler et al. showed a strong inverse correlation between Sequential Organ Failure Assessment (SOFA) scores and serum-S1P levels in septic patients, underlying the potential role of lipids in systemic inflammatory diseases [42].

3. Kidney Involvement in Sepsis Disease: From HDL Target to Modulator

Acute kidney injury (AKI) is a common event in ICU patients, with an estimated incidence of >50% [44]. Furthermore, increasing AKI severity is associated with higher risk of mortality [45]. Sepsis is defined as the major cause of AKI [46], accounting for 45% to 70% of cases, and approximately 25% of sepsis is of intra-abdominal origin [47].

Early diagnosis of AKI in the setting of sepsis is important in order to provide optimal treatment and avoid further kidney injury [48]. Therefore, the use of injury or stress biomarkers, in addition to the clinical assessment of renal function, is required [49][50]. Inflammation appears to play an important role in sepsis-related kidney injury. Interleukin-6 (IL-6) has been described as predictive of AKI, independently of hypotension (e.g., mean arterial pressure, dosage of vasopressors) [51].

Since HDL levels significantly declined in severe sepsis and septic shock, several studies evaluated HDL impact on renal damage as a possible predictive biomarker and therapeutic target [52]. In a blinded, observational cohort study, the authors analyzed HDL amount in 200 adult patients with suspected sepsis at the time of admission [12]. They observed that HDL concentration is a prognostic factor to establish an early efficient therapy to avoid disease progression, multi-organ dysfunction, and renal damage [12]. In another observational small study of 180 patients with clear signs of infections, the authors demonstrated that low HDL levels are associated with increased risk of AKI onset and decreased glomerular filtration rate (GFR) [16]. These results suggest that HDL levels may accurately predict the development and the stages of AKI like serum creatinine levels for the Kidney Disease Improving Global Outcomes (KDIGO) index [16]. Interestingly, low levels of HDL in the early phase of sepsis could represent predictors of worsening renal function at three to twenty-four months after hospital discharge of septic patients with a dyslipidemia profile [16]. These results underline the key role of HDL in affecting renal function and disease progression.

In addition, polymorphisms of genes involved in HDL metabolism have been found strongly associated with increased risk of AKI during sepsis. Accordingly, Genga K.R. et al. retrospectively compared two cohorts of septic patients and demonstrated that one genetic variant in the CETP gene, rs1800777 (allele A), determined low levels of HDL and an increased risk to develop sepsis-related AKI [53].

Most of the current understanding of HDL involvement in sepsis pathogenesis and AKI have been revealed from animal models and in vitro studies. For example, Guo L. et al. demonstrated that higher expression of ApoA-I in transgenic mice increased survival and limited sepsis and renal damage compared to wild type mice [8][9]. HDL might provide renal protection through several mechanisms including pathogens detoxification via SR-BI internalization. As discussed above, SR-BI is expressed by immune cells, endothelial cells, and parenchymal cells such as hepatocytes and renal tubular cells and mediates the cholesterol efflux to circulating HDL. In addition, pathogens molecules as LPS and LTA could bind HDL in the bloodstream and could be transferred to SR-BI for hepatic clearance. Decreased serum levels of LPS or LTA induced lower activation of TLR-4 and TLR-2 respectively, in renal parenchymal cells [7][54][55][56]. Since both TLR-4 and TLR-2 activation is associated with increased renal damage [57][58], it is obviously that HDL levels ameliorate renal function, avoiding tubular damage, interstitial inflammation, and cytokines release.

Endothelial dysfunction is a hallmark of sepsis-associated AKI [59]. In inflammatory conditions, endothelial cells increased inducible nitric oxide synthase activity (iNOS) and decreased eNOS activation [60], leading to vascular impairment and renal parenchymal damage [59]. Increased levels of HDL could activate the eNOS pathway, reducing adhesion molecules expression and leucocytes activation and neutrophil infiltration, assuring reduced renal damage [61].

Renal tubular cells are not only the principal target of renal dysfunction, but they are directly involved in HDL modulation [62]. In the physiological condition, the glomerular filtration barrier is intact and prevents passage of large molecules such as mature HDL. Only lipid poor ApoA-I, ApoA-II, ApoA-IV and enzymes such as LCAT cross the filtration barrier. Then, renal tubular cells express on their surface membrane specific receptors to bind and internalize these particles. In particular, pre- β HDL and lipid poor ApoA-I bind to the cubulin amnion less complex on the surface of renal proximal tubule cells and are endocytosed; this process is accelerated by the binding of another membrane protein, megalin, that is essential for renal proximal tubule reabsorption [62]. After endocytosis, HDL could be degraded in tubular lysosome or transcytosed into intratubular lumen in order to be released into systemic circulation (Figure 1).

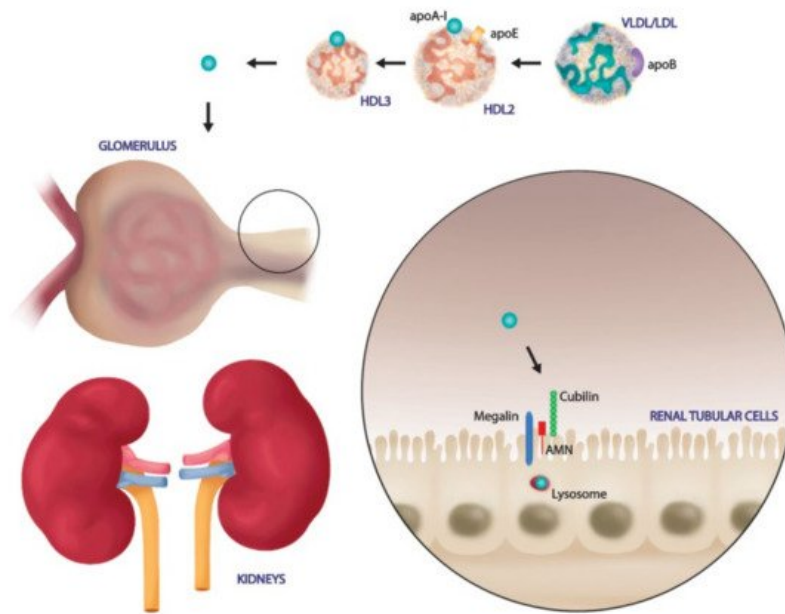


Figure 1. Renal HDL catabolism and transport. Senescent circulating HDLs are filtered in the glomerular capillaries. Renal tubular cells bind pre-B HDL particles or lipid poor ApoA-I via the Cubilin-amnionless complex and Megalin. HDL dissociates from tubulin in their endocytic vesicles and HDL is degraded into lysosome.

In early stages of renal dysfunction, the GFR is impaired and increased levels of Apo A-IV and LCAT are found in urines [63][64]. Certainly, the appearance of tubular damage determines reduced tubular reabsorption function and catabolism, contributing to loss HDL and its components. Accordingly, Aseem O. et al. reported that cubilin deficiency caused impaired renal transcytosis and consequent decreased serum levels of albumin, ApoA-I and HDL3 [65].

Moreover, kidneys can also affect HDL metabolism, influencing extrarenal synthesis and metabolism of HDL components [66]. For example, nephrotoxic syndrome induces a significant reduced expression of hepatic lipase in liver parenchyma, causing decreasing uptake of HDL triglycerides and phospholipids, increasing circulating cholesterol droplets, and impairing HDL maturation [67]. Therefore, augmented renal proteinuria caused profound alteration in plasma and liver enzymes function, avoiding HDL maturation, determining profound impact in several organs including kidneys [67].

It is clear that kidney injury could alter HDL levels and composition, but further studies are needed to evaluate the effects of other receptors, as SR-BI and ABACG1 in renal HDL catabolism or transcytosis.

4. SARS-CoV-2 Exploits the SR-BI Associated HDL to Promote its Entry

Despite the great efforts to elucidate the molecular mechanisms of SARS-CoV2 cell entry, the pathogenetic mechanisms have not been fully clarified and there are no effective therapies to stop or reverse the occurred infection. The partial inhibition obtained by ACE-2 antibodies in contraposition to maximum neutralization efficacy of monoclonal antibodies targeting the SARS-2-S1 N-terminal domain suggested that viral particles exploited additional receptors and channels for host cell entry [68][69][70]. In accordance, besides ACE-2, other receptors, such as Dipeptidyl peptidase 4 (DPP4) or aminopeptidase N, have been proposed to play a role in SARS-CoV2 entry in target cells. Specifically, DPP4, also known as cluster of differentiation 26 (CD26), a serine exopeptidase expressed ubiquitously in lung, kidney, liver, gut, and immune cells, has been investigated to allow virus SARS-CoV-2 cell adhesion/virulence [71].

Among additional mechanisms of enhancement SARS-CoV-2 entry, SR-BI has been recently examined to play a central role in augmenting virus attachment. As previously underlined, SR-BI is a kind of membrane protein with a molecular weight of 82 kDa, that acts as the major HDL scavenger receptor and mediates the selective HDL-cholesterol uptake of cells [72]. This cholesterol delivery system is well studied in hepatocytes as well as steroidogenic cells as adipocytes, adrenal cells but also in fibroblasts, macrophages, ovarian cells, and testicular Leydig cells [73]. Notably, epithelial alveolar type II cells also express SR-BI, in this location, SR-BI is involved in vitamin E intake, specially from HDL [74][75].

In an interesting in vitro study, Wei et al. demonstrated that SARS-COV2 exploited the physiological function of SR-BI to mediate cholesterol-bound HDL intake to promote its cellular entry [70]. In other words, even if SR-BI cannot bind to the SARS-2-S protein directly, the receptor acts as a linker molecule that recruits viral particles to come into proximity with ACE-2 by association with HDL. In accordance, the SR-BI expression conferred the greatest cell susceptibility to SARS-CoV-2 only when co-expressed with ACE-2. Therefore, it emerged that treatment of cultured cells with pharmacological

SR-BI antagonists, could inhibit HDL-enhanced SARS-CoV-2 infection inhibition. Furthermore, also blockade of the cholesterol-binding site on SARS-2-S1 with a monoclonal antibody showed similar effects. In line, ITX 5601, a clinically approved inhibitor of HCV infection, strongly inhibits SARS-CoV-2 infection of cultured cells. Strikingly, besides lung, authors further showed that SR-BI is co-expressed with ACE2, predominately in lower respiratory tissues that are more affected by COVID-19 and in several extrapulmonary tissues as retina, testis, as well as kidney which could indicate an extended tropism for extrapulmonary tissues, thereby to the multiple-organ pathologies of COVID-19 [76].

SARS-COV-2 has a lipid envelope that merges with the host cell through endocytosis, internalizing its components in the cell. Recently, cell experiments showed that the susceptibility of the virus to fusion with the host cell membrane augmented when cholesterol was added to medium culture [77]. For example, cholesterol-supplemented mouse fibroblasts showed increased susceptibility to fusion with murine hepatitis virus [78]. ACE-2 resides mainly within lipid rafts, furthermore, in lipid raft areas, there are also caveolins, clathrins, and dynamin, that are molecules with a central role in the incorporation of viruses [79]. Similarly, uptake mechanisms dependent from these molecules and on the presence of lipid rafts have been also identified for the simian virus (SV40) [77].

In the context of HDL and lipoproteins metabolism, it could be interesting to note the existence of an association between ACE insertion/deletion polymorphism with HDL level and cardiovascular disease risk factors. As recently demonstrated, an ACE polymorphism in men was significantly associated with reduced HDL cholesterol levels [80]. Cholesterol is an essential lipid component of vertebrate cell membranes, mainly through lipid rafts, i.e., microdomains enriched in cholesterol and sphingolipids. Lipid rafts have been suggested to play a fundamental role in several biological processes such as signal transduction, membrane trafficking, cytoskeletal organization [81][82].

Given their unique protein composition, lipid raft microdomains are essential for the endocytosis-mediated process and serve as a platform and docking site for viruses to enter the host cell [79][81]. Emerging evidence revealed that cholesterol diminution from cellular membranes has been shown to hamper SARS-CoV-2 infection [83]. This finding led several researchers to speculate that SARS-CoV-2 may exploit the physiological function of SR-BI to achieve its entry and fusion processes [84].

Furthermore, Meher et al. reported the effect of membrane cholesterol on the structure of the fusion peptide (residues 770–788) of S2 glycoprotein of SARS-CoV. Authors elegantly demonstrated that S2 binding affinity augmented with increasing levels of membrane cholesterol [85]. On the other hand, cholesterol decline disturbs the virion membrane. Through the interference with lipid-dependent attachment to human host cells, sterols and cyclodextrins can reduce the infectivity of CoVs. [86]. In vitro depletion of membrane-bound cholesterol in ACE2-expressing cells led to a reduced infectivity of CoVs, since the binding of the spike protein was reduced by half. The mechanism of employing lipid raft rich in cholesterol together with the utilization of another type of lipid called monosialotetrahexosylganglioside 1 (GM1), to enter mammalian cells in culture is shared by both SARS-CoV and SARS-CoV-2. This concept has emerged by the reduction of infection in cells treated with a compound called methyl- β -cyclodextrin (M β CD) able to deplete cholesterol [87]. The cholesterol removal by M β CD significantly dissociate the number of bonds between ACE-2 membrane protein and viral S glycoproteins [88].

Some studies showed that M β CD treatment dose-dependently reduced expression of ACE-2 in the cell membrane, also reducing the infectivity of SARS-CoV2 [78]. The lipophilic core permits the contact of these molecules with lipid rafts. These harmless macromolecules are able to mimic binding domains for the enveloped virus, competing with host cell attack sites, thereby reducing infection.

Additionally, interaction of phytosterols with lipid raft molecules can lead to a decrease of membrane cholesterol content or destabilization of its structure, thereby affecting viral infectivity [78]. In addition, the viral infectivity is modulated by homeostatic control of cholesterol content and fatty acid metabolism [89]. More recently, Henrich, S. E. et al. elegantly demonstrated that SARS-CoV-2 viral entry is impaired by SR-BI genetic knockdown, suggesting that SR-BI is a co-receptor for SARS-CoV-2. Interestingly, authors also demonstrated that inorganic core nanoparticles around which HDL-associated protein (apolipoprotein A-I) and lipids were clustered (HDL-particles) targeted SR-BI to inhibit SARS-CoV-2 entry. Indeed, the superficial resemblance to HDLs caused these nanoparticles to firmly bind to SR-BI. These nanoparticles are characterized by the ability to strongly inhibit the entry of exosomes, which are extracellular lipid vesicles. Finally, by altering the type of lipids assembled on the surface of these nanoparticles, they could target Gram-negative bacterial LPS and prevent the LPS-mediated release of potent pro-inflammatory signalling.

Based on these findings, HDL nanoparticles could offer a promising strategy to prevent infection with SARS-CoV-2. Strikingly, they could also be investigated as a possible treatment for other cholesterol-dependent viral infections that are

References

1. Khovidhunkit, W.; Kim, M.S.; Memon, R.A.; Shigenaga, J.K.; Moser, A.H.; Feingold, K.R.; Grunfeld, C. Effects of infection and inflammation on lipid and lipoprotein metabolism: Mechanisms and consequences to the host. *J. Lipid Res.* 2004, 45, 1169–1196.
2. Singh, I.M.; Shishehbor, M.H.; Ansell, B.J. High-density lipoprotein as a therapeutic target: A systematic review. *J. Am. Med. Assoc.* 2007, 298, 786–798.
3. Navab, M.; Reddy, S.T.; Van Lenten, B.J.; Fogelman, A.M. HDL and cardiovascular disease: Atherogenic and atheroprotective mechanisms. *Nat. Rev. Cardiol.* 2011, 8, 222–232.
4. Emancipator, K.; Csako, G.; Elin, R.J. In vitro inactivation of bacterial endotoxin by human lipoproteins and apolipoproteins. *Infect. Immun.* 1991, 60, 596–601.
5. Munford, R.S. Detoxifying endotoxin: Time, place and person. *J. Endotoxin Res.* 2005, 11, 69–84.
6. Lee, R.P.; Lin, N.T.; Chao, Y.F.C.; Lin, C.C.; Harn, H.J.; Chen, H.I. High-density lipoprotein prevents organ damage in endotoxemia. *Res. Nurs. Health* 2007, 30, 250–260.
7. Grunfeld, C.; Marshall, M.; Shigenaga, J.K.; Moser, A.H.; Tobias, P.; Feingold, K.R. Lipoproteins inhibit macrophage activation by lipoteichoic acid. *J. Lipid Res.* 1999, 40, 245–252.
8. Guo, L.; Ai, J.; Zheng, Z.; Howatt, D.A.; Daugherty, A.; Huang, B.; Li, X.-A. High density lipoprotein protects against polymicrobe-induced sepsis in mice. *J. Biol. Chem.* 2013, 288, 17947–17953.
9. Guo, L.; Song, Z.; Li, M.; Wu, Q.; Wang, D.; Feng, H.; Bernard, P.; Daugherty, A.; Huang, B.; Li, X.A. Scavenger receptor BI protects against septic death through its role in modulating inflammatory response. *J. Biol. Chem.* 2009, 284, 19826–19834.
10. Feingold, K.R.; Grunfeld, C. Lipids: A key player in the battle between the host and microorganisms. *J. Lipid Res.* 2012, 53, 2487–2489.
11. van Leeuwen, H.J.; Heezius, E.C.J.M.; Dallinga, G.M.; van Strijp, J.A.G.; Verhoef, J.; van Kessel, K.P.M. Lipoprotein metabolism in patients with severe sepsis. *Crit. Care Med.* 2003, 31, 1359–1366.
12. Chien, J.-Y.; Jerng, J.-S.; Yu, C.-J.; Yang, P.-C. Low serum level of high-density lipoprotein cholesterol is a poor prognostic factor for severe sepsis. *Crit. Care Med.* 2005, 33, 1688–1693.
13. Tsai, M.-H.; Peng, Y.-S.; Chen, Y.-C.; Lien, J.-M.; Tian, Y.-C.; Fang, J.-T.; Weng, H.-H.; Chen, P.-C.; Yang, C.-W.; Wu, C.-S. Low serum concentration of apolipoprotein A-I is an indicator of poor prognosis in cirrhotic patients with severe sepsis. *J. Hepatol.* 2009, 50, 906–915.
14. Eggesbø, J.B.; Hjermann, I.; Høstmark, A.T.; Kierulf, P. LPS induced release of IL-1 β , IL-6, IL-8 and TNF- α in EDTA or heparin anticoagulated whole blood from persons with high or low levels of serum HDL. *Cytokine* 1996, 8, 152–160.
15. Morin, E.E.; Guo, L.; Schwendeman, A.; Li, X.A. HDL in sepsis-risk factor and therapeutic approach. *Front. Pharmacol.* 2015, 6.
16. Roveran Genga, K.; Lo, C.; Cirstea, M.; Zhou, G.; Walley, K.R.; Russell, J.A.; Levin, A.; Boyd, J.H. Two-year follow-up of patients with septic shock presenting with low HDL: The effect upon acute kidney injury, death and estimated glomerular filtration rate. *J. Intern. Med.* 2017, 281, 518–529.
17. Zhang, Z.; Datta, G.; Zhang, Y.; Miller, A.P.; Mochon, P.; Chen, Y.F.; Chatham, J.; Anantharamaiah, G.M.; White, C.R. Apolipoprotein A-I mimetic peptide treatment inhibits inflammatory responses and improves survival in septic rats. *Am. J. Physiol. Heart Circ. Physiol.* 2009, 297, H866–H873.
18. Yang, H.; Fogo, A.B.; Kon, V. Kidneys: Key modulators of high-density lipoprotein levels and function. *Curr. Opin. Nephrol. Hypertens.* 2016, 25, 174–179.
19. Li, H.; Liu, L.; Zhang, D.; Xu, J.; Dai, H.; Tang, N.; Su, X.; Cao, B. SARS-CoV-2 and viral sepsis: Observations and hypotheses. *Lancet* 2020, 395, 1517–1520.
20. Remy, K.E.; Brakenridge, S.C.; Francois, B.; Daix, T.; Deutschman, C.S.; Monneret, G.; Jeannet, R.; Laterre, P.F.; Hotchkiss, R.S.; Moldawer, L.L. Immunotherapies for COVID-19: Lessons learned from sepsis. *Lancet Respir. Med.* 2020, 8, 946–949.
21. Sun, P.; Lu, X.; Xu, C.; Sun, W.; Pan, B. Understanding of COVID-19 based on current evidence. *J. Med. Virol.* 2020, 92, 548–551.

22. Perico, L.; Benigni, A.; Remuzzi, G. Should COVID-19 Concern Nephrologists? Why and to What Extent? the Emerging Impasse of Angiotensin Blockade. *Nephron* 2020, 144, 213–221.
23. World Health Organization. Mission China Joint Report of the WHO-China Joint Mission on Coronavirus Disease 2019 (COVID-19); World Health Organization: Geneva, Switzerland, 2020; Volume 2019, pp. 16–24. Available online: (accessed on 7 April 2020).
24. Feingold, K.R. Introduction to Lipids and Lipoproteins; MDText.com, Inc.: South Dartmouth, MA, USA, 2000.
25. Otvos, J.D. Measurement of lipoprotein subclass profiles by nuclear magnetic resonance spectroscopy. *Clin. Lab.* 2002, 48, 171–180.
26. Von Eckardstein, A.; Nofer, J.R.; Assmann, G. High density lipoproteins and arteriosclerosis role of cholesterol efflux and reverse cholesterol transport. *Arterioscler. Thromb. Vasc. Biol.* 2001, 21, 13–27.
27. Suc, I.; Escargueil-Blanc, I.; Trolly, M.; Salvayre, R.; Negre-Salvayre, A. HDL and apoA prevent cell death of endothelial cells induced by oxidized LDL. *Arterioscler. Thromb. Vasc. Biol.* 1997, 17, 2158–2166.
28. Fielding, C.J.; Fielding, P.E. Molecular physiology of reverse cholesterol transport. *J. Lipid Res.* 1995, 36, 211–228.
29. Lewis, G.F.; Rader, D.J. New insights into the regulation of HDL metabolism and reverse cholesterol transport. *Circ. Res.* 2005, 96, 1221–1232.
30. Shah, A.S.; Tan, L.; Long, J.L.; Davidson, W.S. Proteomic diversity of high density lipoproteins: Our emerging understanding of its importance in lipid transport and beyond. *J. Lipid Res.* 2013, 54, 2575–2585.
31. Thompson, P.A.; Berbée, J.F.P.; Rensen, P.C.N.; Kitchens, R.L. Apolipoprotein A-II augments monocyte responses to LPS by suppressing the inhibitory activity of LPS-binding protein. *Innate Immun.* 2008, 14, 365–374.
32. Furlaneto, C.J.; Ribeiro, F.P.; Hatanaka, E.; Souza, G.M.; Cassatella, M.A.; Campa, A. Apolipoproteins A-I and A-II downregulate neutrophil functions. *Lipids* 2002, 37, 925–928.
33. Schuchardt, M.; Tölle, M.; Prüfer, J.; Van Der Giet, M. Pharmacological relevance and potential of sphingosine 1-phosphate in the vascular system. *Br. J. Pharmacol.* 2011, 163, 1140–1162.
34. Kurano, M.; Tsuneyama, K.; Morimoto, Y.; Shimizu, T.; Jona, M.; Kassai, H.; Nakao, K.; Aiba, A.; Yatomi, Y. Apolipoprotein M Protects Lipopolysaccharide-Treated Mice from Death and Organ Injury. *Thromb. Haemost.* 2018, 118, 1021–1035.
35. Karlsson, H.; Leanderson, P.; Tagesson, C.; Lindahl, M. Lipoproteomics I: Mapping of proteins in low-density lipoprotein using two-dimensional gel electrophoresis and mass spectrometry. *Proteomics* 2005, 5, 551–565.
36. Moreno, J.A.; Ortega-Gomez, A.; Rubio-Navarro, A.; Louedec, L.; Ho-Tin-Noé, B.; Caligiuri, G.; Nicoletti, A.; Levoye, A.; Plantier, L.; Meilhac, O. High-density lipoproteins potentiate α 1-antitrypsin therapy in elastase-induced pulmonary emphysema. *Am. J. Respir. Cell Mol. Biol.* 2014, 51, 536–549.
37. Bricarello, D.A.; Mills, E.J.; Petrlova, J.; Voss, J.C.; Parikh, A.N. Ganglioside embedded in reconstituted lipoprotein binds cholera toxin with elevated affinity. *J. Lipid Res.* 2010, 51, 2731–2738.
38. Okamoto, Y.; Wang, F.; Yoshioka, K.; Takuwa, N.; Takuwa, Y. Sphingosine-1-phosphate-specific G protein-coupled receptors as novel therapeutic targets for atherosclerosis. *Pharmaceuticals* 2011, 4, 117–137.
39. Kimura, T.; Tomura, H.; Mogi, C.; Kuwabara, A.; Damirin, A.; Ishizuka, T.; Sekiguchi, A.; Ishiura, M.; Im, D.S.; Sato, K.; et al. Role of scavenger receptor class B type I and sphingosine 1-phosphate receptors in high density lipoprotein-induced inhibition of adhesion molecule expression in endothelial cells. *J. Biol. Chem.* 2006, 281, 37457–37467.
40. Zhang, B.; Tomura, H.; Kuwabara, A.; Kimura, T.; Miura, S.I.; Noda, K.; Okajima, F.; Saku, K. Correlation of high density lipoprotein (HDL)-associated sphingosine 1-phosphate with serum levels of HDL-cholesterol and apolipoproteins. *Atherosclerosis* 2005, 178, 199–205.
41. Tölle, M.; Pawlak, A.; Schuchardt, M.; Kawamura, A.; Tietge, U.J.; Lorkowski, S.; Keul, P.; Assmann, G.; Chun, J.; Levkau, B.; et al. HDL-associated lysosphingolipids inhibit NAD(P)H oxidase-dependent monocyte chemoattractant protein-1 production. *Arterioscler. Thromb. Vasc. Biol.* 2008, 28, 1542–1548.
42. Winkler, M.S.; Nierhaus, A.; Holzmann, M.; Muddersbach, E.; Bauer, A.; Robbe, L.; Zahrt, C.; Geffken, M.; Peine, S.; Schwedhelm, E.; et al. Decreased serum concentrations of sphingosine-1-phosphate in sepsis. *Crit. Care* 2015, 19.
43. Winkler, M.S.; Nierhaus, A.; Poppe, A.; Greiwe, G.; Gräler, M.H.; Daum, G. Sphingosine-1-Phosphate: A Potential Biomarker and Therapeutic Target for Endothelial Dysfunction and Sepsis? *Shock* 2017, 47, 666–672.
44. Hoste, E.A.; Bagshaw, S.M.; Bellomo, R.; Cely, C.M.; Colman, R.; Cruz, D.N.; Edipidis, K.; Forni, L.G.; Gomersall, C.D.; Govil, D.; et al. Epidemiology of acute kidney injury in critically ill patients: The multinational AKI-EPI study. *Intensive Care Med.* 2015, 41, 1411–1423.

45. Fiorentino, M.; Tohme, F.A.; Wang, S.; Murugan, R.; Angus, D.C.; Kellum, J.A. Long-term survival in patients with septic acute kidney injury is strongly influenced by renal recovery. *PLoS ONE* 2018, 13, e0198269.
46. Seymour, C.W.; Liu, V.X.; Iwashyna, T.J.; Brunkhorst, F.M.; Rea, T.D.; Scherag, A.; Rubenfeld, G.; Kahn, J.M.; Shankar-Hari, M.; Singer, M.; et al. Assessment of Clinical Criteria for Sepsis: For the Third International Consensus Definitions for Sepsis and Septic Shock (Sepsis-3). *JAMA* 2016, 315, 762–774.
47. Bagshaw, S.M.; Uchino, S.; Bellomo, R.; Morimatsu, H.; Morgera, S.; Schetz, M.; Tan, I.; Bouman, C.; Macedo, E.; Gibney, N.; et al. Septic acute kidney injury in critically ill patients: Clinical characteristics and outcomes. *Clin. J. Am. Soc. Nephrol.* 2007, 2, 431–439.
48. Peerapornratana, S.; Manrique-Caballero, C.L.; Gómez, H.; Kellum, J.A. Acute kidney injury from sepsis: Current concepts, epidemiology, pathophysiology, prevention and treatment. *Kidney Int.* 2019, 96, 1083–1099.
49. Kashani, K.; Al-Khafaji, A.; Ardiles, T.; Artigas, A.; Bagshaw, S.M.; Bell, M.; Bihorac, A.; Birkhahn, R.; Cely, C.M.; Chawla, L.S.; et al. Discovery and validation of cell cycle arrest biomarkers in human acute kidney injury. *Crit. Care* 2013, 17, R25.
50. Fiorentino, M.; Xu, Z.; Smith, A.; Singbartl, K.; Palevsky, P.M.; Chawla, L.S.; Huang, D.T.; Yealy, D.M.; Angus, D.C.; Kellum, J.A. Serial Measurement of Cell-cycle Arrest Biomarkers [TIMP-2] [IGFBP7] and Risk for Progression to Death, Dialysis or Severe Acute Kidney Injury in Patients with Septic Shock. *Am. J. Respir. Crit. Care Med.* 2020, 202.
51. Chawla, L.S.; Seneff, M.G.; Nelson, D.R.; Williams, M.; Levy, H.; Kimmel, P.L.; Macias, W.L. Elevated plasma concentrations of IL-6 and elevated APACHE II score predict acute kidney injury in patients with severe sepsis. *Clin. J. Am. Soc. Nephrol.* 2007, 2, 22–30.
52. Cirstea, M.; Walley, K.R.; Russell, J.A.; Brunham, L.R.; Genga, K.R.; Boyd, J.H. Decreased high-density lipoprotein cholesterol level is an early prognostic marker for organ dysfunction and death in patients with suspected sepsis. *J. Crit. Care* 2017, 38, 289–294.
53. Genga, K.R.; Trinder, M.; Kong, H.J.J.; Li, X.; Leung, A.K.K.; Shimada, T.; Walley, K.R.; Russell, J.A.; Francis, G.A.; Brunham, L.R.; et al. CETP genetic variant rs1800777 (allele A) is associated with abnormally low HDL-C levels and increased risk of AKI during sepsis. *Sci. Rep.* 2018, 8.
54. Levels, J.H.M.; Abraham, P.R.; Van den Ende, A.; Van Deventer, S.J.H. Distribution and kinetics of lipoprotein-bound endotoxin. *Infect. Immun.* 2001, 69, 2821–2828.
55. Flegel, W.A.; Wolpl, A.; Mannel, D.N.; Northoff, H. Inhibition of endotoxin-induced activation of human monocytes by human lipoproteins. *Infect. Immun.* 1989, 57, 2237–2245.
56. Baumberger, C.; Ulevitch, R.J.; Dayer, J.M. Modulation of endotoxic activity of lipopolysaccharide by high-density lipoprotein. *Pathobiology* 1991, 59, 378–383.
57. Anderberg, S.B.; Luther, T.; Frithiof, R. Physiological aspects of Toll-like receptor 4 activation in sepsis-induced acute kidney injury. *Acta Physiol.* 2017, 219, 573–588.
58. Habib, R. Multifaceted roles of Toll-like receptors in acute kidney injury. *Heliyon* 2021, 7, e06441.
59. Zarbock, A.; Gomez, H.; Kellum, J.A. Sepsis-induced acute kidney injury revisited: Pathophysiology, prevention and future therapies. *Curr. Opin. Crit. Care* 2014, 20, 588–595.
60. Lowry, J.L.; Brovkovich, V.; Zhang, Y.; Skidgel, R.A. Endothelial nitric-oxide synthase activation generates an inducible nitric-oxide synthase-like output of nitric oxide in inflamed endothelium. *J. Biol. Chem.* 2013, 288, 4174–4193.
61. Tran-Dinh, A.; Diallo, D.; Delbosc, S.; Varela-Perez, L.M.; Dang, Q.B.; Lapergue, B.; Burillo, E.; Michel, J.B.; Levoye, A.; Martin-Ventura, J.L.; et al. HDL and endothelial protection. *Br. J. Pharmacol.* 2013, 169, 493–511.
62. Graversen, J.H.; Castro, G.; Kandoussi, A.; Nielsen, H.; Christensen, E.I.; Norden, A.; Moestrup, S.K. A Pivotal Role of the Human Kidney in Catabolism of HDL Protein Components Apolipoprotein A-I and A-IV but not of A-II. *Lipids* 2008, 43, 467–470.
63. Kronenberg, F.; Kuen, E.; Ritz, E.; König, P.; Kraatz, G.; Lhotta, K.; Mann, J.F.E.; Müller, G.A.; Neyer, U.; Riegel, W.; et al. Apolipoprotein A-IV serum concentrations are elevated in patients with mild and moderate renal failure. *J. Am. Soc. Nephrol.* 2002, 13, 461–469.
64. Calabresi, L.; Simonelli, S.; Conca, P.; Busnach, G.; Cabibbe, M.; Gesualdo, L.; Gigante, M.; Penco, S.; Veglia, F.; Franceschini, G. Acquired lecithin: CHolesterol acyltransferase deficiency as a major factor in lowering plasma HDL levels in chronic kidney disease. *J. Intern. Med.* 2015, 277, 552–561.
65. Aseem, O.; Smith, B.T.; Cooley, M.A.; Wilkerson, B.A.; Argraves, K.M.; Remaley, A.T.; Argraves, W.S. Cubilin maintains blood levels of HDL and albumin. *J. Am. Soc. Nephrol.* 2014, 25, 1028–1036.

66. Zhong, J.; Yang, H.; Kon, V. Kidney as modulator and target of “good/bad” HDL. *Pediatric Nephrol.* 2019, 34, 1683–1695.
67. Vaziri, N.D. HDL abnormalities in nephrotic syndrome and chronic kidney disease. *Nat. Rev. Nephrol.* 2016, 12, 37–47.
68. Coughlin, M.M.; Babcook, J.; Prabhakar, B.S. Human monoclonal antibodies to SARS-coronavirus inhibit infection by different mechanisms. *Virology* 2009, 394, 39–46.
69. Chi, X.; Yan, R.; Zhang, J.; Zhang, G.; Zhang, Y.; Hao, M.; Zhang, Z.; Fan, P.; Dong, Y.; Yang, Y.; et al. A neutralizing human antibody binds to the N-terminal domain of the Spike protein of SARS-CoV-2. *Science* 2020, 369, 650–655.
70. Wei, C.; Wan, L.; Yan, Q.; Wang, X.; Zhang, J.; Yang, X.; Zhang, Y.; Fan, C.; Li, D.; Deng, Y.; et al. HDL-scavenger receptor B type 1 facilitates SARS-CoV-2 entry. *Nat. Metab.* 2020, 2, 1391–1400.
71. Strollo, R.; Pozzilli, P. DPP4 inhibition: Preventing SARS-CoV-2 infection and/or progression of COVID-19? *Diabetes Metab. Res. Rev.* 2020, 36, e3330.
72. Zanon, P.; Khetarpal, S.A.; Larach, D.B.; Hancock-Cerutti, W.F.; Millar, J.S.; Cuchel, M.; DerOhannessian, S.; Kontush, A.; Surendran, P.; Saleheen, D.; et al. Rare variant in scavenger receptor BI raises HDL cholesterol and increases risk of coronary heart disease. *Science* 2016, 351, 1166–1171.
73. Shen, W.J.; Azhar, S.; Kraemer, F.B. SR-B1: A Unique Multifunctional Receptor for Cholesterol Influx and Efflux. *Annu. Rev. Physiol.* 2018, 80, 95–116.
74. Kolleck, I.; Sinha, P.; Rüstow, B. Vitamin E as an antioxidant of the lung: Mechanisms of vitamin E delivery to alveolar type II cells. *Am. J. Respir. Crit. Care Med.* 2002, 166, S62–S66.
75. Kolleck, I.; Schlame, M.; Fechner, H.; Looman, A.C.; Wissel, H.; Rüstow, B. HDL is the major source of vitamin E for type II pneumocytes. *Free Radic. Biol. Med.* 1999, 27, 882–890.
76. Kreutz, R.; Algharably, E.A.E.H.; Azizi, M.; Dobrowolski, P.; Guzik, T.; Januszewicz, A.; Persu, A.; Prejbisz, A.; Riemer, T.G.; Wang, J.G.; et al. Hypertension, the renin-angiotensin system, and the risk of lower respiratory tract infections and lung injury: Implications for covid-19. *Cardiovasc. Res.* 2020, 116, 1688–1699.
77. Lajoie, P.; Nabi, I.R. Regulation of raft-dependent endocytosis. *J. Cell. Mol. Med.* 2007, 11, 644–653.
78. Baglivo, M.; Baronio, M.; Natalini, G.; Beccari, T.; Chiurazzi, P.; Fulcheri, E.; Petralia, P.; Michelini, S.; Fiorentini, G.; Miggiano, G.A.; et al. Natural small molecules as inhibitors of coronavirus lipid-dependent attachment to host cells: A possible strategy for reducing SARS-COV-2 infectivity? *Acta Biomed.* 2020, 91, 161–164.
79. Li, G.M.; Li, Y.G.; Yamate, M.; Li, S.M.; Ikuta, K. Lipid rafts play an important role in the early stage of severe acute respiratory syndrome-coronavirus life cycle. *Microbes Infect.* 2007, 9, 96–102.
80. You, C.H.; Hong, Y.S.; Kwak, J.Y.; Kim, N.Y.; Mee, S.R.; Jung, K.Y.; Lee, Y.H.; Kim, J.M.; Kim, J.Y. The relationship between ACE I/D polymorphism and HDL cholesterol. *J. Prev. Med. Public Health* 2006, 39, 505–510.
81. Dou, X.; Li, Y.; Han, J.; Zarlenga, D.S.; Zhu, W.; Ren, X.; Dong, N.; Li, X.; Li, G. Cholesterol of lipid rafts is a key determinant for entry and post-entry control of porcine rotavirus infection. *BMC Vet. Res.* 2018, 14, 45.
82. Korade, Z.; Kenworthy, A.K. Lipid rafts, cholesterol, and the brain. *Neuropharmacology* 2008, 55, 1265–1273.
83. Radenkovic, D.; Chawla, S.; Pirro, M.; Sahebkar, A.; Banach, M. Cholesterol in Relation to COVID-19: Should We Care about It? *J. Clin. Med.* 2020, 9, 1909.
84. Hu, X.; Chen, D.; Wu, L.; He, G.; Ye, W. Low Serum Cholesterol Level Among Patients with COVID-19 Infection in Wenzhou, China. *SSRN Electron. J.* 2020.
85. Meher, G.; Bhattacharjya, S.; Chakraborty, H. Membrane Cholesterol Modulates Oligomeric Status and Peptide-Membrane Interaction of Severe Acute Respiratory Syndrome Coronavirus Fusion Peptide. *J. Phys. Chem. B* 2019, 123, 10654–10662.
86. Garrido, P.F.; Calvelo, M.; Blanco-González, A.; Veleiro, U.; Suárez, F.; Conde, D.; Cabezón, A.; Piñeiro, Á.; García-Fandino, R. The Lord of the NanoRings: Cyclodextrins and the battle against SARS-CoV-2. *Int. J. Pharm.* 2020, 588, 119689.
87. Henrich, S.E.; McMahon, K.M.; Palacio, N.; Bhalla, P.; MacMaster, P.P.; Thaxton, C.S. Targeting scavenger receptor type B-1 (SR-B1) and cholesterol inhibits entry of SARSCoV-2 pseudovirus in cell culture. *BioRxiv* 2020.
88. Glende, J.; Schwegmann-Wessels, C.; Al-Falah, M.; Pfefferle, S.; Qu, X.; Deng, H.; Drosten, C.; Naim, H.Y.; Herrler, G. Importance of cholesterol-rich membrane microdomains in the interaction of the S protein of SARS-coronavirus with the cellular receptor angiotensin-converting enzyme 2. *Virology* 2008, 381, 215–221.
89. Cervin, M.; Anderson, R. Modulation of coronavirus-mediated cell fusion by homeostatic control of cholesterol and fatty acid metabolism. *J. Med. Virol.* 1991, 35, 142–149.

