

Nanoparticles for Biogas Producers

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Nanotechnology has an increasingly large impact on a broad scope of biotechnological, pharmacological and pure technological applications. The novel notion of dosing ions using modified nanoparticles can be used to progress up biogas production in oxygen free digestion processes.

biofuels

bio-methane

environment

nanoparticles

nanotechnology

biosensors

waste activated sludge

1. Introduction

Over the past few decades, industrialization and population growth has led to a significant increase in energy demand. Currently, fossil fuels are the prime source of basic energy production, contributing 80% of total global consumption. Out of this 80% of primary energy produced by fossil fuels, the transport sector is the major consumer with 58% consumption ^{[1][2]}, of which 80% is being produced by Brazil and USA ^[3]. In future, the transportation fuel demand is estimated to increase up to 55% globally by 2030 and this will increase the demand for biofuels ^{[3][4][5]}.

Due to this intensive consumption and increasing demand in the energy sector, fossil fuel resources are depleting at a rapid pace and there is a dire need to explore and identify new and renewable energy sources globally ^[6].

One such renewable energy source is biogas produced by anaerobic digestion (AD), which utilizes various wastes such as animal manure, ^[7] agricultural waste ^[8] and organic wastes ^[9]. Biogas is produced mainly due to the process of AD, resulting in the formation of CO₂ as a byproduct, which is consumed during photosynthesis and retrieved again, for AD, in the form of agricultural waste and animal manures. This consumption of CO₂ takes place in a closed cycle ^[10], as shown in [Figure 1](#).

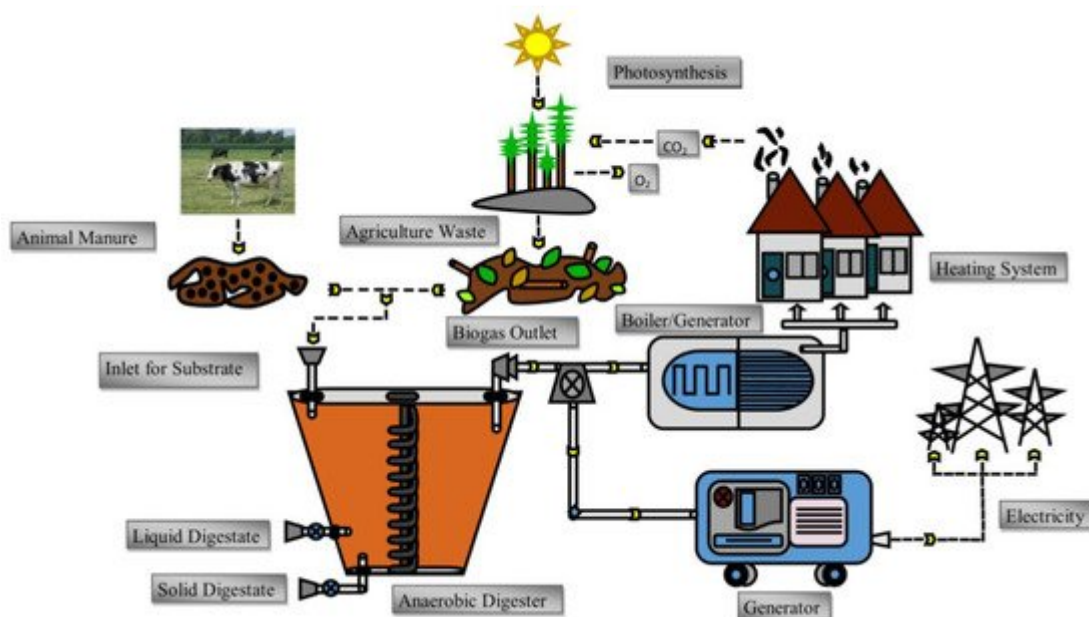


Figure 1. Waste utilization to produce renewable energy.

During AD, four steps are involved in methane production, which include; hydrolysis, acidogenesis, acetogenesis, and methanogenesis. Methane production is a result of the syntrophic microbial relationship. During hydrolyses, bacterial cellulosome and exoenzymes monomerize complex proteins, carbohydrates and fats. In the second step (acidogenesis), along with CO_2 , hydrogen and alcohols, further degradation of monomers into short chain acids takes place. In the third step (acetogenesis), short chain acids are converted into acetate, CO_2 and hydrogen. In the last step (methanogenesis), intermediates are converted into CO_2 and methane by methanogens ^[11] (Figure 2).

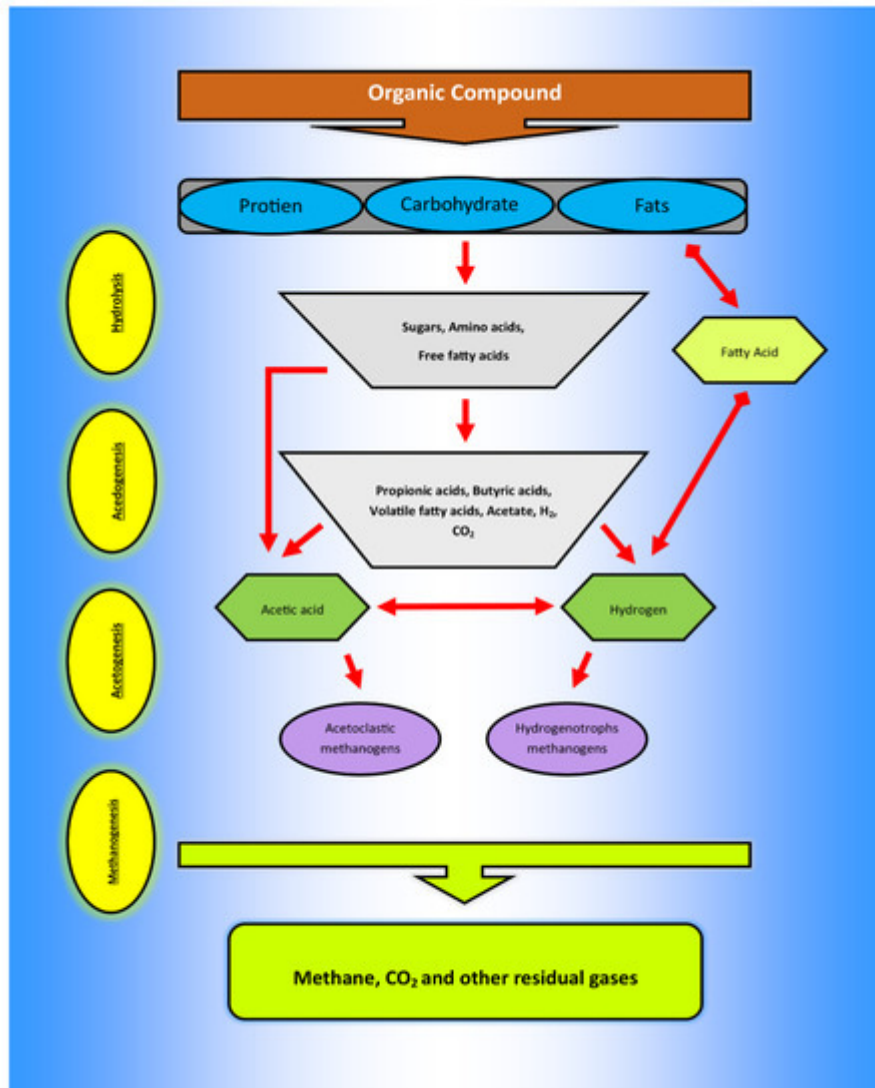


Figure 2. CO₂ and biomethane formation in an anaerobic process.

2. Application of Biosensors in Biogas Monitoring

The assessment of anaerobic digestion is based on the continuous monitoring of organic and volatile fatty acids, resulting in the accumulation of intermediates for unsteady progression conditions [12]. The intensifying public interest in biogas production is a result of the exhaustion of fossil fuels. Anaerobic digestion has the advantage of exploiting industrial waste for energy production and thus treating another modern day problem [13][14]. Efficient methane production and endurance of process stability are resulting outcomes based upon the improvement of several economic and technological aspects. These include a suitable feedstock composition, appropriate biogas purification technologies and ideal conditions for a biogas reactor, which is based upon several physical and biochemical parameters, including pH, alkalinity, gas quality, FOS/TAC (Flüchtige Organische Säuren, i.e., volatile organic acids/Totales Anorganisches Carbonat, i.e., total inorganic carbonate) [15][16][17]. The accumulation of organic acids like formate, lactate and alcohols, and volatile fatty acids (e.g., propionate, acetate, butyrate) results in acidification of the reactor, which clearly indicates process imbalance [18][19][20][21]. The conventional methods for

estimation of acid composition are gas chromatography [22], spectroscopy [23][24] and HPLC (high-performance liquid chromatography) [25][26], which are commonly carried through external sources that cause high cost partanalysis.

3. Metallic Nanoparticles Used for the Enhancement of Bio Gas Production

3.1. Nanoparticles

'Nanomaterials' are the materials with an external dimension or internal or surface structure on a nano scale ranging from 1 to 100 nm in size [27][28]. The chemical origin of nanoparticles is greatly influenced by their chemical origin, which is responsible for their behavior and fate in the environment [29][30]. Nanoparticles are classified into four groups: organic, inorganic, composite and carbon NPs. Nanoparticles possess special chemical, physical and optical characteristics. At the nano scale, properties of the particles change unpredictably, making them behave differently with the same substance at the macro scale. Nanoparticles are ideal in a diversity of areas, such as energy, medical, electronic and commercial products, due to their high reactivity and special features. Using nanoparticles leads to the production of efficient, durable, lighter, firmer, and cleaner products and materials [31].

Different chemical and physical properties of nanoparticles from their macro counterparts make them interesting. The higher chemical reactivity of nanoparticles is due to their high surface area, providing a greater number of reaction sites [32]. Gold (Au) is another example of nanoparticles at the nano scale. Amber does not react with many chemicals at the macro scale and behaves as an inert element, but at the nano scale, gold becomes enormously reactive, behaving as a catalyst to speed up reactions [32]. This extremely reactive property of nanoparticles is due to the ratio between the mass and open area. The human digestive system is a biological example of AD processes being determined by the surface area to volume ratio, microorganism activity aids AD digestion.

3.2. Concentration of Nanoparticles

Nanoparticles have been acquired from both anthropogenic and natural resources. In waste sludge, a very high concentration of NPs could have accumulated. However, the toxicity and the impact of NPs on the sludge treatment stream is still an area that requires a great deal of research [33]. Nguyen determined the effects of ZnO NPs and CeO₂ nanoparticles on the sludge AD process, toxic potential of sludge to plants and bacteria and dewatering process of the sludge.

The concentration of nanoparticles is very important in determining their role for the process of methane and biogas production (Table 1). Not all nanoparticles stimulate the anaerobic digestion system, rather some nanoparticles inhibit the production rate considerably when compared with a controlled sample. Types and concentration of nanoparticles play a vital role in the production rate of the anaerobic digestion system. In comparison with a control sample, the exposure concentration of ZnO at 1000 mg/L resulted in inhibition to 65.3%

biogas volume and 47.7% methane composition. At an endurable exposure concentration of zinc oxide, the inhibition effect could be overcome after an incubation of 14 days [34].

Table 1. Nano additives concentration and their impact on the biogas and methane production rate.

NPs Type	NP size	Concentration	Feedstock	Temperature	Incubation Time	Effect
CeO ₂	192 nm	10 mg/L	Sludge from UASB reactor	30 °C	40	11% Increase in biogas production [33]
Fe ₃ O ₄	7 nm	100 ppm	Waste water Sludge	37 °C	60	180% Increase in biogas production and 234% increase in methane [35]
Fe/SiO ₂	–	105 mol/L	–	55 °C	–	7% Increase in methane production [36]
Pt/SiO ₂	–	105 mol/L	–	55 °C	–	7% Increase in methane production [36]
Co/SiO ₂	–	105 mol/L	–	55 °C	–	48% Increase in methane production [36]
Ni/SiO ₂	–	105 mol/L	–	55 °C	–	70% Increase in methane production [36]
Co	28 nm	1 mg/L	fresh raw manure	37 °C	40	71% increase in biogas production
						45.92% increase in methane production [37]

NPs Type	NP size	Concentration	Feedstock	Temperature	Incubation Time	Effect
Ni	17 nm	2 mg/L	fresh raw manure	37 °C	40	78.53% increase in biogas production
						116.76% increase in methane production [37]
Fe	9 nm	20 mg/L	fresh raw manure	37 °C	40	47.7% increase in biogas production
						67% increase in methane production [37]
Fe ₃ O ₄	7 nm	20 mg/L	fresh raw manure	37 °C	40	73% increase in biogas production
						115.66% increase in methane production [37]
ZnO	140 nm	1 mg/g-TSS	WAS AGS	35 °C	40	No effect [38]
		10 mg/g-TSS			105	No effect [39]
		50 mg/g-TSS				No effect [39]
nZVI	<50 nm	10 mg/g-TSS	WAS	37 °C	30	120% increase in methane production [40]
Fe ₂ O ₃	<30	100 mg/g-	WAS	37 °C	30	117% increase in methane production [41]

2. Nigam, P.S.; Singh, A. Production of liquid biofuels from renewable resources. Prog. Energy Combust. Sci. 2011, 37, 52–68.

NPs Type	NP size	Concentration	Feedstock	Temperature	Incubation Time	Effect	Review
	nm	TSS				production [40]	2–919.
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cobaltite nanoparticles on production and thermostability of cellulases from newly isolated thermotolerant *Eurotium* spp. *NS Appl. Biochem. Biotechnol.* 2014, 174, 1092–1103.

4. An Understanding of Biomass and Their Characteristics

6. Bazmi, A.A.; Zahedi, G. Sustainable energy systems: Role of optimization modeling techniques in power generation and supply—A review. *Renew. Sustain. Energy Rev.* 2013, 15, 3480–3500.

Biomass resources were analyzed on the basis of chemical, biological, physical composition and on the basis of their source. Evaluation of the impact of biomass particle size on the biomass to bioenergy conversion basis was carried out in both biothermal and biochemical aspects. Different biomass types were studied based on pretreatment and particle size. In terms of structure and composition, different effects were produced by different pretreatment methods [41]. For example, lignin can be removed from alkaline pretreatments, and the biomass hemicellulose fraction can be removed by acidic and biothermal pretreatment. Reduction in particle size to increase the biomass specific surface can be carried out to reduce cellulose fiber organization during biomass fibrillation by a milling-based pretreatment, measured by a decline in crystallinity.

7. Bidart, C.; Fröhling, M.; Schultmann, F. Livestock manure and crop residue for energy generation: Macro-assessment at a national scale. *Renew. Sustain. Energy Rev.* 2014, 38, 537–550.

8. Karellas, S.; Boukis, I.; Kontopoulos, G. Development of an investment decision tool for biogas production from agricultural waste. *Renew. Sustain. Energy Rev.* 2010, 14, 1273–1282.

9. Zhang, C.; Su, H.; Baeyens, J.; Fan, T. Reviewing the anaerobic digestion of food waste for biogas production. *Renew. Sustain. Energy Rev.* 2014, 38, 383–392.

10. Al Seadi, T.; Rutz, D.; Prassl, H.; Köttner, M.; Finsterwalder, T.; Volk, S.; Janssen, R. Biogas Enhancement of economically leveraged renewable energy production and energy efficiency can be done with the help of nanotechnology. Nanomaterial Interactions were evident with a few algal biomass species and active sludge. With regards to bioenergy production, inhibition [33] an adverse or increased yield [42] was visible.

11. Deublein, D.; Steinhauser, A. Biogas from Waste and Renewable Resources, Wiley Online Library, Weinheim, Germany, 2008.

12. Röhlen, D.L.; Pilas, J.; Dahmen, M.; Keusgen, M.; Selmer, T.; Schöning, M.J. Toward a Hybrid Biosensor System for Analysis of Organic and Volatile Fatty Acids in Fermentation Processes. *Front. Chem.* 2018, 6, 284.

13. Angelidaki, I.; Ellegaard, L. Co-digestion of manure and organic wastes in centralized biogasplants: Status and future trends. *Appl. Biochem. Biotechnol.* 2003, 109, 95–106.

Nanoparticles have significant effects on microorganisms. NPs strain latent detrimental effects on wastewater microorganisms, according to an overview of their antimicrobial properties. Although, at present, statistical data on the NPs' effect on wastewater microorganisms during aerobic digestion are rather minimal, but it still has a remarkable effect [43][44]. Hence, it is tough to make a particular claim regarding the harmful effect of NPs on wastewater microorganisms. However, minimized efficiency of AS and AD processes, absolute collapse of treatment and environmental pollution from contaminated effluents and utilization of biosolids for changes in soil texture may result due to NPs and microbial community contact [45]

14. Komemoto, K.; Lim, Y.G.; Nagao, N.; Onoue, Y.; Niwa, C.; Toda, T. Effect of temperature on VFA's and biogas production in anaerobic solubilization of food waste. *Waste Manag.* 2009, 29, 2950–2955.

15. Weiland, P. Biogas production: Current state and perspectives. *Appl. Microbiol. Biotechnol.* 2010, 85, 849–860.

16. Andrian, D.; Wresta, A.; Annaja, T.D.; Saepudin, A. A review on optimization production and upgrading biogas through CO2 removal using various techniques. *Appl. Biochem. Biotechnol.* 2014, 172, 1909–1928.

17. Achinas, S.; Achinas, V.; Everink, G.J.W. A technological overview of biogas production from biowaste. *Engineering* 2017, 3, 299–307.

18. Achinas, S.; Achinas, V.; Everink, G.J.W. A technological overview of biogas production from biowaste. *Engineering* 2017, 3, 299–307.

19. Achinas, S.; Achinas, V.; Everink, G.J.W. A technological overview of biogas production from biowaste. *Engineering* 2017, 3, 299–307.

20. Achinas, S.; Achinas, V.; Everink, G.J.W. A technological overview of biogas production from biowaste. *Engineering* 2017, 3, 299–307.

21. Achinas, S.; Achinas, V.; Everink, G.J.W. A technological overview of biogas production from biowaste. *Engineering* 2017, 3, 299–307.

22. Achinas, S.; Achinas, V.; Everink, G.J.W. A technological overview of biogas production from biowaste. *Engineering* 2017, 3, 299–307.

23. Achinas, S.; Achinas, V.; Everink, G.J.W. A technological overview of biogas production from biowaste. *Engineering* 2017, 3, 299–307.

24. Achinas, S.; Achinas, V.; Everink, G.J.W. A technological overview of biogas production from biowaste. *Engineering* 2017, 3, 299–307.

25. Achinas, S.; Achinas, V.; Everink, G.J.W. A technological overview of biogas production from biowaste. *Engineering* 2017, 3, 299–307.

26. Achinas, S.; Achinas, V.; Everink, G.J.W. A technological overview of biogas production from biowaste. *Engineering* 2017, 3, 299–307.

27. Achinas, S.; Achinas, V.; Everink, G.J.W. A technological overview of biogas production from biowaste. *Engineering* 2017, 3, 299–307.

28. Achinas, S.; Achinas, V.; Everink, G.J.W. A technological overview of biogas production from biowaste. *Engineering* 2017, 3, 299–307.

29. Achinas, S.; Achinas, V.; Everink, G.J.W. A technological overview of biogas production from biowaste. *Engineering* 2017, 3, 299–307.

30. Achinas, S.; Achinas, V.; Everink, G.J.W. A technological overview of biogas production from biowaste. *Engineering* 2017, 3, 299–307.

31. Achinas, S.; Achinas, V.; Everink, G.J.W. A technological overview of biogas production from biowaste. *Engineering* 2017, 3, 299–307.

32. Achinas, S.; Achinas, V.; Everink, G.J.W. A technological overview of biogas production from biowaste. *Engineering* 2017, 3, 299–307.

33. Achinas, S.; Achinas, V.; Everink, G.J.W. A technological overview of biogas production from biowaste. *Engineering* 2017, 3, 299–307.

34. Achinas, S.; Achinas, V.; Everink, G.J.W. A technological overview of biogas production from biowaste. *Engineering* 2017, 3, 299–307.

35. Achinas, S.; Achinas, V.; Everink, G.J.W. A technological overview of biogas production from biowaste. *Engineering* 2017, 3, 299–307.

18. Nilsen, K.; De Lencastre, S.; Ahring, B.K. Regulation and optimization of the biogas process, which is a complex system. *Bioprocess Eng.* 2007, 31, 820–830. [\[47\]](#)

19. Boe, K.; Batstone, D.J.; Steyer, J.-P.; Angelidaki, I. State indicators for monitoring the anaerobic digestion process. *Water Res.* 2011, 45, 582–588.

20. Li, J.; He, Q.; Wei, Y.; He, Q.; Peng, X. Early warning indicators for monitoring the process failure of anaerobic digestion system of food waste. *Bioresour. Technol.* 2014, 171, 491–494.

21. Montag, D.; Schink, B. Biogas process parameters—energetics and kinetics of secondary fermentation in methanogenic biomass degradation. *Appl. Microbiol. Biotechnol.* 2016, 100, 2019–2026. (30–40 nm) were compared with their bulk metal salts to evaluate their effects against non-spiked activated sludge (control) [\[48\]](#). This study was conducted using three pilot treatment plants on a pilot scale. In

22. Diamantis, V.; Melidis, P.; Aivasidis, A. Continuous determination of volatile products in anaerobic comparison with the control plant, the specific oxygen uptake rate (SOUR), specific to microbes, increased 200% fermenters by on-line capillary gas chromatography. *Anal. Chim. Acta* 2006, 573–574, 189–194.

23. Falko, H.; Reichling, P.; Andersson, C.; Benz, R. Online monitoring of concentration and dynamics of volatile fatty acids in anaerobic digestion processes with mid-infrared spectroscopy. *Bioresour. Technol.* 2015, 188, 237–249.

24. Stockl, A.; Lichti, F. Near-infrared spectroscopy (NIRS) for a real time monitoring of the biogas influences in nature. *Bioresour. Technol.* 2018, 247, 1249–1252.

25. Zumbusch, P.; Meyer, J.; Ten Brunck, G.; Märkl, H. Online monitoring of organic substances with high-pressure liquid chromatography (HPLC) during the anaerobic fermentation of waste water. *Appl. Microbiol. Biotechnol.* 1994, 42, 140–146.

26. Schiffrs, J.; Baumann, M.E.M.; Selmer, I. Facile analysis of shortchain fatty acids as 4-nitrophenyl esters in complex anaerobic fermentation samples by high performance liquid chromatography. *J. Chromatogr. A* 2011, 1218, 5848–5851.

27. ISO/TS 27687. Nanotechnologies Terminology and Definitions for Nanobiotechnology. Nanoparticle, Nanofibre, Nanoplate; ISO (International Organization for Standardization); Geneva, Switzerland, 2008.

28. ISO/TS 80004-1. International Standardization Organization Technical Standard:

Nanotechnologies Vocabulary Part 1: Core Terms; ISO (International Organization for

7. Phytotoxicity/Ecotoxicity Effect of NPs

29. Farré, M.; Sanchís, J.; Barceló, D. Analysis and assessment of the occurrence, the fate and the behavior of nanomaterials in the environment. *Trend Anal. Chem.* 2011, 30, 517–527.

30. Stone, J.; Nowak, B.; Baun, A.; van der Brink, J.; van der Kammen, I.; Dorsan, K.; Hildebrand, E.; Hassel, M.; Janke, E.; et al. Nanomaterials for environmental studies: Classification and strategies for physico-chemical characterization. *Sci. Total Environ.* 2010, 408, 1745–1754.

31. US Nanoscale Science, Engineering, and Technology Subcommittee of the Committee on Technology, Environmental, Health and Safety Research Strategy, National Science and Technology Council: Washington, DC, USA, 2011.

There has been a significant focus on aquatic, rather than terrestrial, plants in term of ecotoxicity. Toxic effects of nanoparticles, on the germination and root growth of some plant species, have been reported in some studies [56].

32. Nanotechnology Centre for Learning and Teaching, Advancing economic Leadership through Integrated STEM Education; Nanotechnology Centre for Learning and Teaching: Evanston, Chicago, IL, USA, 2015.

One of these studies was designed to compare the effects of five types of commonly used nanoparticles with their corresponding bulk material in regards to biomass, germination and root elongation in the Cucurbita pepo (Zucchini) plant. Effects of CuO and ZnO Nanoparticles on Anaerobic (Digestion) and Toxicity of Cu. To end. Digested Sludge. Master's Thesis, Asian Institute of Technology, Khlong Nuea, Thailand, 2013.

the effect of nanoparticles or bulk Ag concentration on transpiration, Ag content and biomass of the zucchini plant. 34. Luna-del-Risco, M.; Orupold, K.; Dubourguier, H.-C. Particle-size effect of CuO and ZnO on biogas Assessment related to the impacts of nanoparticles on agricultural plants will help find out the potential hazards of and methane production during anaerobic digestion. J. Hazard. Mater. 2011, 189, 603–608. [57].

35. Casals, E.; Barrena, R.; García, A.; González, E.; Delgado, L.; Busquets-Fité, M.; Font, X.; Arbiol, J.; Glatzel, P.; Kvashnina, K.; et al. Programmed iron oxide nanoparticles disintegration in anaerobic digesters boosts biogas production. Small 2014, 10, 2801–2808.

8. Mechanism of Microbial Activity

36. Ram, V.S.; Singh, L.; Suryanarayana, M.V.; Alam, S. Effect of iron, nickel and cobalt on bacterial activity and dynamics during anaerobic oxidation of organic matter. Water Air Soil Pollut. 1999, 117, 305–312.

The focus of this research is mainly on manufacturing of commercialized nanomaterials, which are widely applicable, such as fullerene, metal oxides and CNTs. It is important to assess the fate of nanoparticles on the environment when applying them commercially on a larger scale. Ag nanoparticles are capable of interacting with slurry. Renew. Energy 2016, 87, 592–598.

38. Mullick, Chao, A. The long-term effect of Zn nanoparticles on waste activated sludge anaerobic digestion. Water Res. 2011, 45, 5612–5620. [59]. This change is directly related to concentration, shape and size of the nanoparticles [60]. This influence is confirmed by a study using *Escherichia coli* that affirmed that gaps in the integrity of the player are created by the accumulation of Ag load of zinc oxide nanoparticles during biological wastewater treatment. Environ. Sci. Technol. NPs, which increased the permeability leading to cell death of bacteria [61]. 2012, 46, 5997–6003.

40. Wang, J.; Zhang, D.; Dai, L.; Chen, J.; Dai, X. Effects of Metal Nanoparticles on Methane Production from Waste-Activated Sludge (WAS) Microorganism Community Shift in Anaerobic Granular Sludge. Sci. Rep. 2016, 6, 25857.

Due to the health hazard of nanoparticles, the health upon their release into the environment. So far, the mechanism of toxicity for each nanoparticle is not understood exactly, but various characteristics may result in damage to the exposed organisms. Reactive oxygen species (ROS), such as super oxides (O_2^-), singlet oxygen ($^1\text{O}_2$) and free radicals (OH^\cdot), are generated by nanoparticles that employ various adverse effects on microbes, such as scattered vesicles, disruption of cell wall, enzyme inhibition and protein and sugar membrane leakage [3729].

41. Kumar, P.; Barrett, D.M.; Delwiche, M.J.; Stroeve, P. Methods for pretreatment of lignocellulosic biomass for efficient hydrolysis and biofuel production. Ind. Eng. Chem. Res. 2009, 48, 3713–3729.

42. Victor, F.P.; Ferrer, A.S. Enhancement of Biogas Production in Anaerobic Digesters Using Iron Oxide Nanoparticles; Catalan Institute of Nanoscience and Nanotechnology (ICN2); University Autònoma Barcelona; Barcelona, Spain, 2011.

Significant toxicity was caused by these nanoparticles to the viability of Gram negative bacterial cells by increasing their antibacterial effects. Chen et al. [63] reviewed the toxicity of nanomaterials on biomass and found that the chemical stability of nanoparticles of Ag, TiO_2 , Al_2O_3 and SiO_2 have

43. Batley, G.E.; Kirby, J.R.; McLaughlin, M.J. Fate and risks of nanoparticles in aquatic and terrestrial environments. *Acc. Chem. Res.* 2012, 46, 854–864.
44. Krysanov, E.Y.; Pavlov, D.S.; Demidova, I.B.; Dgebuadze, Y.Y. Effect of nanoparticles on aquatic organisms. *Biol. Bull.* 2010, 37, 406–412.
45. Hoffmann, C.; Christoffi, N. Testing the toxicity of influents to activated sludge plants with the *Vibrio fischeri* bioassay utilizing a sludge matrix. *Environ. Toxicol.* 2001, 16, 422–427.
46. Klasen, H.J. Historical review of the use of silver in the treatment of burns. I. Early uses. *Burns* 2000, 26, 117–130.
47. Li, W.R.; Xie, X.B.; Shi, Q.S.; Zeng, H.Y.; You-Sheng, O.Y.; Chen, Y.B. Antibacterial activity and mechanism of silver nanoparticles on *Escherichia coli*. *Appl. Microbiol. Biotechnol.* 2010, 85, 1115–1122.
48. Eduok, S.; Martin, B.; Villa, R.; Nocker, A.; Jefferson, B.; Coulon, F. Evaluation of engineered nanoparticle toxic effect on wastewater microorganisms: Current status and challenges. *Ecotoxicol. Environ. Saf.* 2013, 95, 1–9.
49. Klaine, S.J.; Alvarez, P.J.; Batley, G.E.; Fernandes, T.F.; Handy, R.D.; Lyon, D.Y.; Mahendra, S.; McLaughlin, M.J.; Lead, J.R. Nanomaterials in the environment: Behavior, fate, bioavailability, and effects. *Environ. Toxicol. Chem.* 2008, 27, 1825–1851.
50. Pelletier, D.A.; Suresh, A.K.; Holton, G.A.; McKeown, C.K.; Wang, W.; Gu, B.; Mortensen, N.P.; Allison, D.P.; Joy, D.C.; Allison, M.R.; et al. Effects of engineered cerium oxide nanoparticles on bacterial growth and viability. *Appl. Environ. Microbiol.* 2010, 76, 7981–7989.
51. Martinez-Gutierrez, F.; Olive, P.L.; Banuelos, A.; Orrantia, E.; Nino, N.; Sanchez, E.M.; Ruiz, F.; Bach, H.; Av-Gay, Y. Synthesis, characterisation, and evaluation of antimicrobial and cytotoxic effect of silver and titanium nanoparticles. *Nanomed. Nanotechnol. Biol. Med.* 2010, 6, 681–688.
52. Martinez-Castanon, G.A.; Nino-Martinez, N.; Martinez-Gutierrez, F.; Martinez-Mendoza, J.R.; Ruiz, F. Synthesis and antibacterial activity of silver nanoparticle with different sizes. *J. Nanopart. Res.* 2008, 10, 1343–1348.
53. Choi, O.; Deng, K.K.; Kim, N.J.; Ross Jr, L.; Surampalli, R.Y.; Hu, Z. The inhibitory effect of silver nanoparticles, silver ions, and silver chloride colloids on microbial growth. *Water Res.* 2008, 42, 3066–3074.
54. Pal, S.; Tak, Y.K.; Song, J.M. Does the antibacterial activity of silver nanoparticles depend on the shape of the nanoparticles? A study of the Gram-negative bacterium *Escherichia coli*. *Appl. Environ. Microbiol.* 2007, 73, 1712–1720.
55. García, A.; Delgado, L.; Torà, J.A.; Casals, E.; González, E.; Puentes, V.; Font, X.; Carrera, J.; Sánchez, A. Effect of cerium dioxide, titanium dioxide, silver, and gold nanoparticles on the

- activity of microbial communities intended in wastewater treatment. *J. Hazard. Mater.* 2012, 199–200, 64–72.
56. Lin, D.; Xing, B. Phytotoxicity of nanoparticles: Inhibition of seed germination and root growth. *Environ. Pollut.* 2007, 150, 243–250.
57. Stampoulis, D.; Sinha, S.K.; White, J.C. Assay-dependent phytotoxicity of nanoparticles to plants. *Environ. Sci. Technol.* 2009, 43, 9473–9479.
58. Brar, S.K.; Verma, M.; Tyagi, R.D.; Surampalli, R.Y. Engineered nanoparticles in wastewater and wastewater sludge: Evidence and impacts. *Waste Manag.* 2010, 30, 504–520.
59. Lazar, V. Quorum sensing in biofilm show to destroy the bacterial citadels or their cohesion/power? *Anaerobe* 2011, 17, 280–285.
60. Lu, Z.; Dai, T.; Huang, L.; Kurup, D.B.; Tegos, G.P.; Jahnke, A.; Wharton, T.; Hamblin, M.R. Photodynamic therapy with a cationic functionalized fullerene rescues mice from fatal wound infections. *Nanomedicine* 2010, 5, 1525–1533.
61. Rai, M.; Kon, K.; Ingle, A.; Duran, N.; Galdiero, S.; Galdiero, M. Broad-spectrum bioactivities of silver nanoparticles: The emerging trends and future prospects. *Appl. Microbiol. Biotechnol.* 2014, 98, 1951–1961.
62. Mu, H.; Chen, Y.; Xiao, N. Effects of metal oxide nanoparticles (TiO₂, Al₂O₃, SiO₂ and ZnO) on waste activated sludge anaerobic digestion. *Bioresour. Technol.* 2011, 102, 10305–10311.
63. Chen, J.L.; Ortiz, R.; Steele, T.W.; Stuckey, D.C. Toxicants inhibiting anaerobic digestion: A review. *Biotechnol. Adv.* 2014, 32, 1523–1534.

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