# **Drought Stress Signaling in Plants**

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Drought stress restricts plant growth and development by altering metabolic activity and biological functions. However, plants have evolved several cellular and molecular mechanisms to overcome drought stress. Drought tolerance is a multiplex trait involving the activation of signaling mechanisms and differentially expressed molecular responses. Broadly, drought tolerance comprises two steps: stress sensing/signaling and activation of various parallel stress responses (including physiological, molecular, and biochemical mechanisms) in plants. At the cellular level, drought induces oxidative stress by overproduction of reactive oxygen species (ROS), ultimately causing the cell membrane to rupture and stimulating various stress signaling pathways (ROS, mitogen-activated-protein-kinase, Ca2+, and hormone-mediated signaling). Drought-induced transcription factors activation and abscisic acid concentration co-ordinate the stress signaling and responses in cotton. The key responses against drought stress, are root development, stomatal closure, photosynthesis, hormone production, and ROS scavenging. The genetic basis, quantitative trait loci and genes of cotton drought tolerance are presented as examples of genetic resources in plants. Sustainable genetic improvements could be achieved through functional genomic approaches and genome modification techniques such as the CRISPR/Cas9 system aid the characterization of genes, sorted out from stress-related candidate single nucleotide polymorphisms, quantitative trait loci, and genes. Exploration of the genetic basis for superior candidate genes linked to stress physiology can be facilitated by integrated functional genomic approaches. We propose a third-generation sequencing approach coupled with genome-wide studies and functional genomic tools, including a comparative sequenced data (transcriptomics, proteomics, and epigenomic) analysis, which offer a platform to identify and characterize novel genes. This will provide information for better understanding the complex stress cellular biology of plants.

Keywords: cellular stress signaling; drought stress responses

#### 1. Introduction

Global warming and climate change adversely affect agricultural production. Erosion of genetic diversity for drought tolerance in major crops is a threat to food security. Abiotic stresses are major threats, and collectively led to 73% decline in cotton production worldwide [1]. Drought refers to low water availability for the long-period of time, and affects crop production [2]. Drought tolerance is a complex trait involving multiple genes associated with cellular signaling pathways which modify several physio-morphological, and molecular responses. Plant cell membranes perceive stress signals and stimulate various self-activated and hormone-dependent signaling mechanisms [3]. Mitogen-activated-protein-kinase (MAPK) networks are involved in stress signaling and activate several stress-responsive proteins [4]. In stress signaling pathways, calcium (Ca<sup>2+</sup>) is a common second messenger, controls many physiological processes in plants. The cytoplasmic Ca<sup>2+</sup> concentration varies in response to drought stress and various hormones such as abscisic acid (ABA), jasmonic acid (JA), and ethylene [5]. Under high concentrations, ABA interacts with SnRK2 proteins, which subsequently initiate molecular and physiological responses to drought stress [6][7][8]. Jasmonic acid (JA) and its derivatives also activate signaling pathways similar to ABA [9]. Overproduction of reactive oxygen species (ROS) also triggers defense mechanisms and excessive amounts of ROS scavenged by enzymatic and non-enzymatic defense machinery in plants [10].

Following the successful transduction of stress signals, plants actively adopt drought recovery mechanisms. Tolerant plants are able to resume growth and overcome the growth deficit induced by drought. Cotton has developed numerous morpho-physiological approaches, such as photosynthetic response  $\frac{[11]}{12}$ , osmotic adjustment, stomatal regulation, low leaf water loss, high relative water contents (RWC), and enlarged tap roots  $\frac{[12]}{12}$ . These features contribute to drought tolerance through a multigenic effect. Genetic statistics and improvements of physio-morphological characters are important to reduce the effects of drought. Alterations in physio-morphological and biochemical traits have vital roles in maintaining favorable water balance in plant cells and tissues.

Genome modification technologies and transgenic approaches have been employed to develop drought-tolerant crops overexpressing transgenes that are important for plant physiology. Targeted genome editing with the CRISPR/Cas9 system has been utilized to modify the genome to obtain more stable and heritable mutations [13]. Genome-wide studies have been performed to explore stress-related candidate regions and genes for drought tolerance. Various drought-related quantitative trait loci (QTL) clusters and hotspots have been mapped in cotton. Several QTLs for abiotic stress, especially drought, have been identified using single nucleotide polymorphisms (SNPs) in genome-wide association studies (GWAS). Meta-analyses can be performed to identify common QTLs for drought-related traits [2]. Whole genome sequencing and re-sequencing of allotetraploid and diploid cotton species provide information in the biologically active states of DNA [14]. Fine- and high-density genetic maps, transcript abundance, epigenetic modifications, and SNP array platforms can also be used, as reported for other model plants (rice and *Arabidopsis*). These approaches serve as a platform for gene mapping, isolation, and cloning for drought tolerance. Moreover, the identification of novel genes can be facilitated by high-throughput marker development for stress tolerance in plants.

This review focuses on the cellular and molecular signaling networks and drought coping adaptations in plants to overcome the impact of drought stress. The use of functional genomics to overcome drought stress is also discussed. Furthermore, this review provides an overview of the genetic basis of drought tolerance in cotton, with a focus on QTLs and candidate abiotic stress tolerance genes in cotton, which might be employed for novel cotton breeding in the future.

### 2. Role of TFs in Drought Stress Signaling Pathways

TFs are the principal regulatory elements for many genes involved in environmental stress responses. TFs have vital roles in signaling pathways, from signal reception to the expression of genes related to drought stress in plants. Genes contain cis-acting components in their promoter regions, which serve as binding sites for TFs to regulate gene expression in signal transduction pathways. Signaling cascades in networks responsive to drought stress are activated via TFs that work together to induce drought tolerance [15]. Approximately, 1500 TFs are involved in the expression of stress related genes in Arabidopsis [16]. Several transcription factor families like MYB, WRKY, ERF, NAC, and bZIP have been characterized and shown to be useful tools for enhancing drought tolerance in plants. In recent studies, TFs involved in stress tolerance were identified in cotton and Arabidopsis (Table 1). Overexpression of GhABF2 in cotton enhanced the activities of catalase (CAT) and superoxide dismutase (SOD), and improved yield in transgenic plants [17]. Another TF related to R2R3-type MYB, GbMYB5, responded positively to drought stress [18]. Ectopic expression of the GhWRKY41 gene in tobacco plants led to increased activity of antioxidant enzymes, lower MDA content, increased stomatal closure, and upregulation of antioxidant-related genes [19]. In Gossypium barbadense, a R2R3-type GbMYB5 TF gene enhanced drought tolerance in transgenic tobacco and cotton. These results suggest the involvement of GbMYB5 in adaptive drought stress responses [18]. GhWRKY59 is an important TF that ensures drought tolerance in cotton (**Figure 1**) [20]. In Upland Cotton, a NAM domain gene termed GhNAC79, improves drought tolerance, and also responds to JA and ethylene treatments. Additionally, its overexpression improved stress tolerance in *Arabidopsis* and cotton [21].

Table 1. Key genes involved in abiotic stress signaling in cotton.

Gene	Туре	Phenotypic Effect/Function	Reference
GhHUB2	Histone H2B monoubiquitinatin E3 ligase encoding gene	Drought tolerance through increased soluble sugar, proline, and leaf relative water contents	[22]
GrMAPKKK and GhMAPKKK	MAPK gene family	Drought and salt responsive	[23]
GhMAP3K1, GhMKK4, and GhMPK6	MAPK signaling gene	Regulates the drought stress response by interacting with GhWRKY59-GhDREB2	[20]
GhMKK3	MAPK signaling gene	Enhanced drought tolerance	[24]
GhMAP3K40	MAPK signaling gene	Salt and drought stress tolerance at the germination stage	[25]
GhMPK4	MAPK signaling gene	Increased sensitivity to ABA, salt, and drought	[26]
GhMPK17	MAPK signaling gene	Osmotic and salt stress tolerance	[27]
GbMPK3	MAPK signaling gene	Enhanced oxidative and drought stress tolerance	[28]
GhMPK6a	MAPK signaling gene	Drought and salinity	[29]

Gene	Туре	Phenotypic Effect/Function	Reference
GhMKK1	MAPK signaling gene	Drought and salinity	[30]
GhMKK5	MAPK signaling gene	Drought and salinity	[ <u>31</u> ]
GhMPK2	MAPK signaling gene	Drought and salinity	[ <u>32</u> ]
GbRLK	Receptor-like kinase	Drought and salinity	[33]
GaHDG11 (HD-ZIP)	Transcription factor	Drought and heat stress	[ <u>34]</u>
GhNAC79	Transcription factor	Improves resistance to drought stress	[21]
GhERF38	Transcription factor	Drought, abscisic acid, and salinity	[ <u>35</u> ]
GhERF2, GhERF3, GhERF6	Transcription factor	Drought, salt, ethylene, and abscisic acid	[ <u>36</u> ]
GhWRKY59	Transcription factor	Activates MAPK signaling gene under drought	[ <u>20</u> ]
GhWRKY25	Transcription factor	Drought and salinity	[37]
GhABF2 (bZIP)	Transcription factor	Enhances the activities of CAT and SOD, regulates gene expression related to ABA	[17]
GhNAC2	Transcription factor	Longer roots, and enhanced salt and drought tolerance	[38]
GhCBF3, GhAREB1, and GhAREB2	ABA-induced gene	Small stomatal aperture, enhanced drought- and high salinity-tolerance via the ABA signaling pathway	[39]
GhNAC7-GhNAC13	Transcription factor	Cold, abscisic acid, drought, and salinity	[ <u>40</u> ]
GbMYB5	Transcription factor	Reduced water loss trough stomatal conductance, and increased proline content and antioxidant enzymes	[ <u>41</u> ]
GhWRKY41	Transcription factor	Lower malondialdehyde content, higher antioxidant activity, and induced stomatal conductance	[19]
GhWRKY17	Transcription factor	Increases sensitivity to ABA and drought stress	[ <u>42</u> ]
GhNAC8-GhNAC17	Transcription factor	Drought, salinity, cold, and ABA	[43]
GhNAC1-GhNAC6	Transcription factor	Drought, cold, salinity, and ABA	[44]
GhDREB	Transcription factor	Drought, cold, and salinity	[ <u>45</u> ]
GhDREB1	Transcription factor	Drought, cold, and salinity	[ <u>46</u> ]
GhDBP2	Transcription factor	Drought, cold, and ABA	[ <u>47]</u>
GhERF1	Transcription factor	ABA production and drought stress signaling regulation	[ <u>48</u> ]
GhERF4	Transcription factor	ABA production and drought stress signaling regulation	[ <u>49</u> ]
GhDREB1L	Transcription factor	Drought, cold, and salinity	[ <u>50</u> ]
GhPYL9-11A	ABA receptor gene	ABA receptor that mediates the response to drought stress	[ <u>51</u> ]
GhSnRK2	Involved in ABA signaling	Drought, salinity, cold, and ABA	[ <u>52</u> ]
GhCDPK35, GhCDPK28, GhCDPK16, GhCDPK14, GhCDPK11 and GhCDPK3	Ca <sup>2+</sup> -activated gene	Drought and salinity stress responsive	[7]
GhCIPK6	Ca <sup>2+</sup> -activated gene	Increased drought, salinity, and ABA stress tolerance	[53]
GhD12G207	CDK gene family	Increased concentration of antioxidant enzymes (POD, SOD, and CAT), cell membrane stability, and chlorophyll content under drought and salt stress	[ <u>54]</u>

Gene	пе Туре		Phenotypic Effect/Function	Reference
G	aMYB62L	Transcription factor	Increased chlorophyll and proline contents, higher germination rate under drought salt stress	<u>[55]</u>
C	GhTPS11	Functional gene	Drought, heat, salinity, ABA, and gibberellin acid	[ <u>56</u> ]
	GhAVP1	Functional gene	Drought and salinity tolerance	[ <u>57]</u>

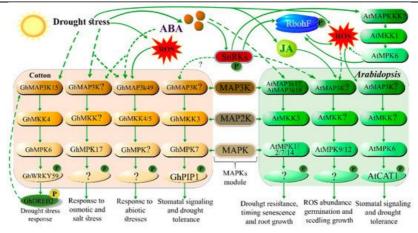


Figure 1. Drought-induced, ABA-dependent, ABA-independent MAPK signaling, and interaction between ABA, ROS, and MAPK signaling under drought stress in plants. ABA-regulates various MAPKs in cotton and Arabidopsis. ABA promotes drought sensing and signaling in plants. The different cascades are represented by different color schemes in the figure. Solid arrow lines denote established signaling mechanisms, while dashed arrow lines denote unestablished signaling pathways. ABA-activated SnRK2s trigger and phosphorylate downstream targets, such as respiratory burst oxidase homolog (RBOH) and various MAPKs. Activation of RBOH induces ROS production. ROS signaling and ABA signaling may overlap with MAPK factors, to interact and regulate drought tolerance. MAP3K17/18-MKK3-MPK1/2/7/14 is an ABAinduced complete MAPK cascade involved in stomatal signaling, senescence, and drought tolerance mechanisms in Arabidopsis. In addition, MKK1 activates MPK6 to positively regulate CATALASE1 (CAT1) for ROS abundance. In cotton, the drought- and ABA-induced MAPK cascade MKK3-MPK7-PIP1 is associated with stomatal signaling and drought tolerance. Another ABA-mediated MAPK module, MAPKKK49-MKK4/MKK5, is associated with abiotic stress responses. GhMPK17 gene is a novel, well-characterized MAPK, which is associated with responses to osmotic and salt stresses in cotton. An ABA-independent and drought-mediated MAPK module (MAP3K15-MKK4-MPK6-WRKY59) regulates drought tolerance in cotton. Drought stress triggers the MAPKKK15 cascade, which phosphorylates the WRKY59 transcriptional factor. Interestingly, WRKY59 binds to the promoter of DREB2 and regulates the expression of drought-sensitive genes. Meanwhile, it positively regulates the expression of MAP3K15 by establishing a feedback loop, which regulates drought tolerance in cotton.

## 3. Cellular and Molecular Responses to Drought Stress in Plants

Drought stress affects plant growth, leaf and stem dry weights, canopy and root growth, plant height, and the number of nodes in plants. Similarly, some physiological properties, such as stomatal conductance, transpiration rate, photosynthetic rate, and water potential decrease under osmotic stress. Finally, osmotic stress limits the accumulation of dry matter by up to 50% under critical water deficiency [12][58]. These traits are potential candidates for drought tolerance in plants. Genetic improvement on the basis of physio-morphological traits is more important because, these traits have vital roles in maintaining a favorable water balance through stomatal closure, reduced transpiration, high water use efficiency, accumulation of proline, trihalose, and polyamines, leaf rolling, wax content, deep root system, and earliness [59].

After successful transduction of signals and sensing the drought stress, plants initiate drought recovery mechanisms through various physio-morphological and biochemical responses (**Figure 2**). Plants have developed various mechanisms to minimize or tolerate multiple stresses. Drought tolerance, drought recovery, drought escape, and drought avoidance are the four important categories of drought tolerance tools  $^{[10]}$ . Tolerant plants subjected to stressful environments adopt an 'escape scenario' by utilizing energy for defense mechanisms, which eventually impacts growth and production. During drought avoidance, plants reduce transpiration and develop deep and vigorous root systems to increase water uptake to help maintain tissue water potential  $^{[60]}$ . Tolerance to drought is the capacity of plants to endure severe dehydration through osmotic adjustment by osmo-protectants  $^{[60][61]}$ . Drought recovery is the ability of plants to restart growth and overcome yield deficits following severe stress. Plants have established numerous morpho-physiological adaptations such as root growth, OA, photosynthetic rate, and stomatal regulation to overcome drought stress (**Figure 2**).

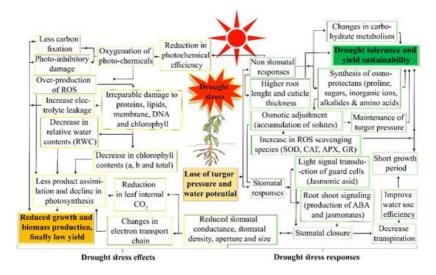


Figure 2. Overall pathways of drought stress effects and plant responses to drought stress.

#### References

- 1. Saranga, Y.; Paterson, A.H.; Levi, A. Bridging Classical and Molecular Genetics of Abiotic Stress Resistance in Cotton. Genet. Genom. Cott. 2009, 3, 337–352.
- 2. Abdelraheem, A.; Esmaeili, N.; O'Connell, M.; Zhang, J. Progress and perspective on drought and salt stress tolerance in cotton. Ind. Crops Prod. 2019, 130, 118–129.
- 3. Ullah, A.; Sun, H.; Yang, X.; Zhang, X. Drought coping strategies in cotton: Increased crop per drop. Plant Biotechnol. J. 2017, 15, 271–284.
- 4. Kazuya Ichimura, M.G.; Ichimura, K.; Shinozaki, K.; Tena, G.; Sheen, J.; Henry, Y.; Champion, A.; Kreis, M.; Zhang, S.; Hirt, H.; et al. Mitogen-activated protein kinase cascades in plants: A new nomenclature. Trends Plant Sci. 2002, 7, 301–308.
- 5. Li, L.; Yu, D.; Zhao, F.; Pang, C.; Song, M.; Wei, H.; Fan, S.; Yu, S. Genome-wide analysis of the calcium-dependent protein kinase gene family in Gossypium raimondii. J. Integr. Agric. 2015, 14, 29–41.
- 6. Dong, T.; Park, Y.; Hwang, I. Abscisic acid: Biosynthesis, inactivation, homoeostasis and signalling. Essays Biochem. 2015, 58, 29–48.
- 7. Yoshida, T.; Mogami, J.; Yamaguchi-Shinozaki, K. ABA-dependent and ABA-independent signaling in response to osmotic stress in plants. Curr. Opin. Plant Biol. 2014, 21, 133–139.
- 8. Danquah, A.; de Zelicourt, A.; Colcombet, J.; Hirt, H. The role of ABA and MAPK signaling pathways in plant abiotic stress responses. Biotechnol. Adv. 2014, 32, 40–52.
- 9. Riemann, M.; Dhakarey, R.; Hazman, M.; Miro, B.; Kohli, A.; Nick, P. Exploring Jasmonates in the Hormonal Network of Drought and Salinity Responses. Front. Plant Sci. 2015, 6, 1–16.
- 10. Fang, Y.; Xiong, L. General mechanisms of drought response and their application in drought resistance improvement in plants. Cell. Mol. Life Sci. 2015, 72, 673–689.
- 11. Chastain, D.R.; Snider, J.L.; Choinski, J.S.; Collins, G.D.; Perry, C.D.; Whitaker, J.; Grey, T.L.; Sorensen, R.B.; van lersel, M.; Byrd, S.A.; et al. Leaf ontogeny strongly influences photosynthetic tolerance to drought and high temperature in Gossypium hirsutum. J. Plant Physiol. 2016, 199, 18–28.
- 12. Hejnák, V.; Tatar; Atasoy, G.D.; Martinková, J.; Çelen, A.E.; Hnilička, F.; Skalický, M. Growth and photosynthesis of upland and pima cotton: Response to drought and heat stress. Plant Soil Environ. 2015, 62, 507–514.
- 13. Long, L.; Guo, D.-D.; Gao, W.; Yang, W.-W.; Hou, L.-P.; Ma, X.-N.; Miao, Y.-C.; Botella, J.R.; Song, C.-P. Optimization of CRISPR/Cas9 genome editing in cotton by improved sgRNA expression. Plant Methods 2018, 14, 85.
- 14. Ashraf, J.; Zuo, D.; Wang, Q.; Malik, W.; Zhang, Y.; Abid, M.A.; Cheng, H.; Yang, Q.; Song, G. Recent insights into cotton functional genomics: Progress and future perspectives. Plant Biotechnol. J. 2018, 16, 699–713.
- 15. Guo, M.; Liu, J.-H.; Ma, X.; Luo, D.-X.; Gong, Z.-H.; Lu, M.-H. The Plant Heat Stress Transcription Factors (HSFs): Structure, Regulation, and Function in Response to Abiotic Stresses. Front. Plant Sci. 2016, 7, 114.
- 16. Lata, C.; Prasad, M. Role of DREBs in regulation of abiotic stress responses in plants. J. Exp. Bot. 2011, 62, 4731–4748.

- 17. Liang, C.; Meng, Z.Z.; Meng, Z.Z.; Malik, W.; Yan, R.; Lwin, K.M.; Lin, F.; Wang, Y.; Sun, G.; Zhou, T.; et al. GhABF2, a bZIP transcription factor, confers drought and salinity tolerance in cotton (Gossypium hirsutum L.). Sci. Rep. 2016, 6, 1–14.
- 18. Chen, T.; Li, W.; Hu, X.; Guo, J.; Liu, A.; Zhang, B. A cotton MYB transcription factor, GbMYB5, is positively involved in plant adaptive response to drought stress. Plant Cell Physiol. 2014, 56, 917–929.
- 19. Chu, X.; Wang, C.; Chen, X.; Lu, W.; Li, H.; Wang, X.; Hao, L.; Guo, X. The Cotton WRKY Gene GhWRKY41 Positively Regulates Salt and Drought Stress Tolerance in Transgenic Nicotiana benthamiana. PLoS ONE 2015, 10, e0143022.
- 20. Li, F.; Li, M.; Wang, P.; Cox, K.L.; Duan, L.; Dever, J.K.; Shan, L.; Li, Z.; He, P. Regulation of cotton (Gossypium hirsutum) drought responses by mitogen-activated protein (MAP) kinase cascade-mediated phosphorylation of GhWRKY59. New Phytol. 2017, 215, 1462–1475.
- 21. Guo, Y.; Pang, C.; Jia, X.; Ma, Q.; Dou, L.; Zhao, F.; Gu, L.; Wei, H.; Wang, H.; Fan, S.; et al. An NAM Domain Gene, GhNAC79, Improves Resistance to Drought Stress in Upland Cotton. Front. Plant Sci. 2017, 8, 1–15.
- 22. Chen, H.; Feng, H.; Zhang, X.; Zhang, C.; Wang, T. An Arabidopsis E3 ligase HUB2 increases histone H2B monoubiquitination and enhances drought tolerance in transgenic cotton. Plant Biotechnol. J. 2019, 17, 556–568.
- 23. Danquah, A.; de Zélicourt, A.; Boudsocq, M.; Neubauer, J.; Frei Dit Frey, N.; Leonhardt, N.; Pateyron, S.; Gwinner, F.; Tamby, J.-P.; Ortiz-Masia, D.; et al. Identification and characterization of an ABA-activated MAP kinase cascade in Arabidopsis thaliana. Plant J. 2015, 82, 232–244.
- 24. Wang, C.; Lu, W.; He, X.; Wang, F.; Zhou, Y.; Guo, X.X.; Guo, X.X. The Cotton Mitogen-Activated Protein Kinase Kinase 3 Functions in Drought Tolerance by Regulating Stomatal Responses and Root Growth. Plant Cell Physiol. 2016, 57, 1629–1642.
- 25. Chen, X.; Wang, J.; Zhu, M.; Jia, H.; Liu, D.; Hao, L.; Guo, X. A cotton Raf-like MAP3K gene, GhMAP3K40, mediates reduced tolerance to biotic and abiotic stress in Nicotiana benthamiana by negatively regulating growth and development. Plant Sci. 2015, 240, 10–24.
- 26. Wang, N.-N.; Zhao, L.; Lu, R.; Li, Y.; Li, X.-B. Cotton mitogen-activated protein kinase4 (GhMPK4) confers the transgenic Arabidopsis hypersensitivity to salt and osmotic stresses. Plant Cell Tissue Organ Cult. 2015, 123, 619–632.
- 27. Zhang, J.; Zou, D.; Li, Y.; Sun, X.; Wang, N.N.; Gong, S.Y.; Zheng, Y.; Li, X.B. GhMPK17, a cotton mitogen-activated protein kinase, is involved in plant response to high salinity and osmotic stresses and ABA signaling. PLoS ONE 2014, 9, 1–12.
- 28. Long, L.; Gao, W.; Xu, L.; Liu, M.; Luo, X.; He, X.; Yang, X.; Zhang, X.; Zhu, L. GbMPK3, a mitogen-activated protein kinase from cotton, enhances drought and oxidative stress tolerance in tobacco. Plant Cell Tissue Organ Cult. 2014, 116, 153–162.
- 29. Li, Y.; Zhang, L.; Wang, X.; Zhang, W.; Hao, L.; Chu, X.; Guo, X. Cotton GhMPK6a negatively regulates osmotic tolerance and bacterial infection in transgenic Nicotiana benthamiana, and plays a pivotal role in development. FEBS J. 2013, 280, 5128–5144.
- 30. Lu, W.; Chu, X.; Li, Y.; Wang, C.; Guo, X. Cotton GhMKK1 Induces the Tolerance of Salt and Drought Stress, and Mediates Defence Responses to Pathogen Infection in Transgenic Nicotiana benthamiana. PLoS ONE 2013, 8, e68503.
- 31. Zhang, L.; Li, Y.; Lu, W.; Meng, F.; Wu, C.; Guo, X. Cotton GhMKK5 affects disease resistance, induces HR-like cell death, and reduces the tolerance to salt and drought stress in transgenic Nicotiana benthamiana. J. Exp. Bot. 2012, 63, 3935–3951.
- 32. Zhang, L.; Xi, D.; Li, S.; Gao, Z.; Zhao, S.; Shi, J.; Wu, C.; Guo, X. A cotton group C MAP kinase gene, GhMPK2, positively regulates salt and drought tolerance in tobacco. Plant Mol. Biol. 2011, 77, 17–31.
- 33. Zhao, J.; Gao, Y.; Zhang, Z.; Chen, T.; Guo, W.; Zhang, T. A receptor-like kinase gene (GbRLK) from Gossypium barbadense enhances salinity and drought-stress tolerance in Arabidopsis. BMC Plant Biol. 2013, 13, 110.
- 34. Chen, E.; Zhang, X.X.; Yang, Z.Z.; Wang, X.; Yang, Z.Z.; Zhang, C.; Wu, Z.; Kong, D.; Liu, Z.; Zhao, G.; et al. Genome-wide analysis of the HD-ZIP IV transcription factor family in Gossypium arboreum and GaHDG11 involved in osmotic tolerance in transgenic Arabidopsis. Mol. Genet. Genom. 2017, 292, 593–609.
- 35. Ma, L.; Hu, L.; Fan, J.; Amombo, E.; Khaldun, A.B.M.; Zheng, Y.; Chen, L. Cotton GhERF38 gene is involved in plant response to salt/drought and ABA. Ecotoxicology 2017, 26, 841–854.
- 36. Jin, L.G.; Li, H.; Liu, J.Y. Molecular Characterization of Three Ethylene Responsive Element Binding Factor Genes from Cotton. J. Integr. Plant Biol. 2010, 52, 485–495.

- 37. Liu, X.; Song, Y.; Xing, F.; Wang, N.; Wen, F.; Zhu, C. GhWRKY25, a group I WRKY gene from cotton, confers differential tolerance to abiotic and biotic stresses in transgenic Nicotiana benthamiana. Protoplasma 2016, 253, 1265–1281.
- 38. Gunapati, S.; Naresh, R.; Ranjan, S.; Nigam, D.; Hans, A.; Verma, P.C.; Gadre, R.; Pathre, U.V.; Sane, A.P.; Sane, V.A. Expression of GhNAC2 from G. herbaceum, improves root growth and imparts tolerance to drought in transgenic cotton and Arabidopsis. Sci. Rep. 2016, 6, 24978.
- 39. Ma, L.-F.; Li, Y.; Chen, Y.; Li, X.-B. Improved drought and salt tolerance of Arabidopsis thaliana by ectopic expression of a cotton (Gossypium hirsutum) CBF gene. Plant Cell Tissue Organ Cult. 2016, 124, 583–598.
- 40. Huang, G.-Q.; Li, W.; Zhou, W.; Zhang, J.-M.; Li, D.-D.; Gong, S.-Y.; Li, X.-B. Seven cotton genes encoding putative NAC domain proteins are preferentially expressed in roots and in responses to abiotic stress during root development. Plant Growth Regul. 2013, 71, 101–112.
- 41. Zhang, X.; Mi, X.; Chen, C.; Wang, H.; Guo, W. Identification on mitogen-Activated protein kinase signaling cascades by integrating protein interaction with transcriptional profiling analysis in cotton. Sci. Rep. 2018, 8, 1–14.
- 42. Yan, H.; Jia, H.; Chen, X.; Hao, L.; An, H.; Guo, X. The Cotton WRKY Transcription Factor GhWRKY17 Functions in Drought and Salt Stress in Transgenic Nicotiana benthamiana Through ABA Signaling and the Modulation of Reactive Oxygen Species Production. Plant Cell Physiol. 2014, 55, 2060–2076.
- 43. Shah, S.T.; Pang, C.; Fan, S.; Song, M.; Arain, S.; Yu, S. Isolation and expression profiling of GhNAC transcription factor genes in cotton (Gossypium hirsutum L.) during leaf senescence and in response to stresses. Gene 2013, 531, 220–234.
- 44. Meng, C.; Cai, C.; Zhang, T.; Guo, W. Characterization of six novel NAC genes and their responses to abiotic stresses in Gossypium hirsutum L. Plant Sci. 2009, 176, 352–359.
- 45. Gao, S.-Q.Q.; Chen, M.; Xia, L.-Q.Q.; Xiu, H.-J.J.; Xu, Z.-S.S.; Li, L.-C.C.; Zhao, C.-P.P.; Cheng, X.-G.G.; Ma, Y.-Z.Z. A cotton (Gossypium hirsutum) DRE-binding transcription factor gene, GhDREB, confers enhanced tolerance to drought, high salt, and freezing stresses in transgenic wheat. Plant Cell Rep. 2009, 28, 301–311.
- 46. Huang, J.-G.; Yang, M.; Liu, P.; Yang, G.-D.; Wu, C.-A.; Zheng, C.-C. GhDREB1 enhances abiotic stress tolerance, delays GA-mediated development and represses cytokinin signalling in transgenic Arabidopsis. Plant Cell Environ. 2009, 32, 1132–1145.
- 47. Huang, B.; Jin, L.; Liu, J.-Y. Identification and characterization of the novel gene GhDBP2 encoding a DRE-binding protein from cotton (Gossypium hirsutum). J. Plant Physiol. 2008, 165, 214–223.
- 48. Qiao, Z.; Huang, B.; Liu, J. Molecular cloning and functional analysis of an ERF gene from cotton (Gossypium hirsutum). Biochim. Biophys. Acta Gene Regul. Mech. 2008, 1779, 122–127.
- 49. Jin, L.G.; Liu, J.Y. Molecular cloning, expression profile and promoter analysis of a novel ethylene responsive transcription factor gene GhERF4 from cotton (Gossypium hirstum). Plant Physiol. Biochem. 2008, 46, 46–53.
- 50. Huang, B.; Jin, L.; Liu, J. Molecular cloning and functional characterization of a DREB1/CBF-like gene (GhDREB1L) from cotton. Sci. China Ser. C Life Sci. 2007, 50, 7–14.
- 51. Liang, C.; Liu, Y.; Li, Y.; Meng, Z.; Yan, R.; Zhu, T.; Wang, Y.; Kang, S.; Ali Abid, M.; Malik, W.; et al. Activation of ABA Receptors Gene GhPYL9-11A Is Positively Correlated with Cotton Drought Tolerance in Transgenic Arabidopsis. Front. Plant Sci. 2017, 8, 1–13.
- 52. Bello, B.; Zhang, X.; Liu, C.; Yang, Z.; Yang, Z.; Wang, Q.; Zhao, G.; Li, F. Cloning of Gossypium hirsutum sucrose non-fermenting 1-related protein kinase 2 gene (GhSnRK2) and its overexpression in transgenic Arabidopsis escalates drought and low temperature tolerance. PLoS ONE 2014, 9, 1–18.
- 53. He, L.; Yang, X.; Wang, L.; Zhu, L.; Zhou, T.; Deng, J.; Zhang, X. Molecular cloning and functional characterization of a novel cotton CBL-interacting protein kinase gene (GhCIPK6) reveals its involvement in multiple abiotic stress tolerance in transgenic plants. Biochem. Biophys. Res. Commun. 2013, 435, 209–215.
- 54. Magwanga, R.O.; Lu, P.; Kirungu, J.N.; Cai, X.; Zhou, Z.; Wang, X.; Diouf, L.; Xu, Y.; Hou, Y.; Hu, Y.; et al. Whole genome analysis of cyclin dependent kinase (CDK) gene family in cotton and functional evaluation of the role of CDKF4 gene in drought and salt stress tolerance in plants. Int. J. Mol. Sci. 2018, 19, 2625.
- 55. Butt, H.I.; Yang, Z.; Chen, E.; Zhao, G.; Gong, Q.; Yang, Z.; Zhang, X.; Li, F. Functional Characterization of Cotton GaMYB62L, a Novel R2R3 TF in Transgenic Arabidopsis. PLoS ONE 2017, 12, e0170578.
- 56. Wang, C.; Zhang, S.; Qi, S.; Zheng, C.; Wu, C. Delayed germination of Arabidopsis seeds under chilling stress by overexpressing an abiotic stress inducible GhTPS11. Gene 2016, 575, 206–212.

- 57. Zhang, H.; Shen, G.; Kuppu, S.; Gaxiola, R.; Payton, P. Creating drought- and salt-tolerant cotton by overexpressing a vacuolar pyrophosphatase gene. Plant Signal. Behav. 2011, 6, 861–863.
- 58. Zafar, S.A.; Patil, S.B.; Uzair, M.; Fang, J.; Zhao, J.; Guo, T.; Yuan, S.; Uzair, M.; Luo, Q.; Shi, J.; et al. DEGENERATED PANICLE AND PARTIAL STERILITY 1 (DPS 1) encodes a cystathionine β-synthase domain containing protein required for anther cuticle and panicle development in rice. New Phytol. 2019, 1, 356–375.
- 59. De Brito, G.G.; Sofiatti, V.; de Andrade Lima, M.M.; de Carvalho, L.P.; da Silva Filho, J.L. Physiological traits for drought phenotyping in cotton Giovani. Acta Sci. Agron. 2011, 33, 117–125.
- 60. Shavrukov, Y.; Kurishbayev, A.; Jatayev, S.; Shvidchenko, V.; Zotova, L.; Koekemoer, F.; de Groot, S.; Soole, K.; Langridge, P. Early Flowering as a Drought Escape Mechanism in Plants: How Can It Aid Wheat Production? Front. Plant Sci. 2017, 8, 1950.
- 61. Luo, L.J. Breeding for water-saving and drought-resistance rice (WDR) in China. J. Exp. Bot. 2010, 61, 3509–3517.

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