Huperzine A and Its Neuroprotective Molecular Signaling

Subjects: Neurosciences

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The neuroprotective effects of HupA treatment result from cholinergic signaling, which increases Wnt signaling, nonamyloidogenic processing of the amyloid precursor protein (APP), neurotrophic factor expression, synaptic activity, antagonism of the N-methyl-D-aspartate (NMDA) receptor (NMDAR), modulation of ROS, iron homeostasis and neuronal survival.

neurodegenerative diseases   huperzine   AChEI   Alzheimer's disease

therapeutic potential   neuroinflammation

1. Introduction

Its IUPAC name is (1R,9S,13E)-1-amino-13-ethylidene-11-methyl-6-azatricyclo[7.3.1.02,7]trideca-2(7),3,10-trien-5-one. With a molecular weight of 242.32 g/mol [1][2], HupA presents a complex array of structural components: a compact tricyclic structure with a bicyclo [3.1.1] skeleton fused with an α-pyridone ring, an exocyclic ethylidene moiety, and a 3-carbon bridge with an -NH 2 group. This complex and unique molecular structure accounts for the interesting biological functions and the high stability of HupA [3], plus it results in the presentation of the correct electrostatic field for binding to AChE [4][5]. The three-dimensional structure of AChE complexed with the nootropic alkaloid [-]-HupA was resolved at the Weizmann Institute of Science, Israel [6]. Notably, chemically synthesized [−]-HupA inhibits AChE with similar effectiveness as the natural molecule [2], and [+] -HupA 's AChE inhibitor (AChEI) activity is at least 50-fold less potent than that of the naturally occurring stereoisomer [7].

The enzyme AChE catalyzes the hydrolysis of the neurotransmitter acetylcholine into choline and acetic acid, regulating the return of cholinergic neurons to a resting state [8]. Most of the cortical AChE activity present in the brain of AD patients is known to be predominantly associated with the amyloid core of senile plaques [9][10]. The globular tetramer (G4) of AChE present in the central nervous system (CNS) [11] (see **Figure 1**), forms a stable toxic complex with the amyloid-β (Aβ) peptide during its assembly into Aβ filaments and increases the aggregation and neurotoxicity of amyloid fibrils [12][13][14].
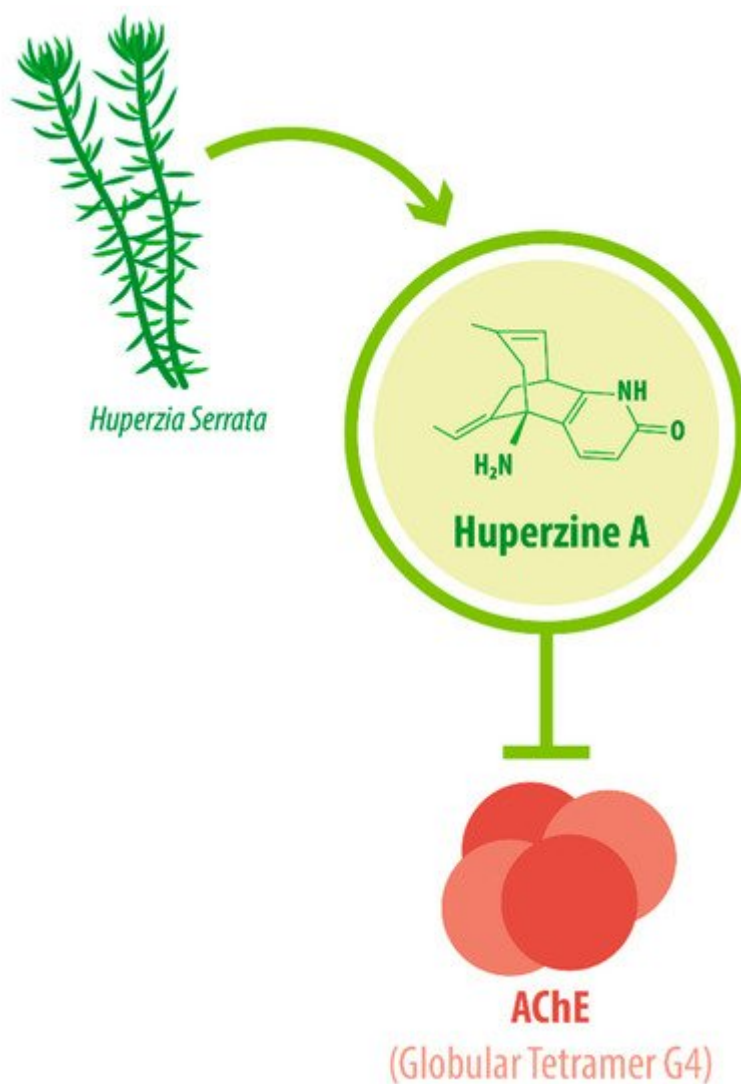
Figure 1. HupA is an acetylcholinesterase inhibitor (AChEI). HupA has a molecular weight of 242.32 g/mol and acts by inhibiting AChE. In the CNS, AChE is present in a tetrameric form anchored to the membrane, or "Globular Tetramer G4" (G4). HupA is the main biological compound in *H. serrata*, which has been used with medicinal purposes in China for centuries. (AChE: acetylcholinesterase; CNS: central nervous system).

Due to its potent AChEI activity, HupA is used as a treatment for AD. According to the World Health Organization (WHO), 135.5 million people worldwide will be living with dementia by the year 2050. AD accounts for 65% of all dementia cases [15]. It is a progressive neurological degenerative disorder affecting memory, among other cognitive functions [16][17][18]. Vascular-associated dementia (VaD), on the other hand, is caused by various cerebrovascular diseases, such as cerebral infarction and hemorrhage [19]. These conditions impose great monetary and emotional costs on patients and their caregivers. HupA is approved as the drug of choice for Alzheimer's disease (AD) treatment in China and as a dietary supplement in the United States [20][21], as intake can improve AD pathogenesis [22].

The biological functions of HupA have been studied both in vitro and in vivo. The present review aims to provide an integrated perspective of the neuroprotective molecular signaling of HupA in dementia, especially in AD.

2. HupA within the CNS: Modulation of Critical Pathological Conditions

One of the hallmarks of AD is the accumulation of Aβ peptide (both oligomers and fibrils) that associate with AChE to form amyloid plaque, producing neuronal cell death [12][13]. The neuroprotective effect of HupA is in part mediated by a dose-dependent reduction in subcellular amyloid-β accumulation in the cortex and hippocampus as shown in a double transgenic mice model carrying the Swedish-mutant amyloid precursor protein (APP) and a deletion in exon 9 of presenilin 1 (PS1) (known as APPswe/PS1 mice) [23], although it is noteworthy that this effect is reportedly absent in TgCRND8 mice [24]. These reductions occur at the protein level through a dose-dependent modulation of the activity of β-site of the APP-cleaving enzyme (BACE1) activity, a reduction in PS1 expression, and a marked increase in α -secretase cleavage, promoting the non-amyloidogenic processing of APP [23][25]. An improvement in spontaneous working memory of 68% has been observed in APPswe/PS1 mice after HupA treatment, although HupA does not seem to improve cognition in wild-type (WT) mice [26]. In primary cortical neuron cultures, HupA ameliorates oligomeric Aβ42-induced neurotoxicity by reducing the intracellular accumulation of Aβ42 [27].

At the same time, HupA treatment attenuates the Aβ load in brain mitochondrial homogenates and ameliorates the mitochondrial swelling observed in APPswe/PS1 mice, although it has no effect on WT mice [26]. It has been reported that Aβ accumulation causes mitochondrial dysfunction, resulting in neurotoxicity. HupA treatment restores adenosine triphosphate (ATP) levels and reduces reactive oxygen species (ROS) levels increased by Aβ42 in neurons, and prevents Aβ42-mediated destabilization of the mitochondrial membrane [27]. Notably, HupA treatment reduces Aβ42 accumulation in mitochondria-enriched cellular fractions, but has no significant effects on membrane or cytosol Aβ42 levels, indicating that it exerts a specifically mitochondrial effect [27].

HupA also ameliorates Aβ25–35-induced apoptosis. A proteomic analysis showed that HupA downregulates 29 proteins, including the tumor suppressor protein p53, which has a direct link with apoptosis [28]. In a co-culture system of neural stem cells and microglia exposed to Aβ1–42, HupA treatment partially reduced the secretion of inflammatory factors interleukin-6 (IL-6), tumor necrosis factor-α (TNF-α), and macrophage inflammatory protein-1α (MIP-1α) [28]. HupA also significantly increases the ratio between B-cell lymphoma-2 (Bcl-2) and Bcl-2-like protein 4 (Bax), resulting in improved cell viability. Similarly, HupA directly acts on microglial cells to reduce the expression of cytokines and chemokines [29]. In primary astrocyte cultures, HupA preincubation reduces Aβ1-42-induced cell damage, preventing peaks in the release of p65 subunit of nuclear factor kappa-light-chain-enhancer of activated B cells (NF-kB) [30].

Several studies have evaluated the potential benefits of HupA in human patients. HupA treatment in human subjects suffering from dementia (AD o VaD) shows evidence of improved cognition [31][19][32][33]. It has also been reported that eight weeks of HupA treatment for AD patients improved task switching and alleviated cognitive impairment [34]. The benefits of HupA treatment are apparent when using measurement tools such as Mini-Mental State Exam (MMSE), the AD Assessment Scale-cognitive subscale (ADAS-Cog)) or the ADAS-noncognitive subscale (non-COG) tests. Measuring with tools such as Hastgawa Dementia Scale (HDS) or Wechsler Memory

Scale (WMS) tests showed no significant improvement from HupA treatment [15][16]. These results were corroborated in a randomized clinical trial in which HupA treatment in VaD patients significantly improved cognitive function according to the MMSE and Activities of Daily Living (ADL) tests but not according to the Clinical Dementia Rating (CDR) scores [15][19]. However, other trials conclude that HupA treatment could be more beneficial than psychotherapy and conventional medicine for VaD patients [31]. A small trial testing the effects of HupA tablets in AD patients showed that the intake of 0.2 mg of HupA improved cognition and memory in 58% of patients with no severe side effects [35], and a phase II multicenter, 3-arm randomized, double-blind placebo-controlled trial, evaluated as having high methodological quality in later reviews [31], showed significant cognitive enhancement in patients receiving 0.4 mg of HupA twice a day. Remarkably, this dose was well tolerated for 24 weeks even though most AD patients reported being unable to tolerate currently marketed AChEIs for a long period of time [32][36]. A more recent analysis concludes that HupA improves cognition in AD patients, and that its effects are dose and duration-dependent [37].

3. Neuroprotective Molecular Signaling of HupA against AD

The neuroprotective effects of HupA treatment result from cholinergic signaling, which increases neurotrophic factor expression, synaptic activity, antagonism of the N-methyl-D-aspartate (NMDA) receptor (NMDAR), modulation of ROS, and neuronal survival [38][1].

Activation of mAChRs and protein kinase C (PKC)/MAPK pathways have been proposed as a mechanism for this effect [7][39][40], as well as HupA mediated increase of phosphorylation of ERK1/2, that can also regulate ADAM10 transcription. Another mechanism for the cholinergic modulation of APP processing is the activation of Wnt signaling. Wnt signaling plays a crucial role in presynaptic assembly, postsynaptic differentiation at vertebrate peripheral neuromuscular synapses, stem cell differentiation, and the development and maintenance of the CNS [41][42][43]. Recent evidence suggests that it also regulates vascular stability, BBB integrity, and inflammation [44]. In the canonical Wnt pathway or Wnt/β-catenin pathway, Wnt ligands bind to receptors of the Frizzled (FZD) and low-density lipoprotein-related protein (LRP) families on the cell surface and activate several cytoplasmic relay components, including GSK-3β and adenomatous polyposis coli (APC), finally signaling to β-catenin. β-catenin enters the nucleus and forms a complex with lymphoid enhancer-binding factor 1 (LEF1) to activate the transcription of Wnt target genes. Wnt signaling pathways that are independent of β-catenin are referred to as non-canonical pathways, which include the planar cell polarity (PCP) pathway, also known as the Wnt/Jun N-terminal kinase (JNK) pathway, and the Wnt/ Ca 2+ pathway [41] (see **Figure 2**).
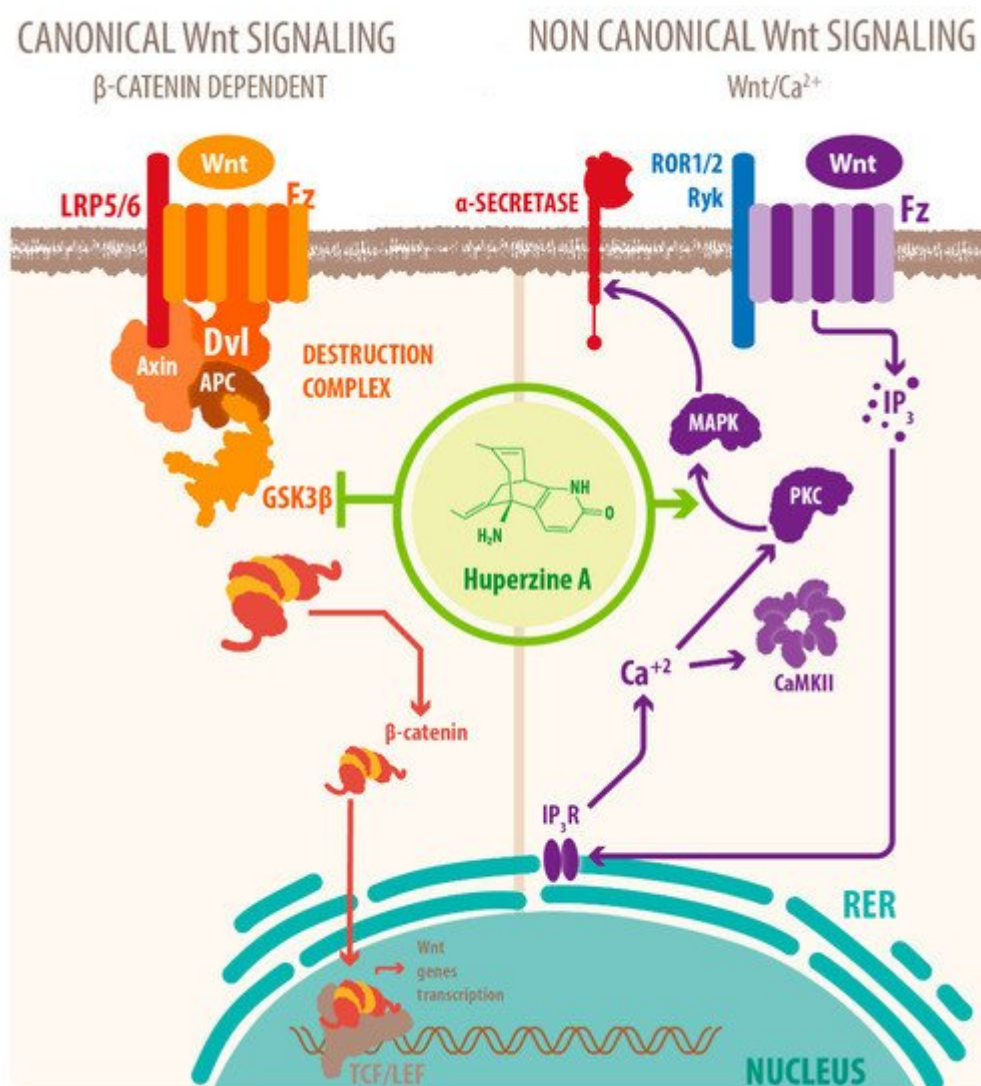
Figure 2. HupA affects Wnt signaling. HupA affects both canonical and noncanonical Wnt signaling. Regarding the canonical signaling, HupA modulates the Wnt/β-catenin pathway by downregulating GSK-3β activity. HupA increases the phosphorylation levels of GSK-3α/β, which stabilizes β-catenin and upregulates expression of its target genes, related to synaptic plasticity and neuronal survival, while preventing tau hyperphosphorylation and Aβ neurotoxicity. HupA also interacts with the noncanonical pathway via PKC/MAPK signaling. PKC activation also inhibits GSK-3β activity and plays a role in upregulating the expression of the α-secretase, preventing Aβ accumulation and neuronal apoptosis (Wnt: wingless-related integration site; GSK3: glycogen synthase kinase 3; PKC: protein kinase C; MAPK: mitogen-activated protein kinase).

Regarding Fe 2+ homeostasis, it has been reported that AD mice models have elevated levels of Fe 2+ in the cortex and hippocampus compared with WT mice. Accordingly, the expression of transferrin receptor protein 1 (TFR1) is upregulated in APPswe/PS1dE9 mice, which explains the increase in Fe 2+ uptake [25]. Fe 2+ -mediated neurodegeneration might originate in the downregulation of α-secretase activity, inhibiting the nonamyloidogenic pathway, with concomitant increase of APP expression due to a functional iron-responsive element (IRE) that has been identified in the APP promoter, which would explain why APP levels increase after Fe 2+ treatment and decrease after chelator treatment. An alternative explanation could be that the translational expression of APP

could be directed via release of a classic repressor interaction between APP mRNA and iron regulatory protein-1 (IRP-1) [25]. HupA might act directly as an Fe 2+ chelator, reducing the capacity of IRP-1 to induce APP translation [25]. HupA also downregulates TFR1 expression in mice in vivo, which reduces the uptake of transferrin-bound iron (TBI) in neurons [25] (see **Figure 3**) (see **Table 1**).
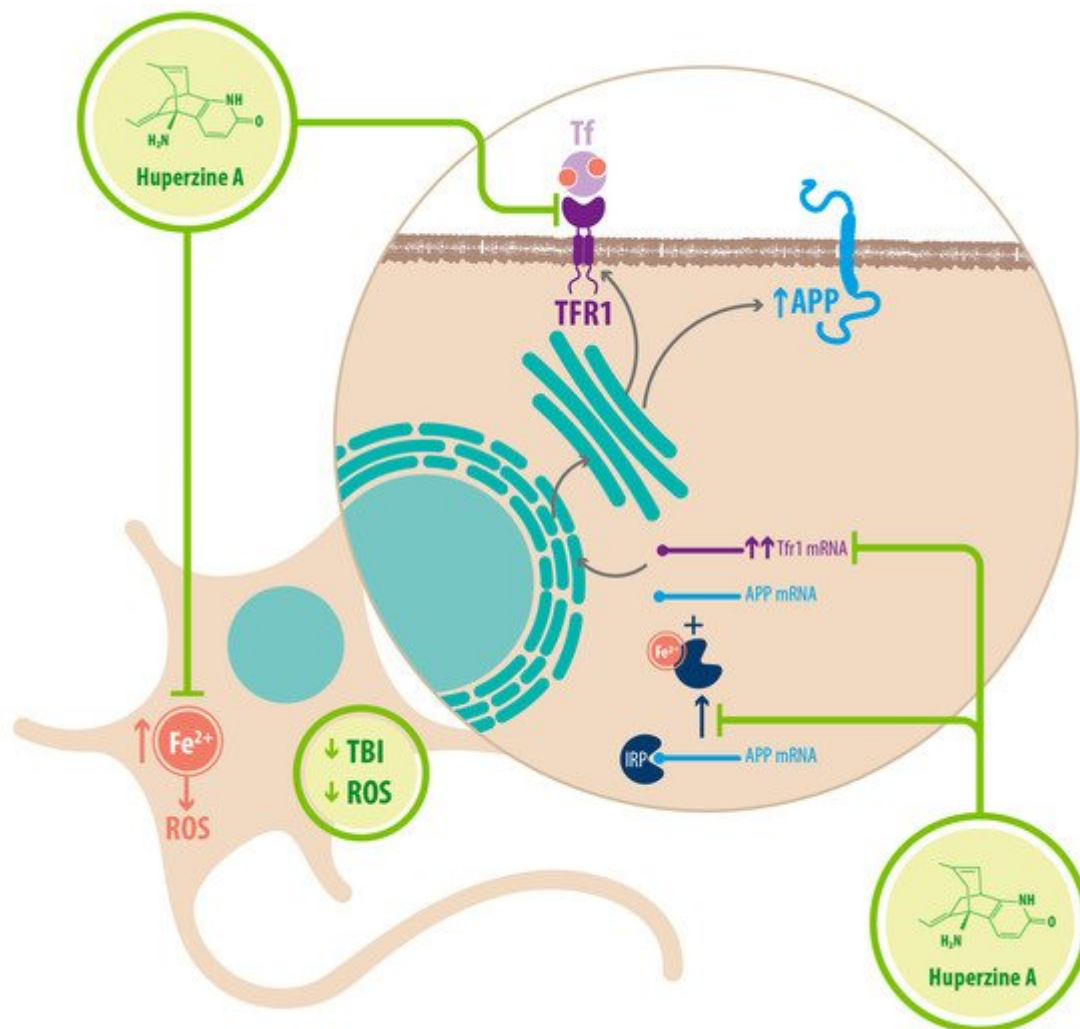
Figure 1. HupA regulates Fe^{2+} homeostasis. HupA prevents the accumulation of Fe^{2+} inside the neurons. It might act as a chelator, while also downregulating the expression of TFR1 and preventing Fe^{2+} uptake into the cell. By reducing Fe^{2+} concentration inside the neuron, HupA reduces APP translation. Fe^{2+} binds the IRP, releasing it from APP mRNA which can then get translated. In the presence of HupA, Fe^{2+} concentration drops, and less IRP is released, meaning less APP is translated. HupA indirectly reduces ROS through all these mechanisms (TFR1: transferrin receptor 1; Tf: transferrin; TBI: transferrin bound iron; ROS: reactive oxygen species; APP: amyloid precursor protein; IRP: iron regulatory protein).

Table 1. Molecular targets of HupA's noncholinergic mechanisms in dementia-related pathologies and effects of treatment.

Mechanism	Molecular Target	Effects of HupA Treatment	References
Mitochondrial protection	Upregulation of oxidative phosphorylation proteins	Recovered energetic homeostasis	[45]
	Inhibition of cytoplasmic translocation of CytoC	Prevented mitochondria-mediated apoptosis	[46]
	Downregulation of AIF	Prevented cas3-independent apoptosis	[45]
	Inhibition of Caspase-3 cleavage	Prevented cas-3 dependent apoptosis	[45]
	Upregulation of GSH-PX SOD CAT	Decreased ROS accumulation Decreased oxidative stress	[45]
Fe^{2+} homeostasis	Chelator/ Reducer of Fe^{2+}	Increased nonamyloidogenic processing of APP Downregulated expression of APP through IRP-1	[25]
	Downregulation of TFR1	Decreased Fe^{2+} uptake in neurons	[25]

nAChRs.

S, results

in suppressed inflammatory cytokine expression and systemic inflammatory responses. The activation of α7nAChR downregulates macrophages, microglia, and astrocytes activation inhibiting the production of proinflammatory cytokines while anti-inflammatory cytokine expression is not altered [2][40][47]. The cholinergic anti-inflammatory effects of HupA downregulates IL-1β, IL-6, and TNF-α secretion and NF-kB signaling, attenuating neuroinflammatory response [48].

(CytoC: cytochrome C; Cas-3: caspase-3; AIF: apoptosis inducing factor; CAT: catalase; GSH-PX: glutathione peroxidase; SOD: superoxide dismutase; TFR1: transferrin receptor 1; ROS: reactive oxygen species; APP: amyloid precursor protein; IRP: iron regulatory protein).

4. Concluding Remarks

HupA has therapeutic benefits for the treatment of AD, but the understanding of molecular interactions involved in the recovery of neuronal function after HupA treatment is still incomplete. What is known is that AChEI activity is crucial through the regulation of Aβ peptide accumulation by activation of α-secretase cleavage and down-regulation of β/γ-secretase, enhancing BDNF/TrkB signaling as well as PI3K/Akt and PI3K /TrkB/mTOR pathways. Concomitant reduction of IL-1β, IL-6, TNF-α, and NF-kB signaling preserves neuronal function. Modulation of Wnt signaling through HupA treatment might be the most promising therapeutic target, as Wnt is involved in neuronal survival and in synaptic plasticity. There are also non-cholinergic neuroprotective effects such as the preservation of mitochondrial structure and function under Aβ insult, as well as the recovery of Fe 2+ homeostasis in the brain. Further research and development of HupA as a common treatment might alleviate the burden related to highly prevalent conditions that are costly in terms of public health policies and devastating to those who bear them and their caretakers.

References

1. Ohba, T.; Yoshino, Y.; Ishisaka, M.; Abe, N.; Tsuruma, K.; Shimazawa, M.; Oyama, M.; Tabira, T.; Hara, H. Japanese Huperzia serrata extract and the constituent, huperzine A, ameliorate the scopolamine-induced cognitive impairment in mice. Biosci. Biotechnol. Biochem. 2015, 79, 1838–1844.

2. Damar, U.; Gersner, R.; Johnstone, J.T.; Schachter, S.; Rotenberg, A. Huperzine A as a neuroprotective and antiepileptic drug: A review of preclinical research. Expert Rev. Neurother. 2016, 16, 671–680.

3. Ferreira, A.; Rodrigues, M.; Fortuna, A.; Falcão, A.; Alves, G. Huperzine A from Huperzia serrata: A review of its sources, chemistry, pharmacology and toxicology. Phytochem. Rev. 2016, 15, 51–85.

4. Ashani, Y.; Grunwald, J.; Kronman, C.; Velan, B.; Shafferman, A. Role of tyrosine 337 in the binding of huperzine A to the active site of human acetylcholinesterase. Mol. Pharmacol. 1994, 45, 555–560.

5. Pang, Y.P.; Kozikowski, A.P. Prediction of the binding sites of huperzine A in acetylcholinesterase by docking studies. J. Comput. Aided Mol. Des. 1994, 8, 669–681.

6. Raves, M.L.; Harel, M.; Pang, Y.P.; Silman, I.; Kozikowski, A.P.; Sussman, J.L. Structure of acetylcholinesterase complexed with the nootropic alkaloid, (-)-huperzine A. Nat. Struct. Biol. 1997, 4, 57–63.

7. Tun, M.K.; Herzon, S.B. The pharmacology and therapeutic potential of (-)-huperzine A. Int. J. Exp. Pharmacol. 2012, 4, 113–123.

8. Colović, M.B.; Krstić, D.Z.; Lazarević-Pašti, T.D.; Bondžić, A.M.; Vasić, V.M. Acetylcholinesterase inhibitors: Pharmacology and toxicology. Curr. Neuropharmacol. 2013, 11, 315–335.

9. Geula, C.; Mesulam, M.M.; Saroff, D.M.; Wu, C.K. Relationship between plaques, tangles, and loss of cortical cholinergic fibers in Alzheimer disease. J. Neuropathol. Exp. Neurol. 1998, 57, 63–75.

10. Reyes, A.E.; Chacón, M.A.; Dinamarca, M.C.; Cerpa, W.; Morgan, C.; Inestrosa, N.C. Acetylcholinesterase-Abeta complexes are more toxic than Abeta fibrils in rat hippocampus: Effect on rat beta-amyloid aggregation, laminin expression, reactive astrocytosis, and neuronal cell loss. Am. J. Pathol. 2004, 164, 2163–2174.

11. Fuentes, M.E.; Inestrosa, N.C. Characterization of a tetrameric G4 form of acetylcholinesterase from bovine brain: A comparison with the dimeric G2 form of the electric organ. Mol. Cell. Biochem. 1988, 81, 53–64.

12. Inestrosa, N.C.; Alvarez, A.; Pérez, C.A.; Moreno, R.D.; Vicente, M.; Linker, C.; Casanueva, O.I.; Soto, C.; Garrido, J. Acetylcholinesterase accelerates assembly of amyloid-b-peptides into Alzheimer's fibrils: Possible role of the peripheral site of the enzyme. Neuron 1996, 16, 881–891.

13. Alvarez, A.; Alarcón, R.; Opazo, C.; Campos, E.O.; Muñoz, F.J.; Calderón, F.H.; Dajas, F.; Gentry, M.K.; Doctor, B.P.; De Mello, F.G.; et al. Stable Complexes Involving Acetylcholinesterase and Amyloid-β Peptide Change the Biochemical Properties of the Enzyme and Increase the Neurotoxicity of Alzheimer's Fibrils. J. Neurosci. 1998, 18, 3213–3223.

14. Bartolini, M.; Bertucci, C.; Cavrini, V.; Andrisano, V. β-Amyloid aggregation induced by human acetylcholinesterase: Inhibition studies. Biochem. Pharmacol. 2003, 65, 407–416.

15. Zhou, X.; Cui, G.; Tseng, H.H.; Lee, S.M.; Leung, G.P.; Chan, S.W.; Kwan, Y.W.; Hoi, M.P. Vascular Contributions to Cognitive Impairment and Treatments with Traditional Chinese Medicine. Evid. Based Complementary Altern. Med. 2016, 9627258.

16. Ha, G.T.; Wong, R.K.; Zhang, Y. Huperzine a as potential treatment of Alzheimer's disease: An assessment on chemistry, pharmacology, and clinical studies. Chem. Biodivers. 2011, 8, 1189–1204.

17. Tapia-Rojas, C.; Inestrosa, N.C. Wnt signaling loss accelerates the appearance of neuropathological hallmarks of Alzheimer's disease in J20-APP transgenic and wild-type mice. J. Neurochem. 2018, 144, 443–465.

18. Rahman, M.; Bajgai, J.; Fadriquela, A.; Sharma, S.; Trinh, T.T.; Akter, R.; Lee, K.J. Therapeutic Potential of Natural Products in Treating Neurodegenerative Disorders and Their Future Prospects and Challenges. Molecules 2021, 26, 5327.

19. Xu, Z.Q.; Liang, X.M.; Juan, W.; Zhang, Y.F.; Zhu, C.X.; Jiang, X.J. Treatment with Huperzine A improves cognition in vascular dementia patients. Cell Biochem. Biophys. 2012, 62, 55–58.

20. Yang, Y.; Wang, Z.; Wu, J.; Chen, Y. Chemical Constituents of Plants from the Genus Phlegmariurus. Chem. Biodivers. 2016, 13, 269–274.

21. Orhan, I.E.; Orhan, G.; Gurkas, E. An overview on natural cholinesterase inhibitors—A multi-targeted drug class-and their mass production. Mini Rev. Med. Chem. 2011, 11, 836–842.

22. Chauhan, P.S.; Yadav, D. Dietary Nutrients and Prevention of Alzheimer's disease. CNS Neurol. Disord. Drug Targets 2021. (E-pub Ahead of Print).

23. Wang, C.Y.; Zheng, W.; Wang, T.; Xie, J.W.; Wang, S.L.; Zhao, B.L.; Teng, W.P.; Wang, Z.Y. Huperzine A activates Wnt/β-catenin signaling and enhances the nonamyloidogenic pathway in an Alzheimer transgenic mouse model. Neuropsychopharmacology 2011, 36, 1073–1089.

24. Yuan, Q.; Lin, Z.X.; Wu, W.; Albert, W.N.; Zee, B.C.Y. Huperzine A in treatment of amyloid-β-associated neuropathology in a mouse model of Alzheimer disease: Abridged secondary publication. Hong Kong Med. J. 2020, 26 (Suppl.8), 34–37.

25. Huang, X.T.; Qian, Z.M.; He, X.; Gong, Q.; Wu, K.C.; Jiang, L.R.; Lu, L.N.; Zhu, Z.J.; Zhang, H.Y.; Yung, W.H.; et al. Reducing iron in the brain: A novel pharmacologic mechanism of huperzine A in

the treatment of Alzheimer's disease. Neurobiol. Aging 2014, 35, 1045–1054.

26. Yang, L.; Ye, C.Y.; Huang, X.T.; Tang, X.C.; Zhang, H.Y. Decreased accumulation of subcellular amyloid-β with improved mitochondrial function mediates the neuroprotective effect of huperzine A. J. Alzheimer's Dis. 2012, 31, 131–142.

27. Lei, Y.; Yang, L.; Ye, C.Y.; Qin, M.Y.; Yang, H.Y.; Jiang, H.L.; Tang, X.C.; Zhang, H.Y. Involvement of Intracellular and Mitochondrial Aβ in the Ameliorative Effects of Huperzine A against Oligomeric Aβ42-Induced Injury in Primary Rat Neurons. PLoS ONE 2015, 10, e0128366.

28. Tao, Y.; Fang, L.; Yang, Y.; Jiang, H.; Yang, H.; Zhang, H.; Zhou, H. Quantitative proteomic analysis reveals the neuroprotective effects of huperzine A for amyloid beta treated neuroblastoma N2a cells. Proteomics 2013, 13, 1314–1324.

29. Zhu, N.; Lin, J.; Wang, K.; Wei, M.; Chen, Q.; Wang, Y. Huperzine A protects neural stem cells against Aβ-induced apoptosis in a neural stem cells and microglia co-culture system. Int. J. Clin. Exp. Pathol. 2015, 8, 6425–6433.

30. Xie, L.; Jiang, C.; Wang, Z.; Yi, X.; Gong, Y.; Chen, Y.; Fu, Y. Effect of Huperzine A on Aβ-induced p65 of astrocyte in vitro. Biosci. Biotechnol. Biochem. 2016, 80, 2334–2337.

31. Yang, G.; Wang, Y.; Tian, J.; Liu, J.P. Huperzine A for Alzheimer's disease: A systematic review and meta-analysis of randomized clinical trials. PLoS ONE 2013, 8, e74916.

32. Rafii, M.S.; Walsh, S.; Little, J.T.; Behan, K.; Reynolds, B.; Ward, C.; Jin, S.; Thomas, R.; Aisen, P.S. A phase II trial of huperzine A in mild to moderate Alzheimer disease. Neurology 2011, 76, 1389–1394.

33. Xing, S.H.; Zhu, C.X.; Zhang, R.; An, L. Huperzine a in the treatment of Alzheimer's disease and vascular dementia: A meta-analysis. Evid. Based Complementary Altern. Med. 2014, 363985.

34. Gul, A.; Bakht, J.; Mehmood, F. Huperzine-A response to cognitive impairment and task switching deficits in patients with Alzheimer's disease. J. Chin. Med Assoc. 2019, 82, 40–43.

35. Xu, S.S.; Gao, Z.X.; Weng, Z.; Du, Z.M.; Xu, W.A.; Yang, J.S.; Zhang, M.L.; Tong, Z.H.; Fang, Y.S.; Chai, X.S.; et al. Efficacy of tablet huperzine-A on memory, cognition, and behavior in Alzheimer's disease. Zhongguo Yao Li Xue Bao 1995, 16, 391–395.

36. Tsai, S. Huperzine-A, a versatile herb, for the treatment of Alzheimer's disease. Crit. Care Med. 2019, 82, 750–751.

37. Ghassab-Abdollahi, N.; Mobasseri, K.; Dehghani Ahmadabad, A.; Nadrian, H.; Mirghafourvand, M. The effects of Huperzine A on dementia and mild cognitive impairment: An overview of systematic reviews. Phytother. Res. 2021, 35, 4971–4987.

38. Wu, T.Y.; Chen, C.P.; Jinn, T.R. Traditional Chinese medicines and Alzheimer's disease. Taiwan J. Obstet. Gynecol. 2011, 50, 131–135.

39. Zhang, H.Y. New insights into huperzine A for the treatment of Alzheimer's disease. Acta Pharmacol. Sin. 2012, 33, 1170–1175.

40. Damar, U.; Gersner, R.; Johnstone, J.T.; Schachter, S.; Rotenberg, A. Huperzine A: A promising anticonvulsant, disease modifying, and memory enhancing treatment option in Alzheimer's disease. Med. Hypotheses 2017, 99, 57–62.

41. Inestrosa, N.C.; Arenas, E. Emerging roles of Wnts in the adult nervous system. Nat. Rev. Neurosci. 2010, 11, 77–86.

42. Oliva, C.A.; Montecinos-Oliva, C.; Inestrosa, N.C. Wnt Signaling in the Central Nervous System: New Insights in Health and Disease. Prog. Mol. Biol. Transl. Sci. 2018, 153, 81–130.

43. Varela-Nallar, L.; Inestrosa, N.C. Wnt signaling in the regulation of adult hippocampal neurogenesis. Front. Cell. Neurosci. 2013, 7, 100.

44. Menet, R.; Lecordier, S.; ElAli, A. Wnt Pathway: An Emerging Player in Vascular and Traumatic Mediated Brain Injuries. Front. Physiol. 2020, 11, 565667.

45. Qian, Z.M.; Ke, Y. Huperzine A: Is it an Effective Disease-Modifying Drug for Alzheimer's Disease? Front. Aging Neurosci. 2014, 6, 216.

46. Lu, H.; Jiang, M.; Lu, L.; Zheng, G.; Dong, Q. Ultrastructural mitochondria changes in perihematomal brain and neuroprotective effects of Huperzine A after acute intracerebral hemorrhage. Neuropsychiatr. Dis. Treat. 2015, 11, 2649–2657.

47. Báez-Pagán, C.A.; Delgado-Vélez, M.; Lasalde-Dominicci, J.A. Activation of the Macrophage α7 Nicotinic Acetylcholine Receptor and Control of Inflammation. J. Neuroimmune Pharmacol. 2015, 10, 468–476.

48. Donat, C.K.; Scott, G.; Gentleman, S.M.; Sastre, M. Microglial Activation in Traumatic Brain Injury. Front. Aging Neurosci. 2017, 9, 208.