

# Ion Channels

Subjects: Virology

Contributor: Jamel Mankouri

This reviews describes the requirement of ion channel function for viral infection. Also discussed is how viral infections can result in acquired channelopathies by altering ion channels. Ion channel modulators may be useful in treating both of these aspects.

Keywords: Ion channel, virus, channelopathy ; virus-host interactions

---

## 1. Introduction

The human “channelome” contains over 300 known channels <sup>[1]</sup> that selectively and rapidly transport ions across biological membranes in response to specific stimuli. Ion channels are present on the plasma membranes and organelles of all cells, where they regulate organelle ion homeostasis, mitochondrial function, inflammasome activation, action potential firing, membrane potential, cell volume, and autophagy <sup>[2][3][4][5]</sup>. Given their importance, it follows that their dysfunctions leads to human diseases, termed channelopathies <sup>[6]</sup>. These include disease states of the nervous <sup>[2]</sup>, musculoskeletal <sup>[5]</sup>, cardiovascular <sup>[7]</sup>, and immune systems <sup>[8]</sup>. This has motivated research on compounds that can modulate ion channel activity; ~19% of current FDA-approved drugs are ion channel modulators, second only to drugs targeting G-protein coupled receptors <sup>[9][10]</sup>.

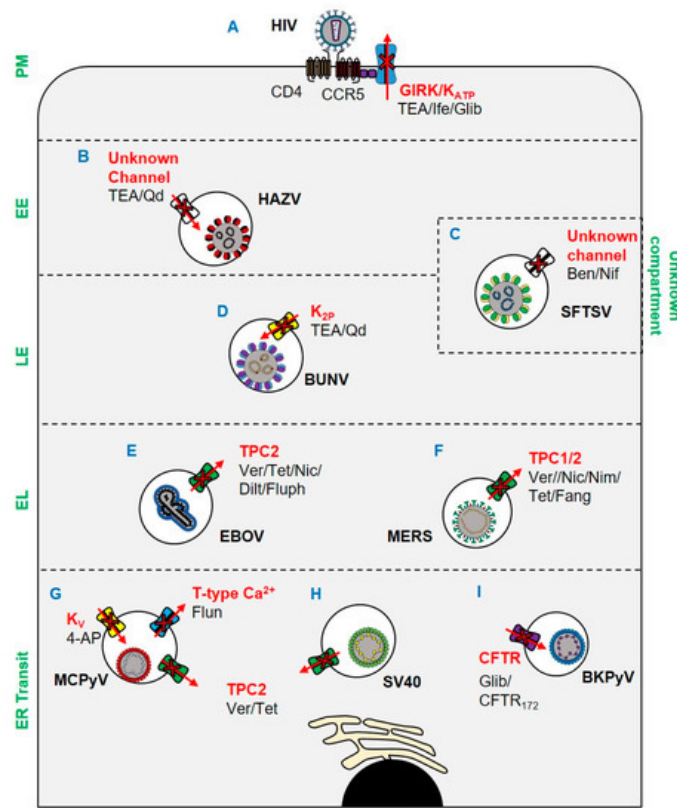
Many viruses encode their own ion channels <sup>[11][12][13]</sup> termed “viroporins,” highlighting the importance of ionic balance during viral infection. This field has spurred intense research and several drugs that target viroporins have emerged (reviewed in <sup>[11]</sup>). More recent evidence highlights how viruses can regulate and/or depend on the ion channels expressed by host cells, highlighting them as new host targets for therapeutic intervention (reviewed by Hover et al., 2017) <sup>[14]</sup>.

## 2. Ion Channels Involved in Viral Entry

### 2.1. Ca<sup>2+</sup> Channels and Viral Entry

The involvement of Ca<sup>2+</sup> channels during viral entry is now well-documented <sup>[15]</sup>. Fujioka et al. showed that influenza virus (IAV) hemagglutinin (HA) triggers intracellular [Ca<sup>2+</sup>] oscillations that are required for viral infection <sup>[16]</sup>. The initial modulation of Ca<sup>2+</sup> by IAV was demonstrated using Förster resonance energy transfer (FRET)-based imaging of the Ca<sup>2+</sup> sensor Yellow Cameleon (YC3.60). These oscillations in Ca<sup>2+</sup> were mediated by a specific voltage-gated Ca<sup>2+</sup> channel (Ca<sub>v</sub>1.2) identified through siRNA silencing approaches. Assays subsequently revealed that IAV directly binds to Ca<sub>v</sub>1.2 via the interaction of HA and a sialylated site on Ca<sub>v</sub>1.2. Accordingly, IAV entry could be inhibited by diltiazem, a clinically available Ca<sup>2+</sup> blocker, highlighting the potential of these compounds for drug repurposing.

Ebola virus (EBOV) also requires Ca<sup>2+</sup> channels for its entry into host cells. EBOV enters cells through endolysosomes positive for both Niemann–Pick C1 (NPC1) and two-pore Ca<sup>2+</sup> channel 2 (TPC2) <sup>[17]</sup>. To further characterise this pathway, Penny et al. expanded the pharmacology of TPCs using a virtual screen of ~1500 FDA-approved drugs. All identified TPC modulators were cross-referenced with two recent anti-EBOV screens, with four dopamine receptor antagonists and five oestrogen receptor modulators identified. As such, it was reasoned that these drugs exert their inhibitory effects on EBOV through the blockade of TPCs (Figure 1E), subsequently confirmed through EBOV virus-like particle (VLP) assays <sup>[18]</sup>. Das et al. further characterised the role of Ca<sup>2+</sup> in EBOV entry using single-molecule FRET (smFRET)-imaging. It was shown that Ca<sup>2+</sup> and pH synergistically induce a conformational change in the EBOV glycoprotein GP2 (a key mediator of receptor binding and viral entry) to form a reversible intermediate state primed for NPC1 binding. NPC1 binding then further promotes the conformational transition into a fusion-ready “primed” state of invading EBOV virions <sup>[19]</sup>.



**Figure 1.** Ion channels implicated in viral entry since 2017. (A) Ifenprodil, glibenclamide, and TEA inhibit HIV through blockade of GIRK channels and  $K_{ATP}$  channels. (B) TEA and quinidine inhibit HAZV escape from EEs via blockade of an unknown channel. (C) Endosomal escape of SFTSV is inhibited by benidipine hydrochloride and nifedipine. (D) BUNV escape from late endosomes is inhibited by  $K_{2P}$  blockade. (E) EBOV escape from lysosomes is TPC2-dependent and can be blocked by verapamil, tetrandrine, nicardipine, diltiazem, and fluphenazine. (F) MERS escape from endolysosomes is prevented by tetrandrine, fangchinoline, verapamil, nimodipine, and nicardipine blockade of TPCs. (G) MCPyV and (H) SV40 ER translocation is TPC2 mediated and can be inhibited by verapamil and tetrandrine. ER translocation of MCPyV is also susceptible to blockade of  $K_V$  and T-type VGCCs by 4-AP and flunarizine respectively. (I) ER trafficking of BKPvV is CFTR dependent and susceptible to blockade by CFTR172 and glibenclamide. Key: PM: plasma membrane; EE: early endosome; LE: late endosome; EL: endolysosome. Ver: verapamil; Tet: tetrandrine; Nic: nicardipine; Dilt: diltiazem; Fluph: fluphenazine; Fang: fangchinoline; Nim: nimodipine; Nif: nifedipine; TEA: tetraethylammonium; Qd: quinidine; lfe: ifenprodil; Glib: glibenclamide; 4-AP: 4-aminopyridine.

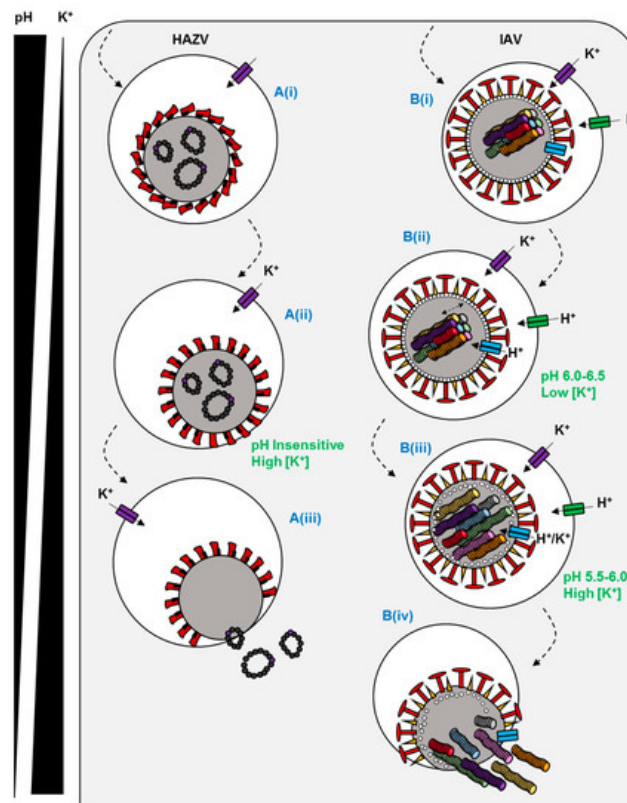
Of importance to the current pandemic, it has been shown that Middle East respiratory syndrome coronavirus (MERS) [20] is dependent on TPCs to escape endosomes [21]. As an enveloped virus, MERS must fuse its envelope with host membranes to enter cells. Following receptor attachment, MERS particles can fuse at either the cell surface or intracellularly in the endosomal network. Fusion is mediated by the proteolytic cleavage of the viral spike (S) protein at its S1/S2 site. At the surface of the cell, this proteolytic event is facilitated by TMPRSS2, which in turn precludes exposure of the fusion loop and coalescence of host and viral membranes [22]. Alternatively, fusion can occur in late endosomes following translocation through the endocytic network and proteolytic processing by proprotein convertases, including furin, in a process regulated by  $Ca^{2+}$  [23][24]. In studies by Gunaratne et al., genetic silencing of TPC1 and TPC2 prevented the entry of pseudotyped MERS (Figure 1F). The dependence of MERS upon TPCs was further demonstrated through its inhibition by tetrandrine and fangchinoline (TPC inhibitors), which prevented a post-internalisation but pre-fusion entry event. The mechanism through which TPC blockade inhibited MERS was multi-faceted: TPC1 and TPC2 silencing impaired furin activity, whilst pharmacological and genetic inhibition of TPC1 impaired endosomal motility. Of note, the related SARS-CoV-2, the causative agent of COVID-19 [25][26], was similarly inhibited by TPC blockade. Specifically, treatment of cells with tetrandrine reduced the entry of a lentiviral vector pseudotyped with the SARS-CoV-2 S (spike) [27].

The bunyavirus severe fever with thrombocytopenia syndrome virus (SFTSV) is an emerging arbovirus with fatality rates of 12–50% and the potential to cause future pandemics [28]. Using a library of FDA-approved drugs, the  $Ca^{2+}$  channel blockers benidipine hydrochloride and nifedipine were shown to inhibit SFTSV infection (Figure 1C), with in vivo activity confirmed using C57BL/6 and humanised mouse models. Through a retrospective analysis of human SFTSV cases, clinical evidence of the efficacy of nifedipine as an anti-SFTSV therapeutic was also demonstrated. A cohort of patients receiving nifedipine prior to and during hospital admission showed enhanced viral clearance and reduced frequency of neurological syndromes, often associated with fatal outcomes [29][30]. The fatality rate of patients receiving nifedipine was

reduced 5-fold compared to untreated patients, which corresponded to abnormal serum  $\text{Ca}^{2+}$  levels at admission. The viral processes through which SFTSV requires  $\text{Ca}^{2+}$  channels were subsequently shown to be during virus internalisation and genome replication.

## 2.2. $\text{K}^+$ Channels and Viral Entry

The involvement of  $\text{K}^+$  channels in viral entry has been extensively characterised for the model bunyaviruses Bunyamwera orthobunyavirus (BUNV); and Hazara orthonairovirus (HAZV), a model for Crimean Congo haemorrhagic fever virus, which causes severe viral haemorrhagic fever outbreaks, with a case fatality rate of up to 40%. Initial work using known  $\text{K}^+$  channel pharmacology suggested that the blockade of two-pore  $\text{K}^+$  channels ( $\text{K}_{2\text{P}}$ ) inhibited the early stages of the BUNV lifecycle (Figure 1D) [31]. Subsequent work identified both acidic pH and  $\text{K}^+$  in endosomes as crucial biochemical cues for the endosomal escape of BUNV [32]. Similar studies in HAZV highlighted a dependence on  $\text{K}^+$  channels for infection, and that  $\text{K}^+$  primarily accumulated in cholesterol-rich endosomes (Figure 1B) [32][33][34]. The  $\text{K}^+$  dependence of HAZV involves the glycoprotein spikes; a change in  $\text{K}^+$  concentration triggers conformational changes in the glycoproteins, as revealed through cryo-electron tomography of HAZV virions incubated with  $\text{K}^+$  that “primed” them for insertion into target membranes (Figure 2A). Moreover, it was shown that both BUNV and HAZV could be “primed” in vitro in buffers containing high  $[\text{K}^+]$ , which expedited entry and subsequent viral gene expression. This phenomenon was analogous to earlier studies for IAV, in which acid bypass in the presence of  $\text{K}^+$  revealed that the exposure of IAV virions to low pH and high  $[\text{K}^+]$  weakened interactions between the M1 matrix protein and ribonucleoprotein (RNP) bundles, a pre-requisite for genome release (Figure 2B). The exposure of IAV virions to  $\text{K}^+$  therefore drives viral uncoating and expedites IAV infection [35].



**Figure 2.** Predicted mechanisms of ion channel dependence for two enveloped viruses. **(A(i))** HAZV is endocytosed by an undefined mechanism. **(A(ii))** Endosomal  $\text{K}^+$  influx triggers a conformational change in the HAZV glycoprotein spikes to a fusion-ready state. **(A(iii))** Host and viral membranes fuse and RNPs are liberated into the cytosol. **(B(i))** IAV is endocytosed via a clathrin-dependent or independent mechanism. **(B(ii))** The virus traffics to late endosomes where the M2 viroporin is activated by acidic pH. **(B(iii))** The influx of  $\text{K}^+$  and  $\text{H}^+$  destabilises matrix-RNP interactions in the core. **(B(iv))** At low pH, a conformational change in HA promotes fusion and RNP release.

Recent work also highlights a requirement for  $\text{K}^+$  channels during human immunodeficiency virus (HIV) infection. Using pharmacological approaches (Figure 1A) [36] HIV entry could be blocked with ifenprodil and the broad spectrum  $\text{K}^+$  channel blocker tetraethylammonium (TEA). Khan et al. also showed that the pharmacological activation of the endolysosome-resident transient receptor potential mucolipin 1 channel (TRPML1) enhanced the degradation of HIV-Tat (a multi-function viral protein involved in transcription, splicing, capping, and translation), which in turn reduced the transition from viral latency [37]. TRPML1 activates large conductance  $\text{Ca}^{2+}$ -activated potassium (BK) channels in endosomes [38], implying that this channel is required for HIV infection.

The reliance of viruses upon ion channels is not restricted to RNA viruses. It was recently shown that both  $K^+$  and  $Ca^{2+}$  channels are important host factors for polyomavirus infection [39]. Using a panel of ion channel modulators, the entry of Merkel cell polyomavirus (MCPyV), the causative agent of Merkel cell carcinoma (MCC), was shown to be sensitive to 4-aminopyridine (4-AP), a blocker of voltage-gated  $K^+$  ( $K_V$ ) channels (Figure 1G). Moreover, both MCPyV and Simian virus 40 (SV40) (Figure 1H) were sensitive to verapamil, a broad-spectrum  $Ca^{2+}$  channel blocker. The identities of the  $Ca^{2+}$  channels involved in polyomavirus entry were further explored, revealing a requirement for transient (T-type, low-voltage activated) channel family members in MCPyV infection but not SV40. Tetrandrine, a TPC blocker, restricted both viruses. The role of TPCs was found to be during endoplasmic reticulum (ER) disassembly and/or ER docking for SV40 [40], which may be explained by the recent demonstration that  $Ca^{2+}$  ions mediate the stabilization of SV40 capsids and contribute to its disassembly [41].

### 2.3. $Cl^-$ Channels and Viral Entry

BK polyomavirus (BKPyV) is a potentially fatal pathogen in patients undergoing solid organ transplantation. Panou et al. demonstrated that the pharmacological and genetic disruption of the cystic fibrosis transmembrane conductance regulator (CFTR)  $Cl^-$  channel could reduce BKPyV infection in primary kidney cell models (Figure 1I) [42]. Time of addition assays using the CFTR inhibitors CFTR<sub>172</sub> and glibenclamide, combined with the assessment of exposure of VP2/VP3 minor capsid proteins, indicated a role for CFTR in the trafficking of BKPyV to the ER. Whilst the mechanism of CFTR involvement in BKPyV ER trafficking remains unclear, it is hypothesised that the channel may be important in the acidification and ER docking of BKPyV-containing endosomes. CFTR has been implicated in the fusion of endosomes [43], and co-localises with vacuolar ATPase (vATPase) to provide the counter-charge during endosomal acidification [44].

---

## References

1. Yu, F.H.; Yarov-Yarovoy, V.; Gutman, G.A.; Catterall, W.A. Overview of molecular relationships in the voltage-gated ion channel superfamily. *Pharmacol. Rev.* 2005, 57, 387–395.
2. Hanna, M.G. Genetic neurological channelopathies. *Nat. Clin. Pr. Neurol.* 2006, 2, 252–263.
3. Netter, R.C.; Amberg, S. M.; Balliet, J. W.; Biscone, M. J.; Vermeulen, A.; Earp, L. J.; White, J. M.; Bates, P. Heptad repeat 2-based peptides inhibit avian sarcoma and leukosis virus subgroup a infection and identify a fusion intermediate. *J. Virol.* 2004, 78, 13430–13439.
4. Verkman, A.S.; Galletta, L. Chloride channels as drug targets. *Nat. Rev. Drug Discov.* 2008, 8, 153–171.
5. Phillips, L.; Trivedi, J.R. Skeletal muscle channelopathies. *Neurotherapeutics* 2018, 15, 954–965.
6. Kim, J.-B. Channelopathies. *Korean J. Pediatr.* 2014, 57, 1–18.
7. Garcia-Elias, A.; Benito, B. Ion channel disorders and sudden cardiac death. *Int. J. Mol. Sci.* 2018, 19, 692.
8. Vaeth, M.; Feske, S. Ion channelopathies of the immune system. *Curr. Opin. Immunol.* 2018, 52, 39–50.
9. Farre, C.; Fertig, N. New strategies in ion channel screening for drug discovery: Are there ways to improve its productivity? *Expert Opin. Drug Discov.* 2014, 9, 1103–1107.
10. McManus, O.B. HTS assays for developing the molecular pharmacology of ion channels. *Curr. Opin. Pharmacol.* 2014, 15, 91–96.
11. Nieva, J.L.; Madan, V.; Carrasco, L. Viroporins: Structure and biological functions. *Nat. Rev. Genet.* 2012, 10, 563–574.
12. Nieva, J.L.; Carrasco, L. Viroporins: Structures and functions beyond cell membrane permeabilization. *Viruses* 2015, 7, 5169–5171.
13. Royle, J.; Dobson, S.J.; Müller, M.; Macdonald, A. Emerging roles of viroporins encoded by DNA viruses: Novel targets for antivirals? *Viruses* 2015, 7, 5375–5387.
14. Hover, S.; Foster, B.; Barr, J.N.; Mankouri, J. Viral dependence on cellular ion channels—An emerging antiviral target? *J. Gen. Virol.* 2017, 98, 345–351.
15. Luzio, J.P.; Bright, N.; Pryor, P. The role of calcium and other ions in sorting and delivery in the late endocytic pathway. *Biochem. Soc. Trans.* 2007, 35, 1088–1091.
16. Fujioka, Y.; Nishide, S.; Ose, T.; Suzuki, T.; Kato, I.; Fukuhara, H.; Fujioka, M.; Horiuchi, K.; Satoh, A.O.; Nepal, P.; et al. A sialylated voltage-dependent  $Ca^{2+}$  channel binds hemagglutinin and mediates influenza A virus entry into mammalian cells. *Cell Host Microbe* 2018, 23, 809–818.

17. Simmons, J.A.; D'Souza, R.S.; Ruas, M.; Galione, A.; Casanova, J.E.; White, J.M. Ebola virus glycoprotein directs fusion through NPC1+ endolysosomes. *J. Virol.* 2015, 90, 605–610.
18. Penny, C.J.; Vassileva, K.; Jha, A.; Yuan, Y.; Chee, X.; Yates, E.; Mazzon, M.; Kilpatrick, B.S.; Muallem, S.; Marsh, M.; et al. Mining of Ebola virus entry inhibitors identifies approved drugs as two-pore channel pore blockers. *Biochim. Biophys. Acta Mol. Cell Res.* 2018.
19. Das, D.K.; Bulow, U.; Diehl, W.E.; Durham, N.D.; Senjobe, F.; Chandran, K.; Luban, J.; Munro, J.B. Conformational changes in the Ebola virus membrane fusion machine induced by pH, Ca<sup>2+</sup>, and receptor binding. *PLoS Biol.* 2020, 18, e3000626.
20. Zumla, A.; Hui, D.S.; Perlman, S. Middle east respiratory syndrome. *Lancet* 2015, 386, 995–1007.
21. Gunaratne, G.S.; Yang, Y.; Li, F.; Walseth, T.F.; Marchant, J.S. NAADP-dependent Ca<sup>2+</sup> signaling regulates Middle East respiratory syndrome-Coronavirus pseudovirus translocation through the endolysosomal system. *Cell Calcium* 2018, 75, 30–41.
22. Park, J.E.; Li, K.; Barlan, A.; Fehr, A.R.; Perlman, S.; McCray, P.B.; Gallagher, T. Proteolytic processing of middle east respiratory syndrome coronavirus spikes expands virus tropism. *Proc. Natl. Acad. Sci. USA* 2016, 113, 12262–12267.
23. Mille, J.K.; Whittaker, G.R. Host cell entry of Middle East respiratory syndrome coronavirus after two-step, furin-mediated activation of the spike protein. *Proc. Natl. Acad. Sci. USA* 2014, 111, 15214–15219.
24. Ruas, M.; Rietdorf, K.; Arredouani, A.; Davis, L.C.; Lloyd-Evans, E.; Koegel, H.; Funnell, T.M.; Morgan, A.J.; Ward, J.A.; Watanabe, K.; et al. Purified TPC isoforms form NAADP receptors with distinct roles for Ca<sup>2+</sup> signaling and endolysosomal trafficking. *Curr. Biol.* 2010, 20, 703–709.
25. Huang, C.; Wang, Y.; Li, X.; Ren, L.; Zhao, J.; Hu, Y.; Zhang, L.; Fan, G.; Xu, J.; Gu, X.; et al. Clinical features of patients infected with 2019 novel coronavirus in Wuhan, China. *Lancet* 2020, 395, 497–506.
26. Zhu, N.; Zhang, D.; Wang, W.; Li, X.; Yang, B.; Song, J.; Zhao, X.; Huang, B.; Shi, W.; Lu, R.; et al. A novel coronavirus from patients with pneumonia in China, 2019. *N. Engl. J. Med.* 2020, 382, 727–733.
27. Ou, X.; Liu, Y.; Lei, X.; Li, P.; Mi, D.; Ren, L.; Guo, L.; Guo, R.; Chen, T.; Hu, J.; et al. Characterization of spike glycoprotein of SARS-CoV-2 on virus entry and its immune cross-reactivity with SARS-CoV. *Nat. Commun.* 2020, 11, 1–12.
28. Li, H.; Zhang, L.K.; Li, S.F.; Zhang, S.F.; Wan, W.W.; Zhang, Y.L.; Xin, Q.L.; Dai, K.; Hu, Y.Y.; Wang, Z.B.; et al. Calcium channel blockers reduce severe fever with thrombocytopenia syndrome virus (SFTSV) related fatality. *Cell Res.* 2019, 29, 739–753.
29. Li, H.; Lu, Q.B.; Xing, B.; Zhang, S.F.; Liu, K.; Du, J.; Li, X.K.; Cui, N.; Yang, Z.D.; Wang, L.Y.; et al. Epidemiological and clinical features of laboratory-diagnosed severe fever with thrombocytopenia syndrome in China, 2011–17: A prospective observational study. *Lancet Infect. Dis.* 2018, 18, 1127–1137.
30. Bao, C.J.; Guo, X.L.; Qi, X.; Hu, J.L.; Zhou, M.H.; Varma, J.K.; Cui, L.B.; Yang, H.T.; Jiao, Y.J.; Klena, J.D.; et al. A family cluster of infections by a newly recognized bunyavirus in Eastern China, 2007: Further evidence of person-to-person transmission. *Clin. Infect. Dis.* 2011, 53, 1208–1214.
31. Hover, S.; King, B.; Hall, B.; Loundras, E.A.; Taqi, H.; Daly, J.; Dallas, M.; Peers, C.; Schnettler, E.; McKimmie, C.; et al. Modulation of potassium channels inhibits bunyavirus infection. *J. Biol. Chem.* 2016, 291, 3411–3422.
32. Hover, S.; Foster, B.; Fontana, J.; Kohl, A.; Goldstein, S.A.; Barr, J.N.; Mankouri, J. Bunyavirus requirement for endosomal K<sup>+</sup> reveals new roles of cellular ion channels during infection. *PLoS Pathog.* 2018, 14, e1006845.
33. Punch, E.K.; Hover, S.; Blest, H.T.; Fuller, J.; Hewson, R.; Fontana, J.; Mankouri, J.; Barr, J.N. Potassium is a trigger for conformational change in the fusion spike of an enveloped RNA virus. *J. Biol. Chem.* 2018, 293, 9937–9944.
34. Charlton, F.W.; Hover, S.; Fuller, J.; Hewson, R.; Fontana, J.; Barr, J.N.; Mankouri, J. Cellular cholesterol abundance regulates potassium accumulation within endosomes and is an important determinant in bunyavirus entry. *J. Biol. Chem.* 2019, 294, 7335–7347.
35. Stauffer, S.; Feng, Y.; Nebioglu, F.; Heilig, R.; Picotti, P.; Helenius, A. Stepwise priming by acidic pH and a high K<sup>+</sup> concentration is required for efficient uncoating of influenza A virus cores after penetration. *J. Virol.* 2014, 88, 13029–13046.
36. Dubey, R.C.; Mishra, N.; Gaur, R. G protein-coupled and ATP-sensitive inwardly rectifying potassium ion channels are essential for HIV entry. *Sci. Rep.* 2019, 9, 1–9.
37. Khan, N.; Lakpa, K.L.; Halcrow, P.W.; Afghah, Z.; Miller, N.M.; Geiger, J.D.; Chen, X. BK channels regulate extracellular Tat-mediated HIV-1 LTR transactivation. *Sci. Rep.* 2019, 9.

38. Cao, Q.; Zhong, X.Z.; Zou, Y.; Zhang, Z.; Toro, L.; Dong, X.P. BK channels alleviate lysosomal storage diseases by providing positive feedback regulation of lysosomal  $\text{Ca}^{2+}$  release. *Dev. Cell* 2015, 33, 427–441.
39. Dobson, S.J.; Mankouri, J.; Whitehouse, A. Identification of potassium and calcium channel inhibitors as modulators of polyomavirus endosomal trafficking. *Antiviral Res.* 2020, 104819.
40. Sakurai, Y.; Kolokoltsov, A.A.; Chen, C.C.; Tidwell, M.W.; Bauta, W.E.; Klugbauer, N.; Grimm, C.; Wahl-Schott, C.; Biel, M.; Davey, R.A. Two-pore channels control Ebola virus host cell entry and are drug targets for disease treatment. *Science* 2015, 347, 995–998.
41. Kawano, M.A.; Xing, L.; Tsukamoto, H.; Inoue, T.; Handa, H.; Cheng, R.H. Calcium bridge triggers capsid disassembly in the cell entry process of simian virus 40. *J. Biol. Chem.* 2009, 284, 34703–34712.
42. Panou, M.M.; Antoni, M.; Morgan, E.L.; Loundras, E.A.; Wasson, C.W.; Welberry-Smith, M.; Mankouri, J.; Macdonald, A. Glibenclamide inhibits BK polyomavirus infection in kidney cells through CFTR blockade. *Antiviral Res.* 2020, 178, 104778.
43. Bradbury, N.A. Intracellular CFTR: Localization and function. *Physiol. Rev.* 1999, 79, S175–S191.
44. Collaco, A.M.; Geibel, P.; Lee, B.S.; Geibel, J.P.; Ameen, N.A. Functional vacuolar ATPase (V-ATPase) proton pumps traffic to the enterocyte brush border membrane and require CFTR. *Am. J. Physiol. Cell Physiol.* 2013, 305, C981–C996.

---

Retrieved from <https://encyclopedia.pub/entry/history/show/3600>