

Ca²⁺-Independent Phospholipase A₂β

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The Ca²⁺-independent phospholipase A₂β (iPLA₂β) is a member of the PLA₂ family that has been proposed to have roles in multiple biological processes including membrane remodeling, cell proliferation, bone formation, male fertility, cell death, and signaling. Such involvement has led to the identification of iPLA₂β activation in several diseases such as cancer, cardiovascular abnormalities, glaucoma, periodontitis, neurological disorders, diabetes, and other metabolic disorders. More recently, there has been heightened interest in the role that iPLA₂β plays in promoting inflammation.

Keywords: iPLA₂β ; macrophages ; T-cells ; inflammation ; eicosanoids ; resolvins

1. Introduction

Phospholipases A₂ (PLA₂s) hydrolyze the sn-2 substituent of glycerophospholipids to release a lysophospholipid and a free fatty acid [1]. Among the family of PLA₂s are the group VI Ca²⁺-independent PLA₂s (iPLA₂s), which include iPLA₂β (VIA), iPLA₂γ (VIB) iPLA₂δ (VIC), iPLA₂ε (VID), iPLA₂ζ (VIE), and iPLA₂η (VIF) [2]. The cytosolic iPLA₂β enzyme, along with the membrane-associated iPLA₂γ, are the most studied of the group VI PLA₂s. Because of its emerging link with inflammation, the focus of this review will be on iPLA₂β.

Encoded by PLA2G6, the iPLA₂β protein (84-88 kDa) has a serine lipase consensus sequence (GTSGT), contains eight N-terminal ankyrin repeats, an ATP binding cassette (GGGVKG), a caspase-3 cleavage site (DVDT), and two calmodulin (CAM) binding domains [2][3]. Crystal structure studies suggest that iPLA₂β forms a dimer through the interaction of the catalytic domains and that CAM binds to the dimer to cause a closed state, denying the access of substrates to the active site [4]. Relief from this inhibitory state is achieved through activation of calmodulin kinase IIβ, which forms a signaling complex with iPLA₂β [5]. Lipidomics-based LC/MS approaches have revealed that among glycerophospholipids, iPLA₂β exhibited the greatest activity towards 1-palmitoyl-2-arachidonoyl-sn-glycero-3-phosphoethanolamine (PAPE) and, among sn-2 substituents, a greater selectivity for linoleate or myristate [6][7]. The currently available selective inhibitors of iPLA₂β include the irreversible S-BEL (S-bromo-enol lactone) and the reversible 1,1,1-trifluoro-6-(naphthalen-2-yl)hexan-2-one (FKGK18) [8][9]. iPLA₂β is widely expressed and has many proposed roles, including signal transduction, and is recognized to contribute to neurodegenerative disorders, cancers, myocardial complications, and metabolic dysfunction (reviewed extensively elsewhere [2][3]).

While localized in the cytosol under basal conditions [10], iPLA₂β mobilizes to subcellular organelles (e.g., the endoplasmic reticulum (ER), mitochondria, nucleus) upon stimulation. Its activation leads to the hydrolysis of the sn-2 fatty acid substituent from membrane glycerophospholipids [11][12][13][14][15][16] to yield a free fatty acid (e.g., arachidonic acid, eicosapentaenoic acid (EPA), docosahexaenoic acid (DHA)) and a 2-lysophospholipid [17]. Subsequent metabolism of arachidonic acid by cyclooxygenases (COX), lipoxygenase (LOX), and cytochrome P450 (CYP) pathways leads to the generation of bioactive oxidized eicosanoids, several of which are proinflammatory [18] and recognized contributors to inflammatory diseases [19][20][21][22][23][24][25][26][27]. Some of the most potent inflammatory eicosanoids [21] are prostaglandin E₂ (PGE₂), leukotrienes (LTs), hydroxyeicosatetraenoic acids (HETEs), and dihydroxyeicosatetraenoic acids (DHETEs), and they contribute to inflammation and autoimmune diseases [22]. It is not unexpected that iPLA₂β activation can play critical roles in the initiation and progression of inflammatory pathways, which if not curbed can lead to the evolution of a variety of disorders. In contrast, other unsaturated fatty acids such as EPA and DHA can be metabolized to generate pro-resolving lipids, designated as specialized resolving mediators (SPMs) [28][29][30]. Among the proinflammatory lipids generated, PGs and LTs are the first to be produced [28]. When there is injury to the tissue, the generated PGs cause pain, swelling, and edema. To resolve the inflammation and clear antigenic debris, SPMs are generated in attempt to restore homeostasis, a process referred to as "lipid mediator class switching" [31][32].

Immune cells express iPLA₂β [33][34][35][36] and iPLA₂β-derived lipids (iDLs) contribute to cell proliferation [37], cell cycle progression [38], cell division [39], monocyte migration [40], and superoxide generation [41]. Inhibition of iPLA₂β reduces reactive oxygen species (ROS) generation [42] and is reported to be effective against autoimmune- [43] and inflammation-

based [44][45][46][47] diseases.

Although extensive literature exists linking a number of secretory PLA2s (sPLA2s) or cytosolic PLA2 α (cPLA2 α) with inflammatory responses, only a few studies have described a link between iPLA2 β and macrophages, and even fewer have considered a link between iPLA2 β and T-cells or B-cells.

2. Protective Consequences of iPLA2 β Activation

2.1. Cancer Development

Inflammation is a key contributor to cancer development [48] and cytokines released by macrophages and T-cells are integral to this process [49]. In view of their earlier observations that iPLA2 β -null mice are more susceptible to various inflammatory-based disorders [50][51][52], Inhoffen et al. [53] assessed the ability of immune cells from iPLA2 β -null mice to produce cytokines following exposure to CD95/FasL, a trigger of proinflammatory cytokine production [54]. They found that iPLA2 β -deficiency increased apoptosis in the liver, spleen, and mesenteric lymph nodes (MLN). Although Kupffer cells (i.e., satellite macrophages in the liver) generated a lower production of proinflammatory cytokines TNF α and IL-6, splenocytes became primed to release proinflammatory Th1-/Th17-related cytokines (IFN γ /IL-17 α). These findings led to the suggestion that iPLA2 β -deficiency can reduce age-related MLN lymphoma development. Mechanistically, they attributed this to the decreased availability of “find-me” and “eat-me” signals derived via iPLA2 β activation. These include LPC, a find-me signal [55], which promotes the clearance of apoptotic debris, the accumulation of which triggers and amplifies subsequent immune responses [56][57] (Figure 1).

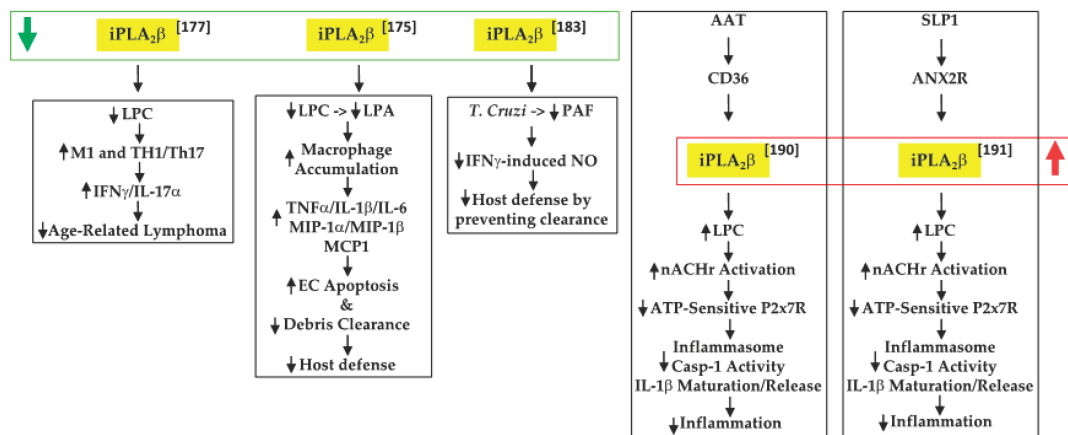


Figure 1. Proposed mechanisms by which iPLA2 β inactivation or activation can have protective outcomes in tumor development and promoting host defenses (green arrow, decreased expression/activity of iPLA2 β ; red arrow, increased expression/activity of iPLA2 β ; black up arrow, indicates increase in; black down arrow, indicates decrease in).

2.2. Inflammatory Bowel Disease (IBD)

An inflammatory disorder such as IBD is a consequence of the dysfunction of the intestinal epithelial barrier and the mucosal immune system [58]. Jiao et al. [51] further explored the role of “find-me” signals derived via iPLA2 β activation in the context of dextran sodium sulfate-induced IBD. They found that iPLA2 β -deficiency promotes the accumulation of infiltrating macrophages and dendritic cells in the colon lamina. These were associated with increased production of inflammatory cytokines (TNF α , IL-1 β , and IL-6), macrophage inflammatory proteins (MIP-1 α and MIP-1 β), and CCL2, leading to intestinal epithelial cell apoptosis and mucus barrier damage. With a concurrent decrease in LPC levels, they speculated that iPLA2 β -deficiency mitigated the availability of a “find-me” signal, which limited the clearance of apoptotic debris and the amplification of immune responses. Further, they predicted that the iPLA2 β -deficiency also reduced LPA levels, impacting the cellular compass, and preventing the optimal phagocytic function of macrophages. Subsequently, Murase et al. [59], comparing the involvement of various PLA2s, suggested that iPLA2 β -deficiency did not worsen the clinical scoring associated with IBD, relative to controls. As they did not reconcile the two studies, it is likely that differences between the source of the iPLA2 β ^{-/-} mice and DSS concentrations used may be partly responsible. Furthermore, while a 7-day DSS regimen was employed by both groups, Jiao’s group maintained the mice for an additional 3 days without DSS, prior to analyses. It may also be noted that Murase’s study did demonstrate similar lower clinical scores in the wild-type (WT) and iPLA2 β -deficient groups between days 1 and 6, relative to mice with deficiencies in cPLA2 α or in a variety of sPLA2s; however, at day 7, the scores in the iPLA2 β -deficient group appeared to be as high as in the other PLA2-deficient groups. As such, the role of iPLA2 β in this model remains to be clarified.

2.3. Chagas Disease

A further link between iPLA2 β and PAF was reported by McHowat's group [60], in the context of Chagas disease. This disease is caused through infection by the protozoan parasite [61] *Trypanosoma cruzi* (*T. cruzi*) and can lead to various cardiac abnormalities [62]. The sequela of infection begins with induction of an inflammatory response and upregulation of endothelial adhesion molecules [63], followed by attempts at resolution through the generation of proinflammatory cytokines and the induction of signaling pathways to promote the chemotaxis of immune cells to mitigate the invasion. McHowat's group reported that infection of iPLA2 β -deficient mice resulted in lowered PAF and NO production by cardiac endothelial cells, but that neither the expression of adhesion molecules nor the development of myocardial inflammation was affected. However, significant increases in parasite pseudocysts were noted in the myocardium of iPLA2 β -deficient mice. The authors suggested that this was due to an impairment in parasite clearance as a consequence of decreased iPLA2 β -mediated LPA production and, as a result, Nox4 expression and NO production. Thus, they surmised that the absence of iPLA2 β mitigates parasite clearance due to the reduced recruitment of inflammatory cells to the infected myocardial areas.

2.4. Negative Modulation of Inflammation by AAT and SLP1

In sequential reports, Grau's group constructed events that participated in regulating IL-1 β activation and release. IL-1 β is critical to host defense against infections, but the generation of excessive active IL-1 β can lead to deleterious inflammatory consequences [64]. The release of IL-1 β occurs via two signals. The first is an external stimulus that induces the synthesis of pro-IL-1 β . The second signal has been suggested to be ATP, which when released from damaged cells activates the purinergic receptor P2 \times 7R, promoting the loss of K⁺ current and triggering the assembly of NLRP3 (NLR family pyrin domain containing 3)-containing inflammasome [65]. This leads to caspase-1 activation, cleavage of pro-IL-1 β , and release of IL-1 β from the immune cell. Present in inflammatory cells, alpha-1 antitrypsin (AAT) is a strong inducer of anti-inflammatory processes and its upregulation during systemic inflammation has been associated with the decreased production of proinflammatory cytokines, including IL-1 β [66]. In examining the potential role of AAT in ATP-dependent regulation of IL-1 β in their first study [67], Siebers et al. found that AAT signaling through the CD36 receptor activates iPLA2 β , which leads to the release of a low molecular weight factor (LMWF). The LMWF is released from the cell and binds to nicotinic acetylcholine receptor (nAChR), leading to inhibition of P2X7R function, prevention of inflammasome assembly, and the processing of pro-IL-1 β to active IL-1 β , and its release from the cell. These outcomes were significantly mitigated with selective inhibition or knockdown of iPLA2 β and were also not evident in PBMCs from iPLA2 β -deficient mice. In the second study [68], Zakrzewicz et al. demonstrated that the secretory leukocyte protease inhibitor (SLP1) also interferes with ATP-dependent regulation of IL-1 β via iPLA2 β activation. SLP1 is also present in inflammatory cells [69] and its actions promote anti-inflammatory outcomes. Their results suggested that the SLP1 signals released through annexin 2, a membrane binding protein for SLP1 [70], activate iPLA2 β , leading to the production of the LMWF and subsequent inhibition of IL-1 β maturation and release. Although both pathways were found to mitigate inflammation, unfortunately neither study explored the identity of the LMWF. Interestingly, AAT administration to recent-onset T1D subjects improved β -cell function, which was correlated with reduced IL-1 β production from monocytes and myeloid dendritic cells [71].

3. Summary

iPLA2 β is a member of the family of PLA2s that hydrolyzes the sn-2 substituent from membrane glycerophospholipids. Thus, activation of iPLA2 β can lead to the production of a variety of bioactive lipid mediators. As eicosanoids generated subsequently to iPLA2 β -mediated hydrolysis of sn-2 arachidonic acid can exhibit profound proinflammatory effects and SPMs are produced from sn-2 substituents EPA and DHA, the impact of iPLA2 β on the inflammatory sequelae is profound. In recent years, the impact of iPLA2 β at the immune cell level is being recognized and those studies, as well as those from our laboratory, suggest that iPLA2 β activation modulates the maturation, polarization, activation, and functionality of macrophages, T-cells, and B-cells. The continuation of studies addressing these actions of iPLA2 β is important and warranted in order to gain a better understanding of the events that lead to the onset, maintenance, and amplification of inflammation. Identifying the relevant selective iDLs with effects on these processes could lead to the development of new strategies to treat autoimmune- and other inflammatory-based diseases.

References

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