

Anti-SARS-CoV-2 effects of plants

Subjects: Plant Sciences

Contributor: Saad Alghamdi

Severe acute respiratory syndrome coronavirus-2 (SARS-CoV-2) has caused a global pandemic and is posing a serious challenge to mankind. As per the current scenario, there is an urgent need for antiviral that could act as a protective and therapeutic against SARS-CoV-2. Plants are a good source of natural medication and an alternative to antibiotic treatment because the excessive use of antibiotics is responsible for drug resistance. Secondly, the main advantage of using plants is that they are economically efficient, have high scalability and safety because the plants can be cultivated in a very low amount, and they also do not support the growth of human pathogens. Plants have produced a large range of antivirals lectins, including griffithsin, cyanovirin and cyanovirin-N-fusion proteins, and also the transgenic rice lines that express griffithsin and cyanovirin-N in the seeds, with or without antibody 2G12. Medicinal plants may be a good source of antivirals, but the accurate dosage and plant parts that have medicinal properties, such as root, shoot, etc., should be known prior to the consumption; otherwise, the adverse effect could also exist.

Keywords: antiviral ; medicinal plants ; SARS-CoV-2

1. SARS-CoV-2

Coronaviruses under the realm of Riboviria belongs to the order Nidovirales, suborder cornidovirineae, family Coronaviridae ^[1]. Orthocoronavirinae the subfamily of coronaviridae is further divided into four genera alpha (a), beta (b), gamma (c), and delta (d) coronavirus. Further, SARS-CoV belongs to Beta coronavirus and sarbecovirus is the subgenus of SARS-CoV-2 ^[2]. It shares similar homology with bat coronavirus and it has been estimated that both have 96.2% sequence homology ^[3].

The two epidemics due to coronavirus have already occurred that were due to SARS-CoV and the Middle East respiratory syndrome (MERS) and both have been the major cause of pneumonia in humans ^[4]. SARS-CoV emerged in 2002 and remained till 2003 during the period it caused 774 death and more than 800 people were infected. MERS-CoV emerged in Saudi Arabia in 2012, which caused more than 800 deaths, and 2500 people became infected ^[5].

Coronaviruses have single-stranded RNA as genetic material that can range from 26 kbs to 32 kbs in length. Coronaviruses possess specific genes in Open Reading Frame ORF1 downstream region that are responsible for encoding the proteins responsible for viral replication, nucleocapsid, and spike formation ^[6]. The structure of SARS-CoV shows the spikes on the outer surface. Furthermore, studies have shown that structural proteins are encoded by the four structural genes, which include spike (S), envelope (E), membrane (M), and nucleocapsid (N) gene. These genes are responsible for viral functionality and structure ^[7].

SARS-CoV-2 genome is 80% similar to the previous human coronavirus SARS-CoV ^[8]. For some of the encoded proteins such as coronavirus main proteinase (3CLpro), papain-like protease (PLpro), and RNA-dependent RNA polymerase (RdRp) the sequence similarity is very high (96%) between SARS-CoV, and SARS-CoV-2 ^[9].

SARS-CoV-2 and SARS-CoV spike protein have 76.5% identical amino acid sequences ^[10]. The 3D structures of spike proteins of SARS-CoV-2 and SARS-CoV as analyzed by computer imaging revealed that both have identical structures in the receptor-binding domain and maintains the Vander Waals force ^[10]. The major amino acid residue of SARS-CoV-2 in the receptor-binding domain is Gln-493, which favors the attachment of spike protein S with human cells more specifically with the lungs therefore the attachment results in respiratory infections in humans ^{[11][12]}. Zhao et al. ^[11] have suggested that 83% of the angiotensin-converting enzyme (ACE-2) expressing cells are alveolar epithelial type II, therefore, these cells can be the reservoir for the viral invasion. Some analysts have suggested that SARS-CoV-2 binds to ACE-2 more efficiently as compared with SARS-CoV, therefore, it has a greater ability to transmit from person to person ^[13].

Initially, for the attachment to the host cells, virus use the spike (S) glycoprotein, which also mediates the host and viral cell membrane fusion during infection ^[14]. The spike (S) glycoprotein has two regions namely S1 and S2. S1 helps in the binding of host cell receptors, and S2 helps in the fusion with the membrane. The receptor-binding domain (RBD) is

located in the S1 region of SARS-CoV and the attachment of the human host cell with the virus is mediated by the RBD protein of the RBD domain with the angiotensin-converting enzyme (ACE-2) as a receptor similar to the one required by SARS-CoV [15][16]. Since SARS-CoV and SARS-CoV-2 are much similar, it is believed that SARS-CoV-2 uses a similar receptor (ACE-2) for entering into human host cells [10][13].

The initial clinical symptoms that appear after SARS-CoV-2 infection are pneumonia, fever, dry cough, headache, dyspnea, and in acute cases leads to respiratory failure and eventually, death occurs [17][18][19]. Although the healing depends upon immunity, pre-existing conditions such as hypertension, diabetes, cardiovascular diseases, or kidney diseases further enhances the severity of the pathogenesis of SARS-CoV-2 [17][19]. Different drugs are available in the market against SARS-CoV-2 but none has shown accurate results. None of the drugs has been approved by FDA against SARS-CoV-2.

Plants have been used as a medicine in the traditional Chinese and Ayurvedic systems [20]. Some of the herbal medicines have safety margins above those of reference drugs, which shows that they can be used against mild SARS-CoV-2 infection [21]. Plants in the form of herbal medicine or dietary components act as immunomodulators and can be a potent antiviral against SARS-CoV-2 infection [22].

Medicinal plants contain diverse secondary metabolites, therefore, they have been the source of drugs against viral, bacterial, and protozoal infection, including cancer [23][24]. The secondary metabolites present in the medicinal plants can interrupt viral proteins and enzymes by binding with them and preventing viral penetration and replication [25][26][27][28]. When consumed without knowing the appropriate dosage, herbal medicines may be toxic [29]. The genetically modified (GM) plants also pose serious health issues because the unnatural change in naturally occurring protein or the metabolic pathways results in toxins or allergens [30]. Therefore, the selection of medicinal plants is also an important aspect.

2. Plants as a Source of Medicine against SARS-CoV-2

2.1. Glycyrrhiza Glabra

Glycyrrhiza glabra belongs to the family Fabaceae and is commonly known as licorice. Dried roots from the plant have a characteristic odor and sweet taste. Glycyrrhizin is the main component isolated from the roots. Glycyrrhizin inhibited 50% hepatitis C virus (HCV) in a dose-dependent manner at a concentration of $14 \pm 2 \mu\text{g}$ [31].

Glycyrrhizin also protects from the influenza A virus and caused a reduction in influenza-infected lung cells [32]. In addition to being effective against hepatitis and influenza virus, it has also shown its effect on SARS CoV [33]. Cinatal et al. [34] showed that glycyrrhizin effectively inhibited the SARS-CoV virus replication at an early stage. They were also of opinion that glycyrrhizin affected cellular signaling pathways such as protein kinase C, casein kinase II, and transcription factors such as protein 1 and nuclear factor B. Furthermore, they also concluded the upregulation of nitrous oxide that inhibits the virus.

Sinha et al. [35] through in silico approach showed that the replication process of SARS-CoV-2 was affected by the two components of licorice (glyasperin A and glycyrrhizic acid). They concluded that the ACE-2 receptor of the virus was disturbed by glycyrrhizic acid and the replication process was inhibited by glyasperin A (Table 1).

Table 1. Plants and their activity against different viral diseases.

Plants	Family	Plant Part	References
<i>Glycyrrhiza glabra</i>	Fabaceae	Roots	Hepatitis C [31] Influenza-A [32] SARS-CoV [34] SARS-CoV-2 [35]
<i>Alnus japonica</i>	Betulaceae	Leaves and bark	Influenza [36] H9N2 Avian influenza virus [37] SARS CoV [38]
<i>Allium sativum</i>	Amaryllidaceae	Bulb	Influenza A and B [39] Cytomegalovirus [40] Herpes simplex virus 1 [41] Herpes simplex virus 2 [42] SARS CoV [43]

Plants	Family	Plant Part	References
<i>Houttuynia cordata</i>	Saururaceae	Leaves, roots	DEN V-2 ^{[44][45]} Avian infection bronchitis virus ^[46] Influenza-A ^[47] Herpes simplex virus-1 ^[48] SARS CoV ^[49]
<i>Lycoris radiata</i>	Amaryllidaceae	Bulb, stem cortex	H5N1 ^[50] Hepatitis C ^[51] Tobacco mosaic virus ^[52] SARS CoV ^[53]
<i>Tinospora cordifolia</i>	Menispermaceae	Stem	HSV-1 ^[54] HIV ^[55] SARS-CoV ^[56]
<i>Vitex trifolia</i>	Verbenaceae	Leaves	HIV ^{[57][58]} SARS-CoV ^{[59][60]}

2.2. *Alnus Japonica*

Alnus japonica, also known as the East Asian alder tree, grows approximately up to 22 m. It is deciduous with a very fast growth rate. It belongs to the family Betulaceae, also known as the birch family, which includes six genera having nut-bearing deciduous trees and shrubs. The extract of leaves and bark of *Alnus japonica* have been used as food that boosts immunity against influenza ^[36]. Studies have shown that methanolic extract of *Alnus japonica* bark possesses strong antiviral activity against the H9N2 subtype avian influenza virus ^[37]. Further, the chromatographic separation revealed the presence of four lupine type triterpenes and one steroid ^[37].

Diarylheptanoids isolated from *Alnus japonica* showed inhibitory activity against papain-like protease (required for replication of SARS-CoV) ^[38]. Nine diarylheptanoids were isolated, namely, platyphyllenone, hirsutenone, hirsutanonol, oregonin, rubranol, rubranoside B, and rubranoside A. Furthermore, hirsutenone, hirsutanonol, oregonin, rubranol, rubranoside B, and rubranoside A showed dose-dependent inhibitory activity against the papain-like protease. They also showed that hirsutenone possessed the most potent inhibitory activity against papain-like protease. Furthermore, the analysis of Park et al. ^[38] showed that catechol and α,β -unsaturated carbonyl moiety were present in the molecule, which are the key requirements for SARS-CoV cysteine protease inhibition.

Furthermore, Kwon et al. ^[61] described that *Alnus japonica* extract is useful for prevention from viral diseases, such as influenza of humans and of other mammalian and avian species. They also said that the extract exhibits very low toxicity in normal cell conditions but has high antiviral activity. Lastly, the extract from *Alnus japonica* could be effectively used in foods and pharmaceutical industries against the influenza virus.

2.3. *Allium Sativum*(Garlic)

Allium sativum belongs to the Amaryllidaceae family. It is a perennial flowering plant that grows from a bulb and comprises a tall flowering stem that attains a height of upto 1 m. *Allium sativum* is commonly called garlic, which is also an ingredient in Indian food. A peculiar smell can be noticed from the bulb of garlic, which is due to the presence of sulfur compounds. In addition to possessing antibacterial activity, *Allium sativum* (garlic) also has antiviral properties. *Allium sativum* extract has an inhibitory effect on avian infectious bronchitis virus, which is a single-stranded RNA and belongs to coronaviruses. It has also shown its effect on other viruses including influenza A and B ^[39], cytomegalovirus ^[40], herpes simplex virus 1 ^[41], and herpes simplex virus-2 ^[42].

Molecular docking studies have revealed that garlic can inhibit SARS-CoV-2 ^[43]. Molecular docking has shown that 17 organosulfur compounds from garlic essential oil can effectively interact with the amino acid of angiotensin-converting enzyme (ACE-2), a receptor in the human body and main protease (PDB6LU7) of SARS-CoV-2. The highest efficacy was observed by allyl disulfide and allyl trisulfide. Furthermore, it was predicted that garlic essential oil or extract can be used as a source of antiviral against SARS-CoV-2 ^[43].

2.4. *Houttuynia Cordata*

Houttuynia cordata belongs to the family Saururaceae and is mostly confined to the moist habitat. It is an aromatic medicinal plant that has a creeping rootstock. Ethyl acetate extract of *Houttuynia cordata* is effective in inhibiting the infectivity of the dengue virus (DENV). Some workers have shown the inhibition of DENV-2 by ethyl acetate extract of *Houttuynia cordata* ^{[44][45]}. *Houttuynia cordata* extract also inhibited the avian infectious bronchitis virus ^[46]. Quercetin

3-rhamnoside from *Houttuynia cordata* showed anti-influenza (inf-A) activity and inhibited virus replication in the initial stages of infection [47]. Stem distillate from the *Houttuynia cordata* has shown inhibitory activity against herpes simplex virus-1, influenza virus, and human immunodeficiency virus-1 without any cytotoxicity [48]. The three major components from the distillate were methyl *n*-nonyl ketone, lauryl aldehyde, and capryl aldehyde.

Houttuynia cordata showed significant inhibition on SARS-CoV. The *Houttuynia cordata* (HC) extract successfully inhibited the SARS-CoV3C such as protease (3CLpro) and RNA-dependent RNA polymerase (RdRp) [49]. Furthermore, it was also observed that *Houttuynia cordata* extract was non-toxic to animal models at an oral administration dose of 16 g/kg. Through flow cytometric analysis, Lau et al. [49] showed that *Houttuynia cordata* extract in mice model significantly increases the CD4⁺ and CD8⁺ T cells with a significant increase in the secretion of IL-2 and IL-10 cytokine in mouse splenic lymphocytes.

2.5. Lycoris Radiata

Lycoris radiata, which is commonly known as red spider lily, hell flower, red magic lily, or sometimes called equinox flower, belongs to the family Amaryllidaceae. *Lycoris radiata* has shown its effect against H5N1 [50]. Lycorine the active component of *Lycoris radiata* successfully inhibited the influenza virus, which is responsible for avian influenza infection [62]. Proteomic analysis revealed that lycorine alters protein expression in avian influenza virus-infected cells and the treatment with lycorine also decreased the levels of nuclear pore complex protein 93(Nup93, E2RSV7), which is a part of nuclear-cytoplasmic transport [62]. Lycoricidine derivatives obtained from the bulb of *Lycoris radiata* have also shown the anti-hepatitis C virus activity [51]. Furthermore, it has also been observed that Lycoricidine derivatives possess inhibitory activity against the tobacco mosaic virus [52].

Ethanol extract of the stem cortex of the plant successfully inhibited the SARS-CoV. [53]. The active component that probably inhibited the SARS-CoV was lycorine (C16H17NO4), which was analyzed by high-performance liquid chromatography (HPLC). Although the mechanism of action of lycorine on the SARS-CoV is still unclear, it has shown promising results as an antiviral. Li et al. [53] also showed that commercially available lycorine inhibited the SARS-CoV.

2.6. Tinospora Cordifolia

Tinospora cordifolia, commonly called Guduchi or Giloy, belongs to the family Menispermaceae. The plant is an herbaceous vine that bears heart-shaped leaves. *Tinospora cordifolia* extract has shown an increase in IFN- α , IL-1, IL-2, and IL-4 levels in the peripheral blood mononuclear cells in the chickens infected with infectious bursal disease virus, which causes infectious bursal disease [63]. Further, the reduction in the mortality rate of chickens treated with *Tinospora cordifolia* extract was also observed when compared with uninfected chickens [63].

The stem extract of *Tinospora cordifolia* has antibacterial, antifungal, and antiviral properties. The stem extract of *Tinospora cordifolia* successfully inhibited the herpes simplex virus by 61.43% in the Vero cell line [54]. Kalikar et al. [55] showed that 60% of the patients who received *Tinospora cordifolia* extract and 20% on placebo showed a decrease in the incidence of various symptoms associated with HIV. However, they did not study all the parameters but hypothesized that *Tinospora cordifolia* extract could be used as an adjunct to HIV management. The in silico approach against SARS-CoV-2 was tested by Sagar and Kumar [56]. The key targets they considered for the docking were surface glycoprotein, receptor-binding domain, RNA-dependent RNA polymerase, and protease. The natural compounds from *Tinospora cordifolia* such as berberine, isocolumbin, magnoflorine, and tinocordiside showed high binding efficacy against all four key targets [56].

2.7. Vitex Trifolia

Vitex trifolia belongs to the family Verbenaceae. The leaves of the plants are compound and oppositely arranged with three linear leaflets ranges between 1–12 cm in length. The upper surface of the leaf is mostly green, whereas the lower surface is grayish.

Fruits of the plants contain essential oils, monoterpenes, diterpenes, beta-sitosterol [64][65][66][67][57][58][68]. The leaves and bark of the plant contain essential oils, flavones, and artemetin [67]. Furthermore, the essential oil of *Vitex trifolia* is used as an anti-inflammatory and against headache, cold, cough, liver disorders, and HIV [57][58]. Extract from the leaves of *vitex trifolia* inhibited TNF- α and IL-1 β production in human U937 macrophages [68]. The components that were isolated from leaf extract were artemetin, casticin, vitexilactone, and maslinic acid. Wee et al. [68] also suggested that artemetin could be the active compound that suppressed TNF- α and IL-1 β production.

Due to the high contents of the phenolic compounds such as phenolic acid, flavones, and flavonols, it was analyzed that methanolic extract contains antioxidant and anticancer properties [69]. *Vitex trifolia* has also been found to block the NK-kB pathways that reduce inflammatory cytokines; this is a similar pathway that is being used in respiratory distress in SARS-CoV [59][60].

References

1. Gorbalenya, A.E.; Baker, S.C.; Baric, R.S.; de Groot, R.J.; Drosten, C.; Gulyaeva, A.A.; Haagmans, B.L.; Lauber, C.; Leontovich, A.M.; Neuman, B.W.; et al. The species severe acute respiratory syndrome-related coronavirus: Classifying 2019-nCoV and naming it SARS-CoV-2. *Nat. Microbiol.* 2020, 5, 536–544.
2. Ansari, M.A.; Almatroudi, A.; Alzohairy, M.A.; AlYahya, S.; Alomary, M.N.; Al-Dossary, H.A.; Alghamdi, S. Lipid-based nano delivery of Tat-peptide conjugated drug or vaccine—promising therapeutic strategy for SARS-CoV-2 treatment. *Expert Opin. Drug Deliv.* 2020, 17, 1671–1674.
3. Chan, J.F.W.; Kok, K.H.; Zhu, Z.; Chu, H.; To, K.K.W.; Yuan, S.; Yuen, K.Y. Genomic characterization of the 2019 novel human-pathogenic coronavirus isolated from a patient with atypical pneumonia after visiting Wuhan. *Emerg. Microbes Infect.* 2020, 9, 221–236.
4. Song, Z.; Xu, Y.; Bao, L.; Zhang, L.; Yu, P.; Qu, Y.; Zhu, H.; Zhao, W.; Han, Y.; Qin, C. From SARS to MERS, thrusting coronaviruses into the spotlight. *Viruses* 2019, 11, 59.
5. Zaki, A.M.; Van Boheemen, S.; Bestebroer, T.M.; Osterhaus, A.D.; Fouchier, R.A. Isolation of a novel coronavirus from a man with pneumonia in Saudi Arabia. *N. Engl. J. Med.* 2012, 367, 1814–1820.
6. van Boheemen, S.; de Graaf, M.; Lauber, C.; Bestebroer, T.M.; Raj, V.S.; Zaki, A.M.; Osterhaus, A.D.M.E.; Haggmans, B.L.; Gorbalenya, A.E.; Snijder, E.J.; et al. Genomic characterization of a newly discovered coronavirus associated with acute respiratory distress syndrome in humans. *MBio* 2012, 3, e00473-12.
7. Schoeman, D.; Fielding, B.C. Coronavirus envelope protein: Current knowledge. *Virology* 2019, 16, 69.
8. Ansari, M.A.; Jamal, Q.M.S.; Rehman, S.; Almatroudi, A.; Alzohairy, M.A.; Alomary, M.N.; Tripathi, T.; Alharbi, A.H.; Adil, S.F.; Khan, M.; et al. TAT-peptide conjugated repurposing drug against SARS-CoV-2 main protease (3CLpro): Potential therapeutic intervention to combat COVID-19. *Arab. J. Chem.* 2020, 13, 8069–8079.
9. Chan, J.F.-W.; Yuan, S.; Kok, K.-H.; To, K.K.-W.; Chu, H.; Yang, J.; Xing, F.; BNurs, J.L.; Yip, C.C.-Y.; Poon, R.W.-S.; et al. A familial cluster of pneumonia associated with the 2019 novel coronavirus indicating person-to-person transmission: A study of a family cluster. *Lancet* 2020, 395, 514–523.
10. Xu, X.; Chen, P.; Wang, J.; Feng, J.; Zhou, H.; Li, X.; Zhong, W.; Hao, P. Evolution of the novel coronavirus from the ongoing Wuhan outbreak and modeling of its spike protein for risk of human transmission. *Sci. China Life Sci.* 2020, 63, 457–460.
11. Zhao, Y.; Zhao, Z.; Wang, Y.; Zhou, Y.; Ma, Y.; Zuo, W. Single-cell RNA expression profiling of ACE2, the putative receptor of Wuhan 2019-nCoV. *Am. J. Respir. Crit. Care Med.* 2020, 202, 756–759.
12. Yin, Y.; Wunderink, R.G. MERS, SARS and other coronaviruses as causes of pneumonia. *Respirology* 2018, 23, 130–137.
13. Wan, Y.; Shang, J.; Graham, R.; Baric, R.S.; Li, F. Receptor recognition by novel coronavirus from Wuhan: An analysis based on decade-long structural studies of SARS. *J. Virol.* 2020, 94, e00127-20.
14. Li, F. Structure, Function, and Evolution of Coronavirus Spike Proteins. *Annu. Rev. Virol.* 2016, 3, 237–261.
15. Song, W.; Gui, M.; Wang, X.; Xiang, Y. Cryo-EM structure of the SARS coronavirus spike glycoprotein in complex with its host cell receptor ACE2. *PLoS Pathog.* 2018, 14, e1007236.
16. Li, W.; Moore, M.J.; Vasilieva, N.; Sui, J.; Wong, S.K.; Berne, M.A.; Somasundaran, M.; Sullivan, J.L.; Luzuriaga, K.; Greenough, T.C.; et al. Angiotensin-converting enzyme 2 is a functional receptor for the SARS coronavirus. *Nature* 2003, 426, 450–454.
17. Gralinski, L.E.; Menachery, V.D. Return of the coronavirus: 2019-nCoV. *Viruses* 2020, 12, 135.
18. Zhou, P.; Yang, X.L.; Wang, X.G.; Hu, B.; Zhang, L.; Zhang, W.; Si, H.-R.; Zhu, Y.; Li, B.; Huang, C.-L.; et al. A pneumonia outbreak associated with a new coronavirus of probable bat origin. *Nature* 2020, 579, 270–273.
19. Huang, C.; Wang, Y.; Li, X.; Ren, L.; Zhao, J.; Hu, Y. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *Lancet* 2020, 395, 497–506.
20. Jaiswal, Y.; Liang, Z.; Zhao, Z. Botanical drugs in Ayurveda and traditional Chinese medicine. *J. Ethnopharmacol.* 2016, 194, 245–259.

21. Silveira, D.; Prieto-Garcia, J.M.; Boylan, F.; Estrada, O.; Fonseca-Bazzo, Y.M.; Jamal, C.M.; Magalhães, P.O.; Pereira, E.O.; Tomczyk, M.; Heinrich, M. COVID-19: Is There Evidence for the Use of Herbal Medicines as Adjuvant Symptomatic Therapy? *Front. Pharmacol.* 2020, 11, 581840.
22. Panyod, S.; Ho, C.-T.; Sheen, L.-Y. Dietary therapy and herbal medicine for COVID-19 prevention: A review and perspective. *J. Trad. Complement. Med.* 2020, 10, 420–427.
23. Gurib-Fakim, A. Medicinal plants: Traditions of yesterday and drugs of tomorrow. *Mol. Asp. Med.* 2006, 27, 1–93.
24. Harvey, A.L. Natural products as a screening resource. *Curr. Opin. Chem. Biol.* 2007, 11, 480–484.
25. Li, T.; Peng, T. Traditional Chinese herbal medicine as a source of molecules with antiviral activity. *Antivir. Res.* 2013, 97, 1–9.
26. Arbab, A.H.; Parvez, M.K.; Al-Dosari, M.S.; Al-Rehaily, A.J. In vitro evaluation of novel antiviral activities of 60 medicinal plants extracts against hepatitis B virus. *Exp. Ther. Med.* 2017, 14, 626–634.
27. Akram, M.; Tahir, I.M.; Shah, S.M.A.; Mahmood, Z.; Altaf, A.; Ahmad, K.; Munir, N.; Daniyal, M.; Nasir, S.; Mehboob, H. Antiviral potential of medicinal plants against HIV, HSV, influenza, hepatitis, and coxsackievirus: A systematic review. *Phytother. Res.* 2018, 32, 811–822.
28. Dhama, K.; Karthik, K.; Khandia, R.; Munjal, A.; Tiwari, R.; Rana, R.; Khurana, S.K.; Ullah, S.; Khan, R.U.; Alagawany, M. Medicinal and therapeutic potential of herbs and plant metabolites/extracts countering viral pathogens-current knowledge and future prospects. *Curr. Drug Metab.* 2018, 19, 236–263.
29. Fatima, N.; Nayeem, N. Toxic effects as a result of herbal medicine intake. In *Toxicology—New Aspects to This Scientific Conundrum*; Intech Open: London, UK, 2016; pp. 193–204.
30. Fagan, J.; Antoniou, M.; Robinson, C. *GMO Myths and Truths*; Earth Open Source: London, UK, 2014.
31. Ashfaq, U.A.; Masoud, M.S.; Nawaz, Z.; Riazuddin, S. Glycyrrhizin as antiviral agent against Hepatitis C Virus. *J. Transl. Med.* 2011, 9, 112.
32. Wolkerstorfer, A.; Kurz, H.; Bachhofner, N.; Szolar, O.H. Glycyrrhizin inhibits influenza A virus uptake into the cell. *Antivir. Res.* 2009, 83, 171–178.
33. Jovic, T.H.; Ali, S.R.; Ibrahim, N.; Jessop, Z.M.; Tarassoli, S.P.; Dobbs, T.D.; Holford, P.; Thornton, C.A.; Whitaker, I.S. Could Vitamins Help in the Fight Against COVID-19? *Nutrients* 2020, 12, 2550.
34. Cinatl, J.; Morgenstern, B.; Bauer, G.; Chandra, P.; Rabenau, H.; Doerr, H.W. Glycyrrhizin, an active component of liquorice roots, and replication of SARS-associated coronavirus. *Lancet* 2003, 361, 2045–2046.
35. Sinha, S.K.; Prasad, S.K.; Islam, M.A.; Gaurav, S.S.; Patil, R.B.; Al-Faris, N.A.; Aldayel, T.S.; AlKehayez, N.M.; Wabaidur, S.M.; Shakya, A. Identification of bioactive compounds from *Glycyrrhiza glabra* as possible inhibitor of SARS-CoV-2 spike glycoprotein and non-structural protein-15: A pharmacoinformatics study. *J. Biomol. Struct. Dyn.* 2020, 1–15.
36. Ra, J.C.; Kwon, H.J.; Kim, B.G.; You, S.H. Anti-Influenza Viral Composition Containing Bark or Stem Extract of *Alnus japonica*. US Patent US 2010/0285160 A1, 11 November 2010.
37. Tung, N.H.; Kwon, H.J.; Kim, J.H.; Ra, J.C.; Kim, J.A.; Kim, Y.H. An anti-influenza component of the bark of *Alnus japonica*. *Arch. Pharm. Res.* 2010, 33, 363–367.
38. Park, J.Y.; Jeong, H.J.; Kim, J.H.; Kim, Y.M.; Park, S.J.; Kim, D.; Park, K.H.; Lee, W.S.; Ryu, Y.B. Diarylheptanoids from *Alnus japonica* inhibit papain-like protease of severe acute respiratory syndrome coronavirus. *Biol. Pharm. Bull.* 2012, 35, 2036–2042.
39. Fenwick, G.R.; Hanley, A.B. *Allium* species poisoning. *Vet. Rec.* 1985, 116, 28.
40. Meng, Y.; Lu, D.; Guo, N.; Zhang, L.; Zhou, G. Anti-HCMV effect of garlic components. *Virol. Sin.* 1993, 8, 147–150.
41. Tsai, Y.; Cole, L.L.; Davis, L.E.; Lockwood, S.J.; Simmons, V.; Wild, G.C. Antiviral properties of garlic: In vitro effects on influenza B, herpes simplex and coxsackie viruses. *Planta Med.* 1985, 5, 460–461.
42. Weber, N.D.; Andersen, D.O.; North, J.A.; Murray, B.K.; Lawson, L.D.; Hughes, B.G. In vitro virucidal effects of *Allium sativum* (garlic) extract and compounds. *Planta Med.* 1992, 58, 417–423.
43. Thuy, B.T.P.; My, T.T.A.; Hai, N.T.T.; Hieu, L.T.; Hoa, T.T.; Loan, H.T.T.P.; Triet, N.T.; Anh, T.T.V.; Quy, P.T.; Tat, P.V.; et al. Investigation into SARS-CoV-2 Resistance of Compounds in Garlic Essential Oil. *ACS Omega* 2020, 5, 8312–8320.
44. Chiow, K.H.; Phoon, M.C.; Putti, T.; Tan, B.K.; Chow, V.T. Evaluation of Antiviral Activities of *Houttuynia Cordata* Thunb. Extract, Quercetin, Quercetrin and Cinanserin on Murine Coronavirus and Dengue Virus Infection. *Asian Pac. J. Trop. Med.* 2016, 9, 1–7.

45. Xie, M.L.; Phoon, M.C.; Dong, S.X.; Tan, B.K.H.; Chow, V.T. *Houttuynia cordata* extracts and constituents inhibit the infectivity of dengue virus type 2 in vitro. *Int. J. Integr. Biol.* 2013, 14, 78–85.
46. Yin, J.; Li, G.; Li, J.; Yang, Q.; Ren, X. In vitro and in vivo effects of *Houttuynia cordata* on infectious bronchitis virus. *Avian Pathol.* 2011, 40, 491–498.
47. Choi, H.J.; Song, J.H.; Park, K.S.; Kwon, D.H. Inhibitory effects of quercetin 3-rhamnoside on influenza A virus replication. *Eur. J. Pharm. Sci.* 2009, 37, 329–333.
48. Hayashi, K.; Kamiya, M.; Hayashi, T. Virucidal effects of the steam distillate from *Houttuynia cordata* and its components on Hsv-1, Influenza-Virus, and HIV. *Planta Med.* 1995, 61, 237–241.
49. Lau, K.M.; Lee, K.M.; Koon, C.M.; Cheung, C.S.; Lau, C.P.; Ho, H.M.; Lee, M.Y.-H.; Au, S.W.-N.; Cheng, C.H.-K.; Lau, C.B.-S.; et al. Immunomodulatory and anti-SARS activities of *Houttuynia cordata*. *J. Ethnopharmacol.* 2008, 118, 79–85.
50. He, J.; Qi, W.B.; Wang, L.; Tian, J.; Jiao, P.R.; Liu, G.Q.; Ye, W.-C.; Liao, M. Amaryllidaceae alkaloids inhibit nuclear-to-cytoplasmic export of ribonucleoprotein (RNP) complex of highly pathogenic avian influenza virus H5N1. *Influenza Other Respir. Viruses* 2012, 7, 922–931.
51. Chen, D.Z.; Jiang, J.D.; Zhang, K.Q.; He, H.P.; Di, Y.T.; Zhang, Y.; Cai, J.-Y.; Wang, L.; Li, S.-L.; Yi, P.; et al. Evaluation of anti-HCV activity and SAR study of (+)-lycoricidine through targeting of host heat-stress cognate 70 (Hsc70). *Bioorg. Med. Chem. Lett.* 2013, 23, 2679–2682.
52. Yang, D.Q.; Chen, Z.R.; Chen, D.Z.; Hao, X.J.; Li, S.L. Anti-TMV Effects of Amaryllidaceae Alkaloids Isolated from the Bulbs of *Lycoris radiata* and Lycoricidine Derivatives. *Nat. Prod. Bioprospect.* 2018, 8, 189–197.
53. Li, S.Y.; Chen, C.; Zhang, H.Q.; Guo, H.Y.; Wang, H.; Wang, L. Identification of natural compounds with antiviral activities against SARS-associated coronavirus. *Antivir. Res.* 2005, 67, 18–23.
54. Pruthvish, R.; Gopinatha, S.M. Antiviral prospective of *Tinospora cordifolia* on HSV-1. *Int. J. Curr. Microbiol. Appl. Sci.* 2018, 7, 3617–3624.
55. Kalikar, M.V.; Thawani, V.R.; Varadpande, U.K.; Sontakke, S.D.; Singh, R.P.; Khiyani, R.K. Immunomodulatory effect of *Tinospora cordifolia* extract in human immuno-deficiency virus positive patients. *Indian J. Pharmacol.* 2008, 40, 107–110.
56. Sagar, V.; Kumar, A.H.S. Efficacy of natural compounds from *Tinospora cordifolia* against SARS-CoV-2 protease, surface glycoprotein and RNA polymerase. *BEMS Rep.* 2020, 6, 6–8.
57. Devi, W.R.; Singh, C.B. Chemical composition, anti-dermatophytic activity, antioxidant and total phenolic content within the leaves essential oil of *Vitex trifolia*. *Int. J. Phytocosmetics Nat. Ingrid.* 2014, 1, 5–10.
58. Suksamrarn, A.; Werawattanamet, K.; Brophy, J.J. Variation of essential oil constituents in *Vitex trifolia* species. *FlavourFragr. J.* 1991, 6, 97–99.
59. Alam, G.; Wahyuono, S.; Ganjar, I.G.; Hakim, L.; Timmerman, H.; Verpoorte, R. Tracheospasmodic activity of viteosin-A and vitexicarpin isolated from *Vitex trifolia*. *Planta Med.* 2002, 68, 1047–1049.
60. Srivastava, R.A.K.; Mistry, S.; Sharma, S. A novel anti-inflammatory natural product from *Sphaeranthus indicus* inhibits expression of VCAM1 and ICAM1, and slows atherosclerosis progression independent of lipid changes. *Nutr. Metab.* 2015, 12, 20.
61. Kwon, J.H.; Cho, S.H.; Kim, S.J.; Ahn, Y.J.; Ra, J.C. Antiviral Composition Comprising *Alnus Japonica* Extracts. US Patent US 2010/0189827 A1, 29 July 2010.
62. Yang, L.; Zhang, J.H.; Zhang, X.L.; Lao, G.J.; Su, G.M.; Wang, L.; Li, Y.L.; Ye, W.C.; He, J. Tandem mass tag-based quantitative proteomic analysis of lycorine treatment in highly pathogenic avian influenza H5N1 virus infection. *Peer J.* 2019, 7, e7697.
63. Sachan, S.; Dhama, K.; Latheef, S.K.; Samad, H.A.; Mariappan, A.K.; Munuswamy, P.; Singh, R.; Singh, K.P.; Malik, Y.S.; Singh, R.K. Immunomodulatory Potential of *Tinosporacordifolia* and CpG ODN (TLR21 Agonist) against the Very Virulent, Infectious Bursal Disease Virus in SPF Chicks. *Vaccines (Basel)* 2019, 7, 106.
64. Cousins, M.M.; Briggs, J.; Gresham, C.; Whetstone, J. Beach *Vitex* (*Vitex rotundifolia*) an invasive coastal species. *Invasive Plant Sci. Manag. (Weed Sci. Soc. Am.)* 1991, 6, 101–103.
65. Zeng, X.; Fang, Z.; Wu, Y.; Zhang, H. Chemical constituents of the fruits of *Vitex trifolia* L. *Zhongguo Zhong Yao Za Zhi* 1996, 21, 167–168.
66. Ono, M.; Sawamura, H.; Ito, Y.; Mizuki, K.; Nohara, T. Diterpenoids from the fruits of *Vitex trifolia*. *Phytochemistry* 2000, 55, 873–877.

67. Chan, E.W.C.; Baba, S.; Chan, H.T.; Kainumaz, M.; Tangah, J. Medicinal plants of sandy shores a short review on *V. trifolia* L. and *lomoea pes caprae*. *Indian J. Nat. Prod. Resour.* 2016, 7, 107–115.
 68. Wee, H.N.; Neo, S.Y.; Singh, D.; Yew, H.C.; Qiu, Z.Y.; Tsai, X.R.C.; How, S.Y.; Yip, K.-Y.C.; Tan, C.-H.; Koh, H.-L. Effects of *Vitex trifolia* L. Leaf Extracts and Phytoconstituents on Cytokine Production in Human U937 Macrophages. *BMC Complement. Med. Ther.* 2020, 20, 91.
 69. Yao, J.L.; Fang, S.M.; Liu, R.; Oppong, M.B.; Liu, E.W.; Fan, G.W.; Zhang, H. A Review on the Terpenes from Genus *Vitex*. *Molecules* 2016, 21, 1179.
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