

Natural Photosensitizers in APDT

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Definition

Antimicrobial photodynamic therapy (APDT) is constantly evolving and can minimize this antimicrobial resistance problem. Reactive oxygen species produced when nontoxic photosensitizers are exposed to light are the main functional components of APDT responsible for microbial destruction; therefore, APDT has a broad spectrum of target pathogens, such as bacteria, fungi, and viruses. Various photosensitizers, including natural extracts, compounds, and their synthetic derivatives, are being investigated. Light, oxygen, and PSs in precise cooperation are the key factors determining APDT efficiency and are responsible for ROS production and the inactivation of the targeted cells.

1. Introduction

After penicillin was identified as a product of *Penicillium notatum* by Alexander Fleming in 1928, its widespread consumption was noted in the early 1940s. In 1944, 50% of the clinical isolates of *Staphylococci* sp. were unexpectedly shown to exhibit resistance to penicillin ^[1]. Extensive antibiotic (mis)use has led to the spread of more resistant bacteria to the environment, and antimicrobial resistance (AMR) is an increasing threat to humans. By 2050, 10 million deaths per year are expected to be related to AMR ^[2]. Antibiotic-resistant pathogens are emerging threats to human life and are classified as methicillin-resistant *Staphylococcus aureus* (MRSA), vancomycin-resistant *Enterococcus faecalis*, multidrug-resistant mycobacteria, Gram-negative pathogens, and fungi ^[1].

Antibiotic resistance and antibiotic pollution are common problems worldwide. Antibiotic resistance in infections potentially results in sepsis and even ineluctable systemic inflammation and organ failure ^[3]. Medicine is not the only source of antibiotic resistance. Aquaculture production, livestock, and pets consume and waste a high rate of antibiotics. Some probiotics and plant extracts containing essential oils have been introduced as alternative treatments to overcome excess antibiotic usage in aquaculture ^[4]. Notably, every use of antibiotics can create selective pressure for mutation and the development of drug resistance ^[5]. Overuse or misuse of antibiotics can increase the rate of AMR. Some country-based strategies have been implemented to overcome AMR. Additionally, in 2014, the World Health Organization (WHO) published a global report on AMR ^[6].

Antimicrobial photodynamic therapy (APDT) is a challenging method to overcome excess antibiotic consumption and limit antibiotic resistance gene transfer. Quorum sensing, vaccines, lectin inhibition, and iron chelation have been used as treatments for drug-resistant microorganisms, and APDT might be considered a favorable technique among them. The photodynamic effect was first described by Oscar Raab in 1904 and first successfully used to treat cancer cells in 1905 ^[7]. By the time PDT focused on cancer cell treatment, APDT was focused on overcoming antibiotic resistance by targeting bacteria, algae, yeasts, and viruses ^[8].

In addition to combating AMR and excess antibiotic consumption, APDT can provide strong antibiotic potency. APDT inhibits a broad spectrum of pathogens because its antimicrobial activity originates from reactive oxygen species (ROS) production induced by a unique photochemical reaction. Viera et al. ^[9] studied APDT efficiency against a broad range of organisms, including bacteria, fungi, and viruses, and reported that APDT is effective against a wide range of organisms. Tissue specificity is another advantage of APDT. Generally, no toxicity is observed in nonphotosensitizer-treated cells or in cells that are not exposed to light. Photosensitizers (PSs) are known to be taken up predominantly by target cells rather than nontarget cells. Only the infected tissue is irradiated, and the PS in unirradiated locations is

pharmacodynamically passive. In addition, Park et al. [10] reported that APDT can be used effectively without damaging resident flora or human tissue.

2. Key Factors in Antimicrobial Photodynamic Therapy (APDT)

Light, oxygen, and PSs in precise cooperation are the key factors determining APDT efficiency and are responsible for ROS production and the inactivation of the targeted cells [11].

2.1. Light Sources

Photobiomodulation has been applied to relieve pain, decrease inflammation, and stimulate the healing of living tissues [12]. Light sources significantly affect PDT. Sunlight with diverse wavelengths activates the PS and causes shallow tissue penetration, thermal effects, and difficulties in controlling the dose [13]. Xenon lamps, light emitting diodes (LEDs), laser beams, and fiber optic devices are alternatives to sunlight and can overcome many problems associated with sunlight exposure. A xenon lamp illuminates a wide range, while a laser beam illuminates a narrow area. Near-infrared (NIR) (700–810 nm), red (600–700 nm), yellow (550–600 nm), green (490–550 nm), blue (400–490 nm), and ultraviolet A (UVA) (330–400 nm) light have been applied for APDT [14]. A broad range of light has been used for APDT; however, longer wavelengths are preferred because of the deeper tissue penetration [15].

LEDs have become attractive light sources because they are easy to operate, safe, and inexpensive. Moreover, some innovative methods have been developed as alternatives to classic light sources. For example, wearable light-emitting fabrics facilitate the delivery of total light over a much longer time with a much lower power density, overcoming the problem of depletion of the available oxygen supply by decreasing the oxygen consumption rate [14]. The sufficient intensity of light for antibacterial photosensitization ranges from 5–1000 W/m², as higher light intensities potentially result in thermal problems. The exposure time varies from seconds to minutes, depending on the light intensity [16].

2.2. Oxygen

PSs and light irradiation are the main mechanisms of photodynamic therapy (PDT), and the ROS generated and the singlet oxygen (¹O₂) converted from molecular oxygen by PSs are responsible for bacterial damage. Singlet oxygen damages organelles and causes programmed cell death in human cells [17].

Molecular oxygen (O₂), a nonpolar small molecule that diffuses across biological membranes, is used by aerobic organisms for oxidation and respiration. After O₂ passes through the membranes, oxidative phosphorylation and adenosine triphosphate generation occur, and oxygen is reduced to produce energy. Respiratory flavoenzymes are the main catalytic redox cofactors that participate directly in ROS formation by transferring e⁻ to O₂ to produce superoxide (O₂^{•-}) and H₂O₂. Afterwards, a Fenton reaction, in which H₂O₂ is oxidized with the available ferrous iron (Fe²⁺) to generate OH[•], occurs [18]. Sodium azide (NaN₃) and histidine are ¹O₂ quenchers, and thiourea and dimethyl sulfoxide are free radical scavengers that have been used in photodynamic inactivation therapies [19].

Singlet oxygen (¹O₂) is an extremely short-lived and reactive form of oxygen that is involved in photochemical reduction processes. In type I reactions, electrons are stripped from biological macromolecules and [•]OH is transformed into hydroxide ions [1]. Superoxide dismutase (SOD) converts O₂^{•-} into HO[•] and O₂, and H₂O₂ participates in the Fenton reaction, resulting in the homolytic fission of the oxygen–oxygen bond in H₂O₂ to yield a hydroxide ion [1]. In type II reactions, ¹O₂ interacts with double bonds, sulfur moieties, and aromatic components of macromolecules in Diels–Alder cycloadditions [1]. Type I and type II reactions involving natural PSs are illustrated in [Figure 1](#), and each step is explained below.

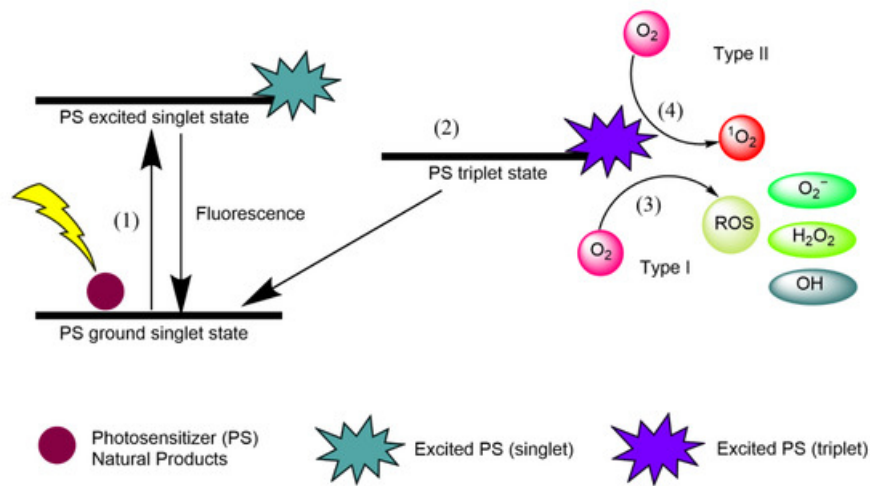


Figure 1. Type I and type II reactions in APDT. (1) Excitation from ground singlet state; (2) intersystem crossing into the triplet excited state; (3) Type I reaction; (4) Type II reaction.

- (1) Natural product PSs are converted from the ground singlet state into the excited singlet state when exposed to a specific wavelength of light.
- (2) If the PS in the excited singlet state does not return to the initial ground state, it can be subjected to intersystem crossing into the triplet excited state.
- (3) A type I reaction comprises transferring a hydrogen atom from PS to an organic molecule to form radicals, and the reduced PS interacts with oxygen through a redox reaction, forming ROS and a superoxide anion radical ($O_2^{\bullet-}$) [20].
- (4) The type II reaction comprises direct energy transfer from the activated PS to molecular oxygen to form singlet oxygen (1O_2) and is simpler than the type I reaction.

2.3. Photosensitizers (PSs)

PSs play an important role in photodynamic reactions as absorbers of light energy [13]. PSs are divided into three subgroups, namely, first-, second-, and third-generation PSs. Water-soluble porphyrins called “hematoporphyrins” are characterized as first-generation PSs, and methylene blue, toluidine blue, photosense[®], Foscan[®], and 5'-aminolevulinic acid (ALA) are examples of second-generation PSs. They possess a higher singlet oxygen quantum yield, chemical purity, and selectivity than first-generation PSs [7]. Third-generation PSs have been investigated recently with the main aims of reducing damage to healthy cells and increasing bioavailability. These systems generally consist of drug delivery systems, gene engineering-based technologies, or monoclonal antibody receptor combinations.

An ideal PS should:

- Have a strong absorption peak in the red to near-infrared spectral region (between 650 and 800 nm) [15];
- Possess a substantial triplet quantum yield leading to good ROS production upon irradiation [21];
- Have high tissue selectivity [22];
- Exhibit no dark toxicity [23];

- Have ideal solubility to maintain lipophilic ability to cross the phospholipid membrane and prevent self-aggregation [24];
- Exhibit high stability under storage conditions [25];
- Kill microorganisms sufficiently without damaging eukaryotic host cells [26];
- Display optimal absorption, distribution, metabolism, and excretion (ADME) [24];
- Have a small size to enable microbial membrane permeation [2]; and
- Have low manufacturing costs [23].

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Keywords

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