Rhipicephalus Tick in Southeast Asia

Subjects: Parasitology Contributor: LI PENG TAN

Rhipicephalus species are distributed globally with a notifiable presence in Southeast Asia (SEA) within animal and human populations. The *Rhipicephalus* species are highly adaptive and have established successful coexistence within human dwellings and are known to be active all year round, predominantly in tropical and subtropical climates existing in SEA.

Keywords: Southeast Asia ; Rhipicephalus tick ; Tick and tick-borne diseases ; Susceptibility Host Responses ; Host Range ; Economical impacts

1. Background

Southeast Asia (SEA) covers about 4.5 million km^2 of landmass, with a human population hovering around 670 million ^[1]. This region comprises 11 countries, and it is a vast Asian region situated east of the Indian subcontinent and South of China (**Figure 1**). All 11 countries fall within the tropical and subtropical climatic zones. The enormous variety of landscapes and climatic complexities have given rise to a considerable diversity of animals throughout the region, including ticks. With the consistent growth in the average annual gross domestic product (GDP), the concurrent expansion of SEA's livestock sector naturally occurred ^[2]. Several adverse effects have accompanied this spectacular change in—the "Livestock Revolution"—the phenomenal rise in demand for foods of animal origin in society ^[3]. Examples include the existing threats of outbreaks of zoonotic diseases that can compromise both animal and human health ^{[4][5]}, cause economic losses due to diseases ^[6], and result in environmental pollutions from the usage of disease control drugs and pesticides ^{[7][8]}. Small-scale livestock farming (i.e., backyard and village farms) remain the predominant practice in most low-income countries in SEA ^[9]. This practice requires intensive contact between livestock and farmers, which creates ideal conditions for cross-transfer of pathogens associated with potential zoonosis, in addition to ticks ^[10].



Figure 1. Geographic depiction of Southeast Asia: SEA comprises countries within the Indo-Chinese peninsula of continental Asia, including Myanmar (Burma), Laos, Vietnam, Thailand, Cambodia, Malaysia, Singapore, Indonesia, Timor-Leste, Brunei and the Philippines (<u>https://aseanup.com/free-maps-asean-southeast-asia/</u>, accessed on 4 January 2021).

Ticks are second only to mosquitoes as vectors of disease of medical and veterinary importance. They transmit the widest variety of pathogens for any known arthropod vector, viz. viruses, bacteria, rickettsia, protozoa, or even certain helminths (microfilaria) ^{[11][12]}. Rhipicephalus. Being the genus most frequently associated with both human and domesticated animals, *Rhipicephalus* is thus the utmost studied genus.

2. Genus Rhipicephalus and Its Common Species in Southeast Asia

Ixodidae, also known as hard ticks, are exclusively parasitic arthropods. *Rhipicephalus* is one of the 12 extant genera of Ixodidae and comprises 84 described species ^{[13][14]}. *Rhipicephalus* falls under the subfamily of Rhipicephalinae (Metastriata) (**Figure 2**).

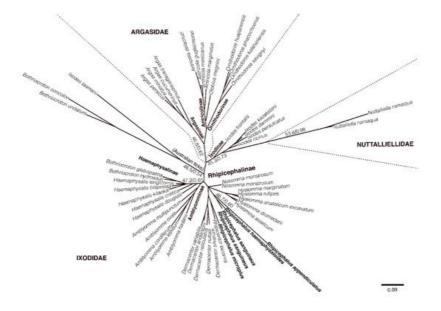


Figure 2. Phylogenetic tree based on maximum-likelihood analysis of the subfamilies of ticks from a 16S ribosomal RNA gene sequence alignment dataset. Branch support value on nodes indicates the bootstrap values of maximum-likelihood and Bayesian posterior probabilities. The highlighted names are *Rhipicephalus spp.* tick sequences from several countries.

Tick species under this genus are found globally even in regions they may not be necessarily 'indigenous' to. Animal trade across the SEA region and other parts of the world enhances the rapid distribution and establishment of tick species such as *Rhipicephalus*. *Rhipicephalus* species are associated with the infestation of livestock or domesticated animals, primarily cattle and dogs ^{[15][16][17][18]} imported into or exported out of the SEA region. They are mainly two- and three-host ticks (*Rhipicephalus*) or one host ticks for all the five species under Boophilus.

Morphology-based taxonomic classification of *R. microplus* and *R. sanguineus* s.l. has been challenging even for the most experienced taxonomists. The intra-species variations within the *R. microplus* species complex led to the description of multiple sub-species. However, many were later considered synonyms to *R. microplus* or *R. australis* ^[19]. In recent years, molecular-based phylogenetic analyses added a great deal of insight into the species diversity within the *R. microplus* species complex. Based on studies of mitochondrial cytochrome c oxidase subunit I (COI) gene marker, there are five different phylogenetic clades within the *R. microplus* species complex viz. *R. annulatus*, *R. australis* and three *R. microplus* sensu stricto (s.s.) clades ^{[18][19][20]}. These species are not possible to be differentiated based on morphology alone. *Rhipicephalus sanguineus* s.l. on the other hand was shown to have two major phylogenetic clades, the northern (tropical) and southern (temperate) lineages ^[21]. Besides, several other phylogenetic clades, or operational taxonomic units (OTUS), also exist, representing separate species and needs to be confirmed in further genetic characterization ^[21].

Rhipicephalus microplus has been reported to occur in Cambodia ^[22], Laos ^{[22][23]}, Myanmar ^[19], Vietnam ^{[24][25]}, Thailand ^{[26][27]}, Malaysia ^[18], the Philippines ^{[28][29]} and Indonesia ^{[30][22]}. *Rhipicephalus microplus* is frequently found on livestock animals such as cattle ^[30], water buffaloes ^[29] and goats ^[18]. *Rhipicephalus microplus* is widely researched as it is a significant pest of cattle with substantial economic impact ^[31]. *Rhipicephalus sanguineus* s.l. refers to a group of closely related species associated with dogs worldwide ^[32]. In SEA, it has been recorded in Laos ^{[23][33]}, Myanmar ^[34], Vietnam ^[35], Thailand ^[36], Malaysia ^{[37][38]}, the Philippines ^[39] and Indonesia ^[40]. So far, the *R. sanguineus* s.l. identified in SEA fall within the tropical lineage ^[37]. Nevertheless, the genetic diversity of *R. microplus* and *R. sanguineus* s.l. ticks in SEA is still largely unexplored. Not to mention that there are other species of *Rhipicephalus* whose molecular work are comparatively lesser than *R. microplus* and *R. sanguineus* s.l. *Rhipicephalus pilans*. For instance, only one nucleotide result was

available in the gene bank after research on the evolution and ecological niches of *Rhipicephalus* was published in the year 2021 [41].

3. Host Range of Rhipicephalus Species in Southeast Asia

The host specificity of *Rhipicephalus* in SEA can be narrowed down based on previous incidences and findings. They are mainly associated with several types of livestock and companion animals (**Table 1**).

Table 1. Host-tick list of *Rhipicephalus* hard tick in Southeast Asia.

Host Type	Country	Tick Species	Host	Reference
	Cambodia	Rhipicephalus microplus	Unknown	[22]
	Camboula	Rhipicephalus australis	Unknown	[42]
		Rhipicephalus australis	Unknown	[42]
		Rhipicephalus haemaphysaloides	Bos taurus Bubalus bubalis Capra aegagrus hircus	[43]
		Rhipicephalus microplus	Bos taurus Bubalus bubalis Capra aegagrus hircus Equus caballus Sus scrofa	[30][43][44]
	Indonesia	Rhipicephalus pilans	Bos taurus Bubalus bubalis Capra aegagrus hircus Equus caballus Ovis aries	[30][43][44]
		Rhipicephalus sanguineus s.l.	Bos taurus Bubalus bubalis Gallus gallus domesticus Sus scrofa domesticus	[44]
		Rhipicephalus haemaphysaloides	Bos sp.	[23]
Livestock	Laos	Rhipicephalus microplus	Bos sp.	[23]
		Rhipicephalus australis	Unknown	[42]
	Malaysia	Rhipicephalus microplus	Bos taurus	[18][45]
		Rhipicephalus microplus	Bos sp.	[19]
	Myanmar	Rhipicephalus microplus	Bos sp. Sus scrofa	[46]
	Singapore	Rhipicephalus microplus	Bos sp. and Bos taurus	[27][47][48]
	Thailand	Rhipicephalus australis	Unknown	[42]
	The Philippines	Rhipicephalus microplus	Bos sp. and Bos indicus Bubalus bubalis Capra aegagrus hircus	[<u>28][29][49]</u>
	Fimppines	Rhipicephalus haemaphysaloides	Bos sp.	[50]
		Rhipicephalus microplus	Bos sp. Capra aegagrus hircus	<u>[50]</u>
	Timor-Leste	Rhipicephalus sanguineus s.l.	Bos taurus	<u>[50]</u>
		Rhipicephalus annulatus	Bos sp.	[51]
		Rhipicephalus microplus	Bos sp.	[24]
	Vietnam	Rhipicephalus sanguineus s.l.	Bos sp.	<u>[52]</u>
		Rhipicephalus haemaphysaloides	Canis lupus familiaris	[43]

Host Type	Country	Tick Species	Host	Reference
	Indonesia	Rhipicephalus sanguineus s.l.	Canis lupus familiaris Felis catus	[53][43][54]
		Rhipicephalus haemaphysaloides	Canis lupus familiaris	[23]
	Laos	Rhipicephalus sanguineus s.l.	Canis lupus familiaris	[33][55]
		Rhipicephalus sanguineus s.l.	Canis lupus familiaris	[37][56][57][58][59][60][61] [62]
	Malaysia	Rhipicephalus sanguineus s.l.	Canis lupus familiaris	<u>[34]</u>
Companion animals	Myanmar	Rhipicephalus sanguineus s.l.	Canis lupus familiaris Felis catus	[53][62][63]
	Singapore	Rhipicephalus sanguineus s.l.	Canis lupus familiaris	[21][36][62]
	Thailand	Rhipicephalus sanguineus s.l.	Canis lupus familiaris Felis catus	[53][29][62]
	The Philippines	Rhipicephalus haemaphysaloides	Canis lupus familiaris	[52]
		Rhipicephalus sanguineus s.l.	Canis lupus familiaris	[21][35][52][62]
	Vietnam	Rhipicephalus haemaphysaloides	Forest rats *	[43]
	Indonesia	Rhipicephalus microplus	Rattus exulans Rattus hoffmanni Rattus rattus	[44]
		Rhipicephalus pilans	Niviventer fulvescens Rattus argentiventer Rattus exulans Rattus rattus Rattus tiomanicus	[43][44][64]
Rodents		Rhipicephalus sp.	Sundamys muelleri	[65]
	Malaysia	Rhipicephalus haemaphysaloides	Pteropus vampirus Rusa unicolor Helarctos malayanus Panthera tigris Varanus salvator Sus scrofa Hylomys suillus	<u>[43][66]</u>
	Indonesia	Rhipicephalus microplus	Bos javanicus Manis javanica Rusa timorensis Rusa unicolor	[43][44]
Wild animals		Rhipicephalus pilans	Crocidura nigripes Hylomys suillus Rusa timorensis Suncus murinus Sus scrofa	[43][67]
		Rhipicephalus sanguineus s.l.	Bos javanicus Rusa unicolor	[43]
		Rhipicephalus haemaphysaloides	Arctictis binturong Cuon alpinus Martes flavigula Neofelis nebulosi	<u>[68]</u>
	Thailand	Rhipicephalus microplus	-	[44]
		Rhipicephalus pilans	-	[44][64]
Luman	Indonesia	Rhipicephalus sanguineus s.l.	-	[43]
Human		Rhipicephalus microplus	-	[68]
	Thailand	Rhipicephalus sanguineus s.l.	-	<u>[69]</u>

* Not being explicitly mentioned on the species in the original article.

4. The Impacts of Ticks and Tick-Borne Diseases

Tick-borne diseases transmitted by *Rhipicephalus* ticks affect cattle production worldwide, including SEA countries ^[70] [^{71]}. ^[72]. Studies have shown the potentially devastating impact of *R. microplus* infestation on developing countries' livestock economies ^[31]. These losses are bothered by developing countries' inability to control and monitor the diseases; hence, it impairs the livestock economy ^[73]. The distribution and prevalence of these diseases across the SEA geopolitical area appear to be quite eco-oriented. Important Rhipicephalus-borne diseases in SEA are babesiosis, anaplasmosis, theileriosis, and ehrlichiosis. Some other pathogens transmitted by *R. sanguineus* s.l. include Hepatozoon canis ^{[39][60][74]} and Coxiella burnetti ^[59], which causes hepatozoonosis and Q-fever, respectively. The host range for these diseases is reasonably consistent, although outliers to the known host range for some tick-borne diseases have also been reported in the SEA. For instance, rare infections in a previously unknown host for Babesia canis, such as in wild rodents, have been reported ^[75] in Thailand. Similarly, Lim et al. ^[76] reported a rare occurrence of human babesiosis (caused by Babesia microti) exported from the USA into Singapore.

Babesiosis affects most warm-blooded animals with high economic and health consequences. *Babesia caballi* and *Theileria equi* collectively cause equine piroplasmosis characterized by fever and jaundice, mainly in horses and other Equidae in SEA $\frac{[77][78]}{R}$. Anaplasma that causes anaplasmosis is a tropical to subtropical rickettsial disease of ruminants and companion animals. Anaplasma marginale and *A. centrale* are the notable species in cattle and buffaloes across SEA $\frac{[79]}{R}$, while *A. platys* occur in dogs $\frac{[74][80]}{R}$.

Currently, tick-borne protozoal and rickettsial diseases are invariably endemic in SEA. Concurrent infectious diseases with Babesia, Theileria, *Anaplasma* and *Ehrlichia spp.* are increasingly reported. The theory of increasing sensitivity of pathogens detection with the help of molecular work could logically fit this scenario. However, it remains unclear why such co-morbidities are consistently challenging to treat, and the ticks are difficult to control in the environment. Hence, an elaborate effort is required to identify the epidemiological patterns of *Rhipicephalus*, the pathogens they transmitted and the rising incidence of resistance to control drugs of this tick in SEA. Molecular detection of the presence of pathogens in squashed ticks is more direct in understanding the host-parasite dynamics for TBDs should be extended further to involve more host species of *Rhipicephalus* in the region. It remains crucial to determine the extent to which *Rhipicephalus* species act as biological, mechanical vectors or both for pathogens of interest.

Tick-borne protozoan diseases cause substantial economic loss in Thailand's dairy and beef industries ^[81]. High mortality rates were noticed in the 50 million USD imported exotic breed of cattle due to tick-borne diseases. The Department also expended over 20 million USD to diagnose, treat and control diseases of animals. However, the exact economic impacts of ticks and tick-borne diseases in SEA are not available due to the lack of farm economic impact study compared to the European and African regions ^[82].

5. Resistant and Susceptibility Host Responses

The complex interaction, mainly due to the host's diverse immune mechanisms and non-immune structural components, has contributed to various responses towards tick feeding ^[83]. Most mammals mount an immunological response to a feeding tick bite. It is often more vital to the host's species with little or no evolutionary experience. Some species or breed appear to be better adapted to the tick bite; for instance, Bos indicus cattle breeds are more resistant to *R. microplus* than *B. taurus* breeds, although considerable variation in resistance exists between and within breeds ^[84]. The pattern of host resistance to ticks in the SEA region is not necessarily different from other parts of the world. Such resistance is often dependent on the commonality of the several species. Resistance is generally believed to be under genetic control ^[85]; thus, highly resistant animals can be selected to progress genetic improvement in tick resistance within a herd.

Overall, resistance to *R. microplus* infestation in cattle has many effector mechanisms. Although some of the mechanisms and modulating factors have been identified and quantified, much remains to be explained. Studying the genetic resistance to ticks among different breeds of cattle can contribute to alternative control methods. Investigations have intensified the crossing of these two groups, aiming to obtain more resistant animals to the conditions found in tropical countries and are also good meat producers. Regarding SEA, in addition, the host-range resistant factors should be expanded to include companion animals, wild animals, and livestock to understand the phenomenon. For future research, potential research of wild cattle in SEA such as Banteng (*Bos javanicus*), Gaur (*Bos gaurus*) and water buffalo (*Bubalus bubalis*) can be explored for conservation and genetic diversification purposes.

6. Controlling and Acaricides Resistance

Rhipicephalus ticks' control mainly depends on conventional acaricides. However, the exhaustive use of these chemicals has resulted in tick populations developing resistance to major acaricide chemical classes ^[86]. Ivermectin, a macrocyclic lactone, is used as an endo-ectoparasiticide. It is used as an acaricide and anthelmintic in goat and sheep farms in Malaysia ^[87], Indonesia ^[88], and Thailand ^[89]. Although there is currently no report of acaricide-resistant *Rhipicephalus* ticks in the SEA region, we cannot discount the possibility of this event. Thus, the application of alternative tick control approaches, including the rotation of acaricide, sterile hybrid ticks, pasture rotation, anti-tick vaccine, development of host resistance to ticks and the use of plant extracts, should be explored in SEA.

The alternation of the use of two or more acaricide with different modes of action could be an advantageous tick control method as well as a measure to prevent cross-resistance ^[86].

The success of mosquito control using genetic control methods $^{[90]}$ rekindled interest in using this method to control *Rhipicephalus* ticks. Osburn and Knipling $^{[91]}$ demonstrated sterile males' production and fertile females through the mating between R. annulatus and R. microplus. The backcrossing of fertile female progenies also produces sterile males and fertile females $^{[91]}$.

The per capita consumption of livestock products among SEA countries is projected to increase in the years to come ^[92] significantly. The increase in demand for livestock products has intensified the race to acquire agricultural land between the livestock and crop farmers. Integrating both cash crop plantations with ruminant cultivation is very much encouraged ^[93].

Since the excessive use of acaricides has been shown to cause the accumulation of chemical residues in milk, meat, and the environment, safer methods have arisen. Vaccination or immunological control is touted as the most promising, environmentally friendly, and sustainable strategy for the management of *Rhipicephalus* infestation ^[94].

Plant extracts or secondary metabolites, including flavonoids, terpenes, spilanthol and coumarins, have been studied comprehensively for their potential to control ticks ^[95].

In essence, livestock farmers in SEA are the most burdened by problems associated with R. microplus infestation. However, due to the structural issues plaguing the SEA livestock industry (such as the high cost of animal feeds, lack of quality breeds, inefficient coordination of agricultural policies and limited industry linkages ^{[96][97][98][99]}, most smallholder farmers resort to using acaricide as it is the most cost-effective method to control tick infestation. Hence, in addition to structural reforms to the agriculture policies by the respective governments, farmers must be educated on sustainable agricultural practices and shown the impact of such practices in improving income levels ^[100]. Besides, there should be more university-industry-farm partnerships for the pilot-testing of newer technologies such as the application of Internet-of-Things and artificial intelligence to improve aspects of livestock farming ^[96].

7. Conclusions

The *Rhipicephalus* species is abundant and widely distributed in SEA. There seems to be no propensity for certain *Rhipicephalus* species in one SEA country over another because of the uniformity in environmental parameters. Thus far, the host range for *Rhipicephalus* is within those animal species of domestic reach (from food animals to companion animals to rodents). The presence and host range of *Rhipicephalus* species in the wild is yet to be studied and understood. There is a realm of unknown ecodynamics for this species. Nevertheless, *Rhipicephalus* pilans were found in some wild animals in Borneo. The distribution in other countries and domestic animals need crucial investigation to factor in this species in the epidemiology of tick-borne diseases in the region. The occurrence of ticks and tick-borne diseases in SEA follows a trend of the countries' affinity for specific domestic species and outbreak incidence. Those with a higher buffalo population, such as Thailand and Cambodia, would have a higher report of *Rhipicephalus* and TBDs prevalence associated with buffaloes, and vice versa for countries that farm cattle or small ruminants more.

Tick-borne diseases in SEA remain poorly characterized, mainly due to limited expertise and insufficient research interest. Base on the works collected from this review paper, we found that the knowledge of *Rhipicephalus* ticks in this region is still somewhat restricted. Reports and studies of these ticks focused primarily on the occurrence and the diseases associated with this parasite. Even though this genus of ticks consists of the two most economically important species, the data on their impacts in both the livestock and pet industry in SEA countries are not available. In some countries, there are absolutely no reports. Therefore, concerted efforts must be mounted to establish a rapporteur system for tick and TBDs in SEA. Babesiosis, anaplasmosis, and theileriosis are the most reported tick-borne disease of animals in SEA. Diagnosis is usually based on clinical signs of anemia, jaundice, fever, and laboratory findings, while treatments range from antibiotics to antiprotozoals. The roles the *Rhipicephalus* plays in the potential mechanical transmission of these diseases remains unclear even as the biological vector status is established.

References

- 1. Worldometers. South-Eastern Asia Population. 2020. Available online: (accessed on 4 January 2021).
- Otte, J.; Pica-Ciamarra, U.; Morzaria, S. A Comparative overview of the livestock-environment interactions in Asia and Sub-saharan Africa. Front. Vet. Sci. 2019, 6, 37.
- Coker, R.J.; Hunter, B.M.; Rudge, J.W.; Liverani, M.; Hanvoravongchai, P. Emerging infectious diseases in Southeast A sia: Regional challenges to control. Lancet 2011, 377, 599–609.
- 4. World Health Organization (WHO). WHO Estimates of the Global Burden of Foodborne Diseases. 2015. Available onlin e: (accessed on 4 January 2021).
- Klous, G.; Huss, A.; Heederik, D.J.; Coutinho, R.A. Human–livestock contacts and their relationship to transmission of z oonotic pathogens, a systematic review of literature. One Health 2016, 2, 65–76.
- Jeanna, B. 13 Animal-to-Human Diseases Kill 2.2 Million People Each Year. 2012. Available online: (accessed on 4 Jan uary 2021).
- 7. Steinfeld, H.; Gerber, P.; Wassenaar, T.; Castel, V.; Rosales, M.; de Haan, C. Livestock's Long Shadow: Environmental Issues and Options; Food and Agriculture Organization of the United Nations (FAO): Rome, Italy, 2006.
- Ilea, R.C. Intensive livestock farming: Global trends, increased environmental concerns, and ethical solutions. J. Agric. Environ. Ethics. 2009, 22, 153–167.
- Food and Agriculture Organization of the United Nations (FAO). Agricultural Transformation of Middle-Income Asian Ec onomies: Diversification, Farm Size and Mechanization; Dawe, D., Ed.; ESA Working Paper No. 15-04; Food and Agric ulture Organization of the United Nations: Rome, Italy, 2015.
- 10. Dantas-Torres, F.; Bruno, B.C.; Otranto, D. Ticks and tick-borne diseases: A One Health perspective. Trends Parasitol. 2012, 28, 437–446.
- 11. de la Fuente, J.; Estrada-Pena, A.; Venzal, J.M.; Kocan, K.M.; Sonenshine, D.E. Overview: Ticks as vectors of pathoge ns that cause disease in humans and animals. Front. Biosci. 2008, 13, 6938–6946.
- 12. Yu, Z.; Wang, H.; Wang, T.; Sun, W.; Yang, X.; Liu, J. Tick-borne pathogens and the vector potential of ticks in China. P arasites Vectors 2015, 8, 24.
- 13. William, L.N.; Sonenshine, D.E.; Noden, B.H.; Brown, R.N. Ticks (Ixodida). In Medical and Veterinary Entomology; Mull en, G.R., Durden, L.A., Eds.; Elsevier Inc.: Amsterdam, The Netherlands, 2019; pp. 603–672.
- 14. Ernieenor, F.C.L.; Ernna, G.; Mariana, A. Phenotypic and genotypic identification of hard ticks of the genus Haemaphys alis (Acari: Ixodidae) in Peninsular Malaysia. Exp. Appl. Acarol. 2017, 71, 387–400.
- Petney, T.N.; Saijuntha, W.; Boulanger, N.; Chitimia-Dobler, L.; Pfeffer, M.; Eamudomkarn, C.; Andrews, R.H.; Ahamad, M.; Putthasorn, N.; Muders, S.V.; et al. Ticks (Argasidae, Ixodidae) and tick-borne diseases of continental Southeast As ia. Zootaxa 2019, 4558, 1–89.
- Irwin, P.J.; Jefferies, R. Arthropod-transmitted diseases of companion animals in Southeast Asia. Trends Parasitol. 200 4, 20, 27–34.
- 17. Colella, V.; Nguyen, V.L.; Tan, D.Y.; Lu, N.; Fang, F.; Zhijuan, Y.; Wang, J.; Liu, X.; Chen, X.; Dong, J.; et al. Zoonotic ve ctorborne pathogens and ectoparasites of dogs and cats in Eastern and Southeast Asia. Emerg. Infect. Dis. 2020, 26, 1 221–1233.
- Low, V.L.; Tay, S.T.; Kho, K.L.; Koh, F.X.; Tan, T.K.; Lim, Y.A.L.; Ong, B.L.; Panchadcharam, C.; Norma-Rashid, Y.; Sofia n-Azirun, M. Molecular characterisation of the tick Rhipicephalus microplus in Malaysia: New insights into the cryptic di versity and distinct genetic assemblages throughout the world. Parasites Vectors 2015, 8, 1–10.
- 19. Roy, B.C.; Estrada-Peña, A.; Krücken, J.; Rehman, A.; Nijhof, A.M. Morphological and phylogenetic analyses of Rhipice phalus microplus ticks from Bangladesh, Pakistan and Myanmar. Ticks Tick Borne Dis. 2018, 9, 1069–1079.
- 20. Csordas, B.G.; Garcia, M.V.; Cunha, R.C.; Giachetto, P.F.; Blecha, I.M.Z.; Andreotti, R. New insights from molecular ch aracterization of the tick Rhipicephalus (Boophilus) microplus in Brazil. Rev. Bras. Parasitol. Vet. 2016, 25, 317–326.
- 21. Dantas-Torres, F.; Latrofa, M.S.; Annoscia, G.; Giannelli, A.; Parisi, A.; Otranto, D. Morphological and genetic diversity o f Rhipicephalus sanguineus sensu lato from the New and Old Worlds. Parasites Vectors 2013, 6, 213.

- 22. Burger, T.D.; Shao, R.; Barker, S.C. Phylogenetic analysis of mitochondrial genome sequences indicates that the cattle tick, Rhipicephalus (Boophilus) microplus, contains a cryptic species. Mol. Phylogenet. Evol. 2014, 76, 241–253.
- 23. Vongphayloth, K.; Brey, P.T.; Robbins, R.G.; Sutherland, I.W. First survey of the hard tick (Acari: Ixodidae) fauna of Nak ai District, Khammouane Province, Laos, and an updated checklist of the ticks of Laos. Syst. Appl. Acarol. 2016, 21, 16 6–180.
- 24. Hai, N.T.; Atsushi, M. Evaluation acaricidal efficacy of Camellia sasanqua thumb seed oil against the cattle tick Rhipice phalus (Boophilus) microplus and the dog tick Rhipicephalus sanguineus. Int. J. Med. Plant Res. 2014, 3, 284–289.
- 25. Kolonin, G.V. Review of the Ixodid Tick Fauna (Acari: Ixodidae) of Vietnam. J. Med. Entomol. 1995, 32, 276-282.
- Muramatsu, Y.; Usaki, N.; Thongchai, C.; Kramomtong, I.; Kriengsak, P.; Tamura, Y. Seroepidemiologic survey in Thaila nd of Coxiella burnetii infection in cattle and chickens and presence in ticks attached to dairy cattle. Southeast Asian J. Trop. Med. Public Health 2014, 45, 1167.
- Kaewmongkol, S.; Kaewmongkol, G.; Inthong, N.; Lakkitjaroen, N.; Sirinarumitr, T.; Berry, C.M.; Jonsson, N.N.; Stich, R.W.; Jittapalapong, S. Variation among Bm86 sequences in Rhipicephalus (Boophilus) microplus ticks collected from c attle across Thailand. Exp. Appl. Acarol. 2015, 66, 247–256.
- Ybanez, A.P.; Sivakumar, T.; Ybanez, R.H.D.; Ratilla, J.C.; Perez, Z.O.; Gabotero, S.R.; Inokuma, H. First molecular ch aracterization of Anaplasma marginale in cattle and Rhipicephalus (Boophilus) microplus ticks in Cebu, Philippines. J. Vet. Med. Sci. 2013, 75, 27–36.
- 29. Portugaliza, H.P.; Bagot, M.A. Different species of lice (Phthiraptera), fleas (Siphonaptera) and ticks (Ixodida) collected from livestock, poultry, reptile and companion animal in Leyte Island, Philippines. Livest. Res. Rural. 2015, 27, 1–10.
- Sahara, A.; Nugraheni, Y.R.; Patra, G.; Prastowo, J.; Priyowidodo, D. Ticks (Acari: Ixodidae) infestation on cattle in vari ous regions in Indonesia. Vet. World 2019, 12, 1755.
- Grisi, L.; Leite, R.C.; Martins, J.R.D.S.; Barros, A.T.M.D.; Andreotti, R.; Cançado, P.H.D.; León, A.A.P.D.; Pereira, J.B.; Villela, H.S. Reassessment of the potential economic impact of cattle parasites in Brazil. Rev. Bras. Parasitol. Vet. 201 4, 23, 150–156.
- Nava, S.; Estrada-Peña, A.; Petney, T.; Beati, L.; Labruna, M.B.; Szabó, M.P.; Venzal, J.M.; Mastropaolo, M.; Mangold, A.J.; Guglielmone, A.A. The taxonomic status of Rhipicephalus sanguineus (Latreille, 1806). Vet. Parasitol. 2015, 208, 2–8.
- Kernif, T.; Socolovschi, C.; Wells, K.; Lakim, M.B.; Inthalad, S.; Slesak, G.; Boudebouch, N.; Beaucournu, J.C.; Newton, P.N.; Raoult, D.; et al. Bartonella and Rickettsia in arthropods from the Lao PDR and from Borneo, Malaysia. Comp. Im munol. Microbiol. Infect. Dis. 2012, 35, 51–57.
- 34. Hmoon, M.M.; Htun, L.L.; Wai, S.S.; Thu, M.J.; Aung, S.T.; Chel, H.M.; Thaw, Y.N.; Win, S.Y.; Soe, N.C.; Bawm, S. Mor phological and molecular identification of ticks infested in stray dogs within Nay Pyi Taw Area, Myanmar. South Asian J. Life Sci. 2018, 6, 41–45.
- 35. Nguyen, V.L.; Colella, V.; latta, R.; Bui, K.L.; Dantas-Torres, F.; Otranto, D. Ticks and associated pathogens from dogs i n northern Vietnam. Parasitol. Res. 2019, 118, 139–142.
- Changbunjong, T.; Buddhirongawatr, R.; Suwanpakdee, S.; Siengsanan, J.; Yongyuttawichai, P.; Cheewajorn, K.; Jangj aras, J.; Sangloung, C.; Ratanakorn, P. A survey of ectoparasitic arthropods on domestic animals in Tak Province, Thail and. Southeast Asian J. Trop. Med. Public Health 2009, 40, 435–442.
- Low, V.L.; Prakash, B.K.; Lim, Y.A.L.; Tan, T.K.; Vinnie-Siow, W.Y.; Sofian-Azirun, M.; AbuBakar, S. Detection of Anaplas mataceae agents and co-infection with other tick-borne protozoa in dogs and Rhipicephalus sanguineus sensu lato tick s. Exp. Appl. Acarol. 2018, 75, 429–435.
- Koh, F.X.; Panchadcharam, C.; Tay, S.T. Vector-borne diseases in stray dogs in peninsular Malaysia and molecular det ection of Anaplasma and Ehrlichia spp. from Rhipicephalus sanguineus (Acari: Ixodidae) ticks. J. Med. Entomol. 2015, 53, 183–187.
- Galay, R.L.; Manalo, A.A.L.; Dolores, S.L.D.; Aguilar, I.P.M.; Sandalo, K.A.C.; Cruz, K.B.; Divina, B.P.; Andoh, M.; Masat ani, T.; Tanaka, T. Molecular detection of tick-borne pathogens in canine population and Rhipicephalus sanguineus (sen su lato) ticks from southern Metro Manila and Laguna, Philippines. Parasites Vectors 2018, 11, 643.
- 40. Hadi, U.K.; Soviana, S.; Pratomo, I.R.C. Prevalence of ticks and tick-borne diseases in Indonesian dogs. J. Vet. Sci. Te chnol. 2016, 7, 330.
- Bakkes, D.K.; Ropiquet, A.; Chitimia-Dobler, L.; Matloa, D.E.; Apanaskevich, D.A.; Horak, I.G.; Mans, B.J.; Matthee, C. A. Adaptive radiation and speciation in Rhipicephalus ticks: A medley of novel hosts, nested predator-prey food webs, o ff-host periods and dispersal along temperature variation gradients. Mol. Phylogenetics Evol. 2021, 162, 107178.

- 42. Estrada-Peña, A.; Venzal, J.M.; Nava, S.; Mangold, A.; Guglielmone, A.A.; Labruna, M.B.; de La Fuente, J. Reinstatem ent of Rhipicephalus (Boophilus) australis (Acari: Ixodidae) with redescription of the adult and larval stages. J. Med. Ent omol. 2012, 49, 794–802.
- 43. Munaf, H.B. Keanekaragaman hospes jenis-jenis caplak marga-marga Amblyomma, Boophilus dan Rhipicephalus (Aca rina: Ixodidae) yang tercatat memarasit kerbau dan sapi di Indonesia. Ber. Biol. 1986, 3.
- 44. Kadarsan, S. Larval Ixodid Ticks of Indonesia (Acarina: Ixodidae). Ph.D. Dissertation, University of Maryland, College P ark, MD, USA, 1971.
- 45. Tay, S.T.; Koh, F.X.; Kho, K.L.; Ong, B.L. Molecular survey and sequence analysis of Anaplasma spp. in cattle and ticks in a Malaysian farm. Trop. Biomed. 2014, 31, 769–776.
- 46. Petney, T.N. A preliminary study of the significance of ticks and tick-borne diseases in South-east Asia. Mitt. Österr. Ge s. Tropenmed. Parasitol. 1993, 15, 33–42.
- Jittapalapong, S.; Thanasilp, S.; Kengradomkit, C.; Sirinarukmit, T.; Kaewmongkol, G.; Stich, R.W. Molecular cloning, s equence analysis, and immune recognition of Bm95 from Thai strains of Rhipicephalus (Boophilus) microplus. Ann. N. Y. Acad. Sci. 2008, 1149, 45–48.
- 48. Kaewhom, P.; Stich, R.W.; Needham, G.R.; Jittapalapong, S. Molecular analysis of calreticulin expressed in salivary gla nds of Rhipicephalus (Boophilus) microplus indigenous to Thailand. Ann. N. Y. Acad. Sci. 2008, 1149, 53–57.
- 49. Swann, P.H.P.; Claveria, F.G. Rhipicephalus (Boophilus) microplus ticks (Family Ixodidae) in goats raised in a small priv ate farm in San Jose del Monte, Bulacan, Central Luzon, Philippines. Philipp. J. Sci. 2017, 146, 493–496.
- 50. Silva, H.R.B.C. Prospecção Parasitológica em Timor. Subsídios para o Estudo da Fauna Parasitológica dos Seus Anim ais Domésticos; Junta de Investigações do Ultramar: Lisboa, Portugal, 1960.
- 51. Chien, N.T.H.; Linh, B.K.; Van Tho, N.; Hieu, D.D.; Lan, N.T. Status of cattle ticks infection in yellow and dairy cows in B a Vi District. In Proceedings of the International Conference on Agriculture Development in the Context of International I ntegration: Opportunities and Challenges, Hanoi, Vietnam, 7–8 December 2016; pp. 115–119.
- 52. Dong, T.L.; Minh, D.B. Determine the presence of pathogens on ticks in the Mekong Delta region. In International Conf erence on the Development of Biomedical Engineering in Vietnam; Springer: Singapore, 2020; pp. 707–713.
- Murrell, A.; Campbell, N.J.H.; Barker, S.C. A total evidence phylogeny of ticks provides insights into the evolution of life cycles and biogeography. Mol. Phylogenet. Evol. 2001, 21, 244–258.
- 54. Sinaga, B.V.; Hariani, N. Prevalensi dan Intensitas ektoparasit pada anjing peliharaan (Canis familiaris) di Kalimantan T imur, Indonesia. J. Bioterdidik 2019, 7, 5.
- 55. Wilson, N. New distributional records of ticks from Southeast Asia and the Pacific (Metastigmata: Argasidae, Ixodidae). Orient. Insects 1970, 4, 37–46.
- 56. Low, V.L.; Prakash, B.K. First genetic characterization of the brown dog tick Rhipicephalus sanguineus sensu lato in Pe ninsular Malaysia. Exp. Appl. Acarol. 2018, 75, 299–307.
- 57. Macadam, I.; Gudan, D.; Timbs, D.V.; Urquhart, H.R.; Sewell, M.M.H. Metazoan parasites of dogs in Sabah, Malaysia. Trop. Anim. Health Prod. 1984, 16, 34–38.
- 58. Latrofa, M.S.; Dantas-Torres, F.; Giannelli, A.; Otranto, D. Molecular detection of tick-borne pathogens in Rhipicephalus sanguineus group ticks. Ticks Tick Borne Dis. 2014, 5, 943–946.
- Watanabe, M.; Nakao, R.; Amin-Babjee, S.M.; Maizatul, A.M.; Youn, J.H.; Qiu, Y.; Watanabe, M. Molecular screening fo r Rickettsia, Anaplasmataceae and Coxiella burnetii in Rhipicephalus sanguineus ticks from Malaysia. Trop. Biomed. 20 15, 32, 390–398.
- 60. Prakash, B.K.; Low, V.L.; Tan, T.K.; Vinnie-Siow, W.Y.; Lim, Y.A.L.; Morvarid, A.R.; Sofian-Azirun, M. Detection of Hepat ozoon canis in the brown dog tick and domestic dogs in Peninsular Malaysia. J. Med. Entomol. 2018, 55, 1346–1348.
- Prakash, B.K.; Low, V.L.; Vinnie-Siow, W.Y.; Tan, T.K.; Lim, Y.A.L.; Morvarid, A.R.; Sofian-Azirun, M. Detection of Babes ia spp. in dogs and their ticks from Peninsular Malaysia: Emphasis on Babesia gibsoni and Babesia vogeli infections in Rhipicephalus sanguineus sensu lato (Acari: Ixodidae). J. Med. Entomol. 2018, 55, 1337–1340.
- 62. Nguyen, V.L.; Colella, V.; Greco, G.; Fang, F.; Nurcahyo, W.; Hadi, U.K.; Venturina, V.; Tong, K.B.Y.; Tsai, Y.L.; Taweeth avonsawat, P.; et al. Molecular detection of pathogens in ticks and fleas collected from companion dogs and cats in Ea st and Southeast Asia. Parasites Vectors 2020, 13, 420.
- 63. Theis, J.H.; Franti, C.E. Changing infestation rates of Rhipicephalus sanguineus (Latreille) (Ixodidae) ticks on dogs on Singapore Island, 1965–1966. J. Med. Entomol. 1971, 8, 23–28.
- 64. Hasan, M.B. Tick fauna of Baluran Wildlife Reserve, Indonesia. Hemera Zoa 1978, 70, 37-44.

- 65. Adrus, M.; Ahamad, M.; Abdullah, M.T. Detection of rickettsiae in engorged ticks from small mammals in Malaysia. Born eo J. Resour. Sci. Technol. 2014, 4, 34–41.
- 66. Anastos, G. Two new species of ticks from Soembawa Island, Indonesia (Acarina: Ixodidae). J. Parasitol. Res. 1956, 4 2, 306–310.
- 67. Walker, J.B.; Keirans, J.E.; Horak, I.G. Genus Rhipicephalus (Acari, Ixodidae). A Guide to the Brown Ticks of the World; Cambridge University Press: Cambridge, UK, 2000.
- 68. Cornet, J.P.; Demoraes, F.; Souris, M.; Kittayapong, P.; Gonzalez, J.P. Spatial distribution of ticks in Thailand: A discuss ion basis for tick-borne virus spread assessment. Int. J. Geoinformat. 2009, 5, 57–62.
- 69. Kitaoka, S.; Suzuki, H. Studies on the parasite fauna of Thailand. Parasitic ticks on mammals and description of Ixodes siamensis sp. n. and Rhipicephalus tetracornus sp. n. (Acarina: Ixodidae). Trop. Med. 1983, 25, 205–219.
- 70. Rahman, W.A.; Lye, Y.P.; Chandrawathani, P. The seroprevalence of bovine babesiosis in Malaysia. Trop. Biomed. 201 0, 27, 301–307.
- 71. Ochirkhuu, N.; Konnai, S.; Mingala, C.N.; Okagawa, T.; Villanueva, M.; Pilapil, F.M.I.R.; Murata, S.; Ohashi, K. Molecula r epidemiological survey and genetic analysis of vector-borne infections of cattle in Luzon Island, the Philippines. Vet. P arasitol. 2015, 212, 161–167.
- 72. Jirapattharasate, C.; Moumouni, P.F.A.; Cao, S.; Iguchi, A.; Liu, M.; Wang, G.; Zhou, M.; Vudriko, P.; Efstratiou, A.; Cha ngbunjong, T.; et al. Molecular detection and genetic diversity of bovine Babesia spp., Theileria orientalis, and Anaplas ma marginale in beef cattle in Thailand. Parasitol. Res. 2017, 116, 751–762.
- 73. Perry, B.; Grace, D. The impacts of livestock diseases and their control on growth and development processes that are pro-poor. Philos. Trans. R. Soc. Lond. B Biol. Sci. 2009, 364, 2643–2655.
- 74. Inpankaew, T.; Hii, S.F.; Chimnoi, W.; Traub, R.J. Canine vector-borne pathogens in semi-domesticated dogs residing i n northern Cambodia. Parasites Vectors 2016, 9, 253.
- 75. Dantrakool, A.; Somboon, P.; Hashimoto, T.; Saito-Ito, A. Identification of a new type of Babesia species in wild rats (Ba ndicota indica) in Chiang Mai Province, Thailand. J. Clin. Microbiol. 2004, 42, 850–854.
- Lim, P.L.; Chavatte, J.M.; Vasoo, S.; Yang, J. Imported human Babesiosis, Singapore, 2018. Emerg. Infect. Dis. 2020, 2 6, 826–828.
- 77. Nugraha, A.B.; Cahyaningsih, U.; Amrozi, A.; Ridwan, Y.; Agungpriyono, S.; Taher, D.M.; Guswanto, A.; Gantuya, S.; Ta yebwa, D.S.; Tuvshintulga, B.; et al. Serological and molecular prevalence of equine piroplasmosis in Western Java, In donesia. Vet. Parasitol. Reg. Stud. Rep. 2018, 14, 1–6.
- 78. Kamyingkird, K.; Yangtara, S.; Desquesnes, M.; Cao, S.; Moumouni, A.; Jittapalapong, S.; Nimsupan, B.; Terkawi, M.A.; Masatani, T.; Nishikawa, Y.; et al. Seroprevalence of Babesia caballi and Theileria equi in horses and mules from North ern Thailand. J. Protozool. Res. 2014, 24, 11–17.
- 79. Ybañez, A.P.; Ybañez, R.H.D.; Claveria, F.G.; Cruz-Flores, M.J.; Xuenan, X.; Yokoyama, N.; Inokuma, H. High genetic diversity of Anaplasma marginale detected from Philippine cattle. J. Vet. Med. Sci. 2014, 76, 1009–1014.
- 80. Faizal, M.D.; Haryanto, A.; Tjahajati, I. Diagnosis and molecular characterization of Anaplasma platys in dog patients in Yogyakarta area, Indonesia. Indones. J. Biotechnol. 2019, 24, 43–50.
- 81. Chansiri, L. Tick-borne diseases in Thailand. Trop. Anim. Health Prod. 1997, 29, 52S.
- 82. Ghosh, S.; Azhahianambi, P.; de la Fuente, J. Control of ticks of ruminants, with special emphasis on livestock farming systems in India: Present and future possibilities for integrated control—A review. Exp. Appl. Acarol. 2006, 40, 49–66.
- Jonsson, N.N.; Piper, E.K.; Constantinoiu, C.C. Host resistance in cattle to infestation with the cattle tick Rhipicephalus microplus. Parasite Immunol. 2014, 36, 551–557.
- 84. Seifert, G. Variations between and within breeds of cattle in resistance to field infestations of the cattle tick (Boophilus microplus). Aust. J. Agric. Res. 1971, 22, 159–168.
- Kongsuwan, K.; Josh, P.; Colgrave, M.L.; Bagnall, N.H.; Gough, J.; Burns, B.; Pearson, R. Activation of several key co mponents of the epidermal differentiation pathway in cattle following infestation with the cattle tick, Rhipicephalus (Boo philus) microplus. Int. J. Parasitol. 2010, 40, 499–507.
- 86. Rodriguez-Vivas, R.I.; Jonsson, N.N.; Bhushan, C. Strategies for the control of Rhipicephalus microplus ticks in a world of conventional acaricide and macrocyclic lactone resistance. Parasitol. Res. 2018, 117, 3–29.
- Basripuzi, H.B.; Sani, R.A.; Ariff, O.M. Anthelmintic resistance in selected goat farms in Kelantan. Mal. J. Anim. Sci. 201 2, 15, 47–56.

- 88. Puspitasari, S.; Farajallah, A.; Erni Sulistiawati, M. Effectiveness of Ivermectin and Albendazole against Haemonchus c ontortus in Sheep in West Java, Indonesia. Trop. Life Sci. Res. 2016, 27, 135–144.
- 89. Kochapakdee, S.; Pandey, V.S.; Pralomkarn, W.; Choldumrongkul, S.; Ngampongsai, W.; Lawpetchara, A. Anthelmintic resistance in goats in southern Thailand. Vet. Rec. 1995, 137, 124–125.
- Bouyer, J.; Culbert, N.J.; Dicko, A.H.; Pacheco, M.G.; Virginio, J.; Pedrosa, M.C.; Garziera, L.; Pinto, A.T.M.; Klaptocz, A.; Germann, J.; et al. Field performance of sterile male mosquitoes released from an uncrewed aerial vehicle. Sci. Ro bot. 2020, 5, eaba6251.
- 91. Osburn, R.L.; Knipling, E.F. The potential use of sterile hybrid Boophilus ticks (Acari: Ixodidae) as a supplemental eradi cation technique. J. Med. Entomol. 1982, 19, 637–644.
- 92. Loong, S.K.; Lim, F.S.; Khoo, J.J.; Lee, H.Y.; Suntharalingam, C.; Ishak, S.N.; Mohd-Taib, F.S.; AbuBakar, S. Culturable pathogenic bacteria in ticks parasitizing farm animals and rodents in Malaysia. Trop. Biomed. 2020, 37, 803–811.
- Wong, C.C.; Moog, F.; Chen, C.P. Forage and ruminant livestock integration in tree crop plantations of Southeast Asia. In Grasslands: Developments Opportunities Perspectives; Reynolds, S., Frame, J., Eds.; Taylor and Francis: London, UK, 2019; pp. 403–431.
- 94. Ndawula, C., Jr.; Tabor, A.E. Cocktail anti-tick vaccines: The unforeseen constraints and approaches toward enhanced efficacies. Vaccines 2020, 8, 457.
- 95. Rodríguez-Molano, C.E.; Torres, S.U.; Monrroy, L.N. Background on the control of the cattle tick R. (B.) microplus and t he use of coumarin substances as an alternative. Pharm. Pharmacol. Int. J. 2020, 8, 215–232.
- 96. Kwanmuang, K.; Pongputhinan, T.; Jabri, A.; Chitchumnung, P. Small-scale farmers under Thailand's smart farming sys tem. FFTC-AP 2020, 2647. Available online: (accessed on 4 March 2021).
- 97. Tran, C.T. Overview of agricultural policies in Vietnam. FFTC-AP 2014, 629. Available online: (accessed on 5 March 20 21).
- Sustiari, R. Livestock development policy in Indonesia. FFTC-AP 2014, 728. Available online: (accessed on 5 March 20 21).
- 99. Hashim, F.A.H. Strategies to strengthen livestock industry in Malaysia. FFTC-AP 2015, 911. Available online: (accesse d on 5 March 2021).
- 100. Altieri, M.A.; Nichols, C.I. Agroecology scaling up for food sovereignty and resiliency. In Sustainable Agriculture Review s; Lichtfouse, E., Ed.; Springer: Berlin, Germany, 2012; pp. 1–29.

Retrieved from https://encyclopedia.pub/entry/history/show/28264