Endocrine Disorders in Autoimmune Rheumatism

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Autoimmune rheumatological diseases' incidence and prevalence have risen over the last decades and they are becoming increasingly important worldwide. Thyroid autoimmune diseases share with them an imbalance in the immune system that lead to a pro-inflammatory environment. Usually this is the result of a multi-factorial process. In fact, it includes not only a possible genetic predisposition, but also environmental causes like microbiota dysbiosis, diet rich in processed foods, exposure to toxicants and infections.

Keywords: rheumatological diseases; thyroid diseases; corticosteroids; cushing's syndrome; withdrawal syndrome

1. Introduction

Autoimmune rheumatological diseases include a wide and heterogeneous spectrum of disorders mainly affecting joints and the anatomical structures associated to them: muscles, bones, tendons, tendon sheaths, ligaments, tendon and ligament insertions, synovial bursa and fasciae [1][2]. They are chronic progressive conditions whose main symptoms include articular pain, stiffness, swelling, redness, warmness and mobility impairment.

Recent data show that both prevalence and incidence of rheumatological diseases have increased over the last decades worldwide and they have become a major public. health challenge [3]. In fact, they represent one of the most frequent causes of work absence, disability, morbidity and, as a result, consistent healthcare expenditures.

Common autoimmune disorders tend to coexist in the same subjects and to cluster in families. Rheumatoid arthritis (RA) is the most common of these diseases with a global prevalence of 0.3-1% and an annual incidence of 0.02-0.05% according to WHO data [4].

The association between RA and thyroid autoimmune diseases (AITDs) like Graves' disease and Hashimoto's thyroiditis is well-known but many aspects remain to be clarified $^{[5]}$.

These conditions share common immunopathogenic mechanisms and it has been reported a relevant influence of genetic susceptibility $^{[\underline{G}][\underline{\mathcal{I}}]}$.

Moreover, patients affected by autoimmune rheumatological diseases are usually treated with corticosteroids and this therapy may last years with consistent cumulative doses. As a result, it is not uncommon to see among these patients cases of endocrine disorders caused by chronic glucocorticoids treatment like iatrogenic Cushing's syndrome or tertiary adrenal insufficiency after steroids' withdrawal.

In this review, we are going to deepen the common aspects of the pathogenicity of rheumatological and thyroid autoimmune diseases and the endocrine dysfunctions related to chronic glucocorticoids treatment.

2. Thyroid Diseases

The prevalence of AITDs, including Hashimoto's thyroiditis, Graves' disease and postpartum thyroiditis, is estimated to be as high as 5% of the general population (abnormal thyroid function varies within 7–9% in females and 1–2% in males across different populations) [8].

The pathogenesis of AITDs, like other autoimmune diseases, is multifactorial, combining genetic, immune, environmental and hormonal influences.

Hashimoto's thyroiditis (HT) is a typical T-cell-mediated autoimmune disease characterized by a diffuse goiter, the presence of anti-thyroid peroxidase (anti-TPO) and/or anti-thyroglobulin (anti-Tg) antibodies in serum (although it can be

seronegative too), varying degree of thyroid hypofunction, and intrathyroidal infiltration of B and T lymphocytes with CD4+ type 1 T helper (Th1) subtype predominance [9].

In Graves' disease, lymphocytic infiltration is mild and involves mainly CD4+ type 2 T helper (Th2) cells, which induce the production of antibodies to bind to the thyroid stimulating hormone (TSH) receptor [10].

Autoimmunity is crucial also for the development of rheumatological diseases, despite the pathogenicity process implies the production of systemic antibodies and not only organ-specific as seen in thyroid diseases. As a result, the diagnosis of these conditions is more challenging and there is a wide spectrum of non-disease specific antibodies associated with rheumatological diseases such as anti-nuclear antibodies (ANA), anti-double stranded DNA antibodies (Anti ds-DNA), anti-Smith (anti SM) antibodies or rheumatoid factor (RF).

The association of autoimmune thyroid diseases and a considerable number of autoimmune diseases including rheumatological diseases such as rheumatoid arthritis, systemic lupus erythematosus and systemic sclerosis, is called Autoimmune Polyendocrine Syndrome type 3 (APS 3).

The correlation between thyroid and rheumatological diseases is that they share in their pathological process both genetic and environmental factors. This is due to both pre-existing and environmental factors. In fact, the onset of autoimmune diseases commonly is in late childhood or late adulthood confirming the fact that phenotype depends not only from genetic factors but also from non-genetic processes like antibody production, epigenetic programming as well as environmental factors like tobacco smoking and the intestinal microbiota [11].

2.1. Genetic Factors

Many genome-wide association studies (GWAS) showed that there are several genetic loci associated with the likelihood of developing both thyroid and rheumatological diseases.

The strongest correlation is with human leukocyte antigen (HLA), the locus of the genes that encode proteins found on cell surface responsible for the regulation of the adaptive immune system. HLAs corresponding to major histocompatibility complex II (MHC II) present antigens from outside the cell to T-helper lymphocytes (or CD4 positive T cells) [12].

Among those HLAs there are autoimmunity-prone haplotypes like HLA-DQ8/DR4 that favour a strong Th-1 and Th-17 proinflammatory response to self-antigens and they are usually found in patients affected by some autoimmune rheumatological diseases such as rheumatoid arthritis, systemic lupus erythematosus, systemic sclerosis and in autoimmune thyroid disease [13][14][15].

A polymorphism of a gene coding protein tyrosine phosphatase non-receptor type (PTPN22) involved in several signaling pathways associated with the immune response was found in other autoimmune disorders like juvenile rheumatoid arthritis, and Graves' disease $\frac{[16]}{}$. However, a Russian study did not found any correlation between the above-mentioned polymorphism and rheumatoid arthritis probably because the prevalence of the former varies in different ethnic groups as reported in previous studies $\frac{[17][18]}{}$.

The cell surface co-receptor cytotoxic T-lymphocyte antigen-4 (CTLA-4) is a critical attenuator of T-cell activation and it is a component of the regulatory systems of peripheral tolerance [19]. Genome wide association studies elucidated that CTLA-4 polymorphisms represent a locus of susceptibility for autoimmune thyroid diseases and rheumatoid arthritis [20][21] [22]

A proof of that is the possible development of Hashimoto's thyroiditis, Graves' disease, arthritis and polymyalgia rheumatica or the flaring up of pre-existing rheumatological conditions like rheumatoid arthritis during treatment with ipilimumab, an immune checkpoint inhibitor drug targeting CTLA-4 [23][24][25][26].

Moreover, rare heterozygous CTLA4 mutations can lead to common variable immunodeficiency (CVID) with non-functional FoxP3+ regulatory T cells (Treg cells) resulting in systemic autoimmunity associated with defective response to infections [27][28][29].

Polymorphisms of interleukin 2 receptor alfa (IL-2 RA or CD 25) that also cause a dysfunction of regulatory T cells are linked to a wide spectrum of rheumatological autoimmune diseases such as systemic lupus erythematosus and rheumatoid arthritis [30].

There are also other promising susceptibility loci linked to the development of autoimmune diseases like IL23R, TYK2 and A20 that are currently being studied [31][32].

2.2. Environmental Factors

Autoimmune thyroid and rheumatological diseases can be associated with several environmental factors including intestinal microbiota, diet components, vitamin D, chemicals including endocrine disruptors, cigarette smoke and infections.

Bacteria that form intestinal microbiota are involved in maintaining the homeostasis of the immune system by the secretion of metabolites. Some of them are short-chain fatty acids (SCAs) like acetate, propionate and butyrate and they are produced through the fermentation of non-digestible carbohydrates.

Their function is to regulate T cells differentiation, stimulate the production of anti-inflammatory or anti-microbial mediators and they are crucial for epithelial barrier function as they prevent a condition characterized by an altered intestine permeability known as the "leaky gut" [33][34][35].

In fact, an abnormal composition of intestinal microbiota, also called gut dysbiosis, alters the expression level of Toll-like receptors (TLRs) of antigen presenting cells, causing an imbalance between Th17 and Treg (Regulatory T cells) and has a major impact of antibodies production [36].

All this process may lead to an increasing number of autoantigens targeted by T cells and to the activation of autoreactive B cells that produce autoantibodies versus a large number of autoantigens leading to autoimmunity [37][38].

A study by Shor et al. found that gastrointestinal-related antibodies are associated to a wide spectrum of autoimmune diseases including thyroid and rheumatological diseases. For instance the titer of antibodies anti Saccharomyces cerevisae (ASCA), which is a yeast usually found in human microbiota, was found to be significantly higher in patients affected by Graves' disease or by systemic lupus erythematosus than in the general population $\frac{[39]}{}$.

Therefore, recent and ongoing studies are focusing on the possibility of using probiotics and fecal transplantation as a treatment of autoimmune disease with promising results $\frac{[40][41][42]}{[40][41][42]}$.

Lately, studies have suggested that low vitamin D concentrations and other conditions which may result in reduced vitamin D function (e.g., certain Vitamin D receptor gene polymorphisms, pathologies of vitamin D gene and its binding protein) may increase the risk of AITDs [43][44]. Vitamin D is known to regulate the adaptive immunity and its deficiency has been linked to the development of Hashimoto's thyroiditis, Graves' disease, rheumatoid arthritis and systemic lupus erythematosus [45][46][47]. In fact, it has been described that the lack of this crucial hormone may cause gut dysbiosis and, a result, a pro-inflammatory environment [48]. The supplementation with the inactive form of vitamin D, cholecalciferol, in animal studies led to the improvement of gut microbiota and intestinal inflammation [49][50]. Moreover, it has been demonstrated that cholecalciferol has beneficial effects on AITDs and on rheumatological manifestations, and these results may be related to a change towards a more favourable microbiota composition [45][51].

Diet is crucial for a healthy and functional microbiota and the change in human nutrition that has occurred since the mid 20th century probably has played a relevant role in the increase of autoimmune diseases' prevalence and incidence. In fact, the need of long-lasting food led to its industrial processing and the adding of compounds like preservatives, artificial sweeteners and emulsifiers that alter the microbiota composition [52][53][54].

However, the consumption of processed food does not affect only microbiota. Usually this type of food has a high content of sodium that causes an increase in hypertonicity that recent studies have linked to an enhancement of Th17 immune response. This causes an imbalance between pro-inflammatory and anti-inflammatory mediators [55][56][57]. A study by Salgado E. et al. with 18,555 partecipants found a significantly higher proportion of patients affected by rheumatoid arthritis between high-salt consumers with a dose dependent relationship [58]. This report was later confirmed in subsequent studies [59][60].

Among chemicals, several toxicants have been described to induce both systemic and organ-specific autoimmune diseases, including rheumatological and thyroid ones.

For instance, exposure to silica and asbestos was found to be associated with the developing of systemic lupus erythematosus, systemic sclerosis, rheumatoid arthritis and vasculitis. The phagocytosis of asbestos and silica crystals

leads to inflammasome activation causing an increase in pro-inflammatory cytokine expression, the generation of reactive oxygen and nitrogen species and the induction of aberrant cell death [61][62][63][64].

Excess iodine ingestion is known to be a contributing factor to the development and the exacerbation of autoimmune thyroiditis. In fact, studies showed that iodine excess can lead to increased thyroid lesions and to the increase in thyroid-specific antibodies [65][66][67].

Chemicals that cause an impairment to the endocrine function of one or more glands are called "endocrine disruptors" and they may contribute to the development of rheumatological and thyroid autoimmune diseases.

Indeed, the increase in the consumption of plasticizers, nitrate and mercury has been linked to the rising number of patients affected by autoimmune thyroid diseases, rheumatoid arthritis and systemic lupus erythematosus $\frac{[68][69][70][71]}{[68][69][70][71]}$.

Moreover, it has been deeply studied that the toxicants contained in cigarette smoke can cause a relevant genetic damage by increasing the oxidative stress and an imbalance between pro-inflammatory and anti-inflammatory immune response $\frac{72}{73}$. As a result, in smokers autoimmune diseases like rheumatoid arthritis, systemic lupus erythematosus and Graves' disease are more frequent and severe $\frac{75}{75}$

To conclude, many studies highlighted a pivotal role for infections in autoimmune diseases. In particular, a strong relationship between autoimmune rheumatological diseases and hepatitis C virus, hepatitis B virus, Herpesviridae, Staphilococcus aureus was found [84][85][86].

For autoimmune thyroid diseases a causative role for infectious agents in humans still has to be established [87][88].

The main mechanisms by which infections can participate in autoimmune diseases' development are molecular mimicry, bystander activation, epitope spreading and polyclonal activation of B cells.

Molecular mimicry is probably the most important of them and it occurs because of the cross-reactivity of microbial and self-antigens. As a consequence, autoreactive T-cells are activated and they trigger an autoimmune reaction.

Bystander activation is the process typical of the chronic inflammation state by which antigen-presenting cells stimulate the proliferation of T and B cells by presenting them self-epitopes of the damaged tissues.

Epitope spreading occurs when the proliferation of T and B cells of bystander activation is directed against a different part of the same protein or against a different protein.

To conclude, the polyclonal activation of B cells is the consequence of the constant activation of immunity that leads to the formation of immune-complexes that cause tissue damage $\frac{[89][90][91]}{[89][90][91]}$.

3. Effects of Chronic Glucocorticoid Treatment

As seen in the previous chapter, an over-activated immune system is crucial for the pathogenesis of rheumatological autoimmune systemic diseases and glucocorticoids (GCs), along with other immunosuppressant drugs, still represent a leading treatment. In fact, they are anti-inflammatory drugs that strongly suppress the immune system.

First, they stimulate the transcription factors coding for anti-inflammatory gene products such as interleukin-1 receptor type II (IL-1R2) and interleukin 10 (IL-10). GCs also inhibit the synthesis of almost all known inflammatory cytokines blocking transcription factors that are essential for the pro-inflammatory response: nuclear factor-kappa-B (NF-kB) and activator-protein 1 (AP-1) [92][93]. Moreover, their activity is also against the secretion of pro-inflammatory cytokines affecting post-translational events [94].

It is estimated that approximately 1% of the Western world receives prolonged therapy with synthetic GCs, resulting in a supraphysiological exposure causing several effects $^{[95]}$. They can be organ-specific, ranging from adrenal glands to thyroid dysfunction. GC-induced impairment of HPA axis will be discussed later. Although thyrocytes express GC receptors that play an important role in differentiation of thyroid cells, an exposure to high doses of GC can cause thyroid function impairment $^{[96]}$. In fact, glucocorticoids decrease serum TSH, reduce TSH response to TRH and impair peripheral conversion of T4 to T3 $^{[97][98]}$. This effect of reducing T3 serum levels is the reason of GC administration during thyroid storm $^{[99]}$. Chronic GC treatment can also cause multi-systemic effects known as Cushing's syndrome $^{[95]}$.

Glucocorticoids are drugs with different properties and some of them are considered as adverse effects of GCs.

A long-lasting treatment with GCs may lead to a severe adverse event, the so-called iatrogenic Cushing's syndrome $\frac{[100]}{}$.

All available forms of GCs are capable of producing Cushing's syndrome with signs and symptoms that are generally related to the dose and the duration of treatment. Although supra-physiological doses (>7.5 mg/d of prednisone) usually are required before patients manifest significant Cushingoid effects, some patients can develop Cushingoid appearance with chronic administration of lower doses (5 mg/d of prednisone) [100][101].

latrogenic Cushing's syndrome is considered as the most common cause of hypercortisolism.

Its clinical presentation is similar to that of the endogenous form and includes plethora, striae rubrae, thin skin and proximal muscles' hypotrophy. These patients are usually obese with a redistribution of body fat to truncal areas, buffalo hump, enlarged dorsocervical and supraclavicular fat pads and the classic moon face.

Moreover, like in the endogenous form, iatrogenic Cushing's syndrome can be linked with systemic diseases and comorbidities [100][101][102].

Among metabolic alterations, GCs increase insulin resistance leading to hyperglycemia, impaired glucose tolerance and eventually diabetes mellitus especially in predisposed patients [100][102][103]. Moreover, the resulting hyperinsulinism, along with hyperphagia related to GCs therapy, contributes to weight gain and obesity.

Dyslipidemia seems to be less frequent than other metabolic comorbidities in human Cushing's syndrome. Nevertheless, it plays an important role in determining the global cardiovascular risk in overt and subclinical Cushing's syndrome. In particular, it is reported an increase of triglycerides and total cholesterol levels whereas HDL levels can vary [104].

Despite their limited mineralocorticoid activity, treatment with high doses of GCs leads to an increased excretion of potassium and sodium absorption resulting in higher blood pressure. Moreover, it has been described that cortisol excess increases levels of clotting factors and impairs fibrinolytic capacity rising the overall risk of acute *cardiovascular* events [104][105]

For what concerns endocrine dysfunction, GCs can cause secondary hypogonadism, GH-deficiency and secondary hypothyroidism [100].

GCs interfere with both intestinal and renal calcium absorption, causing nephrolithiasis and reduced bone density. As a result, many of these patients suffer from osteoporosis that is also provoked by the stimulation of osteoclastogenesis and by the inhibition of the function of osteoblasts operated by these drugs [102][106].

Cushing's syndrome is also characterized by *myopathy* because GCs affect both transcription and translation of enzymes involved in the metabolism of fats and in the process of protein synthesis in the muscle $\frac{[107][108]}{}$.

Although steroids are known to inhibit arachidonic acid and consequently cyclooxygenase 1 (COX-1) that is essential for gastric mucosa protection, the risk of developing gastritis and *gastrointestinal* ulcers during long-term GCs therapy is debated [109][110][111][112].

Corticosteroid treatment can also cause posterior sub-capsular cataract (PSC), as first described in 1960 in 44 patients affected by rheumatoid arthritis chronically treated with GCs. The main mechanism of pathogenicity responsible for this condition is the alteration of eye epithelial proteins transcription [113][114]. Another eye disturb that can be provoked by GCs therapy is glaucoma probably because of the decreased trabecular meshwork outflow that results in an increased intraocular pressure [115][116][117].

For what concerns dermatological manifestations of Cushing's syndrome it is well established that GCs alter collagen, mucopolysaccharides, fibroblast and keratinocytes proliferation leading to vascular fragility and skin atrophy [118].

The use of GCs in predisposed patients can cause a variety of psychiatric conditions, including mania, psychosis, depression, and delirium. Lewis et al. reported severe psychotic reactions in 5.7% of patients taking GCs and mild-to-moderate reactions in 28% of patients [119][120]. Moreover, along with the above-cited disturbs, a recent study described a negative association between cumulative exogenous corticosteroid exposure and the volume of the entire hippocampal region, affecting also dentate gyrus and CA3 regions. This finding can be the explanation of the negative impact of

glucocorticoids on memory retrieval, concentration and efficiency that prompted the investigators to coin the term *steroid dementia*. This disturb showed to be reversible after GC therapy suspension [121].

As previously described, GCs have a strong immuno-suppressant activity so it is has been frequently described an increased infection rate for people with Cushing's syndrome, especially from intracellular and opportunistic pathogens [122] [123]

3.2. Adrenal Insufficiency and Corticosteroids' Withdrawal Syndrome

Chronic exposure to exogenous glucocorticoids also causes a suppression of hypothalamic-pituitary-adrenal axis (HPA) [101]

This condition is called tertiary adrenal insufficiency. While the risk of adrenal insufficiency (AI) is known if GCs therapy is stopped suddenly in chronic users or during acute illness if patients are under GC low doses, its risk is underestimated in other conditions (e.g., GCs tapering, in patients under medium GC doses that experience an acute illness or a surgical intervention). The syndrome that follows reduction or discontinuation of glucocorticoids bears similar but not identical symptoms and signs of adrenal insufficiency and it is called corticosteroids' withdrawal syndrome. They include hypotension, hypoglycemia, fatigue, dizziness, weakness and lethargy [124][125].

The withdrawal syndrome is experienced by the patients after discontinuation of glucocorticoid therapy. It has been considered a withdrawal reaction due to an established physical dependence on supraphysiological glucocorticoid levels [125][126][127]

Although the risk of developing hypoadrenalism/corticosteroids'withdrawal syndrome is higher for patients treated with oral steroids with a dose \geq 20 mg/day of prednisone (or equal dose of other steroids) for \geq 5 days, there is a wide variability of the individual response to exogenous GCs [124][125][126][127][128]. Moreover, the full recovery time of adrenocortical hypoplasia or atrophy induced by GCs can last weeks, months and in some cases years, even with blood tests that show normal ACTH and cortisol secretion [124][125][126][127][128].

In the pathogenesis of corticosteroids'withdrawal syndrome several mediators may be involved, including CRH, vasopressin, POMC, central noradrenergic and dopaminergic systems, cytokines, and prostaglandins $\frac{[126][127][128]}{[127][128]}$.

CRH in the brain not only activates the HPA axis but also mediates stress-related behavioral effects. Moreover, a well-functioning CRH system in the brain seems necessary for adequate mesolimbic dopaminergic function. Thus, central CRH hyposecretion may contribute to anxiety and depression via inadequate stimulation of dopaminergic neurons terminating in the nucleus accumbens. Some investigators feel that hyposecretion of central CRH plays an important role in the pathogenesis of atypical depression. [120][128][129]. In this scenario of both physical and psychophysical symptoms related to corticosteroid's withdrawal, the term "GCs addiction" was coined [130].

Although not being directly related to the hypocortisolism, glucorticosteroids' withdrawal also may lead to a relapse of the underlying autoimmune rheumatological condition treated with GCs $^{[131]}$.

As a result, to avoid that and to prevent the effects of GC withdrawal syndrome, a gradual tapering of glucocorticoid therapy has become the standard of practice.

3.3. Is There a Safe GC Treatment Regimen?

GCs adverse events were shown to be more relevant among patients receiving a moderate (>5 mg and <10 mg of Prednisone Equivalent Dose or PED) or high (>10 mg) daily dose compared to low-dose (<5 mg) users [132]. In fact, a recent study by Mebrahtu et al., including 111,804 patients affected by rheumatological diseases with a follow up period of 5.5 years, found that for every increase of daily dose of 5 mg PED the risk for adrenal insufficiency increased by 7%, 9% for Cushing syndrome and 6% for mortality. The incidence of Cushing's syndrome and adrenal insufficiency were reported to be respectively 0.55 and 0.41 for 1000 people/year [125].

This knowledge led to the recommendation of the rheumatological associations like EULAR (European Alliance of Associations for Rheumatology) to carefully monitor the formers agreeing that the risk of harm is acceptably low for low GC dose users [132].

However, there are other variables besides daily dose that can dictate the extent and severity of adverse events suffered by patients taking corticosteroids.

One factor is the duration of use, since many patients are on long-term GC treatment.

Another aspect of the toxicity of GCs is the cumulative dose. It is a parameter linked to the duration of use but includes also the short-term increments of the daily dose needed when a relapse of the underlying rheumatological disease occurs [133]

Since most randomized trials are too short, we have no valuable information of the risks of long treatment periods. Despite that, a retrospective analysis on patients on low-dose GC treatment of 5 years demonstrated that these patients were prone to a significantly higher prevalence of fragility fractures, arterial hypertension and myocardial infarctions increasing with the duration of treatment [134]. In particular, osteoporosis is a common and severe adverse effect of glucocorticoid excess that occurs even in patients treated with a protracted therapy with very low GC doses. A UK database stated an increase of clinical vertebral fractures by 55% for a prednisone dose of <2.5 mg/day [135]. As a result, osteoporosis represents one of the major limitations to long-term glucocorticoid therapy. The highest rate of bone loss occurs within the first 3–6 months of GC treatment, and a slower decline continues with persistent use. Also high cumulative GC doses (>1 g of prednisone) were shown to increase the risk of fracture, particularly vertebral fracture due to the greater effects of GCs on trabecular bone than on cortical bone [136][137].

Recently, a study by George et al. also found that infection risk in patients treated with daily doses of prednisone 5 mg or less per day was significantly higher [138].

Moreover, as previously described, GC treatment is considered an independent factor for a shortened life expectancy, which is rarely mentioned in overviews over GC toxicity. In particular low-moderate daily dose had a hazard ratio of 2.22 for premature death in a study by Listing et al. [139]. Another study found a total cumulative dose of 40 g as the threshold for increased mortality [140]. That corresponds to 5 mg/day for 21 years or 7.5 mg/day over 14.5 years. Such time periods can easily be reached in the treatment of rheumatological diseases like rheumatoid arthritis.

As described in the last paragraph, after GC withdrawal even patients receiving low doses of GCs may experience AI. If fact, a randomized placebo-controlled withdrawal clinical trial by Pincus T. in patients with rheumatoid arthritis found that in patients treated with prednisone doses around 3 mg/day, it was rarely possible to withdraw the drug [141].

Another critical factor for the development of AI and Cushing's syndrome is cumulative dose. The above-cited study by Mebrahtu et al. reported that the risk of developing AI or Cushing's syndrome for patients with a cumulative dose of 1 g in the previous year was more than doubled $\frac{[125]}{}$.

To conclude, since there is not a safe GC therapy regimen, it seems advisable for physicians to vigilate for glucocorticoid-related adverse events and to counsel patients about possible risks, even among low-dose long-term users.

3.4. Follow up of GC Therapy

Although it is essential to inform patients about the possible adverse effects of chronic GC therapy in order to detect those dysfunctions at the earliest stage possible, prescribers should also estimate their risk of developing a specific GC-induced condition. For example, as described before, GC-induced osteoporosis is more frequent among high GC dose users but also among people with low body weight, low bone mineral density, previous fractures, family history of osteoporosis and among females [95]. Hence, it would be reasonable to perform a bone density scan before starting GC therapy and repeat it basing on the individual risk of the patient. That should also be the basis for the prescription of calcium and vitamin D supplementation, bisphosphonates or other drugs [132][142]. Another example is the need to examine pre-existing hyperglycaemia or diabetes mellitus, schedule tailored blood glucose and glycated haemoglobin tests and where necessary give advice for treatment [143]. It is also important to plan follow-up visits to allow patients to ask questions, to inform physicians about the possible adverse events and for doctors to perform an accurate physical examination to check dysfunctions related to GC therapy (cushingoid appearance, skin lesions, high blood pressure, etc.).

4. Conclusions

Rheumatological and thyroid autoimmune diseases are strongly related and the evidences derive from studies focused on prevalence, genetic and environmental aspects. However, those studies lack of homogeneity and there are probably multiple reasons.

First, systemic rheumatological diseases are heterogeneous and have different physiopathological mechanisms. Second, the prevalence of those diseases is much lower than thyroid diseases' so it is difficult to link them with precision.

In addition, many studies have only focused on patients with an overt thyroid dysfunction so it is reasonable to think that the correlation between them has been underestimated. This is because there is a wide subset of autoimmune thyroid diseases without thyroid function impairment but with typical ultrasound features like a pseudo-nodular pattern, gland hypoechogenicity and heterogeneous echotexture.

Moreover, the design of the majority of the studies available did not include a control group.

To conclude, it is reasonable to consider both a morphologic and functional evaluation of the thyroid gland in all patients affected by rheumatological diseases in order to identify promptly a possible related autoimmune thyroid disease.

The above-described iatrogenic Cushing's syndrome and glucocorticosteroid's withdrawal syndrome related to protracted GCs treatment are still very frequent in rheumatologic disorders despite the rising role of biologic drugs like monoclonal antibodies. In these patients, it is essential to know the importance of prescribing a correct regimen of glucocorticoids' withdrawal to prevent hypocorticosurrenalism and the related symptoms. Moreover, the same attention should be payed to administer GCs at the lowest possible dose to avoid the multi-systemic dysfunctions due to the iatrogenic hypercortisolism without provoking the relapse of the underlying rheumatologic disease.

To sum up, our paper highlights that it is necessary to acknowledge the importance of thyroid diseases as other endocrine disorders in autoimmune rheumatological diseases. Hence, it seems advisable for new studies to focus on treatments for autoimmune rheumatological diseases based on rebalancing the immune system dysfunction in order to avoid or prevent endocrine disorders.

References

- 1. Goldblatt, F.; O'Neill, S.G. Clinical aspects of autoimmune rheumatic diseases. Lancet 2013, 382, 797-808.
- AMRER. Onlus Malattie reumatiche, primo report sull'incidenza delle esenzioni per malattia. Quotid. Sanità 2013, 1, 3–7.
- 3. Safiri, S.; Kolahi, A.A.; Hoy, D.; Smith, E.; Bettampadi, D.; Mansournia, M.A.; Almasi-Hashiani, A.; Ashrafi-Asgarabad, A.; Moradi-Lakeh, M.; Qorbani, M.; et al. Global, regional and national burden of rheumatoid arthritis 1990–2017: A syst ematic analysis of the Global Burden of Disease study 2017. Ann. Rheum. Dis. 2019, 78, 11.
- 4. WHO.int. Available online: (accessed on 11 February 2021).
- 5. Conigliaro, P.; D'Antonio, A.; Pinto, S.; Chimenti, M.S.; Triggianese, P.; Rotondi, M.; Perricone, R. Autoimmune thyroid disorders and rheumatoid arthritis: A bidirectional interplay. Autoimmun. Rev. 2020, 19, 102529.
- 6. Ringold, D.A.; Nicoloff, J.T.; Kesler, M.; Davis, H.; Hamilton, A.; Mack, T. Further evidence for a strong genetic influence on the development of autoimmune thyroid diseases: The California twin study. Thyroid 2002, 12, 647–653.
- 7. Torfs, C.P. Genetic interrelationship between insulin-dependent diabetes mellitus, the autoimmune thyroid diseases and rheumatoid arthritis. Am. J. Hum. Genet. 1986, 38, 170–187.
- 8. Anaya, J.M. Common mechanisms of autoimmune diseases (the autoimmune tautology). Autoimmun. Rev. 2012, 11, 7 81–784.
- 9. Zaletel, K.; Gaberšček, S. Hashimoto's Thyroiditis: From Genes to the Disease. Curr. Genom. 2011, 12, 576-588.
- 10. McIver, B.; Morris, J.C. The pathogenesis of Graves' disease. Endocrinol. Metab. Clin. N. Am. 1998, 27, 73-89.
- 11. Cho, J.H.; Feldman, M. Heterogeneity of autoimmune diseases: Pathophysiologic insights from genetics and implications for new therapies. Nat. Med. 2015, 21, 730–738.
- 12. Medlineplus.gov. Available online: (accessed on 11 February 2021).
- 13. Mangalam, A.K.; Taneja, V.; David, C.S. HLA Class II Molecules Influence Susceptibility vs Protection in Inflammatory D iseases by Determining the Cytokine Profile. J. Immunol. 2013, 190, 513–519.
- 14. Miyadera, H.; Tokunaga, K. Associations of human leukocyte antigens with autoimmune diseases: Challenges in identifying the mechanism. J. Hum. Genet. 2015, 60, 697–702.
- 15. Li, C.W.; Osman, R.; Menconi, F.; Concepcion, E.S.; Tomer, Y. Flexible peptide recognition by HLA-DR triggers specific autoimmune T-cell responses in autoimmune thyroiditis and diabetes. J. Immun. 2017, 76, 1–9.
- 16. Criswell, L.A.; Pfeiffer, K.A.; Lum, R.F.; Gonzales, B.; Novitzke, J.; Kern, M.; Moser, K.L.; Begovich, A.B.; Carlton, V.E.; Li, W.; et al. Analysis of families in the Multiple Autoimmune Disease Genetic Consortium (MADGC) Collection: The PT

- PN22 620W Allele Associates with Multiple Autoimmune Phenothypes. Am. J. Hum. Gen. 2005, 76, 561-571.
- 17. Zhebrun, D.; Kudryashova, Y.; Babenko, A.; Maslyansky, A.; Kunitskaya, N.; Popcova, D.; Klushina, A.; Grineva, E.; Ko stareva, A.; Shlyakhto, E. Association of PTPN22 1858T/T genotype with type 1 diabetes, Graves' disease but not with rheumatoid arthritis in Russian population. Aging 2011, 3, 368–373.
- 18. Mori, M.; Yamada, R.; Kobayashi, K.; Kawaida, R.; Yamamoto, K. Ethnic differences in allele frequency of autoimmune-disease-associated SNPs. J. Hum. Genet. 2005, 50, 264–266.
- 19. Tai, X.; Van Laethem, F.; Pobezinsky, L.; Guinter, T.; Sharrow, S.O.; Adams, A.; Granger, L.; Kruhlak, M.; Lindsten, T.; T hompson, C.B. Basis of CTLA-4 function in regulatory and conventional CD4+ T cells. Blood 2012, 119, 5155–5163.
- 20. Ueda, H.; Howson, J.M.; Esposito, L.; Heward, J.; Chamberlain, G.; Rainbow, D.B.; Hunter, K.M.; Smith, A.N.; Di Geno va, G.; Herr, M.H.; et al. Association of the T-cell regulatory gene CTLA4 with susceptibility to autoimmune disease. Nat ure 2003, 423, 506–511.
- 21. Cooper, J.D.; Simmonds, M.J.; Walker, N.M.; Burren, O.; Brand, O.J.; Guo, H.; Wallace, C.; Stevens, H.; Coleman, G. Seven newly identified loci for autoimmune thyroid disease. Hum. Mol. Genet. 2012, 21, 5202–5208.
- 22. Okada, Y.; Wu, D.; Trynka, G.; Raj, T.; Terao, C.; Ikari, K.; Kochi, Y.; Ohmura, K.; Suzuki, A.; Yoshida, S. Genetics of rhe umatoid arthritis contributes to biology and drug discovery. Nature 2014, 506, 376–381.
- 23. Girotra, M.; Hansen, A.; Farooki, A.; Byun, D.J.; Min, L.; Creelan, B.C.; Callahan, M.K.; Atkins, M.B.; Sharon, E.; Antoni a, S.J.; et al. The current understanding of the endocrine effects from immune checkpoint inhibitors and recommendati ons for management. JNCI Cancer Spectr. 2018, 2, 3.
- 24. Cappelli, L.C.; Gutierrez, A.K.; Baer, A.N.; Albayda, J.; Manno, R.L.; Haque, U.; Lipson, E.J.; Bleich, K.B.; Shah, A.A.; N aidoo, J.; et al. Inflammatory arthritis and sicca syndrome induced by nivolumab and ipilimumab. Ann. Rheum. Dis. 201 7, 76, 43–50.
- 25. Maniu, C.; Kobe, C.; Schlaak, M.; Mauch, C.; Eming, S.A. Polymyalgia rheumatica occurring during treatment with ipili mumab. Eur. J. Dermatol. 2016, 26, 513–514.
- 26. Jaberg-Bentele, N.F.; Kunz, M.; Abuhammad, S.; Dummer, R. Flare up of rheumatoid arthritis by anti CTLA-4 antibody but not with anti PD-1 therapy in a patient with metastatic melanoma. Case Rep. Dermatol. 2017, 9, 65–68.
- 27. Kuehn, H.S.; Ouyang, W.; Lo, B.; Deenick, E.K.; Niemela, J.E.; Avery, D.T.; Schickel, J.N.; Tran, D.Q.; Stoddard, J.; Zh ang, Y.; et al. Immune dysregulation in human subjects with heterozygous germline mutation in CTLA4. Science 2014, 345, 1623–1627.
- 28. Zeissig, S.; Petersen, B.S.; Tomczak, M.; Melum, E.; Huc-Claustre, E.; Dougan, S.K.; Laerdahl, J.K.; Stade, B.; Forster, M.; Schreiber, S.; et al. Early-Onset Crohn's disease and autoimmunity associated with a variant in CTLA-4. Gut 2015, 64, 1889–1897.
- 29. Sun, D.; Heimall, J. Disorders of CTLA-4 expression, how they lead to CVID and dysregulated immune responses. Cur r. Opin. Allergy Clin. Immunol. 2019, 19, 578–585.
- 30. Buckner, J.H. Mechanisms of impaired regulation by CD4+CD25+FOXP3+ regulatory T cells in human autoimmune dis eases. Nat. Rev. Immunol. 2010, 10, 849–859.
- 31. Theofilopoulos, A.N.; Kono, D.H.; Baccala, R. The multiple pathways to autoimmunity. Nat. Immunol. 2017, 18, 716–72 4.
- 32. Inoue, Y. Aploinsufficiency of A20 causes autoinflammatory and autoimmune disorders. J. Allergy Clin. Immunol. 2018, 141, 1485–1488.
- 33. Chen, J.; Li, Y.; Tian, Y.; Huang, C.; Li, D.; Zhong, Q.; Ma, X. Interaction between microbes and host intestinal health: M odulation by dietary nutrients and gut-brain-endocrine-immune axis. Curr. Protein Pept. Sci. 2015, 16, 592–603.
- 34. Cavaglieri, C.R.; Nishiyama, A.; Fernandes, L.C.; Curi, R.; Miles, E.A.; Calder, P.C. Differential effects of short-chain fatt y acids on proliferation and production of pro- and anti-inflammatory cytokines by cultured lymphocytes. Life Sci. 2003, 73, 1683–1690.
- 35. Johnson-Henry, K.C.; Pinnell, L.J.; Waskow, A.M.; Irrazabal, T.; Martin, A.; Hausner, M.; Sherman, P.M. Short-Chain fru cto-oligosaccharide and inulin modulate inflammatory responses and microbial communities in Caco2-bbe cells and in a mouse model of intestinal injury. J. Nutr. 2014, 144, 1725–1733.
- 36. Xu, H.; Liu, M.; Cao, J.; Li, X.; Fan, D.; Xia, Y.; Lu, X.; Li, J.; Ju, D.; Zhao, H. The dynamic interplay between the gut mic robiota and autoimmune diseases. J. Immunol. Res. 2019, 2019, 7546047.
- 37. Liang, B.; Mamula, M.J. Molecular mimicry and the role of B lymphocytes in the processing of autoantigens. Cell. Mol. Life Sci. 2000, 57, 561–568.
- 38. Lanzavecchia, A. How can cryptic epitopes trigger autoimmunity? J. Exp. Med. 1995, 181, 1945–1948.

- 39. Shor, D.B.; Orbach, H.; Boaz, M.; Altman, A.; Anaya, J.M.; Bizzaro, N.; Tincani, A.; Cervera, R.; Espinosa, G.; Stojanovi ch, L.; et al. Gastrointestinal-Associated autoantibodies in different autoimmune diseases. Am. J. Clin. Exp. Immunol. 2 012, 1, 49–55.
- 40. Zamani, B.; Golkar, H.R.; Farshbaf, S.; Emadi-Baygi, M.; Tajabadi-Ebrahimi, M.; Jafari, P.; Akhavan, R.; Taghizadeh, M.; Memarzadeh, M.R.; Asemi, Z. Clinical and metabolic response to probiotic supplementation in patients with rheuma toid arthritis: A randomized, double-blind, placebo-controlled trial. Int. J. Rheum. Dis. 2016, 19, 869–879.
- 41. López, P.; De Paz, B.; Rodríguez-Carrio, J.; Hevia, A.; Sánchez, B.; Margolles, A.; Suárez, A. Th17 responses and natural IgM antibodies are related to gut microbiota composition in systemic lupus erythematosus patients. Sci. Rep. 2016, 6, 24072.
- 42. Zeng, J.; Peng, L.; Zheng, W.; Huang, F.; Zhang, N.; Wu, D.; Yang, Y. Fecal microbiota transplantation for rheumatoid a rthritis: A case report. Clin. Case Rep. 2020, 9, 1–4.
- 43. Mele, C.; Caputo, M.; Bisceglia, A.; Samà, M.T.; Zavattaro, M.; Aimaretti, G.; Pagano, L.; Prodam, F.; Marzullo, P. Immu nomodulatory Effects of Vitamin D in Thyroid Diseases. Nutrients 2020, 12, 1444.
- 44. Muscogiuri, G.; Mitri, J.; Mathieu, C.; Badenhoop, K.; Tamer, G.; Orio, F.; Mezza, T.; Vieth, R.; Colao, A.; Pittas, A. Mech anisms in endocrinology: Vitamin D as a potential contributor in endocrine health and disease. Eur. J. Endocrinol. 2014, 171, R101–R110.
- 45. Kim, D. The Role of Vitamin D in Thyroid Diseases. Int. J. Mol. Sci. 2017, 18, 1949.
- 46. Adorini, L.; Penna, G. Control of autoimmune diseases by the vitamin D endocrine system. Nat. Clin. Pract. Rheumatol. 2008, 4, 404–412.
- 47. Cutolo, M.; Plebani, M.; Shoenfeld, Y.; Adorini, L.; Tincani, A. Vitamin D endocrine system and the immune response in rheumatic diseases. Vitam. Horm. 2011, 86, 327–351.
- 48. Jin, D.; Wu, S.; Zhang, Y.G.; Lu, R.; Xia, Y.; Dong, H.; Sun, J. Lack of Vitamin D Receptor Causes Dysbiosis and Chang es the Functions of the Murine Intestinal Microbiome. Clin. Ther. 2015, 37, 996–1009.
- 49. Wu, S.; Liao, A.P.; Xia, Y.; Li, Y.C.; Li, J.D.; Sartor, R.B.; Sun, J. Vitamin D receptor negatively regulates bacterial-stimul ated NF-kappaB activity in intestine. Am. J. Pathol. 2010, 177, 686–697.
- 50. Jahani, R.; Fielding, K.A.; Chen, J.; Villa, C.R.; Castelli, L.M.; Ward, W.E.; Comelli, E.M. Low vitamin D status througho ut life results in an inflammatory prone status but does not alter bone mineral or strength in healthy 3-month-old CD-1 male mice. Mol. Nutr. Food Res. 2014, 58, 1491–1501.
- 51. Naderpoor, N.; Mousa, A.; Fernanda Gomez Arango, L.; Barrett, H.L.; Dekker Nitert, M.; de Courten, B. Effect of Vitami n D Supplementation on Faecal Microbiota: A Randomised Clinical Trial. Nutrients 2019, 11, 2888.
- 52. Chassaing, B.; Vijay-Kumar, M.; Gewirtz, A.T. How diet can impact gut microbiota to promote or endanger health. Curr. Opin. Gastroenterol. 2017, 33, 417–421.
- 53. Suez, J.; Korem, T.; Zeevi, D.; Zilberman-Schapira, G.; Thaiss, C.A.; Maza, O.; Israeli, D.; Zmora, N.; Gilad, S.; Weinbe rger, A.; et al. Artificial sweeteners induce glucose intolerance by altering the gut microbiota. Nature 2014, 514, 181–18 6.
- 54. Chassaing, B.; Koren, O.; Goodrich, J.K.; Poole, A.C.; Srinivasan, S.; Ley, R.E.; Gewirtz, A.T. Dietary emulsifiers impact the mouse gut microbiota promoting colitis and metabolic syndrome. Nature 2015, 519, 92–96.
- 55. Go, W.Y.; Liu, X.; Roti, M.A.; Liu, F.; Ho, S.N. NFAT5/TonEBP mutant mice define osmotic stress as a critical feature of t he lymphoid microenvironment. Proc. Natl. Acad. Sci. USA 2004, 101, 10673–10678.
- 56. Shapiro, L.; Dinarello, C.A. Osmotic regulation of cytokine synthesis in vitro. Proc. Natl. Acad. Sci. USA 1995, 92, 1223 0–12234.
- 57. Wu, C.; Yosef, N.; Thalhamer, T.; Zhu, C.; Xiao, S.; Kishi, Y.; Regev, A.; Kuchroo, V.K. Induction of pathogenic TH17 cell s by inducible salt-sensing kinase SGK1. Nature 2013, 496, 513–517.
- 58. Salgado, E.; Bes-Rastrollo, M.; de Irala, J.; Carmona, L.; Gomez-Reino, J.J. High sodium intake is associated with self-reported rheumatoid arthritis: A cross sectional and case control analysis within the SUN cohort. Medicine 2015, 94, e9 24.
- 59. Marouen, S.; du Cailar, G.; Audo, R.; Lukas, C.; Vial, G.; Tournadre, A.; Barrat, E.; Ribstein, J.; Combe, B.; Morel, J.; et al. Sodium excretion is higher in patients with rheumatoid arthritis than in matched controls. PLoS ONE 2017, 12, e018 6157.
- 60. Sundstrom, B.; Johansson, I.; Rantapää-Dahlqvist, S. Interaction between dietay sodium and smoking increases the ris k for rheumatoid arthritis: Results from a nested case-control study. Rheumatology 2015, 54, 487–493.

- 61. Pfau, J.C.; Sentissi, J.J.; Li, S.A.; Calderon-Garciduenas, L.; Brown, J.M.; Blake, D.J. Asbestos-induced autoimmunity in C57BL/6 mice. J. Immunotoxicol. 2008, 5, 129–137.
- 62. Brown, J.M.; Archer, A.J.; Pfau, J.C.; Holian, A. Silica accelerated systemic autoimmune disease in lupus-prone New Z ealand mixed mixed. Clin. Exp. Immunol. 2003, 131, 415–421.
- 63. Pfau, J.C.; Brown, J.M.; Holian, A. Silica-Exposed mice generate autoantibodies to apoptotic cells. Toxicology 2004, 19 5, 167–176.
- 64. Hamilton, R.F.; Thakur, S.A.; Holian, A. Silica binding and toxicity in alveolar macrophages. Free Radical Biol. Med. 200 8, 44, 1246–1258.
- 65. Rasooly, L.; Burek, C.L.; Rose, N.R. Iodine-Induced autoimmune thyroiditis in NOD-H-2h4 mice. Clin. Immuno opathol. 1996, 81, 287–292.
- 66. Bournaud, C.; Orgiazzi, J.J. Iodine excess and thyroid autoimmunity. J. Endocrinol. Invest. 2003, 26, 49-56.
- 67. Zois, C.; Stavrou, I.; Svarna, E.; Seferiadis, K.; Tsatsoulis, A. Natural course of autoimmune thyroiditis after elimination of iodine deficiency in northwestern Greece. Thyroid 2006, 16, 289–293.
- 68. Benvenga, S.; Vigo, M.T.; Metro, D.; Granese, R.; Vita, R.; Le Donne, M. Type of fish consumed and thyroid autoimmun ity in pregnancy and postpartum. Endocrine 2016, 52, 120–129.
- 69. Tajtakova, M.; Semanová, Z.; Tomková, Z.; Szökeová, E.; Majoroš, J.; Rádiková, Ž.; Šeböková, E.; Klimeš, I.; Langer, P. Increased thyroid volume and frequency of thyroid disorders signs in schoolchildren from nitrate polluted area. Chem osphere 2006, 62, 559–564.
- 70. Elter, E.; Wagner, M.; Buchenauer, L.; Bauer, M.; Polte, T. Phthalate exposure during the prenatal and lactation period increases the susceptibility to rheumatoid arthritis in mice. Front. Immunol. 2020, 11, 550.
- 71. Ghassabian, A.; Salmon, J.; Karthikraj, R.; Kahn, L.; Mehta-Lee, S.; Buyon, J.; Trasande, L. Phthalate exposure in pregnant women with systemic lupus erythematosus (SLE). Environ. Epidemiol. 2019, 3, 134.
- 72. Pryor, W.A.; Stone, K.; Zang, L.Y.; Bermúdez, E. Fractionation of aqueous cigarette tar extracts: Fractions that contain t he tar radical cause DNA damage. Chem. Res. Toxicol. 1998, 11, 441–448.
- 73. Robbins, C.S.; Dawe, D.E.; Goncharova, S.I.; Pouladi, M.A.; Drannik, A.G.; Swirski, F.K.; Cox, G.; Stampfli, M.R. Cigar ette smoke decreases pulmonary dendritic cells and impacts antiviral immune responsiveness. Am. J. Respir. Cell Mol. Biol. 2004, 30, 202–211.
- 74. Moszczynski, P.; Żabiński, Z.; Moszczyński, P., Jr.; Rutowski, J.; Słowiński, S.; Tabarowski, Z. Immunological findings in cigarette smokers. Toxicol. Lett. 2001, 118, 121–127.
- 75. Hutchinson, D.; Shepstone, L.; Moots, R.; Lear, J.T.; Lynch, M.P. Heavy cigarette smoking is strongly associated with rh eumatoid arthritis (RA), particularly in patients without a family history of RA. Ann. Rheum. Dis. 2001, 60, 223–227.
- 76. Chang, K.; Yang, S.M.; Kim, S.H.; Han, K.H.; Park, S.J.; Shin, J.I. Smoking and rheumatoid arthritis. Int. J. Mol. Sci. 20 14, 15, 22279–22295.
- 77. Criswell, L.A.; Merlino, L.A.; Cerhan, J.R.; Mikuls, T.R.; Mudano, A.S.; Burma, M.; Folsom, A.R.; Saag, K.G. Cigarette s moking and the risk of rheumatoid arthritis among postmenopausal women: Results from the Iowa Women's Health Study. Am. J. Med. 2002, 112, 465–471.
- 78. Majka, D.S.; Kim, D.J.; Peerzada, J.; Lockman, S.; Nobles-Knight, D.; Petri, M.; Karlson, E.W. Cigarette smoking and the erisk of systemic lupus erythematosus and rheumatoid arthritis. Ann. Rheum. Dis. 2006, 65, 561–563.
- 79. Barbhaiya, M.; Tedeschi, S.K.; Lu, B.; Malspeis, S.; Kreps, D.; Sparks, J.A.; Karlson, E.W.; Costenbader, K.H. Cigarette smoking and the risk of systemic lupus erythematosus, overall and by anti-double stranded DNA antibody subtype, in the Nurses' Health Study cohorts. Ann. Rheum. Dis. 2018, 77, 196–202.
- 80. Yan Chua, M.H.; Ng, I.A.T.; Mike, W.C.; Mak, A. Association between cigarette smoking and systemic lupus erythemato sus- an updated multivariate Bayesian metaanalysis. J. Rheumatol. 2019, 47, 1514–1521.
- 81. Holm, I.A.; Manson, J.E.; Michels, K.B.; Alexander, E.K.; Willett, W.C.; Utiger, R.D. Smoking and Other Lifestyle Factors and the Risk of Graves' Hyperthyroidism. Arch. Intern. Med. 2005, 165, 1606–1611.
- 82. Hegediüs, L.; Brix, T.H.; Vestergaard, P. Relationship between cigarette smoking and Graves' ophthalmopathy. J. Endo crinol. Invest. 2004, 27, 265–271.
- 83. Yuksel, N.; Yaman, D.; Pasaoglu, O.T.; Pasaoglu, H. The Effect of Smoking on Mitochondrial Biogenesis in Patients wit h Graves Ophthalmopathy. Ophthalmic. Plast. Reconstr. Surg. 2020, 36, 172–177.
- 84. Barzilai, O.; Sherer, Y.; Ram, M.; Izhaky, D.; Anaya, J.M.; Shoenfeld, Y. Epstein–Barr Virus and Cytomegalovirus in Aut oimmune Diseases. Ann. N. Y. Acad. Sci. 2007, 1108, 567–577.

- 85. Maya, R.; Anaya, J.M.; Barzilai, O.; Izhaky, D.; Katz, B.S.P.; Blank, M.; Shoenfeld, Y. The putative protective role of hep atitis B virus (HBV) infection from autoimmune disorders. Autoimmun. Rev. 2008, 7, 621–625.
- 86. Zahiri Yeganeh, S. Bacteriological and Molecular Assessment of Staphylococcal Enterotoxin E in the Blood of Patients with Rheumatoid Arthritis. Jundishapur J. Microbiol. 2015, 8, e59811.
- 87. Carter, J.K.; Smith, R.E. Rapid induction of hypothyroidism by an avian leukosis virus. Infect. Immun. 1983, 40, 795–80 5.
- 88. Davies, T.F. Infection and Autoimmune Thyroid Disease. J. Clin. Endocrinol. Metab. 2008, 93, 674-676.
- 89. Kivity, S.; Agmon-Levin, N.; Blank, M.; Shoenfeld, Y. Infections and autoimmunity–friends or foes? Trends Immunol. 200 9, 30, 409–414.
- 90. Rioux, J.D. Paths to understanding the genetic basis of autoimmune disease. Nature 2005, 435, 584-589.
- 91. Guilherme, L.; Kalil, J.; Cunningham, M. Molecular mimicry in the autoimmune pathogenesis of rheumatic heart diseas e. Autoimmunity 2006, 39, 31–39.
- 92. Scheinman, R.I.; Cogswell, P.C.; Lofquist, A.K.; Baldwin, A.S. Role of transcriptional activation of I kappa B alpha in me diation of immunosuppression by glucocorticoids. Science 1995, 270, 283.
- 93. Auphan, N.; DiDonato, J.A.; Rosette, C.; Helmberg, A.; Karin, M. Immunosuppression by glucocorticoids: Inhibition of N F-kappa B activity through induction of I kappa B synthesis. Science 1995, 270, 286.
- 94. Rhen, T.; Cidlowski, J.A. Antiinflammatory action of glucocorticoids—New mechanisms for old drugs. N. Engl. J. Med. 2 005, 353, 1711.
- 95. Van Staa, T.P.; Leufkens, H.G.; Abenhaim, L.; Begaud, B.; Zhang, B.; Cooper, C. Use of oral corticosteroids in the Unite d Kingdom. QJM 2000, 93, 105–111.
- 96. Zhang, X.W.; Li, Y.; Wang, Z.L.; Li, P. Glucocorticoid receptor subunit gene expression in thyroid gland and adenomas. Acta Oncol. 2006, 45, 1073–1078.
- 97. Kühn, E.R.; Geris, K.L.; van der Geyten, S.; Mol, K.A.; Darras, V.M. Inhibition and activation of the thyroidal axis by the adrenal axis in vertebrates. Comp. Biochem. Physiol. A Mol. Integr. Physiol. 1998, 120, 169–174.
- 98. Williams, D.E.; Chopra, I.J.; Orgiazzi, J.; Solomon, D.H. Acute effects of corticosteroids on thyroid activity in Graves' dis ease. J. Clin. Endocrinol. Metab. 1975, 41, 354–361.
- 99. Carroll, R.; Matfin, G. Endocrine and metabolic emergencies: Thyroid storm. Ther. Adv. Endocrinol. Metab. 2010, 1, 139 –145.
- 100. Hopkins, R.L.; Leinung, M.C. Exogenous Cushing's syndrome and glucocorticoid withdrawal. Endocrinol. Metab. Clin. N. Am. 2005, 34, 371–384.
- 101. Paragliola, R.M.; Papi, G.; Pontecorvi, A.; Corsello, S.M. Treatment with Synthetic Glucocorticoids and the Hypothalam us-Pituitary-Adrenal Axis. Int. J. Mol. Sci. 2017, 18, 2201.
- 102. Ferraù, F.; Korbonits, M. Metabolic comorbidities in Cushing's syndrome. Eur. J. Endocrinol. 2015, 173, 133-157.
- 103. Ferris, H.A.; Kahn, C.R. New mechanisms of glucocorticoid-induced insulin resistance: Make no bones about it. J. Clin. Invest. 2012, 122, 3854–3857.
- 104. Arnaldi, G.; Mancini, T.; Polenta, B.; Boscaro, M. Cardiovascular risk in Cushing's syndrome. Pituitary 2004, 7, 253–25 6.
- 105. Dal Bo Zanon, R.; Fornasiero, L.; Boscaro, M.; Cappellato, G.; Fabris, F.; Girolami, A. Increased factor VIII associated a ctivities in Cushing's syndrome: A probable hypercoagulable state. Thromb. Haemostas. 1982, 47, 116–117.
- 106. Canalis, E. Mechanisms of glucocorticoid-induced osteoporosis. Curr. Opin. Rheumatol. 2003, 15, 454–457.
- 107. Chaudhry, H.S.; Singh, G. Cushing Syndrome. Available online: (accessed on 29 June 2021).
- 108. Schakman, O.; Kalista, S.; Barbé, C.; Loumaye, A.; Thissen, J.P. Glucocorticoid-induced skeletal muscle atrophy. Int. J. Biochem. Cell Biol. 2013, 45, 2163–2172.
- 109. Conn, H.O.; Poynard, T. Corticosteroids and peptic ulcer: Meta-Analysis of adverse events during steroid therapy. J. Int ern. Med. 1994, 236, 619–632.
- 110. Conn, H.O.; Blitzer, B.L. Nonassociation of adrenocorticosteroid therapy and peptic ulcer. N. Engl. J. Med. 1976, 294, 4
- 111. Guslandi, M. Steroid ulcers: Any news? World J. Gastrointest. Pharmacol. Ther. 2013, 4, 39-40.

- 112. Dorlo, T.P.; Jager, N.G.; Beijnen, J.H.; Schellens, J.H. Concomitant use of proton pump inhibitors and systemic corticos teroids. Ned. Tijdschr. Geneeskd. 2013, 157, A5540.
- 113. Black, R.L.; Oglesby, R.B.; von Sallmann, L.; Bunim, J.J. Posterior subcapsular cataracts induced by corticosteroids in patients with rheumatoid arthritis. JAMA 1960, 174, 150–155.
- 114. Van Venrooj, W.J.; Groeneveld, A.A.; Bloemendal, H.; Benedetti, E.L. Cultured calf lens epithelium. The effect of dexam ethasone. Exp. Eye Res. 1974, 18, 527–536.
- 115. Alward, W.L. The genetics of open-angle glaucoma: The story of GLC1A and myocilin. Eye 2000, 14, 429-436.
- 116. Polansky, J.R.; Nguyen, T.D. The TIGR gene, pathogenic mechanisms, and other recent advances in glaucoma genetic s. Curr. Opin. Ophthalmol. 1998, 9, 15–23.
- 117. Lo, W.R.; Rowlette, L.L.; Caballero, M.; Yang, P.; Hernandez, M.R.; Borrás, T. Tissue differential microarray analysis of dexamethasone induction reveals potential mechanisms of steroid glaucoma. Invest. Ophthalmol. Vis. Sci. 2003, 44, 47 3–485
- 118. Davidovici, B.B.; Orion, E.; Wolf, R. Cutaneous manifestations of pituitary gland diseases. Clin Dermatol. 2008, 26, 288 –295.
- 119. Lewis, D.A.; Smith, R.E. Steroid-induced psychiatric syndromes: A report of 14 cases and a review of the literature. J. A ffect. Disord. 1983, 5, 319–332.
- 120. Warrington, T.P.; Bostwick, J.M. Psychiatric adverse effect of corticosteroids. Mayo Clin. Proc. 2006, 81, 1361–1367.
- 121. Nguyen, D.M.; Yassa, M.A.; Tustison, N.J.; Roberts, J.M.; Kulikova, A.; Nakamura, A.; Ivleva, E.I.; Van Enkevort, E.; Br own, E.S. The Relationship between Cumulative Exogenous Corticosteroid Exposure and Volumes of Hippocampal Su bfields and Surrounding Structures. J. Clin. Psychopharmacol. 2019, 39, 653–657.
- 122. Fareau, G.G.; Vassilopoulou-Sellin, R. Hypercortisolemia and infection. Infect. Dis. Clin. N. Am. 2007, 21, 639-657.
- 123. Bakker, R.C.; Gallas, P.R.J.; Romijn, J.A.; Wiersinga, W.M. Cushing's syndrome complicated by multiple opportunistic i nfections. J. Endocrinol. Invest. 1998, 21, 329–333.
- 124. Broersen, L.H.; Pereira, A.M.; Jørgensen, J.O.L.; Dekkers, O.M. Adrenal insufficiency in corticosteroids use: Systematic review and meta-analysis. J. Clin. Endocrinol. Metab. 2015, 100, 2171–2180.
- 125. Mebrahtu, T.F.; Morgan, A.W.; Keeley, A.; Baxter, P.D.; Stewart, P.M.; Pujades-Rodriguez, M. Dose dependency of iatro genic glucocorticoid excess and adrenal insufficiency and mortality: A cohort study in England. J. Clin. Endocrinol. Meta b. 2019, 104, 3757–3767.
- 126. Amatruda, T.T.; Hurst, A.M.; D'Esopo, N.D. Certain Endocrine and Metabolic Facets of the Steroid Withdrawal Syndrom e. J. Clin. Endocrinol. Metab. 1965, 25, 1207–1217.
- 127. Amatruda, T.T.; Hollingsworth, D.R.; D'esopo, N.D.; Upton, G.V.; Bondy, P.K. A study of the mechanism of the steroid wi thdrawal syndrome. Evidence for integrity of the hypothalamic-pituitary-adrenal system. J. Clin. Endocrinol. Metab. 196 0, 20, 339–354.
- 128. Hochberg, Z.E.; Pacak, K.; Chrousos, G.P. Endocrine withdrawal syndromes. Endocr. Rev. 2003, 24, 523–538.
- 129. Dorn, L.D.; Burgess, E.S.; Friedman, T.C.; Dubbert, B.; Gold, P.W.; Chrousos, G.P. The longitudinal course of psychopa thology in Cushing's syndrome after correction of hypercortisolism. J. Clin. Endocrinol. Metab. 1997, 82, 912–919.
- 130. Giordano, R.; Guaraldi, F.; Mazzoli, M.; Ghigo, E. Do glucocorticoids induce addiction in humans? J. Endocrinol. Invest. 2017, 40, 881–883.
- 131. Vittecoq, O.; Desouches, S.; Kozyreff, M.; Nicolau, J.; Pouplin, S.; Rottenberg, P.; Sens, N.; Lequerre, T.; Avenel, G. Re lapse in rheumatoid arthritis patients undergoing dose reduction and withdrawal of biologics: Are predictable factors mo re relevant than predictive parameters? An observational prospective real-life study. BMJ Open 2019, 9, e031467.
- 132. Strehl, C.; Bijlsma, J.W.; de Wit, M.; Boers, M.; Caeyers, N.; Cutolo, M.; Dasgupta, B.; Dixon, W.G.; Geenen, R.; Huizin ga, T.W.; et al. Defining conditions where long-term glucocorticoid treatment has an acceptably low level of harm to facil itate implementation of existing recommendations: Viewpoints from an EULAR task force. Ann. Rheum. Dis. 2016, 75, 952–957.
- 133. Thiele, K.; Buttgereit, F.; Huscher, D.; Zink, A.; German Collaborative Arthritis Centres. Current use of glucocorticoids in patients with rheumatoid arthritis in Germany. Arthritis Care Res. 2005, 53, 740–747.
- 134. Mazzantini, M.; Talarico, R.; Doveri, M.; Consensi, A.; Cazzato, M.; Bazzichi, L.; Bombardieri, S. Incident comorbidity a mong patients with rheumatoid arthritis treated or not with low-dose glucocorticoids: A retrospective study. J. Rheumato I. 2010, 37, 2232–2236.

- 135. Van Staa, T.P.; Leufkens, H.G.; Abenhaim, L.; Zhang, B.; Cooper, C. Use of oral corticosteroids and risk of fractures. J. Bone Miner. Res. 2000, 15, 993–1000.
- 136. Buckley, L.; Guyatt, G.; Fink, H.A.; Cannon, M.; Grossman, J.; Hansen, K.E.; Humphrey, M.B.; Lane, N.E.; Magrey, M.; Miller, M.; et al. American College of Rheumatology Guideline for the Prevention and Treatment of Glucocorticoid-Induc ed Osteoporosis. Arthritis Rheumatol. 2017, 69, 1521–1537.
- 137. Van Staa, T.P.; Geusens, P.; Pols, H.A.; de Laet, C.; Leufkens, H.G.; Cooper, C. A simple score for estimating the long-t erm risk of fracture in patients using oral glucocorticoids. QJM 2005, 98, 191–198.
- 138. George, M.D.; Baker, J.F.; Winthrop, K.; Hsu, J.Y.; Wu, Q.; Chen, L.; Xie, F.; Yun, H.; Curtis, J.R. Risk for Serious Infection with Low-Dose Glucocorticoids in Patients With Rheumatoid Arthritis: A Cohort Study. Ann. Intern. Med. 2020, 173, 8 70–878.
- 139. Listing, J.; Kekow, J.; Manger, B.; Burmester, G.R.; Pattloch, D.; Zink, A.; Strangfeld, A. Mortality in rheumatoid arthritis: The impact of disease activity, treatment with glucocorticoids, TNFα inhibitors and rituximab. Ann. Rheum. Dis. 2015, 7 4, 415–421.
- 140. Del Rincon, I.; Battaforano, D.F.; Restrepo, J.F.; Erikson, J.M.; Escalante, A. A glucocorticoid dose threshold associated with all-cause and cardiovascular mortality in rheumatoid arthritis. Arthritis Rheum. 2014, 66, 264–272.
- 141. Pincus, T. The clinical efficacy of 3 mg/day prednisone in patients with rheumatoid arthritis: Evidence from a randomize d, double-blind, placebo-controlled withdrawal clinical trial. Clin. Exp. Rheumatol. 2011, 29, S73–S76.
- 142. Teitelbaum, S.L.; Seton, M.P.; Saag, K.G. Should bisphosphonates be used for long-term treatment of glucocorticoid-in duced osteoporosis? Arthritis Rheum. 2011, 63, 325–328.
- 143. Suh, S.; Park, M.K. Glucocorticoid-Induced Diabetes Mellitus: An Important but Overlooked Problem. Endocrinol. Meta b. 2017, 32, 180–189.

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