

Neurotropic Black Yeast *Exophiala dermatitidis*

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The neurotropic and extremophilic black yeast *Exophiala dermatitidis* (*Herpotrichiellaceae*) inhabits diverse indoor environments, in particular bathrooms, steam baths and dishwashers. It can grow at human body temperature, assimilate cyclic hydrocarbons and human neurotransmitters. Accordingly, they are capable to grow in artificial and natural environments, including synthetic/rubber hydrocarbon-rich materials. Their polymorphic nature allows them to survive environmental stress, such as UV radiation, high temperatures, pH fluctuations, low water activity and others. *E. dermatitidis* is known as extremely plastic which has evolutionarily led to adaptation on the human body. It causes numerous infections in almost all human organs, and may also be associated with Alzheimer's disease.

Keywords: neurotropic fungi ; black yeasts ; neurotropism ; hydrocarbons ; extracellular vesicles ; Alzheimer's disease ; *Exophiala dermatitidis*

1. Introduction

Polyphyletic black yeasts, including the genus *Exophiala*, are melanised yeast-like fungi that populate extreme environments dominated by high or low temperatures, high salinity, aridity, low water activity, high UV radiation, fluctuating pH, and oligotrophic conditions [1][2][3][4][5][6][7][8]. They have a distinct extremophilic ecotype, characterized by thick, melanised cell walls, as well as slow, polymorphic growth including hyphae, yeast cells, meristematic clumps, and endoconidiation. Many black yeasts, in particular those within *Chaetothyriales*, fam. *Herpotrichiellaceae*, use the above adaptations and their unusual ability to grow at 37 °C to invade the human body, another extreme environment for fungi [3].

2. Specifics

Exophiala dermatitidis is the most clinically important and thermotolerant species of the genus *Exophiala*. It can cause various medical conditions, from cutaneous and subcutaneous infections to systemic, gastrointestinal, pulmonary, and neurotropic infections [9]. It has been isolated from the ears, sinuses, lungs, mucus of cystic fibrosis patients, blood and different catheters, and most importantly brain infections [3][10][11][12][13][14][15][16][17][18][19][20][21]. Occasionally, it can cause mildly invasive systemic infections that are associated with significant morbidity and mortality. Typical infections are seen in immunocompromised hosts such as transplant recipients, oncology, and pediatric patients, where it manifests itself as a subcutaneous disease and rarely as deep mycoses [9][22][23][24][25][26]. Particularly in East Asia, *E. dermatitidis* was detected in disseminated and neurotropic infections with high mortality [11][27]. Note, however, that despite infections occurring in apparently healthy humans, some authors concluded that *E. dermatitidis* and related species are opportunists rather than pathogens [28][29][30].

Its pathogenicity is probably owing to various virulence factors, including melanin pigmentation, thermotolerance, and polymorphism [9]. Melanin accumulates in the cell wall of *E. dermatitidis*. This has a protective effect against harmful substances and oxidative stress from the environment or the host cell [31]. *Exophiala* species synthesize melanin endogenously from acetate via the pentaketide pathway, leading to 1,8-dihydroxynaphthalene melanin. This pathway can be constrained under controlled growth conditions by addition of inhibitors, thus allowing the study of melanin's relations to morphology, physiology, and pathogenicity [32][33][34].

The polymorphic character and melanin pigmentation of *E. dermatitidis* enable its colonisation under stress conditions. *Exophiala dermatitidis* has adapted to human-made indoor environments, such as steam baths, saunas, public baths [35][36], drainpipes, and drinking water. It is most frequently (and globally) present in domestic dishwashers [37][38][39], where internal rubber seals and plastic parts can harbour up to 10^6 colony forming units/cm² [39]. As people spend more time indoors and the number of immunocompromised people is rapidly increasing, the establishment of *E. dermatitidis* in domestic environments represents an important risk factor for human health [40][41].

Fungi normally populate parts of the human body, for example, skin, mucus of respiratory tract, oral cavity, and mucus of the digestive tract [28]. They most frequently invade through the respiratory tract by inhalation of spores or mycelium, but also enterically via the gastrointestinal tract or via traumatic injuries (e.g., accidents, surgery, interventions) [3]. After the initial infections, the fungi can spread via the haematogenous route. Fungal infections of the central nervous system occur either indirectly via lungs or paranasal sinuses, for example, after near-drowning episodes [42][43][44][45], via ocular orbits, and mastoid region of the temporal bone or retropharyngeal area, or directly as a consequence of trauma, invasive treatments, or brain surgery [41]. The potential mechanism of invasion of *E. dermatitidis* and other neurotropic fungi via the peripheral nervous system has not yet been described, nor have the mechanisms contributing to the spread of fungal infections to the brain.

Exophiala dermatitidis is rare in nature, but its occurrence increases in environments contaminated with cyclic or non-cyclic aromatic hydrocarbons [5][6][8] such as creosote-treated or oil-contaminated railway beams [46][47]. It can also be isolated from the cuticle of ants and ant hills [48][49], but most frequently on artificial rubber seals of dishwashers [37]. The neurotropic potential of black yeasts within *Chaetothyriales*, including *E. dermatitidis*, has been closely associated with their ability to assimilate monoaromatic and polyaromatic hydrocarbons, hypothetically including phenolic and aliphatic metabolic degradation products of catecholamine-like neurotransmitters [6][8][50][51][52]. It is known that disturbed transport or lowered concentrations of neurotransmitters can lead to neurodegenerative diseases, such as Alzheimer's [6][50][53].

According to recent investigations, systemic mycoses could be either one of the causative agents or an additional risk factor for the development of Alzheimer's. Fungal macromolecules were detected in the peripheral vascular system and in the cerebrospinal fluid (CSF) from patients with the disease [54][55], and elevated chitinase levels were also detected in the CSF [56]. Fungal yeast cells and hyphal fragments were detected in different parts of the brain, both inside and outside the neurons. In spite of this and other evidence [57][58], there is still ambiguity regarding the etiological role of fungi in Alzheimer's [57][59].

Fungi have also been associated with other neurological diseases. Recently, an increasing number of opportunistic mycoses of the central nervous system (CNS) has been reported in healthy individuals and, in particular, in patients with sepsis, prolonged ventilation, oncological therapies, organ transplantation, overuse of antibiotics, HIV patients, and others [60]. Opportunistic mycoses of CNS are associated with higher morbidity and mortality [61][62] owing to pathogenic fungi such as *Cryptococcus neoformans*, which are able to cross the otherwise prohibitive blood-brain barrier [63].

Over the last decades, extracellular vesicles (EVs) were identified as potential mediators of intracellular and inter-organism communication in all life kingdoms [64][65]. The first evidence of fungal EVs came from the opportunistic human pathogen *Cryptococcus neoformans* [66]. From that time, studies on the roles of EVs in pathogenicity of other fungi increased significantly (reviewed in Bielska and May) [67]. Fungal EVs are heterogeneous populations of lipid-bilayer nanoparticles that harbour cargo molecules important in modulating virulence, host defence, and host immune function, as well as triggering anti-microbial activities [67]. *C. neoformans* EVs can cross the blood-brain barrier and accumulate as lesions in the brain, facilitating adhesion and transcytosis [68][69][70][71]. Thus EVs show great potential for further research regarding fungal pathogenicity in human brains.

References

1. Sterflinger, K.; De Hoog, G.S.; Haase, G. Phylogeny and ecology of meristematic ascomycetes. *Stud. Mycol.* 1999, 43, 5–22. [Google Scholar]
2. Zalar, P.; De Hoog, G.S.; Gunde-Cimerman, N. Ecology of halotolerant dothideaceous black yeasts. *Stud. Mycol.* 1999, 43, 38–48. [Google Scholar]
3. De Hoog, G.S.; Queiroz-Telles, F.; Haase, G.; Fernandez-Zeppenfeldt, G.; Angelis, D.A.; Ende, A.H.G.G.; Van den Matos, T.; Peltroche-Llacsahuanga, H.; Pizzirani-Kleiner, A.; Rainer, J.; et al. Black fungi: Clinical and pathogenic approaches. *Med. Mycol.* 2000, 38, 243–250. [Google Scholar] [CrossRef] [PubMed]
4. De Hoog, G.S.; Guarro, J.; Gene, J.; Figueras, M.J. *Atlas of Clinical Fungi*, 2nd ed.; De Hoog, G.S., Ed.; Westerdijk Fungal Biodiversity Institute: Utrecht, The Netherlands, 2000; p. 1160. ISBN 9789070351434. [Google Scholar]
5. Prenafeta-Boldú, F.X.; Kuhn, A.; Luykx, D.M.A.M.; Anke, H.; Van Groenestijn, J.W.; De Bont, J.A.M. Isolation and characterisation of fungi growing on volatile aromatic hydrocarbons as their sole carbon and energy source. *Mycol. Res.* 2001, 105, 477–484. [Google Scholar]
6. Prenafeta-Boldú, F.X.; Summerbell, R.; Sybren de Hoog, G. Fungi growing on aromatic hydrocarbons: Biotechnology's unexpected encounter with biohazard? *FEMS Microbiol. Rev.* 2006, 30, 109–130. [Google Scholar] [CrossRef]

7. Zeng, J.S.; Sutton, D.A.; Fothergill, A.W.; Rinaldi, M.G.; Harrak, M.J.; De Hoog, G.S. Spectrum of clinically relevant Exophiala species in the United States. *J. Clin. Microbiol.* 2007, 45, 3713–3720. [Google Scholar] [CrossRef]
8. Prenafeta-Boldú, F.X.; De Hoog, G.S.; Summerbell, R.C. Fungal Communities in Hydrocarbon Degradation. In *Microbial Communities Utilizing Hydrocarbons and Lipids: Members, Metagenomics and Ecophysiology*; McGenity, T.J., Ed.; Springer International Publishing: Cham, Switzerland, 2018; pp. 1–36. ISBN 978-3-319-60063-5. [Google Scholar]
9. Kirchhoff, L.; Olsowski, M.; Rath, P.-M.; Steinmann, J. Exophiala dermatitidis: Key issues of an opportunistic fungal pathogen. *Virulence* 2019, 10, 984–998. [Google Scholar] [CrossRef]
10. Matsumoto, T.; Padhye, A.A.; Ajello, L. Medical significance of the so-called black yeasts. *Eur. J. Epidemiol.* 1987, 3, 87–95. [Google Scholar] [CrossRef]
11. Chang, X.; Li, R.; Yu, J.; Bao, X.; Qin, J. Phaeohyphomycosis of the Central Nervous System Caused by Exophiala dermatitidis in a 3-Year-Old Immunocompetent Host. *J. Child Neurol.* 2009, 24, 342–345. [Google Scholar] [CrossRef]
12. Kondori, N.; Gilljam, M.; Lindblad, A.; Jonsson, B.; Moore, E.R.B.; Wenneras, C. High Rate of Exophiala dermatitidis Recovery in the Airways of Patients with Cystic Fibrosis Is Associated with Pancreatic Insufficiency. *J. Clin. Microbiol.* 2011, 49, 1004–1009. [Google Scholar] [CrossRef]
13. Suzuki, K.; Nakamura, A.; Fujieda, A.; Nakase, K.; Katayama, N. Pulmonary infection caused by Exophiala dermatitidis in a patient with multiple myeloma: A case report and a review of the literature. *Med. Mycol. Case Rep.* 2012, 1, 95–98. [Google Scholar] [CrossRef] [PubMed]
14. Kusenbach, G.; Skopnik, H.; Haase, G.; Friedrichs, F.; Döhmen, H. Exophiala dermatitidis pneumonia in cystic fibrosis. *Eur. J. Pediatr.* 1992, 151, 344–346. [Google Scholar] [CrossRef] [PubMed]
15. Hiruma, M.; Kawada, A.; Ohata, H.; Ohnishi, Y.; Takahashi, H.; Yamazaki, M.; Ishibashi, A.; Hatsuse, K.; Kakihara, M.; Yoshida, M. Systemic phaeohyphomycosis caused by Exophiala dermatitidis. *Mycoses* 1993, 36, 1–7. [Google Scholar] [CrossRef] [PubMed]
16. Horré, R.; De Hoog, G.S. Primary cerebral infections by melanized fungi: A review. *Stud. Mycol.* 1999, 1999, 176–193. [Google Scholar]
17. Kerkemann, M.-L.; Piontek, K.; Mitze, H.; Haase, G. Isolation of Exophiala (Wangiella) dermatitidis in a Case of Otitis Externa. *Clin. Infect. Dis.* 1999, 29, 939–940. [Google Scholar] [CrossRef]
18. Chang, C.L.; Kim, D.-S.; Park, D.J.; Kim, H.J.; Lee, C.H.; Shin, J.H. Acute Cerebral Phaeohyphomycosis due to Wangiella dermatitidis Accompanied by Cerebrospinal Fluid Eosinophilia. *J. Clin. Microbiol.* 2000, 38, 1965–1966. [Google Scholar] [CrossRef]
19. Al-Obaid, I.; Ahmad, S.; Khan, Z.U.; Dinesh, B.; Hejab, H.M. Catheter-associated fungemia due to Exophiala oligosperma in a leukemic child and review of fungemia cases caused by Exophiala species. *Eur. J. Clin. Microbiol. Infect. Dis.* 2006, 25, 729–732. [Google Scholar] [CrossRef]
20. Mukaino, T.; Koga, T.; Oshita, Y.; Narita, Y.; Obata, S.; Aizawa, H. Exophiala dermatitidis infection in non-cystic fibrosis bronchiectasis. *Respir. Med.* 2006, 100, 2069–2071. [Google Scholar] [CrossRef]
21. Taj-Aldeen, S.J.; El Shafie, S.; Alsoub, H.; Eldeeb, Y.; De Hoog, G.S. Isolation of Exophiala dermatitidis from endotracheal aspirate of a cancer patient. *Mycoses* 2006, 49, 504–509. [Google Scholar] [CrossRef]
22. Grenouillet, F.; Cimon, B.; Pana-Katatali, H.; Person, C.; Gainet-Brun, M.; Malinge, M.-C.; Le Govic, Y.; Richaud-Thiriez, B.; Bouchara, J.-P. Exophiala dermatitidis Revealing Cystic Fibrosis in Adult Patients with Chronic Pulmonary Disease. *Mycopathologia* 2018, 183, 71–79. [Google Scholar] [CrossRef]
23. Lang, R.; Minion, J.; Skinner, S.; Wong, A. Disseminated Exophiala dermatitidis causing septic arthritis and osteomyelitis. *BMC Infect. Dis.* 2018, 18, 255. [Google Scholar] [CrossRef] [PubMed]
24. Vasquez, A.; Zavasky, D.; Chow, N.A.; Gade, L.; Zlatanic, E.; Elkind, S.; Litvintseva, A.P.; Pappas, P.G.; Perfect, J.R.; Revankar, S.; et al. Management of an Outbreak of Exophiala dermatitidis Bloodstream Infections at an Outpatient Oncology Clinic. *Clin. Infect. Dis.* 2018, 66, 959–962. [Google Scholar] [CrossRef] [PubMed]
25. Klasinc, R.; Riesenhuber, M.; Bacher, A.; Willinger, B. Invasive Fungal Infection Caused by Exophiala dermatitidis in a Patient after Lung Transplantation: Case Report and Literature Review. *Mycopathologia* 2019, 184, 107–113. [Google Scholar] [CrossRef] [PubMed]
26. Pinheiro, R.L.; Cognielli, R.C.R.; Barros, R.C.; Pinto, T.D.A.; Cunha, M.F.M.; Tahan, T.T.; Voidaleski, M.F.; Gomes, R.R.; Becker, G.N.; Andrade, L.V.; et al. Peritonitis by Exophiala dermatitidis in a pediatric patient. *Med. Mycol. Case Rep.* 2019, 24, 18–22. [Google Scholar] [CrossRef] [PubMed]
27. Wang, C.; Xing, H.; Jiang, X.; Zeng, J.; Liu, Z.; Chen, J.; Wu, Y. Cerebral Phaeohyphomycosis Caused by Exophiala dermatitidis in a Chinese CARD9-Deficient Patient: A Case Report and Literature Review. *Front. Neurol.* 2019, 10, 1–7. [Google Scholar] [CrossRef]

oogle Scholar] [CrossRef]

28. Huffnagle, G.B.; Noverr, M.C. The emerging world of the fungal microbiome. *Trends Microbiol.* 2013, 21, 334–341. [Google Scholar] [CrossRef]
29. Rizzetto, L.; De Filippo, C.; Cavalieri, D. Richness and diversity of mammalian fungal communities shape innate and adaptive immunity in health and disease. *Eur. J. Immunol.* 2014, 44, 3166–3181. [Google Scholar] [CrossRef] [PubMed]
30. Song, Y.; Laureijssen-van de Sande, W.W.J.; Moreno, L.F.; Van den Ende, B.G.; Li, R.; De Hoog, G.S. Comparative Ecology of Capsular Exophiala Species Causing Disseminated Infection in Humans. *Front. Microbiol.* 2017, 8, 1–25. [Google Scholar] [CrossRef]
31. Schnitzler, N.; Peltroche-Llacsahuanga, H.; Bestier, N.; Zündorf, J.; Lütticken, R.; Haase, G. Effect of Melanin and Carotenoids of Exophiala (Wangiella) dermatitidis on Phagocytosis, Oxidative Burst, and Killing by Human Neutrophils. *Infec. Immun.* 1999, 67, 94–101. [Google Scholar] [CrossRef]
32. Taylor, B.E.; Wheeler, M.H.; Szaniszlo, P.J. Evidence for Pentaketide Melanin Biosynthesis in Dematiaceous Human Pathogenic Fungi. *Mycologia* 1987, 79, 320–322. [Google Scholar] [CrossRef]
33. Kogej, T.; Stein, M.; Volkmann, M.; Gorbushina, A.A.; Galinski, E.A.; Gunde-Cimerman, N. Osmotic adaptation of the halophilic fungus Hortaea werneckii: Role of osmolytes and melanization. *Microbiology* 2007, 153, 4261–4273. [Google Scholar] [CrossRef] [PubMed]
34. Kejžar, A.; Gobec, S.; Plemenitaš, A.; Lenassi, M. Melanin is crucial for growth of the black yeast Hortaea werneckii in its natural hypersaline environment. *Fungal Biol.* 2013, 117, 368–379. [Google Scholar] [CrossRef] [PubMed]
35. Nishimura, K.; Miyaji, M.; Taguchi, H.; Tanaka, R. Fungi in bathwater and sludge of bathroom drainpipes. 1. Frequent isolation of Exophiala species. *Mycopathologia* 1987, 97, 17–23. [Google Scholar] [CrossRef] [PubMed]
36. Matos, T.; De Hoog, G.S.; De Boer, A.G.; De Crom, I.; Haase, G. High prevalence of the neurotropic Exophiala dermatitidis and related oligotrophic black yeasts in sauna facilities. *Mycoses* 2002, 45, 373–377. [Google Scholar] [CrossRef]
37. Zalar, P.; Novak, M.; De Hoog, G.S.; Gunde-Cimerman, N. Dishwashers — A man-made ecological niche accommodating human opportunistic fungal pathogens. *Fungal Biol.* 2011, 115, 997–1007. [Google Scholar] [CrossRef]
38. Gümral, R.; Özhak-Baysan, B.; Tümgör, A.; Saracılı, M.A.; Yıldırın, Ş.T.; İlkit, M.; Zupančič, J.; Novak-Babič, M.; Gunde-Cimerman, N.; Zalar, P.; et al. Dishwashers provide a selective extreme environment for human-opportunistic yeast-like fungi. *Fungal Divers.* 2016, 76, 1–9. [Google Scholar] [CrossRef]
39. Zupančič, J.; Novak Babič, M.; Zalar, P.; Gunde-Cimerman, N. The Black Yeast Exophiala dermatitidis and Other Selected Opportunistic Human Fungal Pathogens Spread from Dishwashers to Kitchens. *PLoS ONE* 2016, 11, e0148166. [Google Scholar] [CrossRef]
40. Garber, G. An Overview of Fungal Infections. *Drugs* 2001, 61, 1–12. [Google Scholar] [CrossRef]
41. Sharma, R.R. Fungal infections of the nervous system: Current perspective and controversies in management. *Int. J. Surg.* 2010, 8, 591–601. [Google Scholar] [CrossRef]
42. Mursch, K.; Trnovec, S.; Ratz, H.; Hammer, D.; Horré, R.; Klinghammer, A.; De Hoog, S.; Behnke-Mursch, J. Successful treatment of multiple *Pseudallescheria boydii* brain abscesses and ventriculitis/ependymitis in a 2-year-old child after a near-drowning episode. *Child's Nerv. Syst.* 2006, 22, 189–192. [Google Scholar] [CrossRef]
43. Chen, T.-C.; Ho, M.-W.; Chien, W.-C.; Lin, H.-H. Disseminated *Scedosporium apiospermum* infection in a near-drowning patient. *J. Formos. Med. Assoc.* 2016, 115, 213–214. [Google Scholar] [CrossRef] [PubMed]
44. Wang, X.-Y.; Yu, S.-L.; Chen, S.; Zhang, W.-H. CNS infection caused by *Pseudallescheria boydii* in a near-drowning traveller from a traffic accident. *J. Travel Med.* 2016, 23, tav018. [Google Scholar] [CrossRef] [PubMed]
45. Yamawaki, S.; Nakashima, K.; Suzuki, F.; Otsuki, A.; Watanabe, J.; Takai, M.; Katsurada, M.; Katsurada, N.; Ohkuni, Y.; Misawa, M.; et al. Rice-Field Drowning-Associated Pneumonia in which *Pseudomonas* spp., *Aspergillus fumigatus*, and *Cunninghamella* sp. Are Isolated. *Intern. Med.* 2016, 55, 825–829. [Google Scholar] [CrossRef] [PubMed]
46. Vicente, V.A.; Attili-Angelis, D.; Pie, M.R.; Queiroz-Telles, F.; Cruz, L.M.; Najafzadeh, M.J.; De Hoog, G.S.; Zhao, J.; Pizirani-Kleiner, A. Environmental isolation of black yeast-like fungi involved in human infection. *Stud. Mycol.* 2008, 61, 137–144. [Google Scholar] [CrossRef] [PubMed]
47. Zhao, J.; Zeng, J.; De Hoog, G.S.; Attili-Angelis, D.; Prenafeta-Boldú, F.X. Isolation and Identification of Black Yeasts by Enrichment on Atmospheres of Monoaromatic Hydrocarbons. *Microb. Ecol.* 2010, 60, 149–156. [Google Scholar] [CrossRef]
48. Voglmayr, H.; Mayer, V.; Maschwitz, U.; Moog, J.; Djieto-Lordon, C.; Blatrix, R. The diversity of ant-associated black yeasts: Insights into a newly discovered world of symbiotic interactions. *Fungal Biol.* 2011, 115, 1077–1091. [Google Scholar] [CrossRef]

49. Duarte, A.P.M.; Attili-Angelis, D.; Baron, N.C.; Forti, L.C.; Pagnocca, F.C. Leaf-cutting ants: An unexpected microenvironment holding human opportunistic black fungi. *Antonie Leeuwenhoek Int. J. Gen. Mol. Microbiol.* 2014, **106**, 465–473. [Google Scholar] [CrossRef]
50. Zecca, L.; Zucca, F.A.; Costi, P.; Tampellini, D.; Gatti, A.; Gerlach, M.; Riederer, P.; Fariello, R.G.; Ito, S.; Gallorini, M.; et al. The neuromelanin of human substantia nigra: Structure, synthesis and molecular behaviour. *J. Neural Transm. Suppl.* 2003, **65**, 145–155. [Google Scholar]
51. Lian, X.; De Hoog, G.S. Indoor wet cells harbour melanized agents of cutaneous infection. *Med. Mycol.* 2010, **48**, 622–628. [Google Scholar] [CrossRef]
52. Isola, D.; Selbmann, L.; De Hoog, G.S.; Fenice, M.; Onofri, S.; Prenafeta-Boldú, F.X.; Zucconi, L. Isolation and Screening of Black Fungi as Degraders of Volatile Aromatic Hydrocarbons. *Mycopathologia* 2013, **175**, 369–379. [Google Scholar] [CrossRef]
53. Francis, P.T. The Interplay of Neurotransmitters in Alzheimer’s Disease. *CNS Spectr.* 2005, **10**, 6–9. [Google Scholar] [CrossRef] [PubMed]
54. Alonso, R.; Pisa, D.; Rábano, A.; Carrasco, L. Alzheimer’s disease and disseminated mycoses. *Eur. J. Clin. Microbiol. Infect. Dis.* 2014, **33**, 1125–1132. [Google Scholar] [CrossRef] [PubMed]
55. Alonso, R.; Pisa, D.; Marina, A.I.; Morato, E.; Rábano, A.; Rodal, I.; Carrasco, L. Evidence for Fungal Infection in Cerebrospinal Fluid and Brain Tissue from Patients with Amyotrophic Lateral Sclerosis. *Int. J. Biol. Sci.* 2015, **11**, 546–558. [Google Scholar] [CrossRef]
56. Watabe-Rudolph, M.; Song, Z.; Lausser, L.; Schnack, C.; Begus-Nahrmann, Y.; Scheithauer, M.O.; Rettinger, G.; Otto, M.; Tumani, H.; Thal, D.R.; et al. Chitinase enzyme activity in CSF is a powerful biomarker of Alzheimer disease. *Neurology* 2012, **78**, 569–577. [Google Scholar] [CrossRef]
57. Pisa, D.; Alonso, R.; Juarranz, A.; Rábano, A.; Carrasco, L. Direct visualization of fungal infection in brains from patients with Alzheimer’s disease. *J. Alzheimer’s Dis.* 2015, **43**, 613–624. [Google Scholar] [CrossRef] [PubMed]
58. Kumar, D.K.V.; Choi, S.H.; Washicosky, K.J.; Eimer, W.A.; Tucker, S.; Ghofrani, J.; Lefkowitz, A.; McColl, G.; Goldstein, L.E.; Tanzi, R.E.; et al. Amyloid- β peptide protects against microbial infection in mouse and worm models of Alzheimer’s disease. *Sci. Transl. Med.* 2016, **8**, 340ra72. [Google Scholar] [CrossRef] [PubMed]
59. Pisa, D.; Alonso, R.; Rábano, A.; Rodal, I.; Carrasco, L. Different Brain Regions are Infected with Fungi in Alzheimer’s Disease. *Sci. Rep.* 2015, **5**, 15015–15028. [Google Scholar] [CrossRef] [PubMed]
60. Candoni, A.; Klimko, N.; Busca, A.; Di Blasi, R.; Shadrivova, O.; Cesaro, S.; Zannier, M.E.; Verga, L.; Forghieri, F.; Calore, E.; et al. Fungal infections of the central nervous system and paranasal sinuses in onco-haematologic patients. Epidemiological study reporting the diagnostic-therapeutic approach and outcome in 89 cases. *Mycoses* 2019, **62**, 252–260. [Google Scholar] [CrossRef]
61. Gavito-Higuera, J.; Mullins, C.B.; Ramos-Duran, L.; Olivas Chacon, C.I.; Hakim, N.; Palacios, E. Fungal Infections of the Central Nervous System: A Pictorial Review. *J. Clin. Imaging Sci.* 2016, **6**, 24–30. [Google Scholar] [CrossRef]
62. Bloch, K.C.; Bailin, S.S. Update on fungal infections of the central nervous system. *Curr. Opin. Infect. Dis.* 2019, **32**, 277–284. [Google Scholar] [CrossRef]
63. Swinburne, N.C.; Bansal, A.G.; Aggarwal, A.; Doshi, A.H. Neuroimaging in Central Nervous System Infections. *Curr. Neurol. Neurosci. Rep.* 2017, **17**, 49–63. [Google Scholar] [CrossRef]
64. Yáñez-Mó, M.; Siljander, P.R.-M.; Andreu, Z.; Bedina Zavec, A.; Borràs, F.E.; Buzas, E.I.; Buzas, K.; Casal, E.; Cappello, F.; Carvalho, J.; et al. Biological properties of extracellular vesicles and their physiological functions. *J. Extracell. Vesicles* 2015, **4**, 27066–27126. [Google Scholar] [CrossRef] [PubMed]
65. Raposo, G.; Stahl, P.D. Extracellular vesicles: A new communication paradigm? *Nat. Rev. Mol. Cell Biol.* 2019, **20**, 509–510. [Google Scholar] [CrossRef] [PubMed]
66. Rodrigues, M.L.; Nimrichter, L.; Oliveira, D.L.; Frases, S.; Miranda, K.; Zaragoza, O.; Alvarez, M.; Nakouzi, A.; Feldmesser, M.; Casadevall, A. Vesicular Polysaccharide Export in Cryptococcus neoformans Is a Eukaryotic Solution to the Problem of Fungal Trans-Cell Wall Transport. *Eukaryot. Cell* 2007, **6**, 48–59. [Google Scholar] [CrossRef]
67. Bielska, E.; May, R.C. Extracellular vesicles of human pathogenic fungi. *Curr. Opin. Microbiol.* 2019, **52**, 90–99. [Google Scholar] [CrossRef] [PubMed]
68. Oliveira, D.L.; Nakayasu, E.S.; Joffe, L.S.; Guimarães, A.J.; Sobreira, T.J.P.; Nosanchuk, J.D.; Cordero, R.J.B.; Frases, S.; Casadevall, A.; Almeida, I.C.; et al. Characterization of Yeast Extracellular Vesicles: Evidence for the Participation of Different Pathways of Cellular Traffic in Vesicle Biogenesis. *PLoS ONE* 2010, **5**, e11113. [Google Scholar] [CrossRef] [PubMed]

69. Huang, S.-H.; Wu, C.-H.; Chang, Y.C.; Kwon-Chung, K.J.; Brown, R.J.; Jong, A. Cryptococcus neoformans-Derived Microvesicles Enhance the Pathogenesis of Fungal Brain Infection. *PLoS ONE* 2012, 7, e48570. [Google Scholar] [CrossRef]
70. Wolf, J.M.; Espadas-Moreno, J.; Luque-Garcia, J.L.; Casadevall, A. Interaction of Cryptococcus neoformans Extracellular Vesicles with the Cell Wall. *Eukaryot. Cell* 2014, 13, 1484–1493. [Google Scholar] [CrossRef]
71. Rodrigues, M.L.; Godinho, R.M.C.; Zamith-Miranda, D.; Nimrichter, L. Traveling into Outer Space: Unanswered Questions about Fungal Extracellular Vesicles. *PLOS Pathog.* 2015, 11, e1005240.

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