Immunosuppression in Colorectal Cancer

Subjects: Oncology Contributor: Yang Zhang

More emerging studies are exploring immunotherapy for solid cancers, including colorectal cancer. Besides, checkpoint blockade immunotherapy and chimeric antigen receptor (CAR) -based immune cell therapy have being examined in clinical trials for colorectal cancer patients. However, immunosuppression that leads to the blockage of normal immunosurveillance often leads to cancer development and relapse.

Keywords: Immunosuppression in Colorectal Cancer

1. Introduction

Colorectal cancer (CRC) is the second leading cause of cancer death worldwide, accounting for approximately 880,000 deaths in $2018^{[\underline{1}]}$. Approximately 70–80% of patients with stage I–III CRC can be treated with curative surgical resection^[2], and the 5-year survival rate for early-stage CRC is approximately 90%. However, the survival rate drops to only 10–15% for metastatic (stage IV) CRC^[3]. Patients with stage IV CRC are mainly treated with fluorouracil-based chemotherapies, in combination with epidermal growth factor receptor (EGFR) or vascular endothelial growth factor (VEGF) inhibitors, and the median life expectancy is in the range of 21.3 to 30.1 months ^{[4][5][6][2]}.

CRC is the most common gastrointestinal (GI) cancer. Compared with other tissues, the gastrointestinal tract is home to an enormous number of commensal bacteria and potential pathogens^[8]. Therefore, the immune system in the colon and rectum has to fine-tune the balance of the maintenance of tolerance to self-tissue and immunity against potential pathogens and tumorigenic cells. Disruption of the GI tract's normal immune homeostasis may cause autoimmunity diseases, including inflammatory bowel disease^[9]. In contrast, normal immunosurveillance is critical to eliminate potential pathogens and eradicate sporadic tumorigenic cells to prevent GI diseases, including CRC.

It has been well recognized that the immune system plays a key role in colorectal cancer development and progression^[10]. The presence of CD8⁺ T cells in the tumor bed and invasive margin is strongly associated with diseasefree survival and overall survival in CRC and the "immune score", quantifying the density and location of immune cells within the tumor, has provided prognostic value that may be superior to the American Joint Committee on Cancer/Union for International Cancer Control (AJCC/UICC) tumor (T), nodes (N), and metastases (M) (TNM) classification in CRC^[11] ^[12]. This indicates that adaptive immune response may play a role in CRC progression^[13] and immunotherapy may be promising in CRC treatment. However, the efficacy of immune checkpoint inhibitor monotherapy targeting programmed cell death-1 (PD-1) is only seen in a small subset of patients with microsatellite instability high (MSI-H) status. In contrast, no objective response was observed in microsatellite stable (MSS) CRCs, which account for the majority of the cases^[14]. The mechanisms for resistance to anti-PD-1 immune checkpoint blockade in MSS CRC include lack of tumor neoantigens, immune exclusion, and immunosuppression. Higher tumor mutation burden (10- to 50-fold) and more neoantigens in MSI-H tumors, together with higher levels of multiple checkpoints, including PD-1, cytotoxic T lymphocyteassociated antigen 4 (CTLA-4), and lymphocyte activation gene 3 (LAG3) in MSI-H tumors, may explain in part why anti-PD-1 treatment works more efficiently in MSI-H CRCs than in MSS CRCs^{[15][16]}. With the recent advancement in combination strategies and immunomodulatory monoclonal antibodies (mAbs) targeting other immune checkpoints, there is great enthusiasm in developing novel immunotherapies in CRC.

In this review, we will focus on the systemic review of the mechanisms of immunosuppression and the clinical approaches to circumvent immunosuppression in CRC.

2. Current Immunotherapy Trials for MSS CRC

To reverse the immunosuppressive TME and enhance antitumor immunity, several strategies have been attempted (<u>Table 1</u>), including (1) suppression of the cells or signaling pathways that promote immunosuppression in TME; (2) engagement of immunostimulatory cells through external stimulations or treatments; and (3) replacement of the exhausted immune

cells with engineered re-activated immune cells. Of course, the combination strategy may achieve the most promising result to deter immunosuppression in the TME.

Strategies Drugs/Therapy **Drug Target/Action** Note Pembrolizumab PD-1 FDA-approved PD-1 Durvalumab FDA-approved Not tested alone in PDR001 PD-1 CRC FDA-approved for Atezolizumab PD-L1 other cancers Not tested alone in Tremelimumab CTLA-4 CRC Improve survival of Blocking cells or signaling pathways Bevacizumab VEGF **CRC** patients that mediate immunosuppression Galunisertib Not effective alone TGFβ Not tested alone in **BBI608** STAT3 CRC Not tested alone in Oleclumab **CD73** CRC Not tested alone in NZV930 CD73 CRC Not tested alone in NIR178 Adenosine A2A receptor CRC Standard care for Increase antigen presentation FOLFOX Engagement of immunostimulatory and activate PD-1+ CD8+ T advanced CRC cells through external stimulations or treatments Recruit T cells to TME/tumor CEA-TCB (RG7802, RO6958688) cells Sentinel lymph node May improve OS in Replace exhausted cytotoxic T (SLN)-T lymphocytes metastatic CRC [17] cells ACT Limited efficacy in Replacement of the exhausted Replace exhausted T cells (CAR-) T cell therapy solid cancers^[18] immune cells with engineered reactivated immune cells NKG2D CAR-T cell Promising outcomes in Replace exhausted T cells preclinical study^[19] therapy (CAR)-NK cell therapy NCT02839954 **Replace exhausted NK cells**

Table 1. Current strategies for immunotherapy for CRC.

Strategies	Drugs/Therapy	Drug Target/Action	Note
	Durvalumab + tremelimumab	Target PD-1 and CTLA-4	Moderate improvement in OS for MSS CRC patients
	Bevacizumab + fluorouracil	Target VEGF and increase antigen presentation and activate PD-1 ⁺ CD8 ⁺ T	First- and second-line treatment of metastatic CRC
Combination strategy	Atezolizumab + fluorouracil + bevacizumab	Target PD-L1 and VEGF, and increase antigen presentation and activate PD-1 ⁺ CD8 ⁺ T	No further improvement in efficacy in MSS CRC [20]
	Pembrolizumab + FOLFOX	Target PD-L1 and increase antigen presentation and activate PD-1 ⁺ CD8 ⁺ T	NCT03374254
	Atezolizumab + TCB- CEA	Target PD-L1 and recruit T cells to TME/tumor cells	Increased antitumor activity ^[21]
	Pembrolizumab + BBI608	Target PD-L1 and STAT3	NCT02851004
	Durvalumab + oleclumab	Target PD-1 and CD73	NCT02503774
	PDR001 + NIR178	Target PD-1 and adenosine A2A receptor	NCT03207867
	PDR001+ NZV930 + NIR178	Target PD-1, CD73, and adenosine A2A receptor	NCT03549000
	NKG2D CAR-T cell therapy + FOLFOX	Increase antigen presentation and activate PD-1 ⁺ CD8 ⁺ T, and replace exhausted T cells	NCT03310008

Abbreviations: Folinic acid (leucovorin), Fluorouracil (5-FU), and Oxaliplatin (Eloxatin), FOLFOX; Carcinoembryonic Antigen T-Cell Bispecific antibody, CEA-TCB; adoptive cell transfer, ACT; overall survival, OS.

While anti-PD-1/PD-L1 therapies have demonstrated great efficacy in CRC patients with microsatellite instability high/deficient mismatch repair (MSI-H/dMMR), monotherapy is not effective in patients with advanced MSS CRC ^[14]. The combination strategy of anti-PD-1 or anti-PD-L1 with anti-CTLA-4 has been investigated to enhance antitumor immune response. Durvalumab (anti-PD-L1) plus tremelimumab (anti-CTLA-4) showed moderate improvement in overall survival (OS) in patients with MSS CRC who were previously treated with chemotherapies without a significant *p* value (hazard ratio = 0.7, *p* = 0.07). There is no difference in progression-free survival (PFS) in the immunotherapy group compared to the best supportive care group^[22].

VEGF-targeting therapies, such as bevacizumab, have been shown to attenuate the tumor-induced immunosuppressive microenvironment by decreasing the number of Tregs in both pre-clinical mouse models and patients with CRC^{[23][24]}. However, adding atezolizumab (anti-PD-L1) to fluorouracil and bevacizumab as a first-line maintenance treatment for patients with metastatic MSS CRC did not result in improvement in efficacy in MSS CRC^[20].

The alternative strategy is to block immunosuppressive cytokines that mediate the recruitment of Tregs, MDSCs, and TAMs. For example, TGF- β contributes to the immune exclusion in the TME at later stages of tumor development. Although TGF- β blockade as monotherapy is disappointing^[25], the combination treatment of a small-molecule inhibitor (galunisertib) against TGF- β with anti-PD-1 or anti-PD-L1 agents has being tested in clinical trials of solid tumors (NCT02423343 and NCT02734160) and will likely yield promising outcomes. In addition, clinical trials with anti-PD1/PD-L1 in combination with anti-CD73, anti-adenosine A2A receptor, or triplet therapy are being examined in MSS CRC (NCT02503774, NCT03207867, and NCT03549000). The efficacy and safety of a STAT3 inhibitor (BBI608) in combination with pembrolizumab are also being assessed in a phase Ib/II study for patients with metastatic MSS CRC (NCT02851004).

Chemotherapy, such as FOLFOX, has been shown to induce immunogenic cell death, increase antigen presentation and activate PD-1⁺ CD8⁺ T cells^{[26][27]}. Therefore, immune checkpoint inhibitors (ICIs) in combination with standard-of-care chemotherapies may potentiate ICI efficacy by promoting a more immunogenic TME. In a phase II study, clinical activity was seen in the combination treatment of embrolizumab (anti-PD-1) with FOLFOX for patients with untreated advanced CRC, including those with proficient MMR (70%), even though the FOLFOX dose was reduced due to increased

neutropenia in the initial cohort^[28]. Besides, preliminary efficacy data showed a 70% objective response rate (ORR) for the combination treatment of pembrolizumab with FOLFOX in metastatic CRC patients with MMR-proficient disease in the first-line setting of an ongoing clinical trial (keynote-651 cohort B, NCT03374254)^[29].

To target the immune-excluded TME in MSS CRC, a new strategy is to recruit T cells into the immunosuppressive TME by using T cell bispecific (TCB) antibodies. Bispecific antibody is designed to simultaneously bind two different epitopes or antigens, physically linking two binding specificities that may be temporally or spatially separate^[30]. CEA-TCB (RG7802, RO6958688), a novel bispecific antibody that simultaneously binds to carcinoembryonic antigen (CEA) on tumor cells and CD3 on T cells, selectively engages effector T cells to kill CEA-expressing tumor cells. It was investigated in combination with atezolizumab in a phase 1 trial in patients with MSS CRC. Antitumor activity was observed during dose escalation with CEA-TCB monotherapy, with increased intratumoral CD3 T cell infiltration. Enhanced activity and a manageable safety profile were seen in combination with atezolizumab^[21].

Besides the immune checkpoint inhibitors, adoptive cell transfer (ACT) has been exploited to enhance the activity of immune effector cells in the TME. Cytotoxic T cells and NK cells are the major effector cells in the anti-tumor response. Compared to other strategies, ACT has several advantages, including the allowance of ex vivo expansion of tumor-specific lymphocytes and the infusion of cells that can be genetically engineered. In a phase I/II study, sentinel lymph node (SLN)-T lymphocytes were expanded ex vivo and then transfused to CRC patients who underwent radical or palliative surgery. The 24-month survival rate of the SLN-T lymphocyte group was significantly higher than that of the control group (55.6% versus 17.5%, p = 0.02). This study showed that SLN-T lymphocyte immunotherapy was feasible and safe and may improve OS in metastatic CRC^[17].

Chimeric antigen receptor (CAR)-T cell therapy uses chimeric T cell receptors encompassing an extracellular antigenbinding domain, transmembrane hinge domain, and an intracellular signaling domain with costimulatory molecules (such as CD28) in tandem with a CD3ζ chain. The CAR thereby reactivates the T cell immunity against tumor cells. However, CAR-T cell therapy has limited efficacy for the majority of solid tumors, due to limited trafficking and persistence of T cells into the tumor and an immunosuppressive TME^[18]. Recently, a novel NKG2D CAR-T cell therapy, which uses a non-viral third-generation NKG2D CAR, yielded a promising outcome when tested for its antitumor activity against human CRC cells in vitro and in vivo^[19]. To test the notion in a clinical trial, a phase I study is evaluating the safety and clinical activity of NKG2D CAR-T cells in combination with neoadjuvant FOLFOX in patients with metastatic CRC whose liver metastases are potentially resectable (NCT03310008). Additionally, NK cell-based cell therapy, which uses chimeric antigen receptorexpressing NK cells from the patient for anticancer therapy, has also made it to clinical trials for solid tumors, including CRC (NCT02839954)^{[31][32]}. Further investigations will provide a more definitive conclusion if CAR-T cell therapies can benefit patients with MSS CRC.

References

- 1. Bray, F.; Ferlay, J.; Soerjomataram, I.; Siegel, R.L.; Torre, L.A.; Jemal, A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J. Clin. 2018, 68, 394–424, doi:10.3322/caac.21492.
- 2. Lombardi, L.; Morelli, F.; Cinieri, S.; Santini, D.; Silvestris, N.; Fazio, N.; Orlando, L.; Tonini, G.; Colucci, G.; Maiello, E. Adjuvant colon cancer chemotherapy: Where we are and where we'll go. Cancer Treat. Rev. 2010, 36, S34–S41.
- 3. Siegel, R.L.; Miller, K.D.; Jemal, A. Cancer statistics, 2020. CA Cancer J. Clin. 2020, 70, 7–30.
- 4. Hochster, H.S.; Hart, L.L.; Ramanathan, R.K.; Childs, B.H.; Hainsworth, J.D.; Cohn, A.L.; Wong, L.; Fehrenbacher, L.; Abubakr, Y.; Saif, M.W.; et al. Safety and efficacy of oxaliplatin and fluoropyrimidine regimens with or without bevacizumab as first-line treatment of metastatic colorectal cancer: Results of the TREE Study. J. Clin. Oncol 2008, 26, 3523–3529, doi:10.1200/JCO.2007.15.4138.
- Saltz, L.B.; Clarke, S.; Díaz-Rubio, E.; Scheithauer, W.; Figer, A.; Wong, R.; Koski, S.; Lichinitser, M.; Yang, T.S.; Rivera, F.; et al. Bevacizumab in combination with oxaliplatin-based chemotherapy as first-line therapy in metastatic colorectal cancer: A randomized phase III study. J. Clin. Oncol. 2008, 26, 2013–2019, doi:10.1200/jco.2007.14.9930.
- Hecht, J.R.; Mitchell, E.; Chidiac, T.; Scroggin, C.; Hagenstad, C.; Spigel, D.; Marshall, J.; Cohn, A.; McCollum, D.; Stella, P.; et al. A Randomized Phase IIIB Trial of Chemotherapy, Bevacizumab, and Panitumumab Compared with Chemotherapy and Bevacizumab Alone for Metastatic Colorectal Cancer. J. Clin. Oncol. 2009, 27, 672–680, doi:10.1200/jco.2008.19.8135.
- 7. Yamazaki, K.; Nagase, M.; Tamagawa, H.; Ueda, S.; Tamura, T.; Murata, K.; Eguchi Nakajima, T.; Baba, E.; Tsuda, M.; Moriwaki, T.; et al. Randomized phase III study of bevacizumab plus FOLFIRI and bevacizumab plus mFOLFOX6 as first-line treatment for patients with metastatic colorectal cancer (WJOG4407G). Ann. Oncol. 2016, 27, 1539–1546.

- 8. Ley, R.E.; Lozupone, C.A.; Hamady, M.; Knight, R.; Gordon, J.I. Worlds within worlds: Evolution of the vertebrate gut microbiota. Nat. Rev. Microbiol. 2008, 6, 776–788.
- 9. Caruso, R.; Lo, B.C.; Núñez, G. Host–microbiota interactions in inflammatory bowel disease. Nat. Rev. Immunol. 2020, 20, 411–426.
- 10. Garcia-Lora, A.; Algarra, I.; Garrido, F. MHC class I antigens, immune surveillance, and tumor immune escape. J. Cell. Physiol. 2003, 195, 346–355.
- Mlecnik, B.; Tosolini, M.; Kirilovsky, A.; Berger, A.; Bindea, G.; Meatchi, T.; Bruneval, P.; Trajanoski, Z.; Fridman, W.H.; Pages, F.; et al. Histopathologic-based prognostic factors of colorectal cancers are associated with the state of the local immune reaction. J. Clin. Oncol. 2011, 29, 610–618, doi:10.1200/JCO.2010.30.5425.
- 12. Broussard, E.K.; Disis, M.L. TNM staging in colorectal cancer: T is for T cell and M is for memory. J. Clin. Oncol. 2011, 29, 601–603, doi:10.1200/JCO.2010.32.9078.
- 13. Malka, D.; Lièvre, A.; André, T.; Taïeb, J.; Ducreux, M.; Bibeau, F. Immune scores in colorectal cancer: Where are we? Eur. J. Cancer 2020, 140, 105–118.
- Le, D.T.; Uram, J.N.; Wang, H.; Bartlett, B.R.; Kemberling, H.; Eyring, A.D.; Skora, A.D.; Luber, B.S.; Azad, N.S.; Laheru, D.; et al. PD-1 Blockade in Tumors with Mismatch-Repair Deficiency. N. Engl. J. Med. 2015, 372, 2509–2520, doi:10.1056/NEJMoa1500596.
- 15. Germano, G.; Lamba, S.; Rospo, G.; Barault, L.; Magrì, A.; Maione, F.; Russo, M.; Crisafulli, G.; Bartolini, A.; Lerda, G.; et al. Inactivation of DNA repair triggers neoantigen generation and impairs tumour growth. Nature 2017, 552, 116–120, doi:10.1038/nature24673.
- Marisa, L.; Svrcek, M.; Collura, A.; Becht, E.; Cervera, P.; Wanherdrick, K.; Buhard, O.; Goloudina, A.; Jonchère, V.; Selves, J.; et al. The Balance Between Cytotoxic T-cell Lymphocytes and Immune Checkpoint Expression in the Prognosis of Colon Tumors. JNCI J. Natl. Cancer Inst. 2017, 110, 68–77, doi:10.1093/jnci/djx136.
- Zhen, Y.H.; Liu, X.H.; Yang, Y.; Li, B.; Tang, J.L.; Zeng, Q.X.; Hu, J.; Zeng, X.N.; Zhang, L.; Wang, Z.J.; et al. Phase I/II study of adjuvant immunotherapy with sentinel lymph node T lymphocytes in patients with colorectal cancer. Cancer Immunol. Immunother. 2015, 64, 1083–1093, doi:10.1007/s00262-015-1715-3.
- 18. Abken, H. Adoptive therapy with CAR redirected T cells: The challenges in targeting solid tumors. Immunotherapy 2015, 7, 535–544, doi:10.2217/imt.15.15.
- 19. Deng, X.; Gao, F.; Li, N.; Li, Q.; Zhou, Y.; Yang, T.; Cai, Z.; Du, P.; Chen, F.; Cai, J. Antitumor activity of NKG2D CAR-T cells against human colorectal cancer cells in vitro and in vivo. Am. J. Cancer Res. 2019, 9, 945.
- 20. Grothey, A.; Tabernero, J.; Arnold, D.; De Gramont, A.; Ducreux, M.P.; O'Dwyer, P.J.; Van Cutsem, E.; Bosanac, I.; Srock, S.; Mancao, C.; et al. Fluoropyrimidine (FP) + bevacizumab (BEV) + atezolizumab vs FP/BEV in BRAFwt metastatic colorectal cancer (mCRC): Findings from Cohort 2 of MODUL – a multicentre, randomized trial of biomarkerdriven maintenance treatment following first-line induction therapy. Ann. Oncol. 2018, 29, viii714–viii715, doi:10.1093/annonc/mdy424.020.
- Argilés, G.; Saro, J.; Segal, N.H.; Melero, I.; Ros, W.; Marabelle, A.; Rodriguez, M.E.; Albanell, J.; Calvo, E.; Moreno, V.; et al. Novel carcinoembryonic antigen T-cell bispecific (CEA-TCB) antibody: Preliminary clinical data as a single agent and in combination with atezolizumab in patients with metastatic colorectal cancer (mCRC). Ann. Oncol. 2017, 28, iii151, doi:10.1093/annonc/mdx302.003.
- 22. Chen, E.X.; Jonker, D.J.; Kennecke, H.F.; Berry, S.R.; Couture, F.; Ahmad, C.E.; Goffin, J.R.; Kavan, P.; Harb, M.; Colwell, B.; et al. CCTG CO.26 trial: A phase II randomized study of durvalumab (D) plus tremelimumab (T) and best supportive care (BSC) versus BSC alone in patients (pts) with advanced refractory colorectal carcinoma (rCRC). J. Clin. Oncol. 2019, 37, 481, doi:10.1200/JCO.2019.37.4_suppl.481.
- Terme, M.; Pernot, S.; Marcheteau, E.; Sandoval, F.; Benhamouda, N.; Colussi, O.; Dubreuil, O.; Carpentier, A.F.; Tartour, E.; Taieb, J. VEGFA-VEGFR pathway blockade inhibits tumor-induced regulatory T-cell proliferation in colorectal cancer. Cancer Res. 2013, 73, 539–549.
- 24. Yang, J.; Yan, J.; Liu, B. Targeting VEGF/VEGFR to modulate antitumor immunity. Front. Immunol. 2018, 9, 978.
- 25. Akhurst, R.J.; Hata, A. Targeting the TGFβ signalling pathway in disease. Nat. Rev. Drug Discov. 2012, 11, 790–811, doi:10.1038/nrd3810.
- Dosset, M.; Vargas, T.R.; Lagrange, A.; Boidot, R.; Végran, F.; Roussey, A.; Chalmin, F.; Dondaine, L.; Paul, C.; Marie-Joseph, E.L.; et al. PD-1/PD-L1 pathway: An adaptive immune resistance mechanism to immunogenic chemotherapy in colorectal cancer. Oncolmmunology 2018, 7, e1433981, doi:10.1080/2162402X.2018.1433981.
- 27. Tesniere, A.; Schlemmer, F.; Boige, V.; Kepp, O.; Martins, I.; Ghiringhelli, F.; Aymeric, L.; Michaud, M.; Apetoh, L.; Barault, L.; et al. Immunogenic death of colon cancer cells treated with oxaliplatin. Oncogene 2010, 29, 482–491,

doi:10.1038/onc.2009.356.

- 28. Shahda, S.; Noonan, A.M.; Bekaii-Saab, T.S.; O'Neil, B.H.; Sehdev, A.; Shaib, W.L.; Helft, P.R.; Loehrer, P.J.; Tong, Y.; Liu, Z.; et al. A phase II study of pembrolizumab in combination with mFOLFOX6 for patients with advanced colorectal cancer. J. Clin. Oncol. 2017, 35, 3541, doi:10.1200/JCO.2017.35.15_suppl.3541.
- 29. Kim, R.; Chaves, J.; Kavan, P.; Fakih, M.; Kortmansky, J.; Spencer, K.; Wong, L.; Tehfe, M.; Li, J.J.; Lee, M.; et al. Pembrolizumab (pembro) plus mFOLFOX or FOLFIRI in patients with metastatic colorectal cancer (mCRC): KEYNOTE-651 cohorts B and D. Ann. Oncol. 2019, 30, v229–v230, doi:10.1093/annonc/mdz246.085.
- 30. Labrijn, A.F.; Janmaat, M.L.; Reichert, J.M.; Parren, P. Bispecific antibodies: A mechanistic review of the pipeline. Nat. Rev. Drug Discov. 2019, 18, 585–608, doi:10.1038/s41573-019-0028-1.
- 31. Hodgins, J.J.; Khan, S.T.; Park, M.M.; Auer, R.C.; Ardolino, M. Killers 2.0: NK cell therapies at the forefront of cancer control. J. Clin. Investig. 2019, 129, 3499–3510.
- 32. Mehta, R.S.; Rezvani, K. Chimeric Antigen Receptor Expressing Natural Killer Cells for the Immunotherapy of Cancer. Front. Immunol. 2018, 9, doi:10.3389/fimmu.2018.00283.

Retrieved from https://encyclopedia.pub/entry/history/show/14884