Melatonin and Chronic Pain

Subjects: Neurosciences Contributor: Thomas M Kinfe

Preclinical as well as human studies indicate that melatonin is essential for a physiological sleep state, promotes analgesia and is involved in immunometabolic signaling by regulating neuroinflammatory pathways. Experimental and clinical neuromodulation studies for chronic pain treatment suggest that neurostimulation therapies such as spinal cord stimulation, vagus nerve stimulation and dorsal root ganglion stimulation have an impact on circulating inflammatory mediators in blood, cerebrospinal fluid and saliva.

Keywords: melatonin ; chronic pain ; sleep ; immune-metabolism ; central and peripheral neuroinflammation pathways ; augmentation of neurostimulation therapies

1. Introduction

Melatonin, an indole alkaloid, is chemically N-acetyl-5-methoxytryptamine and is a derivative of tryptophan ^[1]. Melatonin, acting as a pleiotropic hormone, is released from the pineal gland and extra-pineal tissues with a critical role in regulating the circadian rhythms ^[2]. Initially, melatonin was described as a sleep hormone as it is secreted in the dark and induces sleep, but it is now well appreciated that it implicates a wide array of activities encompassing anti-oxidant, anti-inflammatory, anti-apoptotic, anti-sympathetic nerve activation, endothelial cell preservation, neuroprotection, hepatoprotection, immunomodulation, thermoregulation, mood and sexual behavior modulation ^{[1][3][4][5][6][Z][8]}. Although the precise mechanism remains to be elaborated, these implicates a relevant role of the neurobiology of a broad variety of system inflammation driven diseases such as chronic pain syndromes and metabolic associated disorders.

Hence, melatonin has the capability to promote and to interact with relevant and shared pathways of sleep, chronic pain and immunometabolic trafficking due to its anti-inflammatory properties. Melatonin derives its name due to its ability to induce aggregation of melanin granules, which lead to lightening of the skin of the frog due to its α -Melanocyte Stimulating Hormone (α -MSH) antagonistic effects ^{[4][9]}. Besides pineal body, different brain areas, retina, retinal pigment epithelium, lens, iris, ciliary body, lacrimal glands, bone marrow, lymphocytes, skin and gut secrete it ^{[4][9][10]} and is conserved across almost all species from primitive prokaryotes to humans ^[4]. Serum levels of melatonin tend to rise after a few months of birth in neonates, gradually decrease during puberty and eventually decline at old age characterized by rhythmic diurnal release variations exhibiting peak levels at night from midnight to 2 am and lowest from midday to 2 pm ^{[4][10]}.

Over the recent years, non-invasive and invasive neurostimulation of both the central and peripheral nervous systems have become established therapy options in drug-resistant chronic pain disorders of different origins including but not limited to spinal cord stimulation, dorsal root ganglion stimulation and/or vagus nerve stimulation [11][12][13][14][15][16][17][18] [19][20][21][22][23]. So far, outcome of neuromodulation therapies for chronic pain has been determined using score-based parameters and psychometric features (depression, cognition decline, anxiety) neglecting the intra- and inter-individual variability of chronic patients. In view of lacking objective outcome parameters, several uncontrolled and randomized clinical studies have determined the effects of neurostimulation therapies on the neuro-immune axis by quantifying the concentrations of circulating inflammatory mediators and corresponding gene expression levels in different biofluids (blood, cerebrospinal fluid, saliva) in chronic pain patients [24][25][26][27]. Intriguingly, several approaches such as molecular inflammatory phenotyping, electrophysiological recordings, neuroimaging and digital measures have gained increased recognition in order to predict and potentially consolidate the neuromodulation therapy outcome. However, systematic evidence-derived studies assessing the augmentative potential of adjunct anti-inflammatory pharmacological agents such as melatonin, which can modulate the inflammatory pathways, remain under-investigated.

2. Overview of the Melatonin biology

2.1. Synthesis and Metabolism of Melatonin

Tryptophan serves as a precursor of Melatonin. Initially, tryptophan is converted into serotonin, and then serotonin is *N*-acetylated and *O*-methylated to form melatonin by arylalkylamine *N*-acetyltransferase (NAT) and hydroxyindole-*O*-methyltransferase (HIOMT), respectively ^[28]. The suprachiasmatic nucleus of the hypothalamus regulates the synthesis of melatonin by the pineal gland depending upon the input received from retinohypothalamic pathway. During day, the exposure to light induces the breakdown of melanopsin in retinal photoreceptive ganglion cells, which signals through retinohypothalamic pathway to inhibit the synthesis of melatonin, consequently, the levels of melatonin are low during daylight and rise in dark at night ^[3]. The neurotransmitters such as noradrenaline and GABA regulate the synthesis of Melatonin ^[29]. The presence of 5-methoxy and N-acetyl functional groups in the structure of melatonin impart specificity and amphiphilicity, and melatonin can readily cross the plasma membranes affecting almost all the cells of the body ^[3]. In the blood after release from the pineal body, it is transported in combination with albumin and is metabolized in the liver through oxidation by Cytochrome P450 isozymes (CYP1A2, CYP1A1, CYP1B1) into 6-hydroxymelatonin and subsequent conjugation to 6-sulfatoxymelatonin, that is ultimately eliminated via kidneys ^{[3][28][30]}. At the level of brain, substantial quantity of melatonin is biotransformed into kynuramine derivatives, which share antioxidant and anti-inflammatory properties with melatonin (**Figure 1**) ^[30].

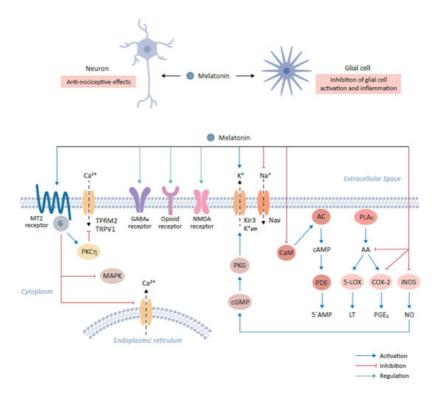


Figure 1. Role of melatonin and its cognate receptors in pain-relieving signal pathways. The binding of melatonin to MT2 receptors expressed on glutamatergic neurons mediates anti-nociceptive effects. This interaction results in the activation of the PKCn signaling pathway and in turn suppresses mitogen-activated protein kinase (MAPK) and calcium signaling. The interaction of melatonin with various neurotransmitter receptors, including gamma-aminobutyric acid (GABA)ergic (GABA_A), produces anti-nociceptive effects. Furthermore, melatonin binds opioid and NMDA receptors to mediate downstream signaling, resulting in anti-nociceptive effects. Additionally, melatonin modulates ion flow by binding to Kir3 K⁺ channel which activates the outflow of K⁺ ions in the central nervous system. Conversely, melatonin inhibits the activities of sodium channels, including NAV1.8 and NAV1.9. Melatonin prevents the influx of Ca²⁺ by inhibiting calmodulin (CaM). CaM modulates intracellular 5'-AMP levels through the step-wise activation of adenylyl cyclase (AC) and cyclic nucleotide phosphodiesterase (PDE) by the intermediate generation of cAMP. Finally, melatonin modulates the inflammatory response and activation of glial cells by inhibiting the formation of arachidonic acid (AA) via the inhibition of phospholipase A₂ (PLA₂), which leads to the subsequent inhibition of 5-lipoxygenase (5-LOX) and cyclooxygenase-2 (COX-2) and the reduction in their targets, leukotriene (LT) and prostaglandin E2 (PGE₂) respectively. Melatonin also decrease the expression of inducible nitric oxide synthase (iNOS), leading to the decline in nitric oxide (NO) expression.

2.2. Melatonin and Its Receptors

The extensive accessibility throughout the body and versatility in the effects of melatonin is brought about by its receptor dependent and independent effects. The anti-oxidative and free radical scavenger effects of melatonin avoids any receptor based interaction and directly stem from melatonin's direct chemical interaction with various free radicals such as hydroxyl radicals, hydrogen peroxide, nitric oxide and others ^[9]. Melatonin is known to upregulate the expression or activity of different antioxidant enzymes such as superoxide dismutase and glutathione peroxidase ^[3]. In addition to these directly mediated effects, two high affinity membrane bound receptors are recognized to mediate the effects of melatonin

and are designated as MT1 and MT2. Both MT1 and MT2 belong to the family of G-protein coupled transmembrane serpentine receptors (GPCRs) ^{[3][9][31]}. A relatively low binding affinity site MT3 for melatonin was in addition recognized intracellularly on a cytosolic enzyme quinone reductase 2, which is responsible for reducing menadione and other quinones ^{[3][31]}. Furthermore, melatonin binds nuclear receptors such as retinoid orphan receptors (ROR) and retinoid Z receptors (RZR) such as ROR α , ROR α 2, RZR α and RZR β ^{[3][9]}.

MT1 and MT2 Receptors

The MT1 receptor, cloned in 1994 as a first mammalian melatonin receptor, consists of 350 amino acids and is linked to G_i , more specifically $G_{i\alpha2}$, $G_{i\alpha3}$ and $G_{q/11}$ ^{[9][32]}. The activation of the MT1 receptor leads to inhibition of adenylyl cyclase leading to a decline in cAMP levels. The MT1 receptors can be coupled to Kir3 potassium channels through G_i and in some cells to $G_{q/11}$, mediating the activation of phospholipase C resulting in increased Ca²⁺ levels through modulation of the calmodulin levels ^{[9][31]}.

The MT1 receptor is found to be expressed in brain, peripheral blood vessels, aorta, heart, testes, ovaries, placenta, mammary glands, skin, liver, spleen, kidney, adrenal cortex, pancreas, and immune system. Inside the brain, expression of MT1 receptors is specifically found in hypothalamus, hippocampus, cerebellum, pars tuberalis, substantia nigra and ventral tegmental area (**Figure 1**) ^{[3][9][31]}.

The MT2 receptor shares 60% homology to MT1 receptor and is composed of 363 amino acids. The MT2 receptor like MT1 receptor is coupled to G_i and leads to inhibition of adenylyl cyclase and soluble guanylyl cyclase activity ^{[9][31]}. The MT2 receptors aredistributed in the gastrointestinal tract, skin, breasts, testes, kidney, adipose tissue, immune system, retina, blood vessels, and inside the brain including the regions such as hypothalamus, suprachiasmatic nucleus and pituitary ^[9]. Both the membrane receptors have been shown to dimerize forming homodimers or heterodimers, with relatively more common homodimers of MT1 or MT1-MT2 heterodimers compared to MT2 homodimers ^[9]. Interestingly, both the receptors may have opposing physiological functions e.g., non-rapid eye movement (NREM) sleep is decreased upon MT1 activation and promoted upon MT2 activation; MT1 activation leads to vasoconstriction, but MT2 activation produce vasodilation; MT1 receptor have implications in depression and anhedonia, whereas MT2 receptor is implicated in anxiety; further MT1 receptors regulate rapid-eye movement (REM) sleep, but MT2 receptors do not regulate it ^[32].

3. Melatonin and Chronic Pain

Pain has been defined as "An unpleasant sensory and emotional experience associated with actual or potential tissue damage or described in terms of such damage" [33]. Pain significantly affects the quality of life and implies increased social costs for its management [34]. A relationship between pain and melatonin has beendelineated, since chronic pain patients have shown lower levels of melatonin in the blood and urine [35]. Even the precursors of melatonin such as L-tryptophan and serotonin were found to be low in patients with fibromyalgia, indicating that the melatonin has a critical role in chronic pain syndromes ^{[35][36]}. Further adding to the evidence, analgesia observed after melatonin administration in both nociceptive and neuropathic pain models also points towards an important role of melatonin in regulating pain [37]. Melatonin affords amelioration of pain through several mechanisms. For instance, Melatonin could ameliorate the nociceptive pain owing to its ability to prevent prostaglandins release, inhibition of the migration of polymorphonuclear cells at the inflammation site, and inhibition of the cyclooxygenase-2 (COX-2) and nitric oxide (NO) synthase. Melatonin provides neuropathic pain relief through its ability to raise the pain threshold and decrease of the thermal hyperalgesia manifestation. Intriguingly, the analgesic effects of melatonin have been shown to be dose dependent and have been observed consistently across various pain induction models relying on thermal, chemical, mechanical and electrical pain induction [37]. Furthermore, the different routes of administration almost provide the same efficacy from a given dose of the melatonin [37]. Analgesic effects of melatonin have been well appreciated across various chronic pain syndromes such as chronic pelvic pain, fibromyalgia, irritable bowel syndrome, tension and cluster headaches, migraine and others including chronic back pain [37][38]. Around 5–10% of the patients with acute back pain progress to subacute back pain category and then, finally develop chronic back pain, which is defined as back pain lasting for more than 3 months [34]. The next section will provide an overview of the melatonin's pain-relieving mechanisms before discussing the melatonin's effects in chronic pain disorders of different origin.

3.1. Mechanisms of Melatonin Action in Relieving Pain

The pain-relieving effects of melatonin either stem from its direct anti-nociceptive effects due to its anti-oxidant, antiinflammatory properties and direct interaction with its cognate receptors or through indirect complex interactions with other systems that can modulate the pain perception pathways ^{[29][37]}. MT2 receptors have been found to be particularly enriched in the brain areas which play an important role in modulating the pain sensation ^[29]. For example, ventromedial nucleus of the thalamus and reticular formation as a part of the ascending nociceptive pathway and ventrolateral periaqueductal gray matter as a part of the descending antinociceptive pathway harbor high densities of the MT2 receptors ^[32]. Initial studies involving the use of melatonin antagonists also confirmed that MT2 receptors are preferentially implicated in mediating melatonin's anti-nociceptive effects ^[32]. Furthermore, selective partial agonists at MT2 receptors such as UCM765 and UCM924 have shown anti-nociceptive effects in experimental models of nociceptive pain ^{[29][37]}. Modulation of the MT2 receptors expressed on glutamatergic neurons in the rostral ventrolateral periaqueductal gray by UCM924 decreased tail flick responses in experimental pain model and modulated the neuronal activity of the pain pathways ^{[29][37]}. An agonist of MT2 receptor, IIK-7, provides relief against neuropathic pain via inhibition of glial cells activation and inhibition of inflammation as exhibited by down regulation of inducible nitric oxide synthase (iNOS) and caspase 3 ^[29]. Intriguingly, MT2 receptor ligation by melatonin reduces neuronal excitability in experimental pain model owing to inhibition of T-type Ca²⁺ channel currents via downstream G_{βy}-mediated PKCη signal pathway (**Figure 1**). In addition, melatonin suppresses mechanical allodynia and thermal hyperalgesia due to the inhibition of the mitogen activated protein kinase (MAPK) and calcium signaling ^[29].

3.2. Interaction with Neurotransmitters (and Their Receptors)

Melatonin interacts with various neurotransmitter systems indirectly to produce nociceptive effects. For instance, melatonin can produce antinociceptive effects by affecting GABAergic (GABA_A receptor), benzodiazepinergic (BZD receptor), opioidergic ($\delta/\mu/\kappa$ receptors), sigma system (sigma receptor), dopaminergic receptor (D₂ receptor), adrenergic (α_2 receptor), serotonergic (5-HT_{2A} receptor), glutamatergic (NMDA receptor) and NO-cyclic GMP-PKG signaling pathway (**Figure 1**) ^[30].

Melatonin can modulate the GABAergic transmission as was evidenced from the observation that Flumazenil, which is a benzodiazepine antagonist and inhibits indirectly the GABAergic neurotransmission via antagonizing the allosteric effects of benzodiazepines receptors on their own receptor subtypes and therefore reversing the anti-nociceptive effects of Melatonin ^{[32][37][38][39]}. Melatonin is known to facilitate GABAergic transmission through various mechanisms such as the increase of the GABA content, increased affinity of GABA for its receptors and density of GABA receptors; even melatonin and its analogues can directly bind to GABA receptors ^{[31][32][37][38][39]}. These mechanisms likely explain the synergistic effects of melatonin and benzodiazepine-GABA agonists, which are independent from direct interaction of melatonin to its cognate receptors (**Figure 1**).

Interestingly, there is evidence that there exists an indirect link between melatonin and the opioidergic system. For instance, the melatonin's antinociceptive effects were antagonized upon Naloxone administration, which is an opiate antagonist [37][38]. Further, melatonin administration could induce the production and release of β -endorphins; and morphine and other opioid agonists could induce the melatonin release from the pineal gland [39]. Furthermore, the analgesic effects of morphine were reduced in pinealectomized mice. Intriguingly, neither melatonin nor any of its analogues were shown to directly interact with opioid receptors, yet there exists an important connection between melatonin and opioids in mediating anti-nociceptive effects [37][38][39].

Glutamate and its NMDA receptors have been implicated in nociceptive sensation pathways ^[29]. Repetitive firing of Cfibers potentiates the spinal nociceptive transmission, an effect known as wind-up, and is dependent on NMDA receptors. Interestingly, melatonin can induce suppression of this wind-up effect in a dose dependent manner through the modulation of intracellular NO levels leading to the activation of cGMP-PKG-ATP sensitive K⁺ channels ^[30]. The antinociceptive effects of melatonin involve modulation of the α_1 -adrenergic, α_2 -adrenergic, muscarinic and nicotinic receptors at the spinal level through the modulation of intracellular cGMP levels (**Figure 1**) ^[30].

3.3. Modulation of Ion Channel Activity

Melatonin has been shown to activate the G-protein coupled Kir3 (inwardly rectifying K⁺) channels at the cellular level, which reduce the rapid firing of action potential trains in neurons ^{[37][38]}. Melatonin activates outward flow of K⁺ ions in various areas of the CNS such as cerebellum, suprachiasmatic nucleus , while in turn it inhibits the activity of voltage gated sodium channels such as Nav1.8 and Nav1.9 to exert anti-thermal hypersensitivity and anti-mechanical allodynia effects ^{[29][37][38][39]}. In the cultured dorsal root ganglion cells of the spinal cord, melatonin has been shown to decrease the neuronal free intracellular Ca²⁺ levels via inhibition of the voltage sensitive Ca²⁺ channels ^{[37][38][39]}. This effect of melatonin to inhibit mobilization of free Ca²⁺ from intracellular storage sites as well as inhibition of inward flow of Ca²⁺ from voltage activated channels has also been demonstrated in various other tissues ^[29]. Melatonin regulates against the Ca²⁺ influx through the desensitization of transient receptor potential vanilloid type 1 (TRPV1) and transient receptor potential melastatin type 2 (TRPM2) channels (**Figure 1**) ^[29].

Melatonin is known to bind calmodulin and directly affect the calcium signaling and modulate the activity of the enzymes such as adenylate cyclase and phosphodiesterase or structural proteins ^[28].

It is well established that Ca^{2+} channels play a vital role in the development and maintenance of central sensitization that is associated with inflammation and neuropathic pain, melatonin, therefore provides pain relief through modulation of the activity of Ca^{2+} channels [37][38].

3.4. Melatonin and Inflammatory Mediators

The inflammation is upregulated by various lipid mediators generated from arachidonic acid by the action of Phospholipase A₂ (PLA₂), and subsequent activity of 5-Lipoxygenase (5-LOX) and Cyclooxygenase-2 (COX-2). Melatonin can inhibit arachidonic acid formation, acting as a negative regulator of PLA₂, inhibit the expression of 5-LOX and COX-2 leading to decreased synthesis of chemoattractant leukotrienes and prostaglandins, respectively ^{[32][39]}. Melatonin is also known to decrease the expression of iNOS (inducible nitric oxide synthase) and COX-2, which subsequently leads to a decline in NO and PGE₂ expression, respectively ^[39]. These inflammatory mediators are involved in inflammatory pain perception and therefore, the inhibition of these inflammatory mediators contribute to the analgesic effects of melatonin (**Figure 1**).

4. Melatonin at the Crossroads of Different Chronic Pain Disorders, Sleep and Inflammation

Back pain is a commonly encountered musculoskeletal disorder that is often perceived to be related to disability. Most of the patients with acute back pain have good recovery, however, a small portion of the patients suffer from this pain over an extended period of time than acute back pain (more than 12 weeks) and are referred to have chronic back pain. Since, the incidence of acute back pain is higher, consequently the number of patients with chronic back pain is also getting higher ^[40]. It is estimated that 30–60% of the population in developed countries suffer from chronic back pain and it affects mostly the people of working age with a peak prevalence in the age group of 30–60 years. This represents a high cost to the society and contributes to disability ^{[37][40]}. It is of note that chronic back pain is responsible for the most years lived with disability when compared to any other medical illness ^[40].

Furthermore, melatonin provides pain relief in several other chronic pain conditions such as migraine, chronic cluster and tension type headaches, fibromyalgia, irritable bowel syndrome and chronic pelvic pain. Intriguingly, melatonin administration to chronic back pain patients has been shown to significantly relieve the pain both at rest and during motion ^{[37][38]}. In a clinical study, the addition of melatonin at a dosage of 3 mg/day, 30–40 min before sleeping at night, to the standard therapy consisting of administering glucosamine hydrochloride and chondroitin sulfate alone or in combination with diclofenac or only diclofenac was found to significantly reduce the pain intensity during movement and at rest as assessed by Visual Analog Scale (VAS) and related scoring batteries ^{[37][38]}. Melatonin addition to the standard regimens also provided additional benefits such as reduced pain-associated anxiety, mood stabilization and sleep quality improvement, which are often impaired in the chronic back pain patients ^[37]. These beneficial effects of melatonin might be due to the restoration of the circadian rhythm disturbed by the chronic pain, improvement in sleep and return of the body's adaptive potential to normal besides the intrinsic analgesic properties of the melatonin administration. Intriguingly, in this study, the analgesic effects of melatonin were observed earlier than the sleep normalization effects, supporting the notion that the melatonin has its own intrinsic antinociceptive effects apart from similar effects brought about through complex interactions with other systems ^[37].

On one hand, the pain substantially affects the sleep and on the other hand, deprivation of restful sleep escalates the pain susceptibility $^{[41]}$. So, both conditions can aggravate each other suggesting a bidirectional and reciprocal relationship, further adding to the suffering of the patient $^{[42]}$. Chronic pain can result in sleep deprivation for extended periods of time and can affect the physical, emotional and behavioral wellbeing of the patients $^{[41]}$. It is estimated that around 50–80% of the patients with chronic pain experience sleep disturbances $^{[42]}$. Interestingly, a study involving chronic back pain patients showed that 53% of the patients met the criteria for insomnia compared to 3% of the healthy pain free controls $^{[42]}$. Additionally, the severity of insomnia showed a positive association with pain intensity, sensory pain ratings, affective pain ratings, general anxiety, health anxiety and general depression. Clinical studies have shown that chronic pain patients reporting sleep disturbance have higher pain perception, more fatigue, poor mood, and increased levels of stress and disability. Alternatively, healthy subjects who were deprived of sleep showed an increased pain probably due to an increase in the release of pro-inflammatory cytokines and reduced pain tolerance $^{[42]}$.

Melatonin is shown to improve the quality of sleep in chronic inflammatory pain patients and other chronic pain disorders in addition to its analgesic effects ^{[29][37]}. Repeated administration of melatonin progressively improves the sleep and

reduces anxiety, thereby leading to lower levels of pain ^[43]. Intriguingly, in an experimental chronic constriction injury rat model, deprivation of sleep was shown to further aggravate the neuropathic pain and was associated with increased activation of microglia and lower levels of serum melatonin ^[44]. However, when melatonin was administered, it attenuated the microglial activation, reduced the levels of pro-inflammatory cytokines ultimately leading to a reduction in neuropathic pain ^[44]. A study involving the administration of melatonin to chronic back pain patients also showed a significant improvement in the sleep quality in addition to a reduction in back pain ^[45].

A variety of inflammatory mediators are liberated upon tissue damage or inflammation including prostaglandins (PGE₂), leukotrienes, bradykinin, substance P and inflammatory cytokines, which either directly activate nociceptors or release local allogenic agents capable of further sensitizing the nociceptors leading to enhanced pain transmission ^{[30][46]}. These inflammatory mediators setting the nociceptive system on fire are often produced by the immune cells. The coordination between nociceptive neuronal pathways and immune response represents a potential advantage to the host in protecting against danger (**Figure 2**). Interestingly, neuroimmune communication in pain is bidirectional, for instance immune cell secreted mediators sensitize peripheral and central pain pathways and in turn, nociceptive neurons actively release neuropeptides via their peripheral nerve terminals to shape the response of innate and adaptive immune cells ^[46].

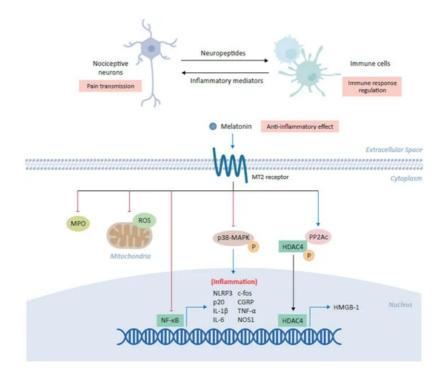


Figure 2. Anti-inflammatory role of melatonin in alleviating chronic pain. Melatonin plays a central role in the coordination between nociceptive neurons and immune cells in which its binding the cross-talk between inflammation and pain transmission. The binding of melatonin to MT2 receptors expressed on immune cells inhibits myeloperoxidase (MPO) activity and NF- κ B, as well as downregulating the production of mitochondrial reactive oxygen species (ROS). Melatonin has also been shown to inhibit the NLRP3 (NOD-, LRR- and pyrin domain-containing protein 3) inflammasome and modulate the downstream expression of several inflammatory genes, including c-fos, calcitonin-gene-related peptide (CGRP), IL-1 β , IL-6, TNF- α and NOS1. In various cell types, melatonin inhibits the phosphorylation of p38 mitogenactivated protein kinase (MAPK). Finally, melatonin increases expression of the catalytic subunit of phosphatase 2A (PP2Ac) which in turn mediates the dephosphorylation of HDAC4, driving its nuclear accumulation to repress the expression of high mobility group box-1 (HMGB-1) and ultimately alleviating chronic pain.

Accumulating evidence suggests that melatonin can modulate the activation of immune system leading to a reduction in acute and chronic inflammation ^[3]. Melatonin demonstrates anti-inflammatory effects through the inhibition of the inflammatory cells activation by inhibiting myeloperoxidase activity ^[47]. Melatonin can produce anti-inflammatory effects through direct interaction with binding sites on macrophages and lymphocytes ^[39]. Melatonin inhibits the release of pro-inflammatory cytokines and downregulates the expression of NFkB in various experimental studies. Consequently, melatonin administration was found to inhibit the release of TNF- α from peritoneal macrophages obtained from mice (**Figure 2**) ^[39]. Melatonin has been shown to inhibit NLRP3 (nucleotide-binding oligomerization domain, leucine-rich-containing family, pyrin domain-containing-3) inflammasome activation under various conditions ^[47]. Interestingly, in an intervertebral disc degeneration (IVDD) rat model of low back pain, melatonin administration prevented IVDD progression and IVDD associated back pain, which was found to result from melatonin induced decrease in NLRP3, p20, and IL-1 β levels due to inhibition of NFkB signaling and down modulation of mitochondrial ROS (reactive oxygen species)

production ^[48]. In another neuropathic pain study, it was revealed that melatonin via MT2 receptors in the DRG (Dorsal root ganglion) could alleviate the allodynia and hyperalgesia in a sciatic nerve cuffing model of mice (**Figure 2**) ^[36]. Interestingly, melatonin inhibited the activation of peptogenic neurons and neuroinflammation in DRG through downregulation of inflammatory genes such as c-fos, calcitonin-gene related peptide (CGRP), TNF- α , IL-1 β and nitric oxide synthase 1 (NOS1) ^[36].

In an Oxaliplatin induced neuropathic pain model, melatonin administration attenuated the astrocyte mediated spinal neuroinflammation and consequently, reduced pain hypersensitivity [49]. Another study found that melatonin administration significantly reduced the neuropathic pain in a chronic constrictive injury with sleep deprivation experimental model and exhibited significant inhibition of the microglial activation along with lower levels of inflammatory cytokines such as TNF-α, IL-1 β and IL-6 (Figure 2) [44]. Interestingly, chronic constrictive injury of the median nerve in the cuneate nucleus led to the phosphorylation of p38 mitogen activated protein kinase (p38-MAPK) and microglial activation, which was associated with behavioral hypersensitivity ^[50]. Administration of melatonin reduced behavioral hypersensitivity, phosphorylation of p38-MAPK in microglia and consequently, reduced inflammatory cytokines secretion ^[50]. Intriguingly, a spinal nerve ligation (SNL) pain model was used to unveil the melatonin's underlying epigenetic mechanisms of chronic pain alleviation by controlling the inflammatory genes expression [51]. SNL resulted in the reduced expression of the catalytic subunit of phosphatase 2A (PP2Ac) and increased histone deacetylase 4 (HDAC4) phosphorylation and cytoplasmic accumulation, which epigenetically relieved HDAC4-suppressed hmgb1 gene transcription, resulting in enhanced high mobility group box-1 (HMGB-1) expression in the ipsilateral dorsal horn of the rats (Figure 2). Notably, melatonin acting via MT2 receptors upon intrathecal administration increased PP2Ac expression, HDAC4 dephosphorylation and nuclear accumulation, restored the HDCA4 mediated suppression of hmgb1, and thus, alleviated the SNL induced behavioral pain (Figure 2) [51].

All this evidence supports that melatonin can modulate the triad of chronic pain, sleep disturbance and inflammation. However, detailed clinical studies assessing the impact of Melatonin on levels of various inflammatory mediators in chronic back pain patients with disrupted sleep are lacking.

References

- 1. Arnao, M.B. Phytomelatonin: Discovery, Content, and Role in Plants. Adv. Bot. 2014.
- 2. Hardeland, R. Melatonin and inflammation—Story of a double-edged blade. J. Pineal. Res. 2018, 65, e12525.
- Tarocco, A.; Caroccia, N.; Morciano, G.; Wieckowski, M.R.; Ancora, G.; Garani, G.; Pinton, P. Melatonin as a master re gulator of cell death and inflammation: Molecular mechanisms and clinical implications for newborn care. Cell Death Di s. 2019, 10, 317.
- 4. Brennan, R.; Jan, J.E.; Lyons, C.J. Light, dark, and melatonin: Emerging evidence for the importance of melatonin in oc ular physiology. Eye 2007, 21, 901–908.
- 5. Ohashi, N.; Ishigaki, S.; Isobe, S. The pivotal role of melatonin in ameliorating chronic kidney disease by suppression o f the renin-angiotensin system in the kidney. Hypertens Res. 2019, 42, 761–768.
- Welin, A.-K.; Svedin, P.; Lapatto, R.; Sultan, B.; Hagberg, H.; Gressens, P.; Kjellmer, I.; Mallard, C. Melatonin Reduces I nflammation and Cell Death in White Matter in the Mid-Gestation Fetal Sheep Following Umbilical Cord Occlusion. Pedi atric Res. 2007, 61, 153–158.
- Grigorov, I.; Bogojević, D.; Jovanović, S.; Petrović, A.; Ivanović-Matić, S.; Zolotarevski, L.; Poznanović, G.; Martinović, V. Hepatoprotective effects of melatonin against pronecrotic cellular events in streptozotocin-induced diabetic rats. J. P hysiol. Biochem. 2014, 70, 441–450.
- 8. Maestroni, G.J.M. Melatonin and the Immune System Therapeutic Potential in Cancer, Viral Diseases, and Immunodefi ciency States. In The Pineal Gland and Cancer; Springer: Berlin/Heidelberg, Germany, 2001; pp. 384–394.
- Slominski, R.M.; Reiter, R.J.; Schlabritz-Loutsevitch, N.; Ostrom, R.S.; Slominski, A.T. Melatonin membrane receptors i n peripheral tissues: Distribution and functions. Mol. Cell Endocrinol. 2012, 351, 152–166.
- 10. Almughrabi, O.M.; Marzouk, K.M.; Hasanato, R.M.; Shafik, S.S. Melatonin levels in periodontal health and disease. J. P eriodontal Res. 2013, 48, 315–321.
- 11. Timothy, D.R.; Jain, S.; Hunter, C.; Chakravarthy, K. Neurostimulation for Intractable Chronic Pain. Brain Sci. 2019, 9, 2 3.
- 12. Deer, T.R.; Falowski, S.; Arle, J.E.; Vesper, J.; Pilitsis, J.; Slavin, K.V.; Hancu, M.; Grider, J.S.; Mogilner, A.Y. A Systema tic Literature Review of Brain Neurostimulation Therapies for the Treatment of Pain. Pain Med. 2020, 21, 1415–1420.

- Deer, T.R.; Pope, J.E.; Lamer, T.J.; Grider, J.S.; Provenzano, D.; Lubenow, T.R.; FitzGerald, J.J.; Hunter, C.; Falowski, S.; Sayed, D.; et al. The neuromodulation appropriateness consensus committee on best practices for dorsal root gangl ion stimulation. Neuromodulation 2019, 22, 1–35.
- Deer, T.R.; Levy, R.M.; Kramer, J.; Poree, L.; Amirdelfan, K.; Grigsby, E.; Staats, P.; Burgher, A.H.; Scowcroft, J.; Golov ac, S.; et al. Comparison of paresthesia coverage of patient's pain: Dorsal root ganglion vs. spinal cord stimulation. An ACCURATE study sub-analysis. Neuromodulation Technol. Neural Interface 2019, 22, 930–936.
- 15. Pope, J.E.; Falowski, S.; Deer, T.R. Advanced waveforms and frequency with spinal cord stimulation: Burst and high-fre quency energy delivery. Expert Rev. Med. Devices 2015, 12, 431–437.
- 16. Lendvai, I.S.; Maier, A.; Scheele, D.; Hurlemann, R.; Kinfe, T.M. Spotlight on cervical vagus nerve stimulation for the tre atment of primary headache disorders: A review. J. Pain Res. 2018, 11, 1613–1625.
- 17. Grazzi, L.; Tassorelli, C.; de Tommaso, M.; Pierangeli, G.; Martelletti, P.; Rainero, I.; Gepetti, P.; Ambrosini, A.; Sarchiell i, P.; Liebler, E.; et al. Practical and clinical utility of non-invasive vagus nerve stimulation (nVNS) for the acute treatmen t of migraine: A post hoc analysis of the randomized, sham-controlled, double-blind PRESTO trial. J. Headache Pain 20 18, 19, 98.
- Silberstein, S.D.; Calhoun, A.H.; Lipton, R.B.; Grosberg, B.M.; Cady, R.K.; Dorlas, S.; Simmons, K.A.; Mullin, C.; Lieble r, E.J.; Goadsby, P.J.; et al. Chronic migraine headache prevention with non-invasive vagus nerve stimulation: The EVE NT study. Neurology 2016, 87, 1–10.
- Silberstein, S.D.; Mechtler, L.L.; Kudrow, D.B.; Calhoun, A.H.; McClure, C.; Saper, J.R.; Liebler, E.J.; Engel, E.R.; Tepp er, S.J.; ACT1 Study Group. Non-Invasive Vagus Nerve Stimulation for the ACute Treatment of Cluster Headache: Findi ngs From the Randomized, Double-Blind, Sham-Controlled ACT1 Study. Headache 2016, 56, 1317–1332.
- Goadsby, P.J.; de Coo, I.F.; Silver, N.; Tyagi, A.; Ahmed, F.; Gaul, C.; Jensen, R.H.; Diener, H.C.; Solbach, K.; Straube, A.; et al. Non-invasive vagus nerve stimulation for the acute treatment of episodic and chronic cluster headache: A rand omized, double-blind, sham-controlled ACT2 study. Cephalalgia 2018, 38, 959–969.
- 21. Gaul, C.; Diener, H.C.; Silver, N.; Magis, D.; Reuter, U.; Andersson, A.; Liebler, E.J.; Straube, A.; PREVA Study Group. Non-invasive vagus nerve stimulation for PREVention and Acute treatment of chronic cluster headache (PREVA): A ran domized controlled study. Cephalalgia 2015, 36, 534–546.
- 22. Gaul, C.; Magis, D.; Liebler, E.; Straube, A. Effects of non-invasive vagus nerve stimulation on attack frequency over ti me and expanded response rates in patients with chronic cluster headache: A post hoc analysis of the randomized, con trolled PREVA study. J. Headache Pain 2017, 18, 22.
- 23. O'Connell, N.E.; Marston, L.; Spencer, S.; DeSouza, L.H.; Wand, B.M. Non-invasive brain stimulation techniques for ch ronic pain. Cochrane Database Syst. Rev. 2018, 3, CD008208.
- 24. Kinfe, T.M.; Asif, M.; Chakravarthy, K.V.; Deer, T.R.; Kramer, J.M.; Yearwood, T.L.; Hurlemann, R.; Hussain, M.S.; Mota meny, S.; Wagle, P.; et al. Unilateral L4-dorsal root ganglion stimulation evokes pain relief in chronic neuropathic postsu rgical knee pain and changes of inflammatory markers: Part II whole transcriptome profiling. J. Transl. Med. 2019, 17, 1 –13.
- 25. Chaudhry, S.R.; Lendvai, I.S.; Muhammad, S.; Westhofen, P.; Kruppenbacher, J.; Scheef, L.; Boecker, H.; Scheele, D.; Hurlemann, R.; Kinfe, T.M. Inter-ictal assay of peripheral circulating inflammatory mediators in migraine patients under adjunctive cervical non-invasive vagus nerve stimulation (nVNS), A proof-of-concept study. Brain Stimul. 2019, 12, 643 –651.
- 26. Gravius, N.; Chaudhry, S.R.; Muhammad, S.; Boström, A.; Gravius, S.; Randau, T.; Scheele, D.; Westhofen, P.; Kruppe nbacher, J.; Stoffel-Wagner, B.; et al. Selective L4 Dorsal Root Ganglion Stimulation Evokes Pain Relief and Changes of Inflammatory Markers, Part I Profiling of Saliva and Serum Molecular Patterns. Neuromodulation 2019, 22, 44–52.
- 27. Chakravarthy, K.V.; Xing, F.; Bruno, K.; Kent, A.R.; Raza, A.; Hurlemann, R.; Kinfe, T.M. A Review of Spinal and Periphe ral Neuromodulation and Neuroinflammation: Lessons Learned Thus Far and Future Prospects of Biotype Developmen t. Neuromodulation 2019, 22, 235–243.
- 28. Brzezinski, A. Melatonin in humans. N. Engl. J. Med. 1997, 336, 186–195.
- 29. Xie, S.; Fan, W.; He, H.; Huang, F. Role of Melatonin in the Regulation of Pain. J. Pain Res. 2020, 13, 331–343.
- Srinivasan, V.; Lauterbach, E.C.; Ho, K.Y.; Acuna-Castroviejo, D.; Zakaria, R.; Brzezinski, A. Melatonin in antinociceptio n: Its therapeutic applications. Curr. Neuropharmacol. 2012, 10, 167–178.
- 31. Ng, K.Y.; Leong, M.K.; Liang, H.; Paxinos, G. Melatonin receptors: Distribution in mammalian brain and their respective putative functions. Brain Struct. Funct. 2017, 222, 2921–2939.
- 32. Posa, L.; De Gregorio, D.; Gobbi, G.; Comai, S. Targeting Melatonin MT2 Receptors: A Novel Pharmacological Avenue for Inflammatory and Neuropathic Pain. Curr. Med. Chem. 2018, 25, 3866–3882.

- 33. Bonica, J.J. The need of a taxonomy. Pain 1979, 6, 247-248.
- 34. Park, B.Y.; Lee, J.J.; Kim, H.J.; Woo, C.W.; Park, H. A neuroimaging marker for predicting longitudinal changes in pain i ntensity of subacute back pain based on large-scale brain network interactions. Sci. Rep. 2020, 10, 1–11.
- 35. Almay, B.G.; von Knorring, L.; Wetterberg, L. Melatonin in serum and urine in patients with idiopathic pain syndromes. Psychiatry Res. 1987, 22, 179–191.
- Kuthati, Y.; Lin, S.-H.; Chen, I.-J.; Wong, C.S. Melatonin and their analogs as a potential use in the management of Ne uropathic pain. J. Formos. Med. Assoc. 2019, 118, 1177–1186.
- 37. Danilov, A.; Kurganova, J. Melatonin in Chronic Pain Syndromes. Pain Ther. 2016, 5, 1–17.
- Kurganova, Y.M.; Danilov, A.B. The Role of Melatonin in the Treatment of Chronic Back Pain. Neurosci. Behav. Physiol. 2016, 46, 737–742.
- Ambriz-Tututi, M.; Rocha-González, H.I.; Cruz, S.L.; Granados-Soto, V. Melatonin: A hormone that modulates pain. Life Sci. 2009, 84, 489–498.
- 40. Koes, B.W.; Backes, D.; Bindels, P.J.E. Pharmacotherapy for chronic non-specific low back pain: Current and future opt ions. Expert Opin. Pharmacother. 2018, 19, 537–545.
- 41. Moawad, H. Sleep Disturbances as a Sequalae of Chronic Pain. Psychiatr. Times 2020, 37, 22–27.
- 42. Cheatle, M.D.; Foster, S.; Pinkett, A.; Lesneski, M.; Qu, D.; Dhingra, L. Assessing and Managing Sleep Disturbance in Patients with Chronic Pain. Anesthesiol. Clin. 2016, 34, 379–393.
- 43. Wilhelmsen, M.; Amirian, I.; Reiter, R.J.; Rosenberg, J.; Gögenur, I. Analgesic effects of melatonin: A review of current e vidence from experimental and clinical studies. J. Pineal Res. 2011, 51, 270–277.
- 44. Huang, C.T.; Chiang, R.P.; Chen, C.L.; Tsai, Y.J. Sleep deprivation aggravates median nerve injury-induced neuropathic pain and enhances microglial activation by suppressing melatonin secretion. Sleep 2014, 37, 1513–1523.
- 45. Barchas, J.; Dacosta, F.; Spector, S. Acute Pharmacology of Melatonin. Nature 1967, 214, 919–920.
- 46. Baral, P.; Udit, S.; Chiu, I.M. Pain and immunity: Implications for host defence. Nat. Rev. Immunol. 2019, 19, 433–447.
- 47. Hardeland, R. Melatonin in aging and disease -multiple consequences of reduced secretion, options and limits of treat ment. Aging Dis. 2012, 3, 194–225.
- 48. Chen, F.; Jiang, G.; Liu, H.; Li, Z.; Pei, Y.; Wang, H.; Pan, H.; Cui, H.; Long, J.; Wang, J.; et al. Melatonin alleviates inter vertebral disc degeneration by disrupting the IL-1β/NF-κB-NLRP3 inflammasome positive feedback loop. Bone Res. 20 20, 8, 1–13.
- 49. Wang, Y.S.; Li, Y.Y.; Cui, W.; Li, L.B.; Zhang, Z.C.; Tian, B.P.; Zhang, G.S. Melatonin Attenuates Pain Hypersensitivity a nd Decreases Astrocyte-Mediated Spinal Neuroinflammation in a Rat Model of Oxaliplatin-Induced Pain. Inflammation 2017, 40, 2052–2061.
- 50. Chiang, R.P.; Huang, C.T.; Tsai, Y.J. Melatonin reduces median nerve injury-induced mechanical hypersensitivity via inh ibition of microglial p38 mitogen-activated protein kinase activation in rat cuneate nucleus. J. Pineal Res. 2013, 54, 232 –244.
- 51. Lin, T.B.; Hsieh, M.C.; Lai, C.Y.; Cheng, J.K.; Wang, H.H.; Chau, Y.P.; Chen, G.D.; Peng, H.Y. Melatonin relieves neuro pathic allodynia through spinal MT 2-enhanced PP 2Ac and downstream HDAC 4 shuttling-dependent epigenetic modif ication of hmgb1 transcription. J. Pineal Res. 2016, 60, 263–276.

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