

# Lignans from *Bursera fagaroides*

Subjects: Plant Sciences

Contributor: Mayra Antunez-Mojica

*Bursera fagaroides* is a medicinal tree endemic to México, it belongs to the Burseraceae family and has proven antitumor activity. Modern research, performed principally with the bark extracts, have indicated that lignans are the main active constituents of *B. fagaroides*, with a high content of aryltetralin, aryldihydronaphtalene, dibenzylbutirolactone, and dibenzylbutane type lignans as the constituents of the active extracts. In general, lignans from *B. fagaroides* exhibited potent anti-cancer activity, although antitumor, anti-bacterial, anti-protozoal, anti inflammatory, and anti-viral properties have also been described.

Keywords: lignans ; tubulin ; cytotoxicity

## 1. Introduction

Cancer is well known as one of the most important causes of morbidity and mortality worldwide, its effects in both more and less economically developed countries, and its likelihood to rank as the leading cause of death in the 21st century. It is estimated that this public health problem caused 9.6 million deaths worldwide in 2018 <sup>[1][2]</sup>. Currently, there are various types of therapies for cancer treatment, but chemotherapy is one of the most widely used. However, because of the severe side effects exhibited by commercially available drugs to treat cancer, as well as the drug resistance in tumor cells, it remains a challenge for medicinal chemistry to develop novel agents and treatment strategies to attack this public health problem <sup>[3][4]</sup>. According to the Food and Drug Administration (FDA), 246 anti-cancer drugs were approved between 1940 and 2014 and around 38% of them are natural products (or derived from them) <sup>[5]</sup>. In this context, one of the most important sources of anti-cancer secondary metabolites is the *Bursera* genus because it has been reported that these plants are effective against different types of cancer <sup>[6]</sup>. The genus *Bursera* Jacq. ex L. (Burseraceae) consists of about 105 shrubs and trees with a geographical distribution extending from the Southern U.S. to Peru and the Caribbean. In Mexico, the *Bursera* species grow principally in the tropical dry forests, where about 92 species have been described and most of them (~85%) are endemic <sup>[7][8]</sup>. *Bursera* has been divided into two subgenera (subg.), or sections: *B. subg. Bursera* and *B. subg. Elaphrium* (previously known as *Bullockia*) <sup>[8]</sup>; the bark, among other traits, mainly differentiates between them. The species of section *Bursera* have colorful trunks and peeling bark, while species of *Elaphrium* have rough and non-peeling bark <sup>[9]</sup>. Several *Bursera* species are recognized because of their characteristic production of an aromatic resin (exuded) known as “copal” that provides a chemical defense against specialized herbivores <sup>[10]</sup>. Since ancient times, copal resin has been commonly used in México and Central America as incense in religious activities <sup>[11][12]</sup>. The chemical profile of the species of *Bursera* includes flavonoids <sup>[13][14]</sup>, triterpenes <sup>[15][16]</sup>, sesquiterpenes <sup>[16][17]</sup>, diterpenes <sup>[18]</sup>, and lignans <sup>[19][20]</sup>. Most of the *Bursera* species that produce lignans are widely used in México as traditional, natural medicine due to their pharmacological properties, including analgesic, anti-inflammatory, and antitumoral properties. Also, they can help treat different illnesses, such as colds, polyps, and venereal diseases <sup>[6][21]</sup>. In general, lignans from the *Bursera* genus are secondary metabolites, known for their antioxidant, apoptotic, anti-cancer, anti-inflammatory, anti-bacterial, anti-viral, anti-fungal, and anti-protozoal properties. In particular, lignans from *B. fagaroides* have been reported to have an important anti-cancer effect <sup>[6]</sup>. This review aims to summarize literature findings on the Mexican *B. fagaroides*, such as uses in medicinal folk, pharmacological effects of its extracts and chemistry, and the biological activities of its lignans. This review focuses on the biosynthesis, chemical aspects, anti-cancer effects, and molecular mechanisms of lignans from *B. fagaroides*. The information reported in this work results from a search in ScienceDirect, PubMed, and Scifinder databases.

## 2. *Bursera fagaroides*: Description, Distribution, and Uses in the Mexican Traditional Medicine

*Bursera fagaroides* (*B. subg. Bursera*) (**Figure 1**), also identified as *Elaphrium fagaroides*, *Amyris fagaroides*, and *Terebinthus fagaroides*, is a Mexican medicinal plant locally known as “copalillo”, “aceitillo”, “copal”, “sarzafrás”, “xixote”, “cuajote amarillo” “jiote”, “palo del diablo”, “papelillo”, and “xixote” <sup>[17][22][23][24]</sup>. It is an aromatic bush or tree of about 0.5–

8 m high, distributed from the Southwestern United States of America to the Isthmus of Tehuantepec in México; it grows mainly at altitudes from 300 to 2200 m [8][17][23]. *B. fagaroides*, as traditional natural medicine, have been popularly used to treat inflammation, hits, tumors, cancer, and stomach disorders [20][22][25]. These medicinal properties have served as inspiration for various cancer research groups, as described below.



**Figure 1.** *Bursera fagaroides*, specimens from Morelos State (México). From right to left: leaves, bark, and fruits.

### 3. Anti-Cancer Studies of Extracts of *B. Fagaroides*

This plant species has been studied principally for its anti-cancer properties, although its antimicrobial and antiangiogenic effects also have been reported. An overview of the anti-cancer biological studies performed on this plant species shows that the only parts examined have been the bark and the exudate resin from the tree trunk. In vivo and in vitro studies on the extracts of these plant parts have shown important cytotoxic activities.

For instance, in 1969, Bianchi and Cole [26] found that the chloroform extract displayed a 32% reduction in the *in vivo* Walker carcinoma 256 tumor system (WA16). Further, the ethanol extract from the dried exudates of *B. fagaroides* showed a concentration-dependent inhibitory effect on cell proliferation against the human colon cell line HT-29, with an IC<sub>50</sub> value of  $0.41 \pm 0.01$  µg/mL at 72 h [27].

Another in vivo study by Rojas-Sepulveda [19] reported that the intraperitoneal administration of 100 mg/Kg of the hydroalcoholic extract from the bark on mice, inoculated with L5178Y lymphoma cells, increased the survival time and cured 26% ( $p < 0.001$ ) of the treated mice. This extract also significantly inhibited the proliferation of KB (nasopharyngeal, ED<sub>50</sub> =  $9.6 \times 10^{-2}$  µg/mL), PC-3 (prostate, ED<sub>50</sub> =  $2.5 \times 10^{-1}$  µg/mL), HF-6 (colon, ED<sub>50</sub> =  $7.1 \times 10^{-3}$  µg/mL), and MCF-7 (mama, ED<sub>50</sub> = 6.6 µg/mL) tumor cell lines [19]. Later, Acevedo et al. (2015) [28] described the cytotoxicity of the n-hexane and chloroform extracts measured by the sulforhodamine B protein staining assay using KB, HF-6, MCF-7, and PC-3 cancer cell lines, along with a normal skin fibroblast cell line. The results indicated that both extracts displayed an important antiproliferative effect on all the studied cells, including normal cells, corroborating the results obtained previously [28].

In another *in vivo* study, the hydroalcoholic extract from the bark of *B. fagaroides* does not affect the number of Histone H3 phosphorylated at serine 10 (H3S10ph)-positive nuclei, with respect to the control without treatment, when measured in whole 24 h post-fertilization (hpf) zebrafish embryos; this indicated that the extract does not induce mitotic cells in the embryos [29]. This result contrasts with the strong in vivo antitumor activity against L5178Y lymphoma in mice and the authors attributed this to the poor bioavailability because of the low concentration of the active compounds present in the studied extract [19]. Further, chromatographic fractionation afforded two rich-lignans fractions that induced a high amount of cells in mitotic arrest in zebrafish embryos [29].

### 4. Lignans from *B. Fagaroides*

The chemical study of *B. fagaroides* through the years allowed the characterization of 19 lignan structures named: podophyllotoxin (1), β-peltatin-A-methylether (2), 5'-desmethoxy-β-peltatin-A-methylether (3), desoxypodophyllotoxin (4), acetyl podophyllotoxin (5) [1], morelensin (6) [1], burseranin (7) [1], acetylpicropodophyllotoxin (8) [2], desmethoxy-yatein (9), yatein (10) [1], hinokinin (11) [40], 7',8'-dehydropodophyllotoxin (12), 7',8'-dehydroacetyl podophyllotoxin (13), 7',8'-dehydro-*trans*-*p*-cumaroylpodophyllotoxin (14) [3], 9-acetyl-9-pentadecanoildihydroclusin (15), 2,3-demethoxy-secoisolintetralin diacetate (16), dihydroclusin diacetate (17), 2,3 demethoxy-secoisolintetralin monoacetate (18) dihydroclusin mono acetate (19) [4]. Eight of these are aryltetralin (1–8), three are dibenzylbutyrolactone (9–11), three are arylidihydronaphthalene (12–14), and five are dibenzylbutane lignans (15–19).

The lignans **1–19** have been isolated from specimens of *B. fagaroides*, collected in México, principally from the bark (Michoacán state), two reports from Oaxaca state [14] and one from Guerrero state [15] analyzed the resin. The polarity of the used extracts was chloroform (CHCl<sub>3</sub>), dichloromethane (CH<sub>2</sub>Cl<sub>2</sub>), ethanol (CH<sub>3</sub>CH<sub>2</sub>OH) [27], CH<sub>3</sub>CH<sub>2</sub>OH 80% (previously treated with hexane) [15], and methanol (CH<sub>3</sub>OH) 70% [19,29,39]. In general, the purification of these extracts was carried out by bioassay-guided chromatographic methods. The extracts were fractionated and the components were separated by repeated column chromatography, eluting with gradients or isocratic mixtures [14, 15] of organic solvents through preparative thin layer chromatography (TLC), semi-preparative reverse phase HPLC with a diode array detection system, flash chromatography, or preparative reverse phase TLC, as required. The yields and purity of isolated compounds were based on the peak areas of the HPLC chromatograms [2].

In addition, their <sup>13</sup>C NMR (CDCl<sub>3</sub>) spectra show the occurrence of carbon resonances, ascribable to carboxyl ester groups ( $\delta$  C ~ 167.7–173.7); signals at  $\delta$  ~ 171.1 and  $\delta$  ~ 21.0 also confirm the presence of acetyl groups; carbons at  $\delta$  C7' ~ 147.2–147.9 and  $\delta$  C8' ~ 118.7 in lignans **12**, **13**, and **14** were characteristics of a tetrasubstituted double bond. Other signals also were assigned: methylenedioxy group in ring A [ $\delta$  C ~ 102.0–103.1], methoxyl groups [ $\delta$  C ~ 58.6 - 61.2], aliphatic methylene group [ $\delta$  C ~ 69.6–70.1], aliphatic methine groups (C7)  $\delta$  ~ 74.2–75.2 for lignans **1–8** and **12–14**, aliphatic methine groups [ $\delta$  C8 ~ 41.7–43.9], characteristic signals of carbon atoms bearing oxygen at  $\delta$  ~ 63.95 - 64.3, aromatic carbons for ring B [ $\delta$  C1 ~ 128.2–130.6,  $\delta$  C2 ~ 129.1–132.0,  $\delta$  C3 ~ 110.0–110.2,  $\delta$  C4 ~ 150–153,  $\delta$  C5 ~ 147.5–148.4, and  $\delta$  C6 ~ 104.8–105.8] and those corresponding to substituted aromatic carbons for ring E [ $\delta$  C1' ~ 129.1–131.2,  $\delta$  C2' ~ 109.6–110.0,  $\delta$  C3' ~ 153.0–153.7,  $\delta$  C4' ~ 135.7–139.5,  $\delta$  C5' ~ 153.0–154.0, and  $\delta$  C6' ~ 109.6–110.3], respectively [19][20][25][26][27][29][30]. On the other hand, the absolute configurations of lignans **4**, **5**, **7**, **11**, and **12** were determined using vibrational circular dichroism [27], while that of lignan **1** was determined by chemical correlation with *D*-phenylalanine [31] and by X-ray diffraction analysis of 2'-bromopodophyllotoxin [32].

The molecular studies of lignans isolated from *B. fagaroides* are diverse and all head to anti-cancer activity. Cancer is a worldwide health problem with 9.6 million cancer deaths in 2018 [2]; due to this, lignans are eye-catching secondary metabolites from medicinal plant research implicated in cancer. **Figure 5** summarizes all the assays performed for aryl tetralin and arylidihydronaphtalene lignans isolated from *B. fagaroides*, from the cytotoxic *in vitro* results, *in vivo* assays in mice and zebrafish models, and to the molecular recognition by NMR. Lignans **8**, **10**, **11**, **15–19** do not report any activity.

*In vivo* studies with Walker carcinoma 256 (intramuscular) were performed for **2** and **3**, they exhibited an important antitumoral activity at a level of 10% T/C at 12.5 mg/kg and 20% T/C at 100 mg/kg, respectively [26]. Also, the *in vivo* model using zebrafish embryos was performed to observe disruption of cell behavior; the results showed a delay cell migration in actin filaments for **1**, **2**, **5**, and **9**. Also the same compounds presented a microtubule depolymerization in the same model by  $\alpha$ -tubulin immunofluorescence [29].

The cytotoxicity (IC<sub>50</sub>) of 14 lignans were reported in 9 cancer cell lines: KB, PC-3, MCF-7, MDA-MB-231, BT-549, HF-6, A549, A2780, and SiHa. According to the results, PC-3 and KB cell lines were the most sensitive with IC<sub>50</sub> values between 2.29 and 4.43  $\times 10^{-6}$   $\mu$ M, respectively [19][20][33][30].  $\beta$ -peltatin-A-methylether (**2**) and podophyllotoxin (**1**) were the most active compounds in KB cells with an IC<sub>50</sub> of 4.43  $\times 10^{-6}$  and 4.61  $\times 10^{-6}$   $\mu$ M, respectively [19]. It should be highlighted that the importance of these results is due to prostate cancer occupying the first place in mortality cancer in males [34].

In general, lignans obtained from *B. fagaroides* are important secondary metabolites with promising pharmacological anti-cancer effects and it could be interesting to explore them as antivirals. These compounds can act by the same mechanism of action of podophyllotoxin and can be considered in clinical trials for cancer.

## References

1. Mbaveng, A.T.; Manekeng, H.T.; Nguenang, G.S.; Dzotam, J.K.; Kuete, V.; Effertha, E. Cytotoxicity of 18 Cameroonian medicinal plants against drug sensitive and multi-factorial drug resistant cancer cells. *J. Ethnopharmacol.* 2018, 222, 21–33.
2. Bray, F.; Ferlay, J.; Soerjomataram, I.; Siegel, R.L.; Torre, L.A.; Jemal, A. Global Cancer Statistics 2018: GLOBOCAN Estimates of Incidence and Mortality Worldwide for 36 Cancers in 185 Countries. *CA Cancer J. Clin.* 2018, 68, 394–424.
3. Roodhart, J.M.L.; Daenen, L.G.M.; Stigter, E.C.A.; Prins, H.J.; Gerrits, J.; Houthuijzen, J.M.; Gerritsen, M.G.; Schipper, H.S.; Backer, M.J.G.; van Amersfoort, M.; et al. Mesenchymal Stem Cells Induce Resistance to Chemotherapy through the Release of Platinum-Induced Fatty Acids. *Cancer Cell* 2011, 20, 370–383.

4. Atmaca, H.; Çamli, Ç.; Sert, S. Ethanol Extract of *Pinus nigra* ssp. *pallasiana* var. *şeneriana* Inhibits Human Breast Cancer Cell Viability through Induction of Apoptosis. *Celal Bayar Univ. J. Sci.* 2018, 14, 35–40.
5. Newman, D.J.; Cragg, G.M. Natural Products as Sources of New Drugs from 1981 to 2014. *J. Nat. Prod.* 2016, 79, 629–661.
6. Marcotullio, M.C.; Curini, M.; Becerra, J.X. An Ethnopharmacological, Phytochemical and Pharmacological Review on Lignans from Mexican *Bursera* spp. *Molecules* 2018, 23, 1976.
7. Becerra, J.X.; Noge, K.; Olivier, S.; Venable, L. The monophyly of *Bursera* and its impact for divergence times of Burseraceae. *Taxon* 2012, 61, 333–343.
8. Rzedowski, J.; Medina, R.; Calderón, G. Inventario del conocimiento taxonómico, así como de la diversidad y del endemismo regionales de las especies mexicanas de *Bursera* (Burseraceae). *Acta Botánica Mexicana* 2005, 70, 85–111.
9. Becerra, J.X. Evolution of Mexican *Bursera* (Burseraceae) inferred from ITS, ETS, and 5S nuclear ribosomal DNA sequences. *Mol. Phylogenetics Evol.* 2003, 26, 300–309.
10. Becerra, J.X.; Venable, D.L.; Evans, P.H.; Bowers, W.S. Interactions between Chemical and Mechanical Defenses in the Plant Genus *Bursera* and Their Implications for Herbivores. *Am. Zool.* 2001, 41, 865–876.
11. Case, R.J.; Tucker, A.O.; Maciarello, M.J.; Wheeler, K.A. Chemistry and Ethnobotany of Commercial Incense Copals, Copal Blanco, Copal Oro, and Copal Negro of North America. *Econ. Bot.* 2003, 57, 189–202.
12. Linares, E.; Bye, R. El copal en México. *Biodiversitas* 2008, 78, 8–11.
13. Nakanishi, T.; Inatomi, Y.; Satomi, A.; Yamada, T.; Fukatsu, H.; Murata, H.; Inada, A.; Matsuura, N.; Ubukata, M.; Murata, J.; et al. New luteolin 3-O-acylated rhamnosides from leaves of *Bursera graveolens*. *Heterocycles* 2003, 60, 2077–2083.
14. Souza, M.P.; Machado, M.I.L.; Braz-Filho, R. Six flavonoids from *Bursera leptophloeos*. *Phytochemistry* 1989, 28, 2467–2470.
15. Romero-Estrada, A.; Maldonado-Magaña, A.; González-Christen, J.; Marquina, S.; Garduño-Ramírez, M.L.; Rodríguez-López, V.; Alvarez, L. Anti-inflammatory and antioxidative effects of six pentacyclic triterpenes isolated from the Mexican copal resin of *Bursera copallifera*. *BMC Complement. Altern. Med.* 2016, 16, 422–431.
16. Columba-Palomares, M.C.; Villarreal, M.L.; Marquina, S.; Romero-Estrada, A.; Rodríguez-López, V.; Zamilpa, A.; Alvarez, L. Antiproliferative and Anti-inflammatory Acyl Glucosyl Flavones from the Leaves of *Bursera copallifera*. *J. Mex. Chem. Soc.* 2018, 62, 214–224.
17. Noge, K.; Becerra, J.X. Germacrene D, A Common Sesquiterpene in the Genus *Bursera* (Burseraceae). *Molecules* 2009, 14, 5289–5297.
18. García-Gutiérrez, H.A.; Cerda-García-Rojas, C.M.; Hernández-Hernández, J.D.; Román-Marín, L.U.; Joseph-Nathan, P. Oxygenated verticillene derivatives from *Bursera suntui*. *Phytochemistry* 2008, 69, 2844–2848.
19. Rojas-Sepúlveda, A.M.; Mendieta-Serrano, M.; Mojica, M.Y.; Salas-Vidal, E.; Marquina, S.; Villarreal, M.L.; Puebla, A.M.; Delgado, J.I.; Alvarez, L. Cytotoxic Podophyllotoxin Type-Lignans from the Steam Bark of *Bursera fagaroides* var. *fagaroides*. *Molecules* 2012, 17, 9506–9519.
20. Antunez-Mojica, M.; León, A.; Rojas-Sepúlveda, A.M.; Marquina, S.; Mendieta-Serrano, M.; Salas-Vidal, E.; Villarreal, M.L.; Alvarez, L. Aryldihydronaphthalene-type lignans from *Bursera fagaroides* var. *fagaroides* and their antimitotic mechanism of action. *RSC Adv.* 2016, 6, 4950–4959.
21. Purata, S.E. (Ed.) *Uso y Manejo de los Copales Aromáticos: Resinas y Aceites*; CONABIO/RAISES: Mexico, 2008; pp. 1–60. Available online: <https://bioteca.biodiversidad.gob.mx/janium-bin/detalle.pl?Id=20210712091900> (accessed on 9 June 2021).
22. Puebla-Pérez, A.M.; Huacuja-Ruiz, L.; Rodríguez-Orozco, G.; Villaseñor-García, M.M.; Miranda-Beltrán, M.; Celis, A.; Sandoval-Ramírez, L. Cytotoxic and Antitumour Activity from *Bursera fagaroides* Ethanol Extract in Mice with L5178Y Lymphoma. *Phytother. Res.* 1998, 12, 545–548.
23. Rzedowski, J. Las especies de *Bursera* (Burseraceae) en la cuenca superior del río Papaloapan (México). *Acta Botánica Mexicana* 2004, 66, 23–151.
24. Castañeda-Miranda, A.G.; Chaparro, M.A.E.; Pacheco-Castro, A.; Chaparro, M.A.E.; Böhnelt, H.N. Magnetic biomonitoring of atmospheric dust using tree leaves of *Ficus benjamina* in Querétaro (México). *Environ. Monit. Assess.* 2020, 192, 291–382.
25. Morales-Serna, J.A.; Cruz-Galicia, E.; García-Ríos, E.; Madrigal, D.; Gaviño, R.; Cárdenas, J.; Salmón, M. Three new diarylbutane lignans from the resin of *Bursera fagaroides*. *Nat. Prod. Res.* 2013, 27, 824–829.

26. Bianchi, E.; Sheth, K.; Cole, J.R. Antitumor agents from *Bursera fagaroides* (Burseraceae) ( $\beta$ -peltatin-A-methylether and 5'-desmethoxy- $\beta$ -peltatin-A-methylether). *Tetrahedron Lett.* 1969, 32, 2759–2762.
27. Velázquez-Jiménez, R.; Torres-Valencia, J.M.; Cerda-García-Rojas, C.M.; Hernández-Hernández, J.D.; Román-Marín, L.U.; Manríquez-Torres, J.J.; Gómez-Hurtado, M.A.; Valdez-Calderón, A.; Motilva, V.; García-Mauriño, S.; et al. Absolute configuration of podophyllotoxin related lignans from *Bursera fagaroides* using vibrational circular dichroism. *Phytochemistry* 2011, 72, 2237–2243.
28. Acevedo, M.; Nuñez, P.; González-Maya, L.; Cardoso-Taketa, A.; Villarreal, M.L. Cytotoxic and Anti-inflammatory Activities of *Bursera* species from Mexico. *J. Clin. Toxicol.* 2015, 5, 2–8.
29. Antúñez-Mojica, M.; Rojas-Sepúlveda, A.M.; Mendieta-Serrano, M.A.; Leticia Gonzalez-Maya, L.; Marquina, S.; Salas-Vidal, E.; Alvarez, L. Lignans from *Bursera fagaroides* Affect in vivo Cell Behavior by Disturbing the Tubulin Cytoskeleton in Zebrafish Embryos. *Molecules* 2019, 24, 8.
30. Antunez-Mojica, M.; Rodríguez-Salarichs, J.; Redondo-Horcajo, M.; León, A.; Barasoain, I.; Canales, A.; Cañada, F.J.; Jiménez-Barbero, J.; Alvarez, L.; Díaz, J.F. Structural and Biochemical Characterization of the Interaction of Tubulin with Potent Natural Analogues of Podophyllotoxin. *J. Nat. Prod.* 2016, 79, 2113–2121.
31. Schrecker, A.W.; Hartwell, J.L. Components of podophyllin XX. The absolute configuration of podophyllotoxin and related lignans. *J. Org. Chem.* 1956, 21, 381–382.
32. Petcher, T.J.; Weber, H.P.; Kuhn, M.; Von Wartburg, A. Crystal structure and absolute configuration of 2'-bromopodophyllotoxin—0.5 ethyl acetate. *J. Chem. Soc. Perkin Trans.* 1973, 2, 288–292.
33. Peña-Morán, O.; Villareal, M.L.; Álvarez, L.; Meneses-Acosta, A.; Rodríguez-López, V. Cytotoxicity, Post-Treatment Recovery, and Selectivity Analysis of Naturally Occurring Podophyllotoxins from *Bursera fagaroides* var. *fagaroides* on Breast Cancer Cell Lines. *Molecules* 2016, 21, 1013.
34. Cancer Facts & Figures 2020. 2020. Available online: <https://www.cancer.org/research/cancer-facts-statistics/all-cancer-facts-figures/cancer-facts-figures-2020.html> (accessed on 26 April 2021).

---

Retrieved from <https://encyclopedia.pub/entry/history/show/31389>