# **Metabolic Phenotypes in Asthmatic Adults**

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Bronchial asthma is a chronic respiratory disease that belongs to all ages. Transforming involves chronic airway inflammation and symptoms of varying magnitude over time, which include dyspnea, chest tightness, and coughing. It has a high prevalence, high morbidity and considerable levels of mortality. According to the Global Initiative for Asthma (GINA), "Asthma is a heterogeneous disease with different underlying pathological processes. Recognizable groups of demographic, clinical and/or pathophysiological characteristics are called 'asthma phenotypes'. In fact, several studies show that various asthma subtypes may be reflected in external manifestations of the disease, which are designated as "phenotypes", and may involve clinical and inflammatory characteristics, among others. However,

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## 1. Overview

Bronchial asthma is a chronic disease that affects individuals of all ages. It has a high prevalence and is associated with high morbidity and considerable levels of mortality. However, asthma is not a single disease, and multiple subtypes or phenotypes (clinical, inflammatory or combinations thereof) can be detected, namely in aggregated clusters. Most studies have characterized asthma phenotypes and groups of phenotypes using mainly clinical and inflammatory parameters. These studies are important because they can have clinical and prognostic implications and can also help define personalized treatment approaches. In addition, several metabolomics studies have helped to better define the metabolic characteristics of asthma, using electronic noses or targeted and non-targeted approaches. In addition to discriminating between asthma and a healthy state, metabolomics can detect the metabolic signatures associated with some asthma subtypes, namely, eosinophilic and non-eosinophilic phenotypes or the obese asthma phenotype, and this can be very useful in application to the site of attendance. In addition, metabolomics also discriminates between asthma and other "phenotypes" of chronic obstructive airway diseases, such as chronic obstructive pulmonary disease (COPD) or Asthma-COPD Overlap (OAC). However, there are still several aspects that need to be further investigated in the context of asthma phenotypes in properly designed, homogeneous and multicentric studies, using appropriate tools and integrating metabolomics in a multilevel approach. namely, eosinophilic and non-eosinophilic phenotypes or the obese asthma phenotype, and this can be very useful in the application at the point of care. In addition, metabolomics also discriminates between asthma and other "phenotypes" of chronic obstructive airway diseases, such as chronic obstructive pulmonary disease (COPD) or Asthma-COPD Overlap (OAC). However, there are still several aspects that need to be further investigated in the context of asthma phenotypes in properly designed, homogeneous and multicentric studies, using appropriate tools and integrating metabolomics in a multilevel approach. namely, eosinophilic and non-eosinophilic phenotypes, or the obese asthma phenotype, and this can be very useful in point-of-care application. In addition, metabolomics also discriminates between asthma and other "phenotypes" of chronic obstructive airway diseases, such as chronic obstructive pulmonary disease (COPD) or Asthma-COPD Overlap (OAC). However, there are still several aspects that need to be further investigated in the context of asthma phenotypes in properly designed, homogeneous and multicentric studies, using appropriate tools and integrating metabolomics in a multilevel approach. such as chronic obstructive pulmonary disease (COPD) or asthma-COPD overlap (COC). However, there are still several aspects that need to be further investigated in the context of asthma phenotypes in properly designed, homogeneous and multicentric studies, using appropriate tools and integrating metabolomics in a multilevel approach. such as chronic obstructive pulmonary disease (COPD) or asthma-COPD overlap (COC). However, there are still several aspects that need to be further investigated in the context of asthma phenotypes in properly designed, homogeneous and multicentric studies, using appropriate tools and integrating metabolomics in a multilevel approach.

## 2. Bronchial asthma

Bronchial asthma is a chronic respiratory disease that affects individuals of all ages. It usually involves chronic airway inflammation and symptoms of variable magnitude over time, which include dyspnoea, chest tightness, and  $cough^{[1]}$ . It has a high prevalence, high morbidity and considerable levels of mortality<sup>[2]</sup>. According to the Global Initiative for Asthma (GINA), "Asthma is a heterogeneous disease with different underlying pathological processes. Recognizable groups of demographic, clinical and/or pathophysiological characteristics are often referred to as 'asthma phenotypes'<sup>[1]</sup>. In fact, several studies have shown that various asthma subtypes can be reflected in external manifestations of the disease, which are referred to as "phenotypes". ", and may involve clinical and inflammatory features, in addition to others<sup>[3]</sup>. However, since asthma phenotypes do not imply any specific underlying pathophysiological mechanisms, asthma can also be classified into subtypes known as "endotypes"<sup>[4]</sup>, which are based on specific pathophysiological mechanisms at cellular and molecular levels<sup>[5][G][7]</sup>.

The detection of biomarkers is necessary to obtain more robust definitions of asthma phenotypes or endotypes<sup>[8][9][10]</sup>. This further helps to classify patients and may allow for a more tailored therapeutic approach for each phenotype or endotype<sup>[11]</sup>. Although different types of biomarkers have been described, metabolic pathways also have components that are different between a healthy state and disease, and which may also be relevant as asthma biomarkers. Thus, the complete analysis of small molecules such as amino acids, lipids, organic acids and nucleotides through metabolomic studies carried out in different biological samples - exhaled air condensate (EBC), peripheral blood or urine - can be very important in the management of asthma in diagnosis, monitoring, personalized treatment and prognosis, but many issues still need to be addressed. In fact, more specifically, metabolomics-associated biomarkers can be very useful for understanding the pathophysiology of asthma, as well as several other aspects of the disease, including the prediction of exacerbation and response to treatment.

Metabolomics uses high-throughput analytical techniques that are combined with bioinformatics to obtain a complete and detailed overview of various metabolites in biological sources, thus being able to characterize health status and diseaserelated metabolic signatures. Fast and targeted metabolomics and non-targeted metabolomics are the two main study strategies in the field of metabolomics<sup>[12][13]</sup>. Both provide important information about changes in metabolism and quantification of metabolites in many chronic pathological environments, with applications in the diagnosis, pathophysiology and treatment of diseases, including asthma<sup>[14]</sup>. If, on the one hand, targeted metabolomics is concerned only with identifying and quantifying partially or totally the predefined metabolites of interest, the untargeted strategy offers much more comprehensive results regarding the identification and quantification of metabolites, since it does not restrict the analysis to previously defined target molecules<sup>[12]</sup>. The latter is possibly the best way to characterize a disease from a metabolic point of view and identify new biomarkers [12][15]. However, the untargeted metabolomic strategy can be problematic because it identifies a wide range of metabolites that can be difficult to interpret and constitute a confounding factor. In fact, identification and validation of relevant metabolites using undirected metabolomics requires careful analysis, as only a subset of all metabolite characteristics can be positively attributed to a molecular structure<sup>[16][17]</sup>. Furthermore, a high level of big data computational analysis is crucial for an adequate and standardized analysis and interpretation of results that can avoid or significantly minimize the possibility of producing erroneous results<sup>[16][18][19][20]</sup>. This is very important because the metabolome can be influenced or confused by many aspects such as age, sex, circadian rhythm, medications and other xenobiotics, microbiota, physical exercise, diet or even air pollution, both in healthy states and in diseases. In addition, sample source and types, sample collection and storage aspects, analytical procedure aspects as well as data analysis also influence the results. Finally, external validation using results from different patient cohort samples is crucial to make the results robust and generalizable; however, this aspect is absent in many studies.

Methodologically, metabolomics strategies can be supported by several techniques, namely nuclear magnetic resonance (NMR) "spectroscopy"<sup>[21]</sup>, liquid chromatography-mass spectrometry (LC-MS) and gas chromatography-mass spectrometry (GC) -MS) <sup>[20][22]</sup>. LC-MS has possibly been the most used technique as it offers greater sensitivity in identifying metabolites<sup>[15][23]</sup>. In any case, NMR is also an extremely useful technique, and the best and most complete metabolomic approach would likely involve a combination of both the techniques. In fact, the joint use of LC-MS and NMR (LC-NMR-MS systems) allows the combination of high throughput (via NMR) with high sensitivity and resolution levels (via LC-MS)<sup>[24][25]</sup>.

Electronic nose (eNose) devices can be used for global characterization of metabolites, detecting complex mixtures of Volatile Organic Compounds (VOCs) in exhaled breath and providing associated respiratory impressions of such mixtures. eNose technologies are cheaper, non-invasive, and provide faster features, allowing early detection of metabolite changes compared to conventional methods based on analytical chemistry<sup>[26][27][28][29]</sup>.

In the specific context of respiratory diseases, eNoses can detect changes in VOC mixtures in asthma<sup>[30][31][32]</sup>, COPD<sup>[33]</sup> <sup>[34][35]</sup>, as well as in several other respiratory diseases, including cystic fibrosis or tuberculosis<sup>[36][37][38][39]</sup>. Dualtechnology eNoses are similar to conventional chemical identification approaches in that they have chemical analysis capabilities that allow them to identify VOCs as disease-specific biomarkers<sup>[30][31][32][33][34][35]</sup>. Finally, in this context, the use of Application-specific database libraries of VOC biomarkers may favor early disease detection<sup>[29][40][41]</sup>.

Overall, several metabolomics studies, focusing on small molecule metabolites in urine, peripheral blood or EBC, including VOCs, have shown that the expression of metabolites can discriminate between (a) asthmatic and non-asthmatic individuals<sup>[32][42][43][44][45][46][47][48][49][50][51]</sup>; (b) asthmatic patients and patients with chronic obstructive pulmonary disease (COPD)<sup>[52]</sup>; (c) asthma exacerbations and stable asthma<sup>[53]</sup>, (d) severe and non-severe asthma<sup>[54][55]</sup> [<sup>56][57][58]</sup>, (e) different asthma phenotypes<sup>[59][60][61]</sup> and (f) evaluation of treatment responses and effects, including responsiveness or not to corticosteroids<sup>[13]</sup>.

Most studies on biomarkers and phenotypes have been carried out mainly in children and non-elderly asthmatic adults. In fact, phenotyping studies in elderly asthmatics are scarce and, as far as we know, no metabolomic approach has been used in this subgroup of patients. This constitutes a great knowledge gap, as, in the last twenty years, there has been an evident increase in the percentage of the elderly population<sup>[62]</sup>. In addition, asthma is not always easy to diagnose or treat in these patients, due to multiple comorbidities, polypharmacy, partially different clinical manifestations, less awareness of symptoms, medication non-compliance, or other problems<sup>[62][63]</sup>. Thus, having metabolomic biomarkers that can increase the diagnostic, prognostic and therapeutic capacity in the approach of personalized medicine becomes of great importance in all age groups, especially in the elderly.

### 3. Conclusions

#### [64][65][66]

Several studies have shown that metabolomics can help distinguish between asthma and a healthy state, between severe and non-severe asthma, and between asthma and other chronic obstructive respiratory diseases. In particular, and as mentioned earlier in this review and also reviewed in depth by others<sup>[49][64][65][66]</sup>, some metabolic pathways appear to be altered more consistently in asthma than in a healthy state.

Furthermore, it also seems clear that some of the inflammatory phenotypes of asthma (eg, eosinophilic asthma) may be preferentially associated with certain metabolic signatures. However, fully demonstrable and reproducible asthma-related metabolic "phenotypes" cannot be robustly defined, with the possible exception of obesity-related asthma, which may constitute an endotype of its own and also involve a clearer metabolic phenotype with characteristics specific underlyings - that is, a "metabolic endotype".

Thus, it is now probably more appropriate to mention the metabolic signatures of asthma than the actual metabolic phenotypes or endotypes. In any case, the situation will be better clarified once some of the future challenges are resolved. This may involve aspects such as the actual definition of clear molecular metabolic phenotypes, based on integrated, multi-level and unbiased cluster analyses. In addition, proper assessment of the reliable relationships between metabolic phenotypes and integrated groups of multiparameter asthma phenotypes will be relevant in the hope that noninvasive and timely assessment of the metabolic aspects of asthma can accurately reflect the specificities of various clustered asthma of phenotypes and endotypes. For this to occur, more multicentric multinational metabolic studies are needed, using the same techniques and similar targeted and non-targeted approaches. In addition, the reproducibility of asthma metabolic signatures needs to be better defined in different scenarios, as well as over time, in further longitudinal studies, so that the limits of variability and stability are understood for the most relevant metabolites and pathways. In addition, at least some additional aspects that can affect the expression of asthma and the metabotypes related to the asthma phenotype should also be studied, namely the nutritional aspects [so that the limits of variability and stability are understood for more metabolites and pathways. relevant. In addition, at least some additional aspects that can affect the expression of asthma and the metabotypes related to the asthma phenotype should also be studied, namely the nutritional aspects [so that the limits of variability and stability are understood for more metabolites and pathways. relevant. In addition, at least some additional aspects that may affect the expression of asthma and the metabotypes related to the asthma phenotype should also be studied, namely nutritional aspects<sup>[67]</sup>, metabolic aspects associated with the microbiome<sup>[68]</sup> or air pollution parameters<sup>[69][70]</sup>.

Further research is warranted and the integration of metabolomics with multifunctional and multifunctional parameters, with subsequent algorithm-based analysis, based on "big data" artificial intelligence (AI), can allow for a more complete and complete analysis of phenotype global/integrative clusters not only of asthma, but also in the context of chronic

obstructive respiratory diseases, thus allowing for greater diagnostic yield, personalized approaches and prognostic capacity.

#### References

- 1. Global Initiative for Asthma. Global Strategy for Asthma Management and Prevention. Updated 2021. Available online: Ginasthma.org/wp-content/uploads/2021/05/GINA-Main-Report-2021-V2-WMS.pdf (accessed on 2 March 2021).
- Global Asthma Network. The Global Asthma Report 2018. New Zealand 2018. Available online: http://www.globalasthmareport.org/ (accessed on 6 March 2021).
- Silkoff, P.E.; Strambu, I.; Laviolette, M.; Singh, D.; FitzGerald, J.M.; Lam, S.; Kelsen, S.; Eich, A.; Ludwig-Sengpiel, A.; Hupp, G.C.; et al. Asthma characteristics and biomarkers from the Airways Disease Endotyping for Personalized Therapeutics (ADEPT) longitudinal profiling study. Respir. Res. 2015, 16, 142. [Google Scholar] [CrossRef]
- 4. Anderson, G.P. Endotyping asthma: New insights into key pathogenic mechanisms in a complex, heterogeneous disease. Lancet 2008, 372, 1107–1119. [Google Scholar] [CrossRef]
- Lötvall, J.; Akdis, C.A.; Bacharier, L.B.; Bjermer, L.; Casale, T.B.; Custovic, A.; Lemanske, R.F., Jr.; Wardlaw, A.J.; Wenzel, S.E.; Greenberger, P.A. Asthma endotypes: A new approach to classification of disease entities within the asthma syndrome. J. Allergy Clin. Immunol. 2011, 127, 355–360. [Google Scholar] [CrossRef]
- Chung, K.F. Precision medicine in asthma: Linking phenotypes to targeted treatments. Curr. Opin. Pulm. Med. 2018, 24, 4–10. [Google Scholar] [CrossRef] [PubMed]
- Kuruvilla, M.E.; Lee, F.E.; Lee, G.B. Understanding asthma phenotypes, endotypes, and mechanisms of disease. Clin. Rev. Allergy Immunol. 2019, 56, 219–233. [Google Scholar] [CrossRef] [PubMed]
- Rufo, J.; Taborda-Barata, L.; Lourenço, O. Serum biomarkers in elderly asthma. J. Asthma 2013, 50, 1011–1019. [Google Scholar] [CrossRef] [PubMed]
- Breiteneder, H.; Peng, Y.-Q.; Agache, I.; Diamant, Z.; Eiwegger, T.; Fokkens, W.J.; Tradil-Hoffmann, C.; Nadeau, K.; O'Hehir, R.E.; O'Mahony, L.; et al. Biomarkers for diagnosis and prediction of therapy responses in allergic diseases and asthma. Allergy 2020, 75, 3039–3068. [Google Scholar] [CrossRef]
- Lee, Y.; Quoc, Q.L.; Park, H.S. Biomarkers for severe asthma: Lessons from longitudinal cohort studies. Allergy Asthma Immunol. Res. 2021, 13, 375–389. [Google Scholar] [CrossRef] [PubMed]
- 11. Ozdemir, C.; Kucuksezer, U.C.; Akdis, M.; Akdis, C.A. The concepts of asthma endotypes and phenotypes to guide current and novel treatment strategies. Expert Rev. Respir. Med. 2018, 12, 733–743. [Google Scholar] [CrossRef]
- 12. Ribbenstedt, A.; Ziarrusta, H.; Benskin, J.P. Development, characterization and comparisons of targeted and nontargeted metabolomics methods. PLoS ONE 2018, 13, e0207082. [Google Scholar] [CrossRef]
- Zhu, Z.; Camargo, C.A., Jr.; Hasegawa, K. Metabolomics in the prevention and management of asthma. Expert Rev. Respir. Med. 2019, 13, 1135–1138. [Google Scholar] [CrossRef] [PubMed]
- Peel, A.M.; Wilkinson, M.; Sinha, A.; Loke, Y.K.; Fowler, S.J.; Wilson, A.M. Volatile organic compounds associated with diagnosis and disease characteristics in asthma—A systematic review. Respir. Med. 2020, 169, 105984. [Google Scholar] [CrossRef]
- Gertsman, I.; Barshop, B.A. Promises and pitfalls of untargeted metabolomics. J. Inherit. Metab. Dis. 2018, 41, 355– 366. [Google Scholar] [CrossRef] [PubMed]
- Johnson, C.H.; Ivanisevic, J.; Siuzdak, G. Metabolomics: Beyond biomarkers and towards mechanisms. Nat. Rev. Mol. Cell Biol. 2016, 17, 451–459. [Google Scholar] [CrossRef] [PubMed]
- 17. Viant, M.R.; Kurland, I.J.; Jones, M.R.; Dunn, W.B. How close are we to complete annotation of metabolomes? Curr. Opin. Chem. Biol. 2017, 36, 64–69. [Google Scholar] [CrossRef]
- Johnson, C.H.; Patterson, A.D.; Idle, J.R.; Gonzalez, F.J. Xenobiotic metabolomics: Major impact on the metabolome. Annu. Rev. Pharmacol. Toxicol. 2012, 52, 37–56. [Google Scholar] [CrossRef]
- Eghbalnia, H.R.; Romero, P.R.; Westler, W.M.; Baskaran, K.; Ulrich, E.L.; Markley, J.L. Increasing rigor in NMR-based metabolomics through validated and open source tools. Curr. Opin. Biotechnol. 2017, 43, 56–61. [Google Scholar] [CrossRef]
- 20. Dominick, T.M.; Gill, E.L.; Vedam-Mai, V.; Yost, R.A. Mass spectrometry-based cellular metabolomics: Current approaches, applications and future directions. Anal. Chem. 2021, 93, 546–566. [Google Scholar] [CrossRef]

- 21. Crook, A.A.; Powers, R. Quantitative NMR-based biomedical metabolomics: Current status and applications. Molecules 2020, 25, 5128. [Google Scholar] [CrossRef]
- 22. Alves, S.; Paris, A.; Rathahao-Paris, E. Chapter Four—Mass spectrometry-based metabolomics for an in-depth questioning of human health. Adv. Clin. Chem. 2020, 99, 147–191. [Google Scholar] [CrossRef]
- Schrimpe-Rutledge, A.C.; Codreanu, S.G.; Sherrod, S.D.; McLean, J.A. Untargeted metabolomics strategieschallenges and emerging directions. J. Am. Soc. Spectrom. 2016, 27, 1897–1905. [Google Scholar] [CrossRef]
- 24. Dunn, W.B.; Ellis, D.I. Metabolomics: Current analytical platforms and methodologies. Trends Anal. Chem. 2005, 24, 285–294. [Google Scholar]
- Zhang, A.; Sun, H.; Wang, P.; Han, Y.; Wang, X. Modern analytical techniques in metabolomics analysis. Analyst 2012, 137, 293–300. [Google Scholar] [CrossRef] [PubMed]
- 26. Wilson, A.D. Advances in electronic-nose technologies for the detection of volatile biomarker metabolites in the human breath. Metabolites 2015, 5, 140–163. [Google Scholar] [CrossRef]
- 27. Wilson, A.D. Biomarker metabolite signatures pave the way for electronic-nose applications in early clinical disease diagnoses. Curr. Metab. 2017, 5, 90–101. [Google Scholar] [CrossRef]
- Wilson, A.D.; Baietto, M. Advances in electronic-nose technologies developed for biomedical applications. Sensors 2011, 11, 1105–1176. [Google Scholar] [CrossRef] [PubMed]
- 29. Wilson, A.D. Noninvasive early disease diagnosis by electronic-nose and related VOC-detection devices. Biosensors 2020, 10, 73. [Google Scholar] [CrossRef] [PubMed]
- 30. Lärstad, M.A.E.; Torén, K.; Bake, B.; Olin, A.-C. Determination of ethane, pentane and isoprene in exhaled air-effects of breath-holding, flow rate and purified air. Acta. Physiol. 2007, 189, 87–98. [Google Scholar] [CrossRef] [PubMed]
- Smith, A.D.; Cowan, J.O.; Filsell, S.; McLachlan, C.; Monti-Sheehan, G.; Jackson, P.; Robin Taylor, D. Diagnosing asthma: Comparisons between exhaled nitric oxide measurements and conventional tests. Am. J. Respir. Crit. Care Med. 2004, 169, 473–478. [Google Scholar] [CrossRef] [PubMed]
- 32. Cavaleiro Rufo, J.; Madureira, J.; Oliveira Fernandes, E.; Moreira, A. Volatile organic compounds in asthma diagnosis: A systematic review and meta-analysis. Allergy 2016, 71, 175–188. [Google Scholar] [CrossRef] [PubMed]
- 33. Corradi, M.; Majori, M.; Cacciani, G.C.; Consigli, G.F.; de'Munari, E.; Pesci, A. Increased exhaled nitric oxide in patients with stable chronic obstructive pulmonary disease. Thorax 1999, 54, 572–575. [Google Scholar] [CrossRef]
- 34. Binson, V.A.; Subramoniam, M.; Mathew, L. Discrimination of COPD and lung cancer from controls through breath analysis using a self-developed e-nose. J. Breath Res. 2021. [Google Scholar] [CrossRef] [PubMed]
- 35. Ratiu, I.A.; Ligor, T.; Bocos-Bintintan, V.; Mayhew, C.A.; Buszewski, B. Volatile Organic Compounds in exhaled breath as fingerprints of lung cancer, asthma and COPD. J. Clin. Med. 2020, 10, 32. [Google Scholar] [CrossRef]
- Balint, B.; Kharitonov, S.A.; Hanazawa, T.; Donnelly, L.E.; Shah, P.L.; Hodson, M.E.; Barnes, P.J. Increased nitrotyrosine in exhaled breath condensate in cystic fibrosis. Eur. Respir. J. 2001, 17, 1201–1207. [Google Scholar] [CrossRef] [PubMed]
- Kamboures, M.A.; Blake, D.R.; Cooper, D.M.; Newcomb, R.L.; Barker, M.; Larson, J.K.; Meinardi, S.; Nussbaum, E.; Rowland, F.S. Breath sulfides and pulmonary function in cystic fibrosis. Proc. Natl. Acad. Sci. USA 2005, 102, 15762– 15767. [Google Scholar] [CrossRef] [PubMed]
- Barker, M.; Hengst, M.; Schmid, J.; Buers, H.-J.; Mittermaier, B.; Klemp, D.; Koppmann, R. Volatile organic compounds in the exhaled breath of young patients with cystic fibrosis. Eur. Respir. J. 2006, 27, 929–936. [Google Scholar] [CrossRef] [PubMed]
- Syhre, M.; Manning, L.; Phuanukoonnon, S.; Harino, P.; Chambers, S.T. The scent of Mycobacterium tuberculosis— Part II breath. Tuberculosis 2009, 89, 263–266. [Google Scholar] [CrossRef]
- 40. Wilson, A.D. Applications of electronic-nose technologies for noninvasive early detection of plant, animal and human diseases. Chemosensors 2018, 6, 45. [Google Scholar] [CrossRef]
- 41. Wilson, A.D. Developing electronic-nose technologies for clinical practice. J. Med. Surg. Pathol. 2018, 3, 4. [Google Scholar] [CrossRef]
- 42. Paredi, P.; Kharitonov, S.A.; Barnes, P.J. Elevation of exhaled ethane concentration in asthma. Am. J. Respir. Crit. Care Med. 2000, 162, 140–1454. [Google Scholar] [CrossRef]
- Montuschi, P.; Santonico, M.; Mondino, C.; Pennazza, G.; Mantini, G.; Martinelli, E.; Capuano, R.; Ciabattoni, G.; Paolesse, R.; Di Natale, C.; et al. Diagnostic performance of an electronic nose, fractional exhaled nitric oxide, and lung function testing in asthma. Chest 2010, 137, 790–796. [Google Scholar] [CrossRef] [PubMed]

- 44. Ibrahim, B.; Basanta, M.; Cadden, P.; Singh, D.; Douce, D.; Woodcock, A.; Fowler, S.J. Non-invasive phenotyping using exhaled volatile organic compounds in asthma. Thorax 2011, 66, 804–809. [Google Scholar] [CrossRef] [PubMed]
- Schivo, M.; Seichter, F.; Aksenov, A.A.; Pasamontes, A.; Peirano, D.J.; Mizaikoff, B.; Kenyon, N.J.; David, C.E. A mobile instrumentation platform to distinguish airway disorders. J. Breath Res. 2013, 7, 98–106. [Google Scholar] [CrossRef] [PubMed]
- 46. van der Schee, M.P.; Palmay, R.; Cowan, J.O.; Taylor, D.R. Predictive steroid responsiveness in patients with asthma using exhaled breath profiling. Clin. Exp. Allergy 2013, 43, 1217–1225. [Google Scholar] [CrossRef] [PubMed]
- Jung, J.; Kim, S.-H.; Lee, H.S.; Choi, G.S.; Jung, Y.-S.; Ryu, D.H.; Park, H.-S.; Hwang, G.-S. Serum metabolomics reveals pathways and biomarkers associated with asthma pathogenesis. Clin. Exp. Allergy 2013, 43, 425–433. [Google Scholar] [CrossRef]
- Checkley, W.; Deza, M.P.; Klawitter, J.; Romero, K.M.; Klawitter, J.; Pollard, S.L.; Wise, R.A.; Christians, U.; Hansel, N.N. Identifying biomarkers for asthma diagnosis using targeted metabolomics approaches. Respir. Med. 2016, 121, 59–66. [Google Scholar] [CrossRef]
- Kelly, R.S.; Dahlin, A.; McGeachie, M.J.; Qiu, W.; Sordillo, J.; Wan, E.S.; Wu, A.C.; Lasky-Su, J. Asthma metabolomics and the potential for integrative omics in research and the clinic. Chest 2017, 151, 262–277. [Google Scholar] [CrossRef]
- Esteves, P.; Blanc, L.; Celle, A.; Dupin, I.; Maurat, E.; Amoedo, N.; Cardouat, G.; Ousova, O.; Gales, L.; Bellvert, F.; et al. Crucial role of fatty acid oxidation in asthmatic bronchial smooth muscle remodelling. Eur. Respir. J. 2021. [Google Scholar] [CrossRef]
- Jiang, T.; Dai, L.; Li, P.; Zhao, J.; Wang, X.; An, L.; Liu, M.; Wu, S.; Wang, Y.; Peng, Y.; et al. Lipid metabolism and identification of biomarkers in asthma by lipidomic analysis. Biochim. Biophys. Acta. Mol. Cell Biol. Lipids 2021, 1866, 158853. [Google Scholar] [CrossRef]
- 52. Maniscalco, M.; Paris, D.; Melck, D.J.; Molino, A.; Carone, M.; Ruggeri, P.; Caramori, G.; Motta, A. Differential diagnosis between newly diagnosed asthma and COPD using exhaled breath condensate metabolomics: A pilot study. Eur. Respir. J. 2018, 51, 1701825. [Google Scholar] [CrossRef]
- Saude, E.J.; Skappak, C.D.; Regush, S.; Cook, K.; Ben-Zevi, A.; Becker, A.; Moqbel, R.; Sykes, B.D.; Roew, B.H.; Adamkp, D.J. Metabolomic profiling of asthma: Diagnostic utility of urine nuvlear magnetic resonance spectroscopy. J. Allergy Clin. Immunol. 2011, 127, 757–764. [Google Scholar] [CrossRef]
- 54. Comhair, S.A.A.; McDunn, J.; Bennett, C.; Fettig, J.; Erzurum, S.C.; Kalhan, S.C. Metabolic endotype of asthma. J. Immunol. 2015, 195, 643–650. [Google Scholar] [CrossRef] [PubMed]
- 55. Loureiro, C.C.; Oliveira, A.S.; Santos, M.; Rudnitskaya, A.; Todo-Bom, A.; Bousquet, J.; Rocha, S.M. Urinary metabolomic profiling of asthmatics can be related to clinical characteristics. Allergy 2016, 71, 1362–1365. [Google Scholar] [CrossRef] [PubMed]
- 56. Carraro, S.; Giordano, G.; Reniero, F.; Carpi, D.; Stocchero, M.; Sterk, P.J.; Baraldi, E. Asthma severity in childhood and metabolomic profiling of breath condensate. Allergy 2013, 68, 110–117. [Google Scholar] [CrossRef] [PubMed]
- Reinke, S.N.; Gallart-Ayala, H.; Gomez, C.; Checa, A.; Fauland, A.; Naz, S.; Kamleh, M.A.; Djukanovic, R.; Hinks, T.S.C.; Wheelock, C.E. Metabolomics analysis identifies different metabotypes of asthma severity. Eur. Respir. J. 2017, 49, 1601740. [Google Scholar] [CrossRef]
- 58. Ntontsi, P.; Ntzoumanika, V.; Loukides, S.; Benaki, D.; Gkikas, E.; Mikros, E.; Bakakos, P. EBC metabolomics for asthma severity. J. Breath Res. 2020, 14, 036007. [Google Scholar] [CrossRef]
- 59. Ibrahim, B.; Marsden, P.; Smith, J.A.; Custovic, A.; Nilsson, M.; Fowler, S.J. Breath metabolomic profiling by nuclear magnetic resonance spectroscopy in asthma. Allergy 2013, 68, 1050–1056. [Google Scholar] [CrossRef]
- Mattarucchi, E.; Baraldi, E.; Guillou, C. Metabolomics applied to urine samples in childhood asthma, differentiation between asthma phenotypes and identification of relevant metabolites. Biomed. Chromatogr. 2012, 26, 89–94. [Google Scholar] [CrossRef] [PubMed]
- Maniscalco, M.; Paris, D.; Melck, D.J.; D'Amato, M.; Zedda, A.; Sofia, M.; Stellato, C.; Motta, A. Coexistence of obesity and asthma determines a distinct respiratory metabolic phenotype. J. Allergy Clin. Immunol. 2017, 139, 1536–1547. [Google Scholar] [CrossRef]
- 62. Skloot, G.S.; Busse, P.J.; Braman, S.S.; Kovacs, E.J.; Dixon, A.E.; Vaz Fragoso, C.A.; Ragless, B.B. An official American Thoracic Society Workshop Report: Evaluation and management of asthma in the elderly. Ann. Am. Thorac. Soc. 2016, 13, 2064–2077. [Google Scholar] [CrossRef]
- 63. Maricoto, T.; Santos, D.; Carvalho, C.; Teles, I.; Correia-de-Sousa, J.; Taborda-Barata, L. Assessment of poor inhaler technique in older patients with asthma or COPD: A predictive tool for clinical risk and inhaler performance. Drugs

Aging 2020, 37, 605-616. [Google Scholar] [CrossRef]

- 64. Agache, I.; Rocha, C.; Beltran, J.; Song, Y.; Posso, M.; Solà, I.; Alonso-Coello, P.; Akdis, C.; Akdis, M.; Canonica, G.W.; et al. Efficacy and safety of treatment with biologicals (benralizumab, dupilumab and omalizumab) for severe allergic asthma: A systematic review for the EAACI guidelines—Recommendations on the use of biologicals in severe asthma. Allergy 2020, 75, 1043–1057. [Google Scholar] [CrossRef] [PubMed]
- 65. Sparvero, L.J.; Tian, H.; Amoscato, A.A.; Sun, W.-Y.; Anthonymuthu, T.S.; Tyurina, Y.Y.; Kapralov, O.; Javadov, S.; He, R.-R.; Watkins, S.C.; et al. Direct mapping of phospholipid ferroptotic death signals in cells and tissues by Gas Cluster Ion Beam Secondary Ion Mass Spectrometry (GCIB-SIMS). Angew. Chem. Int. Ed. Engl. 2021, 60, 11784–11788. [Google Scholar] [CrossRef]
- 66. Pité, H.; Morais-Almeida, M.; Rocha, S.M. Metabolomics in asthma: Where do we stand? Curr. Opin. Pulm. Med. 2018, 24, 94–103. [Google Scholar] [CrossRef]
- 67. Acevedo, N.; Alhamwe, B.A.; Caraballo, L.; Ding, M.; Ferrante, A.; Garn, H.; Garssen, J.; Hii, C.S.; Irvine, J.; Llinás-Caballero, K.; et al. Perinatal and early-life nutrition, epigenetics and allergy. Nutrients 2021, 13, 724. [Google Scholar] [CrossRef] [PubMed]
- 68. Lee-Sarwar, K.; Lasky-Su, J.; Kelly, R.S.; Litonjua, A.A.; Weiss, S.T. Gut microbial-derived metabolomics of asthma. Metabolites 2020, 10, 97. [Google Scholar] [CrossRef]
- Nassan, F.L.; Kelly, R.S.; Kosheleva, A.; Koutrakis, P.; Vokonas, P.S.; Lasky-Su, J.A.; Schwartz, J.D. Metabolomic signatures if the long-term exposure to air pollution and temperature. Environ. Health. 2021, 20, 3. [Google Scholar] [CrossRef] [PubMed]
- 70. Jeong, A.; Fiorito, G.; Keski-Rahkonen, P.; Imboden, M.; Kiss, A.; Robinot, N.; Gmuender, H.; Vlaanderen, J.; Vermeulen, R.; Kyrtopoulos, S.; et al. Perturbation of metabolic pathways mediates the association of air pollutants with asthma and cardiovascular diseases. Environ. Int. 2018, 119, 334–345. [Google Scholar] [CrossRef] [PubMed]

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