

Arabidopsis MLO4 Interacts with CaM/CML

Subjects: **Plant Sciences**

Contributor: Lei Zhu

Here, we report the functional characterization of the MLO4 protein in *Arabidopsis* roots. The MLO4 was identified as interacting with CML12 in a screening for the interaction between the proteins from *Arabidopsis* MLO and calmodulin/calmodulin-like (CaM/CML) families using yeast two hybrid (Y2H) assays. Then, the interaction between MLO4 and CML12 was further verified by Luciferase Complementation Imaging (LCI) and Bimolecular Fluorescence Complementation (BiFC) assays. Genetic analysis showed that the *mlo4*, *cml12*, and *mlo4 cml12* mutants displayed similar defects in root gravity response. These results imply that the MLO4 might play an important role in root gravity response through interaction with CML12. Moreover, our results also demonstrated that the interaction between the MLO and CaM/CML families might be conservative.

calmodulin

calmodulin-like proteins

CML12

MLO protein

1. Introduction

Mildew resistance locus O (MLO) was first identified as being resistant against powdery mildew (PM) infection in barley [1]. Then, the MLO proteins were found in many plant species. *Arabidopsis* has at least 15 MLO proteins that diverged into several clades and were involved in different physiological processes, such as sexual reproduction, PM infection, and root thigmomorphogenesis [2][3][4][5]. MLO7/NORTIA (NTA), a downstream component of the receptor-like kinase FERONIA (FER), is involved in pollen tube reception and PM infection [6]. Study showed that its function is dependent on NTA homooligomerization and its carboxyl-terminal tail identity [7]. MLO5/9/15 played roles in pollen tube responses to ovular signals, and MLO5/9 selectively recruit Ca^{2+} channel CNGC18-containing vesicles to the plasma membrane [8]. The MLO1, MLO2, MLO3, MLO6, MLO8, MLO9, MLO10, and MLO13 are expressed in discrete domains during reproductive development [4]. The MLO2, MLO4, and MLO6 modulate defense responses against PM fungi and a number of other phytopathogens [9]. Furthermore, the MLO2 also function as a negative regulator in plant ROS responses involving biotic and abiotic stress [10]. Mutations in *MLO4* and *MLO11* exhibit abnormal root thigmomorphogenesis and gravity sensitivity [3][11]. Genetical complementation with MLO4 domains indicate that the C-terminal cytoplasmic domain of MLO4 is necessary for regulation of asymmetrical root growth [11].

All MLO proteins are predicted to share a conserved calmodulin-binding domain (CaMBD) in C-terminal cytoplasmic tail [12][13]. The gel overlay assays demonstrated in vitro Ca^{2+} -dependent binding of CaM to the CaMBD of the MLOs from barley [14]. However, little is known about the interaction of individual MLO with the specific calmodulins (CaMs) or calmodulin-like proteins (CML) in *Arabidopsis* or other plants.

The *Arabidopsis* genome encodes 7 CaMs and 50 CMLs, which are presumed to sense and transduce Ca^{2+} signals [15]. Functional studies of CaM/CML in plants revealed that this protein family converts calcium signals into transcriptional responses, protein phosphorylation, or metabolic changes to regulate plant development responding to the ever-changing environment [16]. Some CaM/CML proteins are involved in plant growth and development, such as AtCaM7 regulating light-induced seedling development [17][18], AtCaM2 functioning in pollen germination [19], AtCML42 for trichome morphogenesis [20], and AtCML23 and AtCML24 in flowering [21][22]. Some CaM/CML proteins are indispensable for plants to respond to abiotic and biotic stress. The mutations in AtCaM3 lead to reduction in thermotolerance of the mutant plants [23]. AtCaM4 negatively regulates freezing tolerance by interacting with CaM-binding protein PATL1 in a CBF-independent manner [24]. A knockout mutant of AtCML9 enhances its tolerance to drought and salinity stress [25]. AtCML20 is a negative regulator in guard cell ABA signaling during drought tolerance [26]. AtCML8 and AtCML9 positively regulate plant immunity in response to *Pst* inoculation [27]. Less NtCaM13 expression causes more susceptibility to virulent bacteria and fungi in tobacco [28]. Overexpression of pepper CaM1 and AtCML43 confers enhanced resistance to pathogens [29]. All these data suggest that the CaM/CML-mediated defense signaling pathways involve a complex regulatory network.

Both MLO and CaM/CML proteins are important for plants. They have potential to interact with each other in between the two families through CaMBD [12]. However, for now, it is just predicted by physical structure of the proteins, and not solid evidence, to prove their interaction or even their related biological function.

2. The Expression Patterns of MLO and CaM/CML Genes

The co-expression patterns of *MLO* and *CaM/CML* genes were analyzed using the e-FP data from TAIR website (www.arabidopsis.org, accessed on 19 May 2021) (Figure 1). The results showed that the *MLO* and *CaM/CML* genes are expressed in various *Arabidopsis* tissues. The *MLO1* was expressed more highly in seeds than in other tissues. The *CaM/CML* genes, which exhibit similar expression in seeds, included 12 members, namely, CaM4, CML14, CML20, CML24, CML27, CML32, CML33, CML34, CML43, CML48, CML49, and CML50. The *MLO5*, *MLO9*, and *MLO14* genes were highly expressed in pollen. In the pollen, 17 *CaM/CML* genes, including CaM2, CaM3, CML2, CML3, CML6, CML7, CML13, CML15, CML16, CML21, CML26, CML28, CML29, CML31, CML39, CML42, and CML49, were found to be highly co-expressed. In flowers, four *MLO* genes (*MLO4*, *MLO6*, *MLO8*, and *MLO11*) and nine *CML* genes (CML5, CML24, CML27, CML36, CML40, CML41, CML44, CML46, and CML47) were highly co-expressed. The four *MLO* genes, *MLO2*, *MLO3*, and *MLO8*, were found to be highly expressed in leaves; similarly, five *CML* genes were highly expressed in leaves, namely, CML9, CML10, CML18, CML35, and CML46. In roots, *MLO12* gene was highly co-expressed, together with five *CaM/CML* genes (CaM1, CaM5, CaM7, CML8, and CML19). *MLO4* and *CML12* were expressed in many tissues, including roots. Although the highest expression levels of both *MLO4* and *CML12* did not present in roots, they were highly co-expressed in roots, implying both of them may function in roots.

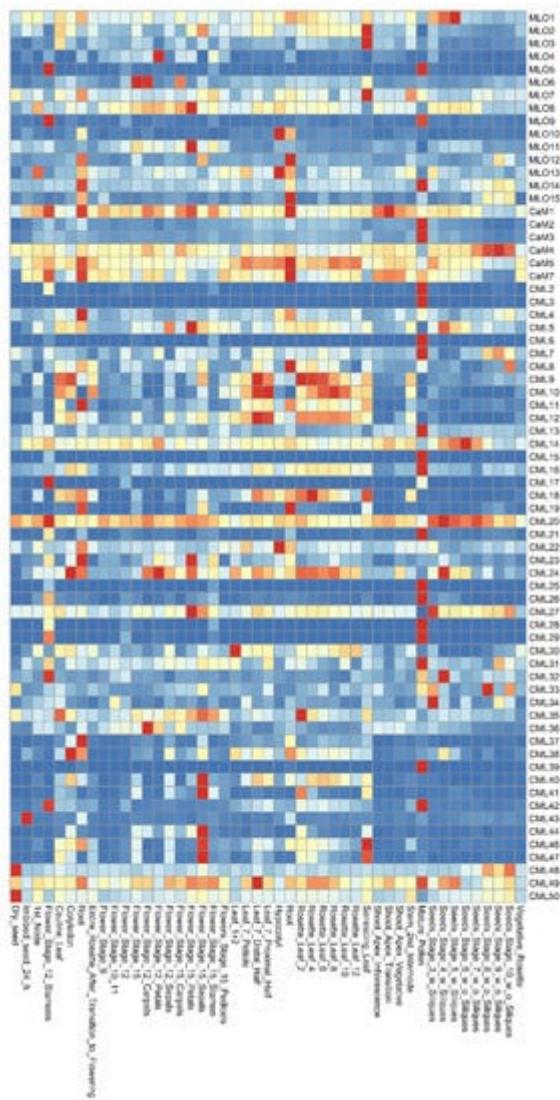


Figure 1. Expression pattern of *Arabidopsis* MLO and CaM/CML family genes.

3. Screening for Interactive Pairs of MLO and CaM/CML Proteins

The MLO proteins contain a CaMBD in the C-terminus region [12]. Therefore, the C-terminus regions of the 15 MLO proteins were used to characterize the interaction of MLO and CaM/CML proteins by Y2H. The results showed that each MLO protein may interact with at least one CML protein in yeast cells (Table 1). In particular, the NTA/MLO7, which was involved in pollen tube reception, could interact with up to 11 candidate CML proteins, including CML8-12, CML26, CML30, CML37, CML38, CML40, and CML40. Similar results were also obtained from the assays for the three closely-related proteins MLO4, MLO11, and MLO14, which function in root thigmomorphogenesis [11]. MLO4 interacted with CML12, CML40, and CML44. MLO11 interacted with CML47. MLO14 interacted with CML17, CML18, and CML49.

Table 1. Interactive protein pairs between MLO and CaM/CML family.

MLOs	CaM/CML Proteins that Interacted with Corresponding MLO Proteins
MLO1C	CaM2, CML29
MLO2C	CML9, CML18
MLO3C	CML18, CML20, CML23, CML26, CML32, CML40, CML44
MLO4C	CML12, CML40, CML44
MLO5C	CML18, CML26, CML40, CML44
MLO6C	CML8, CML10, CML20, CML26, CML73, CML40, CML44, CML49
MLO7C	CML8, CML9, CML10, CML11, CML12, CML26, CML30, CML37, CML38, CML40, CML44
MLO8C	CML8, CML9, CML23, CML26, CML37, CML40, CML44
MLO9C	CML10, CML42, CML44, CML49
MLO10C	CML8, CML9, CML10, CML26, CML37, CML44
MLO11C	CML47
MLO12C	CML10, CML12, CML30, CML35, CML36, CML37, CML44
MLO13C	CML32, CML40, CML41
MLO14C	CML17, CML18, CML49
MLO15C	CML10, CML44

4. LCI and BiFC Assays Further Demonstrated That MLO4 Interacted with CML12

At first, MLO4 and CML12 were chosen to verify the results of Y2H screening. Then, the LCI results showed that CML12 interacted with the full length of MLO4 in tobacco cells (Figure 2a). BiFC results demonstrated that the CML12 interacted with the cytoplasmic domain of MLO4, in concert to Y2H result. Furthermore, when the full length of MLO4 was used for the assays, the interactive signal was detected only on the plasma membrane, which was co-localized with the staining signals of the membrane dye FM 4-64 (Figure 2b). This result was consistent to the ones obtained from the membrane localization of MLO4 with a signal peptide and seven transmembrane domains [11].

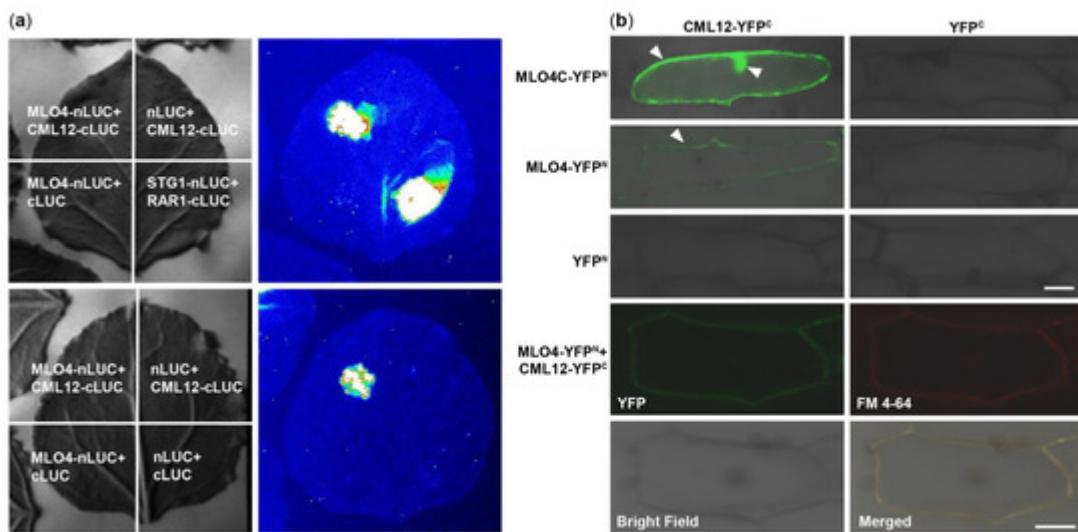


Figure 2. CML12 interacted with MLO4. **(a)** LCI results showed CML12 interacted with full length of MLO4 in tobacco cells. **(b)** BiFC results showed that CML12 interacted with cytoplasmic domain of MLO4 in onion cells, and the signal was detected in nucleus and periphery. When CML12 interacted with full length of MLO4, the signal was only detected on the plasma membrane, colocalized with FM 4-64. The YFP signals are indicated by pointed triangles in each figure. Bars = 50 μ m.

5. MLO4 and CML12 Displayed a Similar Expression Pattern

To understand the relationship between *MLO4* and *CML12*, we further investigated whether *MLO4* and *CML12* have a co-expression pattern *in vivo*. First, GUS staining of pCML12::CML12-GUS transgenic lines showed that *CML12* was expressed in most of the vegetable tissues, such as seedlings, roots, root hairs, leaves, trichomes, stems, siliques, calyces, and chalazal end of ovules but not in pollen (Figure 3a–j). In addition, pCML12::CML12-GFP transgenic lines showed that the CML12-GFP fusion protein was detected in the root cap (Figure 3k). Real-time PCR results showed that *MLO4* and *CML12* shared a similar expression pattern. Both genes were expressed constitutively in many tissues, including seedlings, roots, stems, leaves, inflorescence, and siliques. Expression levels of both genes were closely similar in seedlings. *MLO4* was expressed with twice the *CML12* expression in roots (Figure 3l). A previous study revealed that the pMLO4::MLO4-GFP was also expressed in the roots, and MLO4-GFP fusion protein was detected in the root epidermal cell [11]. Therefore, both CML12 and MLO4 proteins were expressed in roots, suggesting that they may function coordinately.

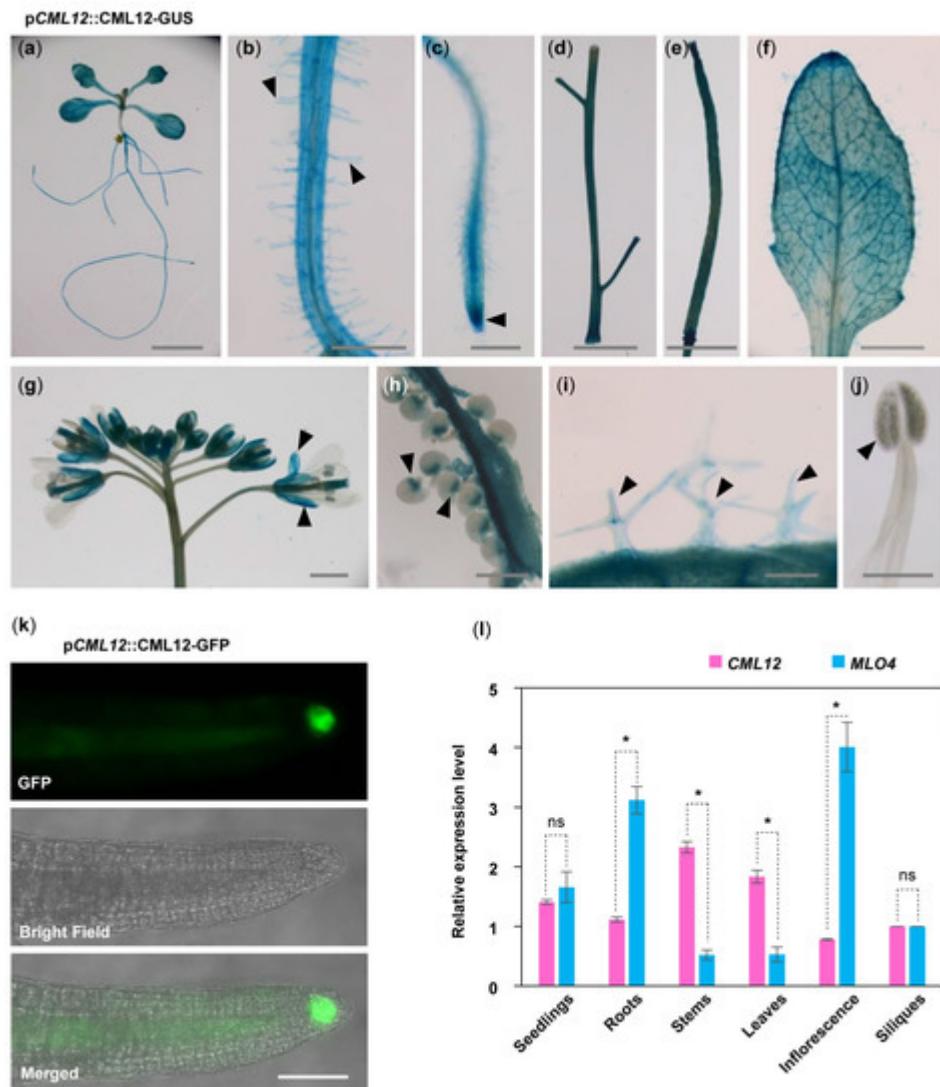


Figure 3. Expression pattern of *CML12* Gene. GUS staining of *pCML12::CML12-GUS* transgenic lines showed *CML12* was expressed in most of the vegetable tissues: Seedling (a), root hairs (b), root tip (c), stems (d), siliques (e), leaf (f), sepals (g), chalazal end of ovules (h), and trichomes (i), not in pollen (j), as indicated by pointed triangles in each figure. Bars = 100 μ m. (k) *pCML12::CML12-GFP* transgenic lines showed *CML12-GFP* was detected in root cap. Bar = 20 μ m. (l) Real-time PCR showed that *CML12* and *MLO4* genes both were expressed in various tissues of *Arabidopsis*. ns: not significant; the * above bars indicate the *t*-test results at significant level of 0.05.

6. The *mlo4* and *cml12* Mutants Shared Similar Phenotypes

The *MLO4* has been characterized as regulating root thigmomorphogenesis, including gravitropism in roots [11]. Previous characterization of *cml12* revealed that the mutations in *CML12* exhibited a similar phenotype [30]. To study the genetic relationship between *MLO4* and *CML12*, we further characterized their roles in root growth.

The *cml12-3* mutant (SALK_122731) was obtained from Dr. Sherryl R. Bisgrove (Department of Biological Sciences, Simon Fraser University, Burnaby, BC, Canada). Real-time PCR showed that *cml12-3* was a knock-out

mutant, as reported by Gleeson et al. [31]. To test its gravity response, *cml12-3* seedlings were vertically grown on the agar minimal medium (0.25% sucrose) for 7 d (16 h light/8 h dark cycle); then, the plates were rotated by 90° in the clockwise direction. The roots bent down in response to gravity change until root tips became parallel with new gravity vector (Figure 4a). Compared to the time (12.60 ± 0.66 h) for the wild-type roots to form a bend, the *cml12-3* roots took less time (9.33 ± 0.75 h) to form a bend (Figure 5a), suggesting that *cml12-3* roots were more sensitive to gravity response.

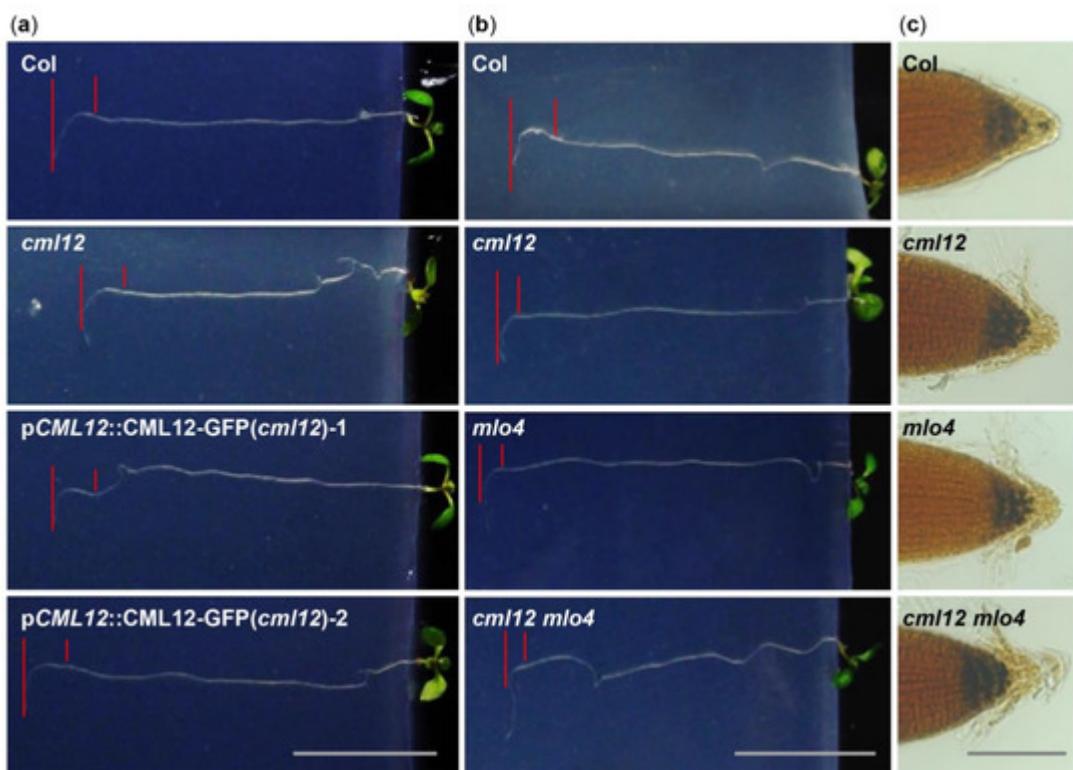


Figure 4. Phenotype analysis of *cml12-3* and *mlo4-5* mutants. *cml12-3* and *mlo4-5* mutants were indicated, respectively, as *cml12* and *mlo4*, for short. (a) 7-day-old seedlings growing through the agar medium were rotated by 90° in the clockwise direction. Roots bent down to respond to gravity change. Red lines point to tip position at the time of rotation and the location where root growth became parallel with new gravity vector. Bar = 1 cm. (b) Roots bent down to respond to gravity change. Red lines point to tip position at the time of rotation and the location where root growth became parallel with new gravity vector. Bar = 1 cm. (c) Starch was detected using KI-I₂ stain. Starch in single mutants *cml12-3* and *mlo4-5* and double mutant *cml12-3 mlo4-5* was obviously more than that in wild-type root tips. Bar = 100 μ m.

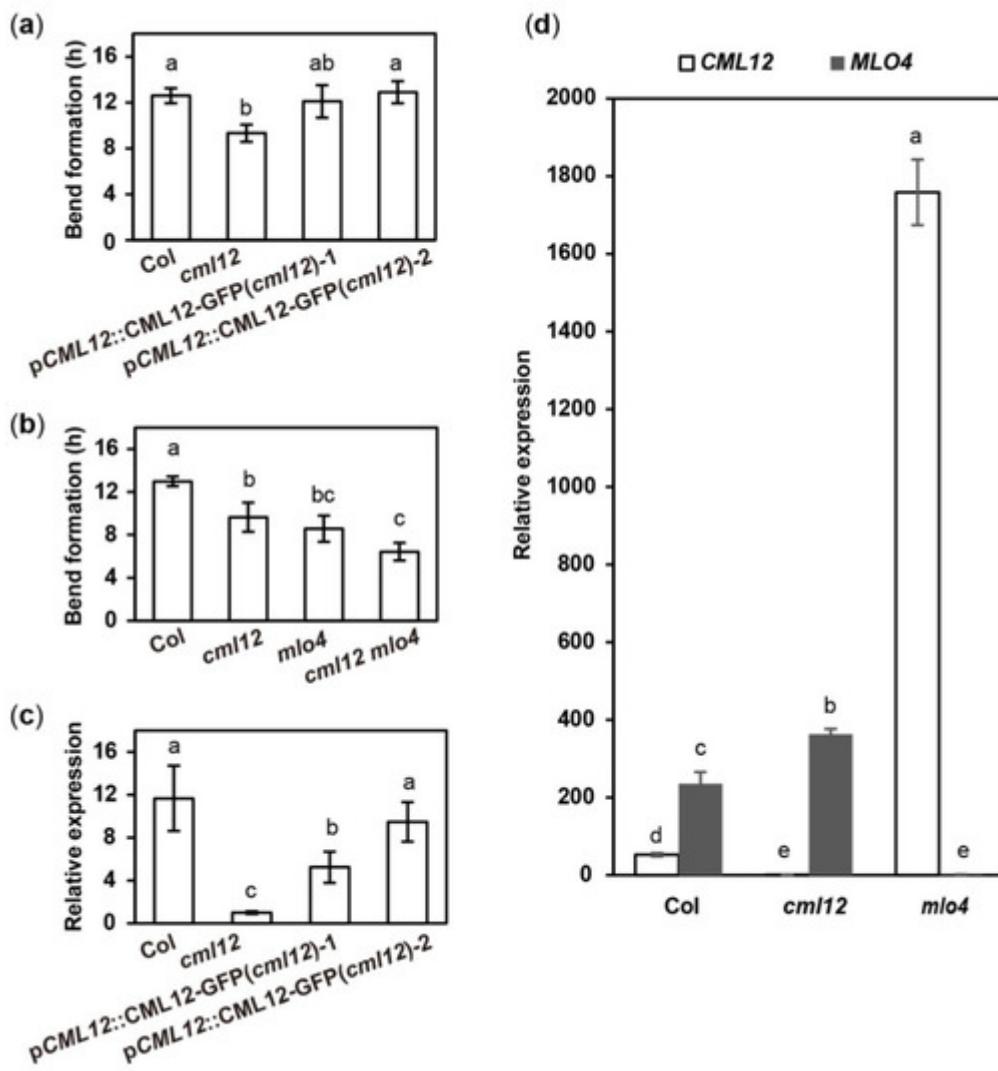


Figure 5. Quantitative analysis of bend formation and gene expression in *cml12-3* and *mlo4-5* mutants. *cml12-3* and *mlo4-5* mutants were indicated, respectively, as *cml12* and *mlo4* for short. (a) The time taken to form a bend in Col, *cml12-3*, pCML12::CML12-GFP (*cml12-3*-1), and pCML12::CML12-GFP (*cml12-3*-2) were 12.60 ± 0.66 , 9.33 ± 0.75 , 12.10 ± 1.42 , and 12.90 ± 0.96 hours, respectively, $80 < n < 150$. (b) The time taken to form a bend in Col, *cml12-3*, *mlo4-5*, and *cml12-3 mlo4-5* were 12.97 ± 0.45 , 9.63 ± 1.36 , 8.57 ± 1.22 , and 6.43 ± 0.83 h, respectively, $80 < n < 150$. (c) Real-time PCR showed that expression of *CML12* gene was restored in pCML12::CML12-GFP (*cml12-3*-) plants. (d) Real-time PCR showed that relative expression of *CML12* increased significantly in *mlo4-5* mutant. Different letters above the bars indicate significant differences ($P < 0.05$, Bonferroni correction) in a pairwise comparison using the least significant difference (LSD) method.

To verify the phenotype of *cml12-3* in gravity response, a complementation assay was performed using the construct pCML12::CML12-GFP in PCAMBIA 1300 vector containing the native promoter (1500 bp upstream ATG) and coding sequence of *CML12* (975 bp). Two pCML12::CML12-GFP transgenic *cml12-3* mutant lines were chosen to further characterize their root phenotype. The average times for the roots from each pCML12::CML12-GFP (*cml12-3*-) transgenic lines to form a bend were 12.10 ± 1.42 h and 12.90 ± 0.96 h, respectively, similar to that of wild-type roots (12.60 ± 0.66 h) (Figure 4a). The real-time PCR results showed that *CML12* expression was

higher in transgenic roots than that in *cml12-3* mutant roots (Figure 5a). Therefore, *CML12* was able to restore the defect of *cml12-3* mutants in gravity response, further demonstrating that *CML12* is involved in gravity response.

The previous study showed that *mlo4-1*, *mlo4-3*, and *mlo4-4* mutants exhibited different growth patterns on the reclined medium than wild-type [11]. In this study, we identified another Salk allele *mlo4-5* in which the T-DNA was inserted in the last exon of *MLO4*, different from the previously-reported alleles. Real-time PCR showed that *mlo4-5* was a knock-out mutant and expressed truncated protein. At first, an allelic analysis was performed. The F1 progeny of *mlo4-5* from the crosses with *mlo4-4* exhibited a resembled phenotype in growth pattern. The loops in each root of *mlo4-4/+*; *mlo4-5/+* plant were denser with shorter wavelengths, and its phenotype was similar to that of single mutant-*mlo4-4/-* and *mlo4-5/-*. The result demonstrated that *mlo4-5* also was an effective mutant of *MLO4* gene. Therefore, the *mlo4-5* was used for further characterization.

The phenotypic comparison in root gravity response was performed with the single mutant *cml12-3* (*cml12-3/-*) and *mlo4-5* (*mlo4-5/-*), as well as the double mutant *cml12-3 mlo4-5* (*cml12-3/-; mlo4-5/-*). The time for bend formation was 12.97 ± 0.45 h in wild-type roots. In contrast, the roots of the single mutant *cml12-3*, *mlo4-5* took 9.63 ± 0.45 h and 8.57 ± 1.22 h to form bends, respectively. The roots of the double mutant *cml12-3 mlo4-5* took relatively shorter time (6.43 ± 0.83 h) to form a bend (Figure 4b, Figure 5b), indicating that the phenotype of double mutant was a little stronger than those of the single mutants.

Starch grains in root tips are sensors of gravity response [30]. Therefore, the starch stain assays were further performed to compare the starch contents in the mutants and wild-type root tips using KI-I₂ staining. Contents of the starch grains in the single mutant *cml12-3* and *mlo4-5*, as well as the double mutant *cml12-3 mlo4-5*, were obviously higher than that in wild-type root tips (Figure 4c).

In addition, *cml12-3*, *mlo4-5*, and *cml12-3 mlo4-5* mutant roots shared similar growth patterns with denser loops and shorter wavelengths on the reclined agar medium. The phenotype of the double mutant *cml12-3 mlo4-5* was not obviously aggravated.

7. Mutation in MLO4 Affected the Expression of CML12

CaM/CMLs bound calcium and were supposed to regulate target proteins downstream [16]. To further study the possible regulatory relationship between *CML12* and *MLO4*, their relative expression levels in each other mutants were analyzed. The real-time PCR assay showed the expression of *CML12* was significantly elevated in *mlo4-5* mutant, far more than *MLO4* expression changing in *cml12-3* mutant (Figure 5d). Thus, *MLO4* likely modulates the expression of *CML12*.

References

1. Miklis, M.; Consonni, C.; Bhat, R.A.; Lipka, V.; Schulze-Lefert, P.; Panstruga, R. Barley MLO modulates actin-dependent and actin-independent antifungal defense pathways at the cell periphery. *Plant Physiol.* 2007, 144, 1132–1143.
2. Acevedo-Garcia, J.; Kusch, S.; Panstruga, R. Magical mystery tour: MLO proteins in plant immunity and beyond. *New Phytol.* 2014, 204, 273–281.
3. Bidzinski, P.; Noir, S.; Shahi, S.; Reinstadler, A.; Gratkowska, D.M.; Panstruga, R. Physiological characterization and genetic modifiers of aberrant root thigmomorphogenesis in mutants of *Arabidopsis thaliana* MILDEW LOCUS O genes. *Plant Cell Environ.* 2014, 37, 2738–2753.
4. Davis, T.C.; Jones, D.S.; Dino, A.J.; Cejda, N.I.; Yuan, J.; Willoughby, A.C.; Kessler, S.A. *Arabidopsis thaliana* MLO genes are expressed in discrete domains during reproductive development. *Plant Reprod.* 2017, 30, 185–195.
5. Kusch, S.; Pesch, L.; Panstruga, R. Comprehensive phylogenetic analysis sheds light on the diversity and origin of the MLO family of integral membrane proteins. *Genome Biol. Evol.* 2016, 8, 878–895.
6. Kessler, S.A.; Shimosato-Asano, H.; Keinath, N.F.; Wuest, S.E.; Ingram, G.; Panstruga, R.; Grossniklaus, U. Conserved molecular components for pollen tube reception and fungal invasion. *Science* 2010, 330, 968–971.
7. Jones, D.S.; Yuan, J.; Smith, B.E.; Willoughby, A.C.; Kumimoto, E.L.; Kessler, S.A. Mildew Resistance Locus O function in pollen tube reception is linked to its oligomerization and subcellular distribution. *Plant Physiol.* 2017, 175, 172–185.
8. Meng, J.S.; Liang, L.; Jia, P.F.; Wang, Y.C.; Li, H.C.; Yang, W.C. Integration of ovular signals and exocytosis of a Ca²⁺ channel by MLOs in pollen tube guidance. *Nat. Plant* 2020, 6, 143–153.
9. Consonni, C.; Humphry, M.E.; Hartmann, H.A.; Livaja, M.; Durner, J.; Westphal, L.; Vogel, J.; Lipka, V.; Kemmerling, B.; Schulze-Lefert, P.; et al. Conserved requirement for a plant host cell protein in powdery mildew pathogenesis. *Nat. Genet.* 2006, 38, 716–720.
10. Cui, F.; Wu, H.; Safronov, O.; Zhang, P.; Kumar, R.; Kollist, H.; Salojarvi, J.; Panstruga, R.; Overmyer, K. *Arabidopsis* MLO2 is a negative regulator of sensitivity to extracellular reactive oxygen species. *Plant Cell Environ.* 2018, 41, 782–796.
11. Chen, Z.; Noir, S.; Kwaaitaal, M.; Hartmann, H.A.; Wu, M.J.; Mudgil, Y.; Sukumar, P.; Muday, G.; Panstruga, R.; Jones, A.M. Two seven-transmembrane domain MILDEW RESISTANCE LOCUS O proteins cofunction in *Arabidopsis* root thigmomorphogenesis. *Plant Cell* 2009, 21, 1972–1991.
12. Panstruga, R.; Schulze-Lefert, P. Corruption of host seven-transmembrane proteins by pathogenic microbes: A common theme in animals and plants? *Microbes Infect.* 2003, 5, 429–437.

13. La Verde, V.; Dominici, P.; Astegno, A. Towards understanding plant calcium signaling through calmodulin-like proteins: A biochemical and structural perspective. *Int. J. Mol. Sci.* **2018**, *19*, 1331.
14. Kim, M.C.; Panstruga, R.; Elliott, C.; Muller, J.; Devoto, A.; Yoon, H.W.; Park, H.C.; Cho, M.J.; Schulze-Lefert, P. Calmodulin interacts with MLO protein to regulate defence against mildew in barley. *Nature* **2002**, *416*, 447–451.
15. McCormack, E.; Tsa, Y.C.; Braam, J. Handling calcium signaling: Arabidopsis CaMs and CMLs. *Trends Plant Sci.* **2005**, *10*, 383–389.
16. Perochon, A.; Aldon, D.; Galaud, J.P.; Ranty, B. Calmodulin and calmodulin-like proteins in plant calcium signaling. *Biochimie* **2011**, *93*, 2048–2053.
17. Kushwaha, R.; Singh, A.; Chattopadhyay, S. Calmodulin7 plays an important role as transcriptional regulator in *Arabidopsis* seedling development. *Plant Cell* **2008**, *20*, 1747–1759.
18. Abbas, N.; Maurya, J.P.; Senapati, D.; Gangappa, S.N.; Chattopadhyay, S. Arabidopsis CAM7 and HY5 physically interact and directly bind to the HY5 promoter to regulate its expression and thereby promote photomorphogenesis. *Plant Cell* **2014**, *26*, 1036–1052.
19. Landoni, M.; De Francesco, A.; Galbiati, M.; Tonelli, C. A loss-of-function mutation in Calmodulin2 gene affects pollen germination in *Arabidopsis thaliana*. *Plant Mol. Biol.* **2010**, *74*, 235–247.
20. Dobney, S.; Chiasson, D.; Lam, P.; Smith, S.P.; Snedden, W.A. The calmodulin-related calcium sensor CML42 plays a role in trichome branching. *J. Biol. Chem.* **2009**, *284*, 31647–31657.
21. Delk, N.A.; Johnson, K.A.; Chowdhury, N.I.; Braam, J. CML24, regulated in expression by diverse stimuli, encodes a potential Ca²⁺ sensor that functions in responses to abscisic acid, daylength, and ion stress. *Plant Physiol.* **2005**, *139*, 240–253.
22. Tsai, Y.C.; Delk, N.A.; Chowdhury, N.I.; Braam, J. Arabidopsis potential calcium sensors regulate nitric oxide levels and the transition to flowering. *Plant Signal. Behav.* **2007**, *2*, 446–454.
23. Zhang, W.; Zhou, R.G.; Gao, Y.J.; Zheng, S.Z.; Xu, P.; Zhang, S.Q.; Sun, D.Y. Molecular and genetic evidence for the key role of AtCaM3 in heat-shock signal transduction in *Arabidopsis*. *Plant Physiol.* **2009**, *149*, 1773–1784.
24. Chu, M.; Li, J.; Zhang, J.; Shen, S.; Li, C.; Gao, Y.; Zhang, S. AtCaM4 interacts with a Sec14-like protein, PATL1, to regulate freezing tolerance in *Arabidopsis* in a CBF-independent manner. *J. Exp. Bot.* **2018**, *69*, 5241–5253.
25. Magnan, F.; Ranty, B.; Charpenteau, M.; Sotta, B.; Galaud, J.P.; Aldon, D. Mutations in AtCML9, a calmodulin-like protein from *Arabidopsis thaliana*, alter plant responses to abiotic stress and abscisic acid. *Plant J. Cell Mol. Biol.* **2008**, *56*, 575–589.
26. Wu, X.; Qiao, Z.; Liu, H.; Acharya, B.R.; Li, C.; Zhang, W. CML20, an *Arabidopsis* calmodulin-like protein, negatively regulates guard cell ABA signaling and drought stress tolerance. *Front. Plant*

Sci. 2017, 8, 824.

27. Zhu, X.; Perez, M.; Aldon, D.; Galaud, J.P. Respective contribution of CML8 and CML9, two *Arabidopsis* calmodulin-like proteins, to plant stress responses. *Plant Signal. Behav.* 2018, 12, e1322246.

28. Takabatake, R.; Karita, E.; Seo, S.; Mitsuhashi, I.; Kuchitsu, K.; Ohashi, Y. Pathogen-induced calmodulin isoforms in basal resistance against bacterial and fungal pathogens in tobacco. *Plant Cell Physiol.* 2007, 48, 414–423.

29. Chiasson, D.; Ekengren, S.K.; Martin, G.B.; Dobney, S.L.; Snedden, W.A. Calmodulin-like proteins from *Arabidopsis* and tomato are involved in host defense against *Pseudomonas syringae* pv. *tomato*. *Plant Mol. Biol.* 2005, 58, 887–897.

30. Vitha, S.; Yang, M.; Sack, F.D.; Kiss, J.Z. Gravitropism in the starch excess mutant of *Arabidopsis thaliana*. *Am. J. Bot.* 2007, 94, 590–598.

31. Gleeson, L.; Squires, S.; Bisgrove, S.R. The microtubule associated protein END BINDING 1 represses root responses to mechanical cues. *Plant Sci.* 2012, 187, 1–9.

Retrieved from <https://encyclopedia.pub/entry/history/show/26423>