

TNBC Histological Subtypes with a Favourable Prognosis

Subjects: **Oncology**

Contributor: Anikó Kovács

Triple-negative breast cancers (TNBC), as a group of tumours, have a worse prognosis than stage-matched non-TNBC and lack the benefits of routinely available targeted therapy. However, TNBC is a heterogeneous group of neoplasms, which includes some special type carcinomas with a relatively indolent course.

acinic cell carcinoma

adenoid cystic carcinoma

fibromatosis like metaplastic carcinoma

low-grade adenosquamous carcinoma

mucoepidermoid carcinoma

secretory carcinoma

tall cell carcinoma with reversed polarity

triple negative breast cancer

1. Introduction

Our understanding of the biology of human breast cancer (BC) has evolved exponentially since the first documented written reference to BC in the Edwin Smith papyrus in 3000 B.C. and ancient Greek times when Hippocrates perceived the disease as a single mortal one ^[1]. BC is now a collective term for a heterogeneous group of diseases, variously classified according to the putative cell of origin, histological type, tumour grade, molecular markers, tumour stage and other clinical and biological variables that correlate with outcome and indicate potential response to specific therapies.

Since the seminal work of Perou and colleagues, BC has been classified into molecular (intrinsic) subtypes according to their gene expression profiles (GEPs) ^[2]. The different molecular subtypes of BC are associated with different prognoses ^{[3][4]} with luminal A tumours having the best outcome and basal-like carcinomas demonstrating the worst survival ^{[3][4]}. In daily practice immunohistochemistry (IHC) is used as a surrogate classification system to categorise BCs, according to oestrogen receptor (ER), progesterone receptor (PR) and human epidermal receptor growth factor 2 (HER2) expression, into luminal (ER+ and/or PR+), HER2 positive and triple-negative tumours. Whilst there is considerable overlap between basal-like tumours and triple-negative breast cancers (TNBCs), these tumours are not identical ^[5] and the surrogate classification has a number of mismatches when compared to the GEP based classification ^[6]. As a group, TNBCs have worse overall survival (OS) than stage-matched non-TNBCs. An analysis of the Survival, Epidemiology and End Results database with 18,855 TNBCs and 139,503 non-TNBCs grouped according to their hormone receptor (HR) and HER2 statuses into HR+HER2-, HR+HER2+ and HR-HER2+, showed that the OS of TNBCs was significantly worse for all substages and groups of non-TNBCs with

hazard ratios between 1.40 and 4.19 (except for substages IA and IB of HR-HER2+ cancers, where the hazard ratios of 1.21 and 1.76 failed to be significant) [7].

It is now recognized that TNBC constitutes a heterogeneous group of tumours at the molecular level [8] with six subgroups originally identified [9][10], later refined to four: basal-like 1 and 2 tumours, that differ in their immune-response, mesenchymal and luminal androgen receptor tumours, recognised in the 5th edition of the World Health Organisation (WHO) Classification of Breast Tumours [11]. At present, no clinically verified molecular assay exists for the optimal classification of TNBC [11] and the prognostic and predictive relevance of this classification of TNBC has yet to be defined.

At the morphological level, TNBC is also a heterogeneous disease with high- and low-grade variants and a corresponding spectrum of biological behaviour. However, due to its frequent association with high histological grade and aggressive biological behaviour, the management of TNBC is a major focus in the field of medical oncology due to the lack of options for targeted therapy [12]. Although the prognosis of non-metastatic TNBC is influenced by a number of variables besides the triple negative phenotype [13][14][15], it is not an unexceptional practice to recommend adjuvant systemic chemotherapy (CT) to (nearly) all patients with TNBC to improve survival.

2. Acinic Cell Carcinoma (ACC) of the Breast

2.1. Definition, Main Features, Diagnostic Clues and Differential Diagnosis

Acinic cell carcinoma (ACC) of the breast is a subtype of TNBC, morphologically similar to ACC of the salivary glands (**Figure 1A**). It mainly affects adult women, presenting as a nodule/mass ranging in size from 10 to 52 mm (average 28 mm; summary in [Supplementary Table S1](#)) [16][17][18][19][20][21][22][23][24][25][26][27][28][29][30][31][32][33][34][35][36][37].

ACC can show a wide spectrum of architectural patterns. At one end of the spectrum, ACC is composed of a proliferation of small round glands (microglandular pattern), lined by a single layer of cuboidal to columnar epithelial cells resembling the acinar cell structure of salivary glands and microglandular adenosis of the breast. Eosinophilic/amphophilic secretions may be present in the glandular lumina. An in situ component may be associated. Tumour cells are polygonal, sometimes with clear (hypernephroid), PAS-positive, finely granular eosinophilic or basophilic cytoplasm and prominent nucleoli after diastase digestion. The eosinophilic zymogen-type cytoplasmic granules may be large and coarse, resembling intestinal Paneth cells. Mitoses are variably present [11][38]. Tumour cells show serous differentiation verified by a positive immunoreaction for amylase, lysozyme and α -1 antichymotrypsin. Epithelial membrane antigen (EMA), S-100, low molecular weight cytokeratins (CKs) are usually positive; and gross cystic disease fluid protein 15 (GCDFFP-15) may be positive. No pathognomonic genetic alterations have been identified in three breast ACCs analysed by whole-exome sequencing. The alterations identified were similar to those seen in conventional TNBC of NST [39]. On the basis of

the small series analysed, commonly occurring mutations affect the following genes: *PIK3CA*, *KMT2D*, *ERBB4/ERBB3*, *NEB*, *BRCA1*, *MTOR*, *CTNNB1*, *INPP4B* and *FGFR2* [11][34][40].

At the other end of the spectrum, ACC is composed of circumscribed solid nests of variable size with comedo-type necrosis, prominent nuclear atypia and increased mitotic activity. ACC may also be admixed with NST or metaplastic elements. ACC may show intra-tumour heterogeneity with both well-differentiated microglandular and less-differentiated solid areas. Thorough sampling of ACC is important to identify components that are likely to change the perceived risk of these tumours and may explain the clinical events reported in some studies (see below).

2.2. Clinical Correlations, Treatment and Outcome Data

ACC was originally described in 1996 [16], with a limited number of reported cases in the English literature. Foschini et al. [41] listed 45 ACCs with a further 2 cases since reported [36][37]. Some studies include very limited or no follow-up data. [Supplementary Table S1](#) is an adapted version of the table reported by Foschini et al. [41] including only those patients for whom follow-up data were available (follow-up range 3 to 184.8 months, average 73 months) and updated with recent literature. In approximately half of the patients (16/35) follow-up was less than 2 years ([Supplementary Table S1](#)).

At presentation most patients with ACC regardless of its subtype (24/35, 68.6%) had pT2 tumours. Axillary lymph node metastases were detected in 8/31 (26%), only one of which was associated with a pT1c tumour. In most patients, regional axillary metastases were limited to 1 or 2 lymph nodes. One patient had a high axillary metastatic burden [34] and died of disease 24 months after presentation [34]. Local recurrences were observed in 3 patients [18][34][35], one of whom had a large tumour treated with local excision only [18] and one who developed recurrence despite aggressive treatment [35]. No data on surgical treatment were reported for the remaining patients [34]. Distant metastases (lung, bone, liver) are reported in 3 patients, two of whom died from their disease [20][23][25]. Sixteen patients (comprising all with distant metastases) were treated with adjuvant therapies ([Supplementary Table S1](#)). ACC, when associated with other types of high-grade conventional BC, can show aggressive behaviour mimicking that of the higher grade component. Sardana et al. [42] reported a patient with ACC, associated with NST and metaplastic BC components who developed meningeal metastases and died from her disease.

It is difficult to draw firm conclusions regarding the prognosis of breast ACC from available literature in view of morphological variation within and between tumours (pure and mixed types) in the published studies. It is our opinion that low-grade pure ACC is a bland tumour type that overlaps with microglandular adenosis and is associated with indolent biological behaviour, and is, therefore, unlikely to benefit from aggressive adjuvant chemotherapy. ACCs with high-grade areas or admixed with other BC types are likely to behave more aggressively and may account for some of the reported events in the literature. Currently, there is no evidence to support withholding systemic chemotherapy in ACC with high-grade features if clinically indicated. Further data on their response to therapy is needed.

3. Classic Adenoid Cystic Carcinoma (CAdCC)

3.1. Definition, Main Features, Diagnostic Clues and Differential Diagnosis

Adenoid cystic carcinoma (AdCC) is a rare salivary gland type tumour in the breast, first described as a cylindroma of the breast by Billroth in 1856 [43][44]. According to the current WHO classification of breast tumours three subtypes of AdCC are recognized: classic, solid basaloid, and AdCC with high-grade transformation [11].

Classic AdCC (CAdCC) is usually unifocal [44] but may be multifocal [45]. Approximately half of all cases arise in the subareolar region [46]. Microscopically, the tumour is composed of epithelial (luminal) and myoepithelial (abluminal) neoplastic cells arranged in tubular and cribriform patterns with intervening lumina containing mucin and pseudolumina containing reduplicated basement membrane material (**Figure 1B**) [11]. A minor solid growth pattern is occasionally observed in the classical subtype but is more common in the solid basaloid variant and AdCC with high grade transformation. A solid growth pattern and high-grade features are histological signs of likely aggressive biological behaviour in AdCC.

The cells of the epithelial component are positive for CK7, CK5/6, CK 8/18 and CD117 [47]. The myoepithelial/abluminal cells express p63, smooth muscle actin and basal CKs: CK5/6, CK14, CK17. CAdCC generally displays a triple-negative immunophenotype, although ER-positive cases also occur ([Supplementary Table S2](#)) [46][48][49][50][51][52][53][54][55][56][57][58][59][60][61][62][63][64][65][66][67][68]. CAdCC frequently expresses EGFR, an immunohistochemical marker of the “basal-like” phenotype, without gene amplification [69]. CAdCC may also express a truncated form of the ER receptor-alpha, which is not detected by antibodies in general use [70] and the significance of which is not known. CAdCC has a low proliferative fraction [11], in keeping with the overall good prognosis associated with this tumour.

Most AdCCs investigated to date have harboured the *MYB-NFIB* fusion gene. AdCCs lacking the *MYB-NFIB* fusion gene may show *MYBL1* rearrangements or *MYB* amplification [71]. The most frequently mutated genes in AdCC include *MYB*, *BRAF*, *FBXW7*, *SMARCA5*, *SF3B1*, and *FGFR2*. AdCCs appear to lack somatic mutations in the *TP53*, *PIK3CA*, *RB1*, *BRCA1* and *BRCA2* genes which are often mutated in TNBC, NST and basal-like breast cancers [72].

CAdCC shows morphological overlap with a variety of benign, atypical and good-prognosis malignant breast lesions including collagenous spherulosis, syringomatous adenoma, adenomyoepithelioma, cribriform ductal carcinoma in situ (DCIS) and invasive cribriform carcinoma (ICC) [47]. CD117, which is positive in the luminal cells of CAdCC, is negative in collagenous spherulosis. Calponin and smooth muscle myosin heavy chain, negative in abluminal CAdCC cells, are strongly positive in collagenous spherulosis [73]. Syringomatous adenoma lacks cytologic atypia [47]. The myoepithelial cells in adenomyoepithelioma are positive for calponin while the abluminal cells in CAdCC are negative [47]. Cribriform DCIS shows regular contours with peripheral myoepithelial cells but no internal reactivity for myoepithelial cell markers [47]. The stroma of ICC tends to be desmoplastic, whereas in

CAdCC the stroma shows myxoid change around the cribriform islands. ICCs are typically ER-positive, with 69% also PR positive, and HER2 negative [11].

3.2. Clinical Correlations, Treatment and Outcome Data

A review of the published literature identified database studies and small case series in addition to single case reports concentrating on tumours associated with an unfavourable outcome ([Supplementary Table S2](#)) [46][49][50][51][52][53][54][55][56][57][58][59][60][61][62][63][64][65][66][67][68]. The reported series often included a mixture of AdCC tumours with CAdCC rarely studied in isolation. This may be related to the fact, that prior to the 2019 edition, the WHO classification of breast tumours did not categorically distinguish between the three morphological variants of AdCC, although the solid form had been described in the 4th edition [74].

Taking account of the publications to date, the median age of patients with AdCC ranged from 58 to 66 years. Data on the ethnical differences are limited, but larger registry-based series suggest that approximately 85% of patients diagnosed in the USA were whites [46][48][50][52][53]. The majority of patients presented with a palpable mass (85.7%) and underwent lumpectomy, breast-conserving surgery (BCS) or mastectomy. Lymph node status was assessed in all reviewed cases, and nodal involvement ranged from 0 to 15% with the exception of a small series ($n = 19$), where it reached 27% [60]. Although grading was applied to 57% of the collected cases in [Supplementary Table S2](#), the methods were different and included the original Bloom and Richardson system, the Nottingham (modified Bloom and Richardson) system, nuclear grade (NG1-3), a three-tiered grading for salivary gland AdCC (G1-G3), and low grade vs. high grade.

Adjuvant radiotherapy was administered to 17–66% of the patients, all of whom had BCS. The percentage of patients who received chemotherapy ranged from 4 to 66%, not exceeding 25% in most studies ([Supplementary Table S2](#)).

The literature on rare entities including CAdCC is limited and of questionable value without a detailed analysis of pathological characteristics. Slodkowska et al. presented a precise pathological approach with predictors of clinical outcome [68] and confirmed that pure CAdCC has an excellent prognosis. In contrast, solid-basaloid AdCC is an aggressive variant as supported by survival analysis [68]. Slodkowska et al. also confirmed that the Nottingham grading system was a strong predictor of the behaviour of breast AdCC. Perineural invasion was more often seen in tumours that recurred (19% overall and 50% in recurring cases). Neovascularisation also emerged as a new independent predictor of aggressive behaviour in this disease [68].

Therefore, CAdCC in its pure form, without solid or transformed components has a good prognosis and patients with this entity are unlikely to derive benefit from adjuvant chemotherapy.

4. Fibromatosis-Like Metaplastic Carcinoma (FLMC)

4.1. Definition, Main Features, Diagnostic Clues and Differential Diagnosis

Fibromatosis-like metaplastic carcinoma (FLMC) represents a rare low-grade subtype of metaplastic breast carcinoma (MBC). With a total of 70 cases reported in the peer-reviewed English language literature ([Supplementary Table S3](#)) [\[75\]\[76\]\[77\]\[78\]\[79\]\[80\]\[81\]\[82\]\[83\]\[84\]\[85\]\[86\]\[87\]\[88\]](#), FLMC is likely to account for significantly less than 1% of all MBCs. Due to its bland morphology, FLMC is diagnostically challenging and is likely to account for a higher relative percentage of tumours in referral centres, diagnosed at the time of primary presentation or on review following a recurrence of an incompletely excised FLMC originally (mis)diagnosed as a benign entity.

FLMC is characterized by a proliferation of spindled fibroblast-like cells and stellate myofibroblast-like cells, which compose more than 95% of the total tumour area, and histologically resembles fibromatosis. The neoplastic cells are cytologically bland with absent to minimal nuclear atypia and pale eosinophilic cytoplasm. Within the same tumour, there may be a gradual transition from cells with thin, slender, spindled nuclei with tapered ends to cells with plump, round to oval nuclei with discrete nucleoli within finely distributed chromatin. High nuclear grade is not seen [\[11\]\[75\]\[76\]](#). Mitotic figures are either completely absent or rare [\[75\]\[76\]](#). Necrosis is not identified. Neoplastic squamous or glandular epithelial elements may be admixed with the spindle cells but should account for less than 5% of the total tumour area and FLMC should not contain any other differentiated component (**Figure 1C**) [\[11\]](#). The epithelial nature of the spindle cells is often only recognisable on immunohistochemistry and may be focal. The use of a panel of CKs is advisable to confirm the diagnosis, including broad-spectrum, high and low molecular weight CKs e.g., CK AE1/AE3, MNF116, 34betaE12, CK8/18, CK5/6, and CK14. Most tumours also express p63. Expression of vimentin and other myoepithelial markers is variable. ER, PR and HER2 are negative [\[75\]\[76\]\[89\]\[90\]\[91\]](#).

FLMC and low-grade adenosquamous carcinoma (LGASC) may represent a spectrum, composed of similar histological components (spindle, glandular and squamous), with the glandular and squamous components accounting for less than 5% in FLMC and a significantly greater percentage of LGASC. It is our experience that most of the missed LGASC lack glandular or squamous components and just feature bland-looking spindle cell proliferation that express epithelial markers. Both FLMC and LGASC may incorporate sclerosing papillary or adenomyepitheliomatous foci with which they share molecular alterations including mutations in the *PIK3CA* and *SF3B1* genes, suggesting a possible pathogenetic link.

In clinical practice pathologists often differentiate FLMC from fibromatosis using IHC markers. Fibromatosis lacks CK expression and usually shows abnormal nuclear localization of beta-catenin. The two entities are also different at the molecular level, with exon 3 *CTNNB1* mutations being highly specific for desmoid type fibromatosis among spindle cell lesions of the breast, and *APC* mutations also being relatively common in fibromatosis [\[92\]](#). FLMC should also be distinguished from some more aggressive malignant lesions with overlapping morphology. Spindle cell MBC is associated with recognisable nuclear atypia and is usually larger. Pure FLMC should not display intermediate/high-grade nuclear features or be admixed with other BC types such as NST or lobular carcinoma. The presence of one or more of the latter components in a tumour with low-grade spindle cell morphology excludes a diagnosis of FLMC [\[11\]\[89\]](#).

4.2. Clinical Correlations, Treatment and Outcome Data

Since the first detailed description of the histological criteria for the diagnosis of FLMC by Gobbi et al. in 1999 (adopted by the current WHO classification), a total of 70 cases have been published in the English-language literature with follow-up data available for 41. Overall, FLMC shows clinically indolent behaviour with a high tendency for local recurrence (14/41 cases), but with a low potential for regional lymph node (3/41 cases) or distant metastases (5/41 cases) ([Supplementary Table S3](#)) [75][76][77][78][79][80][81][82][83][84][85][86][87][88].

Our review of the literature suggests that the risk of local recurrence is at least partly related to inadequate local resection [75][76][80]. Lack or incomplete excision of these tumours may lead to progressive growth and acquisition of additional mutations and more aggressive behaviour. It is our view that the metastatic events reported in the literature for cases of FLMC likely reflect the inclusion of tumours with a high-grade spindle cell component (MBC). The association between tumour diameter and the risk of distant metastasis assumed by individual authors is not supported by data in the published literature ([Supplementary Table S3](#)).

There are no evidence-based treatment guidelines for FLMC. Regarding local therapy, wide excision appears to prevent local recurrence [75][76][80]. The available data do not support the benefit of adjuvant radiotherapy or chemotherapy in reducing the risk of local recurrence or metastasis ([Supplementary Table S3](#)) [75][76][77][78][79][80][81][82][83][84][85][86][87][88]. However, some authors, including the present authors, advocate the use of adjuvant radiotherapy for more bulky lesions [90]. Because of the very low risk of regional lymph node metastases, several authors argue against axillary lymph node dissection in pure FLMC particularly in the management of small tumours [75][89].

5. Low-Grade Adenosquamous Carcinoma (LGASC)

5.1. Definition, Main Features, Diagnostic Clues and Differential Diagnosis

Low-grade adenosquamous carcinoma (LGASC) of the breast is a rare variant of MBC with a favourable outcome. Histologically, these tumours are composed of well-developed glandular and tubular formations intimately admixed with solid nests of squamous cells arranged in a haphazard, infiltrative pattern in a spindle cell background, typically associated with a peripheral lymphocytic infiltrate (**Figure 1D**) [11]. Cytological atypia is mild and proliferative activity, measured by mitotic activity and Ki-67 index, is low [93].

LGASC constitutes a distinct genetic entity among MBCs, characterised by high rates of *PIK3CA* mutations, lack of *TP53* mutations, a triple-negative phenotype and lack of androgen receptor (AR) expression [93]. There is generally expression of luminal (CK7, CK8) and basal (CK5, CK14) CKs and squamous (myoepithelial) markers p63 and p40.

LGASC should be distinguished from other TNBC and basal-like carcinomas that are associated with aggressive biological behaviour, i.e., high-grade MBCs that may have areas of squamous metaplasia and spindle cells. Nuclear pleomorphism, mitoses, necrosis, a prominent malignant appearing sarcomatous spindle cell component

and solid nests of atypical squamous cells support classification as spindle cell MBC and/or squamous carcinoma rather than as LGASC.

5.2. Clinical Correlations, Treatment and Outcome Data

Rosen and Ernsberger ^[94] described this entity in 1987 emphasising that, despite the presence of metaplastic elements, this tumour displays low-grade histological features. In keeping with their low-grade morphological features, the majority of LGASCs have an excellent prognosis with a low incidence of lymph node metastases.

A review of the published English literature identified all reports of LGASC of the breast. Case reports/series without data on treatment and outcome were excluded. A total of 15 publications dating from 1987 to the present describing 92 cases of LGASC with data on treatment and outcome were identified ([Supplementary Table S4](#)) ^{[93][94][95][96][97][98][99][100][101][102][103][104][105][106][107]}. Patient age ranged from 19 to 88 years. The tumours were unilateral apart from one patient with bilateral LGASC. The majority of patients presented with a palpable mass (83%). Patients underwent BCS or excisional biopsy (67%) or mastectomy (33%). Lymph node status was assessed in 34 patients (37%) with only one incidence of nodal metastasis reported. Adjuvant radiotherapy was administered to 18 patients, all of whom had BCS or excisional biopsy, and seven patients received chemotherapy.

Early studies ^{[94][107]} reported relatively high rates of local recurrence, in 36% and 20% of patients respectively all of whom had been treated initially by excisional biopsy only. In more recent literature, only a single patient had subsequent progression of the disease ^[101].

Although the literature on this rare tumour is limited, the available data strongly suggest that this is a malignant tumour with an indolent course and a favourable prognosis. Recent studies demonstrate a low rate of local recurrence with no nodal or distant metastases, anyway the definitive and optimal treatment for LGASC has yet to be determined. The benefit of adjuvant radiotherapy has not been studied yet. Chemotherapy does not appear to be warranted and adjuvant hormonal treatment is not indicated because of the triple negative phenotype of these tumours. At present, the clinical management of LGASC requires complete surgical excision followed by adjuvant radiotherapy in case of conservative surgery.

6. Mucoepidermoid Carcinoma (MEC) of the Breast

6.1. Definition, Main Features, Diagnostic Clues and Differential Diagnosis

Mucoepidermoid carcinoma (MEC) of the breast is an invasive carcinoma composed of mucoid, epidermoid and intermediate cells analogous to the tumour of the same name encountered in salivary glands (**Figure 1E**) ^[11]. In contrast to minor salivary glands, where this entity is the most common malignancy, breast MEC is rare, with less than 50 cases reported to date in the English literature.

Breast MEC affects middle-aged and elderly women. It presents as a unilateral nodule, sometimes with a cystic component. On ultrasound examination, MEC can simulate a benign lesion with enhancement rather than

shadowing [\[108\]](#).

Morphologically, it is similar to salivary gland MEC with a great variety of patterns. Histological grade ranges from low to high and grading can be performed by applying the systems for both the breast and the salivary gland [\[109\]](#).

Low-grade MEC is more frequently cystic, composed of mucoid, epidermoid and basaloid cells. Mucoid cells usually line the cystic spaces and may be intermingled with columnar cells devoid of intracellular mucous. The epidermoid cells have eosinophilic or clear cytoplasm. True keratinization with squamous pearls is not seen and, when present, should direct the diagnosis toward adenosquamous carcinoma rather than MEC. Basaloid cells are usually located at the periphery of the neoplastic cysts and nests. High-grade MEC is more frequently solid and is composed of the same cell types as low-grade MEC, but with a higher degree of nuclear atypia and a higher mitotic count. Necrosis may be present. Rare cases of intermediate grade MEC have been reported [\[41\]\[109\]](#).

Immunohistochemistry assists the diagnosis as each cell type has a characteristic profile. Specifically: mucoid cells are positive for low molecular weight CKs, e.g., CK7, while high molecular weight CKs, e.g., CK14, CK5, and p63 stain the epidermoid and basaloid cells. All reported breast MECs to date have shown a triple negative phenotype.

A few cases have been studied with molecular analyses, demonstrating *CRTC1-MAML2* rearrangement, similar to that observed in the salivary gland counterpart [\[110\]\[111\]\[112\]\[113\]](#).

6.2. Clinical Correlations, Treatment and Outcome Data

Histological grading is very important for prognostic purposes [\[114\]](#). In a recent review, Ye et al. identified 42 cases of breast MEC published from 1979 when Patchefsky et al. first described this tumour type in the breast [\[115\]](#), of which 19 were classified as low grade, 3 as intermediate grade, 17 as high and 3 had no grade reported. Their table has been completed with treatment information and used to summarize the low and intermediate grade cases ([Supplementary Table S5](#)) [\[109\]\[111\]\[115\]\[116\]\[117\]\[118\]\[119\]\[120\]\[121\]\[122\]\[123\]](#). In the non-high grade MEC group, 7 patients had BCS (6 low grade and 1 with intermediate grade) and 16 had mastectomy (13 low grade and 3 intermediate grade) [\[108\]](#). Axillary nodal status was reported in 11 patients with low-grade MEC, with lymph node metastases (3/18) in only one (9.1%; 95%CI (confidence interval): 0.5–42.9%) [\[118\]](#). Of the 2 patients with intermediate MEC and known nodal status, one had a positive sentinel lymph node [\[111\]](#). Follow-up data were available in 19 patients with low-grade MEC, ranging from 3 to 156 (median 48) months, and in two patients with intermediate MEC with 3- and 8-months follow-up, respectively. At present, none of the patients with low or intermediate grade MEC has developed metastases or died. There is one report of a patient with low grade breast MEC who developed a high grade MEC recurrence [\[122\]](#) but was alive and well at 156-month follow-up. In contrast, distant metastases and progression to death occurred in 4 patients with high-grade MEC [\[108\]](#).

According to the data reported, low-grade MEC patients have a good overall prognosis, even though adjuvant chemotherapy was not administered to some of them. The review of the data, therefore, supports that adjuvant chemotherapy is not indicated in these patients simply on the basis of the triple-negative phenotype.

7. Secretory Carcinoma (SC)

7.1. Definition, Main Features, Diagnostic Clues and Differential Diagnosis

Secretory carcinoma (SC) is a rare tumour accounting for less than 0.02% of all BCs [11][124]. The name “secretory” was assigned due to the presence of eosinophilic, intra- and extracellular secretory material. The first case may have been described by Levings in 1917, but the first series and its recognition as a childhood cancer (hence the original name “juvenile breast carcinoma”) was by McDivitt and Stewart [125]. As the tumour is not restricted to the paediatric population and more commonly occurs in adults, Tavassoli and Norris re-named this tumour ‘secretory carcinoma’ in 1980 [126]. The average age at presentation is 53 years (3–87 years). It mainly occurs in females [11] but has also been reported in males, summarized by Ghilli et al. [127], in whom it may exhibit a more aggressive clinical course [128]. A similar tumour type may be seen in salivary glands (previously referred to as mammary analogue secretory carcinoma, MASC) and occasionally in the thyroid gland, skin and respiratory tract [129]. Rare cases of secretory carcinoma in situ have also been reported [130][131].

Grossly, the typical appearance is that of a rounded, circumscribed, greyish-white mass, sometimes with tan to yellow discolouration [11]. This appearance may mimic benign lesions, e.g., fibroadenoma, on imaging.

The tumour cells have vacuolated cytoplasm with abundant intracellular material that is Periodic Acid Schiff (PAS), PAS diastase or alcian blue positive. Extracellular secretory material is also seen. The nuclei of the tumour cells are small and bland. Mitotic activity is low. The architectural arrangement varies with solid, ductal/tubular, microcystic (“honeycomb”, mimicking thyroid follicles) and mixed patterns (**Figure 1F**). A case with a predominant papillary pattern has also been reported [132].

Immunohistochemically, most SCs are triple negative and may express basal markers (e.g., cytokeratin 5/6 and EGFR). Weak ER and PR positivity are observed in some tumours. SCs consistently exhibit positivity for S-100 and α -lactalbumin [11][133].

SC is characterised by a recurrent chromosomal rearrangement, t(12;15)(p13;q25), that is detected in 75–92% of tumours. This leads to fusion between E26 transformation specific translocation variant 6 (*ETV6*) and neurotrophic receptor tyrosine kinase 3 (*NTRK3*) [134] with potential for targeted therapy using larotrectinib and entrectinib. This fusion may also be detected in congenital fibrosarcoma, cellular mesoblastic nephroma, acute myeloid leukaemia, ALK-negative inflammatory myofibroblastic tumour and radiation-induced papillary thyroid carcinoma [130][135] but appears to be specific for SC in the breast context. Harrison et al. suggested the use of pan-TRK (tropomyosin receptor kinase) IHC for all TNBCs to identify *NTRK* rearrangements [136] but the specificity of this approach is questionable [137]. Additional mutations like *TERT* promoter mutations and loss of *CDKN2A/B* have also been reported and may be associated with an aggressive course in SC but further studies are needed to identify predictive markers of a more aggressive clinical course [130].

The differential diagnosis of SC includes invasive apocrine carcinoma, ACC, juvenile papillomatosis with apocrine metaplasia, microglandular adenosis and other breast lesions with secretory type features including cystic

hypersecretory hyperplasia, cystic hypersecretory in situ carcinoma, collagenous spherulosis, microglandular adenosis and lactational type change. Careful evaluation of morphology and judicious use of immunohistochemistry greatly assists accurate diagnosis. Apocrine carcinomas can generally be distinguished by their characteristic androgen receptor expression. Microglandular adenosis also expresses S100 but has specific morphological features with a typically infiltrative pattern.

7.2. Clinical Correlations, Treatment and Outcome Data

Since the description of the initial series by McDivitt and Stewart, several hundred SCs have been reported, mostly as single case reports, small single institutional series and two larger database analyses [\[138\]\[139\]](#). These are summarized in [Supplementary Table S6](#) [\[124\]\[125\]\[127\]\[129\]\[130\]\[131\]\[132\]\[138\]\[139\]\[140\]\[141\]\[142\]\[143\]\[144\]\[145\]\[146\]\[147\]\[148\]](#). In 2018, Garlick reviewed 89 cases previously published and these are included in the table on the basis of his review [\[144\]](#). The largest series, from the National Cancer Database, includes 246 SCs [\[139\]](#). Data on racial differences are limited, but one series suggested a slightly greater incidence among African American women in the USA [\[139\]](#). It is noteworthy that 64% of the tumours with known receptor status were ER-positive. Although this may cast doubt on the appropriateness of histological typing in some cases, it is consistent with the observation that, while most SCs are triple negative, it is not uncommon to see weak ER and PR expression [\[11\]](#). Several examples of SCs with a proven diagnostic translocation present have demonstrated ER positivity.

The cases and series listed in [Supplementary Table S6](#) represent only those reported in the English language literature. About half of the patients with SC were treated with mastectomy, including simple, modified radical and radical, the latter reflecting accepted practice at the time this tumour was first described. Lymph node involvement is influenced by tumour size and lymphovascular invasion, and ranged between 0 and 50% in reports, with a rate of 32% in the National Cancer Database series of 246 patients [\[138\]](#) and 15–30% in the series reported by Altundag et al. [\[148\]](#).

Patients treated with BCS were likely to receive adjuvant radiotherapy, although some did not [\[144\]](#).

Fifty-five years ago, when the first series was published, adjuvant therapy was not widely administered. Of seven patients treated with surgery alone and a median follow-up of 10 years, only one developed two consecutive local recurrences leading to mastectomy and had an uneventful course after two years of follow-up. More recent case reports and series have documented the use of adjuvant chemotherapy, but more than half of the cases summarized in [Supplementary Table S6](#) did not receive systemic treatment. De-escalating systemic treatment has also been advocated [\[143\]](#). Rare tumours may demonstrate high-grade transformation similar to translocation driven salivary gland-type carcinomas [\[129\]](#) and may benefit from targeted anti-TRK treatment [\[141\]\[149\]](#).

In general, the follow-up data suggest that SC generally pursues an indolent clinical course, even in patients with lymph node-positive disease [\[128\]\[130\]](#), with 5- and 10-year disease-free survival rates of over 90% [\[136\]](#). Breast cancer-specific survival at 5 and 10 years were 94% and 91%, respectively, in the Surveillance, Epidemiology and End Results (SEER) analysis of 83 patients [\[138\]](#). The data, therefore, support that SC does not require adjuvant systemic therapy simply because of the TNBC phenotype.

8. Tall Cell Carcinoma with Reversed Polarity (TCCRP)

8.1. Definition, Main Features, Diagnostic Clues and Differential Diagnosis

Tall cell carcinoma with reversed polarity (TCCRP) is a rare special type of BC. It is characterized by tall columnar cells arranged in nests with a predominant solid papillary pattern and demonstrates reversed nuclear polarity, i.e., the nuclei are located at the apical rather than at the basal aspect of the cells [\[11\]](#). The absence of myoepithelial cells at the periphery of the tumour nests is also an essential diagnostic criterion. An in situ component, as reported in some papers [\[150\]\[151\]\[152\]](#), does not exclude the diagnosis.

The histological appearances may vary with both structural (papillae, dense eosinophilic colloid-like material in follicle-like structures and occasional calcifications) and nuclear (grooves, pseudo-inclusions, tall columnar cells) similarities to the tall cell variant of thyroid papillary carcinoma observed; the first name of this entity was based on this resemblance (**Figure 1G**) [\[150\]](#). The neoplastic cells have eosinophilic and granular cytoplasm, rich in mitochondria. The stroma between the tumour cell nests is generally dense, and as in most solid papillary carcinomas, the nests are often surrounded by a delicate rim of capillaries. Basement membrane and smooth muscle stains on immunohistochemistry may mimic a myoepithelial cell layer or an epithelium-bound basement membrane. In keeping with a low-grade appearance, mitotic figures are rare and the Ki67 proliferation index is low.

Most TCCRPs are triple negative although some may demonstrate weak hormone receptor positivity [\[11\]](#), a phenotype that more closely resembles hormone receptor negativity than positivity [\[153\]](#). TCCRP may also express androgen receptors. Co-expression of high- and low-molecular-weight CKs is listed as a desirable diagnostic criterion in the WHO classification [\[11\]](#). However, expression of CK5 may show a mosaic-like pattern and lead to a mistaken diagnosis of a benign, hyperplastic proliferation [\[154\]](#). Unlike typical hyperplastic proliferations, however, CK14 is not expressed in TCCRP [\[152\]](#). Neuroendocrine and thyroid markers are negative in these lesions. Breast markers are expressed in TCCRP, and breast origin may be proven by the use of multiple markers e.g., GATA-3, GCDFP-15 and mammaglobin. The immunostaining with anti-mitochondrial antibody is strongly positive, especially at the basal aspect of the neoplastic cells, evidencing the reverse polarization [\[155\]](#).

The most characteristic molecular alteration in TCCRP is the presence of a p.R172 hotspot mutation in the *IDH2* gene [\[156\]\[157\]\[158\]\[159\]\[160\]](#) which is very uncommon in other breast tumours. A p.R120 mutation of the *IDH2* gene has also been described [\[161\]](#). *PIK3CA* mutations, common in many types of BC, are also frequent in TCCRP [\[157\]\[159\]](#). PRUNE2 mutations and *ATM* mutations have also been described in 6/9 and 2/3 cases, respectively [\[154\]\[158\]](#). In keeping with the IHC profile excluding a thyroid origin, *RET/PTC* rearrangements and *BRAF* mutations have not been reported in TCCRP.

Differential diagnostic problems may arise especially in core needle biopsies, when the full architecture cannot be evaluated.

The main differential diagnosis of TCCRP is metastatic thyroid papillary carcinoma, with which the morphology overlaps [\[162\]](#). The immunohistochemistry and molecular features described above, including breast marker

positivity, exclude this entity. As a triple-negative tumour of the breast, it must be separated from other triple-negative/basal-like carcinomas that are associated with aggressive behaviour, i.e., metaplastic or basal-like carcinomas which also express CK5 or CK5/6. The typical low grade and thyroid-like nuclear features, in addition to the reversed nuclear polarity, assist this distinction.

The thin vascular channels around the tumour cell nests, when highlighted by basement membrane or smooth muscle immunostains, may mimic myoepithelium and therefore an in situ or benign process [155]. The use of IHC markers that do not label vessels (e.g., p63, CD10) may help to exclude the presence of myoepithelial cells.

The low-grade nuclear pattern, the papillary growth and the CK5 staining pattern may point to a usual type hyperplastic proliferation [154] which may be excluded by the absence of CK14 staining and lack of a mosaic pattern of ER and PR expression.

8.2. Clinical Correlations, Treatment and Outcome Data

The low-grade nuclear features, infrequent mitoses and low proliferative activity all suggest a favourable prognosis. Indeed, since the first description of the entity by Eusebi et al. in 2003 [150], most reported tumours have been associated with an indolent biological course.

Zhang et al. have summarized the treatment and follow-up information of the 73 TCCRP published up to February 2021 [161]. Seventy-two tumours with documented size had a median size of 12 mm; 12 tumours were pT2 and the remainder were pT1. Lymph node status was known in 31 patients and metastases were reported in 3 (9.7%; 95%CI: 2.5–26.9%). Thirty-seven and five patients were treated with BCS and mastectomy, respectively; no data were given for the remainder.

Adjuvant treatment details were less well documented, as many reports concentrated on the histopathology and/or molecular aspects of these tumours. There were no data regarding adjuvant treatment in 42 patients. Five patients received adjuvant radiotherapy only, two received chemotherapy (one neoadjuvant) and four received both chemo- and radiotherapy. Tamoxifen was also administered to some patients with ER-positive tumours. Chemotherapy included carboplatin + paclitaxel ($n = 2$) [156], cyclophosphamide + doxorubicin + 5-fluoro-uracil ($n = 1$) [151] or was not specified further ($n = 2$) [155][161]. Neoadjuvant chemotherapy (unspecified) and trastuzumab were given to a patient for a contralateral BC which regressed, but the TCCRP showed no signs of regression [154]. No adjuvant therapy was administered in 21 cases.

Of 34 patients with reported outcome and follow-up ranging from 3 to 132 months (median 28.5 months), only 2 relapsed (5.9%; 95%CI: 1.0–21.1%). One patient (pT1c pN0(sn)) treated with BCS without adjuvant therapy) relapsed locally and regionally with 1/10 lymph nodes involved after 60 months; recurrences were surgically excised and the patient remained disease-free for a further 48 months [155]. The second patient developed bone metastases at 32 months following mastectomy and axillary dissection (pT2 pN3 M0) and sequential adjuvant chemotherapy, radiotherapy and tamoxifen [151].

In conclusion, TCCRP is usually a TNBC with generally favourable outcome, even without administration of adjuvant systemic therapy.

References

1. Hajdu, S.I. A note from history: Landmarks in history of cancer, part 1. *Cancer* 2011, 117, 1097–1102.
2. Perou, C.M.; Sørlie, T.; Eisen, M.B.; van de Rijn, M.; Jeffrey, S.S.; Rees, C.A.; Pollack, J.R.; Ross, D.T.; Johnsen, H.; Akslen, L.A.; et al. Molecular portraits of human breast tumours. *Nature* 2000, 406, 747–752.
3. Sorlie, T.; Perou, C.M.; Tibshirani, R.; Aas, T.; Geisler, S.; Johnsen, H.; Hastie, T.; Eisen, M.B.; van de Rijn, M.; Jeffrey, S.S.; et al. Gene expression patterns of breast carcinomas distinguish tumor subclasses with clinical implications. *Proc. Natl. Acad. Sci. USA* 2001, 98, 10869–10874.
4. Sorlie, T.; Tibshirani, R.; Parker, J.; Hastie, T.; Marron, J.S.; Nobel, A.; Deng, S.; Johnsen, H.; Pesich, R.; Geisler, S.; et al. Repeated observation of breast tumor subtypes in independent gene expression data sets. *Proc. Natl. Acad. Sci. USA* 2003, 100, 8418–8423.
5. Badve, S.; Dabbs, D.J.; Schnitt, S.J.; Baehner, F.L.; Decker, T.; Eusebi, V.; Fox, S.B.; Ichihara, S.; Jacquemier, J.; Lakhani, S.R.; et al. Basal-like and triple-negative breast cancers: A critical review with an emphasis on the implications for pathologists and oncologists. *Mod. Pathol.* 2011, 24, 157–167.
6. Prat, A.; Carey, L.A.; Adamo, B.; Vidal, M.; Tabernero, J.; Cortés, J.; Parker, J.S.; Perou, C.M.; Baselga, J. Molecular features and survival outcomes of the intrinsic subtypes within HER2-positive breast cancer. *J. Natl. Cancer Inst.* 2014, 106, dju152.
7. Li, X.; Yang, J.; Peng, L.; Sahin, A.A.; Huo, L.; Ward, K.C.; O'Regan, R.; Torres, M.A.; Meisel, J.L. Triple-negative breast cancer has worse overall survival and cause-specific survival than non-triple-negative breast cancer. *Breast Cancer Res. Treat.* 2017, 161, 279–287.
8. Abramson, V.G.; Lehmann, B.D.; Ballinger, T.J.; Pietersen, J.A. Subtyping of triple-negative breast cancer: Implications for therapy. *Cancer* 2015, 121, 8–16.
9. Lehmann, B.D.; Bauer, J.A.; Chen, X.; Sanders, M.E.; Chakravarthy, A.B.; Shyr, Y.; Pietersen, J.A. Identification of human triple-negative breast cancer subtypes and preclinical models for selection of targeted therapies. *J. Clin. Invest.* 2011, 121, 2750–2767.
10. Lehmann, B.D.; Jovanović, B.; Chen, X.; Estrada, M.V.; Johnson, K.N.; Shyr, Y.; Moses, H.L.; Sanders, M.E.; Pietersen, J.A. Refinement of triple-negative breast cancer molecular subtypes: Implications for neoadjuvant chemotherapy selection. *PLoS ONE* 2016, 11, e0157368.

11. WHO Classification of Tumours Editorial Board (Ed.) WHO Classification of Tumours, 5th ed.; Breast Tumours; IARC: Lyon, France, 2019.
12. Howlader, N.; Cronin, K.A.; Kurian, A.W.; Andridge, R. Differences in breast cancer survival by molecular subtypes in the United States. *Cancer Epidemiol. Biomark. Prev.* 2018, 27, 619–626.
13. Loi, S.; Drubay, D.; Adams, S.; Pruner, G.; Francis, P.A.; Lacroix-Triki, M.; Joensuu, H.; Dieci, M.V.; Badve, S.; Demaria, S.; et al. Tumor-infiltrating lymphocytes and prognosis: A pooled individual patient analysis of early-stage triple-negative breast cancers. *J. Clin. Oncol.* 2019, 37, 559–569.
14. Hortobagyi, G.; Connolly, J.L.; D’Orsi, C.J.; Edge, S.B.; Mittendorf, E.A.; Rugo, H.S.; Solin, L.J.; Weaver, D.L.; Winchester, D.J.; Giuliano, A. Breast. In *AJCC Cancer Staging Manual*, 8th ed.; Amin, M.B., Edge, S.B., Greene, F.L., Byrd, D.R., Brookland, R.K., Washington, M.K., Gershenwald, J.E., Compton, C.C., Hess, K.R., Sullivan, D.C., et al., Eds.; Springer: New York, NY, USA, 2017; pp. 587–628.
15. Sejbien, A.; Nyári, T.; Zombori, T.; Cserni, G. Comparison of nottingham prognostic index, PREDICT and PrognostILs in triple negative breast cancer—A retrospective cohort study. *Pathol. Oncol. Res.* 2020, 26, 2443–2450.
16. Roncaroli, F.; Lamovec, J.; Zidar, A.; Eusebi, V. Acinic cell-like carcinoma of the breast. *Virchows Arch.* 1996, 429, 69–74.
17. Shimao, K.; Haga, S.; Shimizu, T.; Imamura, H.; Watanabe, O.; Kinoshita, J.; Nagumo, H.; Utada, Y.; Okabe, T.; Kajiwara, T.; et al. Acinic cell adenocarcinoma arising in the breast of a young male: A clinicopathological, immunohistochemical and ultrastructural study. *Breast Cancer* 1998, 5, 77–81.
18. Damiani, S.; Pasquinelli, G.; Lamovec, J.; Peterse, J.L.; Eusebi, V. Acinic cell carcinoma of the breast: An immunohistochemical and ultrastructural study. *Virchows Arch.* 2000, 437, 74–81.
19. Schmitt, F.C.; Ribeiro, C.A.; Alvarenga, S.; Lopes, J.M. Primary acinic cell-like carcinoma of the breast—A variant with good prognosis? *Histopathology* 2000, 36, 286–289.
20. Coyne, J.D.; Dervan, P.A. Primary acinic cell carcinoma of the breast. *J. Clin. Pathol.* 2002, 55, 545–547.
21. Hirokawa, M.; Sugihara, K.; Sai, T.; Monobe, Y.; Kudo, H.; Sano, N.; Sano, T. Secretory carcinoma of the breast: A tumour analogous to salivary gland acinic cell carcinoma? *Histopathology* 2002, 40, 223–229.
22. Kahn, R.; Holtveg, H.; Nissen, F.; Holck, S. Are acinic cell carcinoma and microglandular carcinoma of the breast related lesions? *Histopathology* 2003, 42, 195–196.

23. Peintinger, F.; Leibl, S.; Reitsamer, R.; Moinfar, F. Primary acinic cell carcinoma of the breast: A case report with long-term follow-up and review of the literature. *Histopathology* 2004, 45, 645–648.
24. Tanahashi, C.; Yabuki, S.; Akamine, N.; Yatabe, Y.; Ichihara, S. Pure acinic cell carcinoma of the breast in an 80-year-old Japanese woman. *Pathol. Int.* 2007, 57, 43–46.
25. Huo, L.; Bell, D.; Qiu, H.; Sahin, A.; Wu, Y.; Sneige, N. Paneth cell-like eosinophilic cytoplasmic granules in breast carcinoma. *Ann. Diagn. Pathol.* 2011, 15, 84–92.
26. Choh, C.T.; Komar, V.; Courtney, S.P. Primary acinic cell carcinoma of the breast: A rare lesion with good prognosis. *Breast J.* 2012, 18, 610–611.
27. Sakuma, T.; Mimura, A.; Tanigawa, N.; Takamizu, R. Fine needle aspiration cytology of acinic cell carcinoma of the breast. *Cytopathology* 2013, 24, 403–405.
28. Zhao, Y.; Li, W.; Lang, R.; Yang, Y.; Gao, X.; Zheng, Y.; Zhang, C.; Fu, X.; Fu, L. Primary acinic cell carcinoma of the breast: A case report and review of the literature. *Int. J. Surg. Pathol.* 2014, 22, 177–181.
29. Shingu, K.; Ito, T.; Kaneko, G.; Itoh, N. Primary acinic cell carcinoma of the breast: A clinicopathological and immunohistochemical study. *Case Rep. Oncol. Med.* 2013, 2013, 372947.
30. Osako, T.; Takeuchi, K.; Horii, R.; Iwase, T.; Akiyama, F. Secretory carcinoma of the breast and its histopathological mimics: Value of markers for differential diagnosis. *Histopathology* 2013, 63, 509–519.
31. Winkler, N.; Morrell, G.; Factor, R.E. Invasive carcinoma with acinic cell-like features of the breast. *Breast J.* 2013, 19, 334–335.
32. Ripamonti, C.B.; Colombo, M.; Mondini, P.; Siranoush, M.; Peissel, B.; Bernard, L.; Radice, P.; Carcangiu, M.L. First description of an acinic cell carcinoma of the breast in a BRCA1 mutation carrier: A case report. *BMC Cancer* 2013, 13, 46.
33. Limite, G.; Di Micco, R.; Esposito, E.; Sollazzo, V.; Cervotti, M.; Pettinato, G.; Varone, V.; Benassai, G.; Amato, B.; Pilone, V.; et al. Acinic cell carcinoma of the breast: Review of the literature. *Int. J. Surg.* 2014, 12 (Suppl. 1), S35–S39.
34. Guerini-Rocco, E.; Hodi, Z.; Piscuoglio, S.; Ng, C.K.; Rakha, E.; Schultheis, A.M.; Marchiò, C.; da Cruz, P.A.; De Filippo, M.R.; Martelotto, L.G.; et al. The repertoire of somatic genetic alterations of acinic cell carcinomas of the breast: An exploratory, hypothesis-generating study. *J. Pathol.* 2015, 237, 166–178.
35. Conlon, N.; Sadri, N.; Corben, A.D.; Tan, L.K. Acinic cell carcinoma of breast: Morphologic and immunohistochemical review of a rare breast cancer subtype. *Hum. Pathol.* 2016, 51, 16–24.

36. Li, H.; Wang, F.; Shen, P.; Zhou, F. Pure acinic cell carcinoma of the breast. *Medicine* 2017, 96, e8866.
37. Sen, R.; Bhutani, N.; Kamboj, J.; Dahiya, S. Primary acinic cell carcinoma of the breast: A case report with clinicopathological and immunohistochemical study of a rare breast cancer. *Ann. Med. Surg.* 2018, 35, 137–140.
38. Schnitt, S.J.; Collins, L.C. *Biopsy Interpretation of the Breast*, 3rd ed.; Wolters Kluwer—Lippincott, Williams & Wilkins: Philadelphia, PA, USA, 2018.
39. Beca, F.; Lee, S.S.K.; Pareja, F.; Da Cruz, P.A.; Selenica, P.; Ferrando, L.; Gularte-Mérida, R.; Wen, H.Y.; Zhang, H.; Guerini-Rocco, E.; et al. Whole-exome sequencing and RNA sequencing analyses of acinic cell carcinomas of the breast. *Histopathology* 2019, 75, 931–937.
40. Geyer, F.C.; Berman, S.H.; Marchiò, C.; Burke, K.A.; Guerini-Rocco, E.; Piscuoglio, S.; Ng, C.K.; Pareja, F.; Wen, H.Y.; Hodi, Z.; et al. Genetic analysis of microglandular adenosis and acinic cell carcinomas of the breast provides evidence for the existence of a low-grade triple-negative breast neoplasia family. *Mod. Pathol.* 2016, 4, 529–553.
41. Foschini, M.P.; Morandi, L.; Asioli, S.; Giove, G.; Corradini, A.G.; Eusebi, V. The morphological spectrum of salivary gland type tumours of the breast. *Pathology* 2017, 49, 215–227.
42. Sardana, R.; Parwani, A.V.; Cui, X.; Balakrishna, J. Unusual cerebrospinal fluid finding of intracytoplasmic granules in metaplastic carcinoma of the breast with acinar differentiation. *Diagn. Cytopathol.* 2021, 49, E152–E155.
43. Wang, H.; Liu, F.; Gu, R.; Li, Y.; Su, F. Rare imaging appearance of adenoid cystic carcinoma of the breast: A case report. *Mol. Clin. Oncol.* 2017, 7, 473–475.
44. Boujelbene, N.; Khabir, A.; Boujelbene, N.; Jeanneret Sozzi, W.; Mirimanoff, R.O.; Khanfir, K. Clinical review-breast adenoid cystic carcinoma. *Breast* 2012, 21, 124–127.
45. Franceschini, G.; Terribile, D.; Scafetta, I.; Magno, S.; Fabbri, C.; Chiesa, F.; Di Leone, A.; Moschella, F.; Scaldaferri, A.; Fragomeni, S.; et al. Conservative treatment of a rare case of multifocal adenoid cystic carcinoma of the breast: Case report and literature review. *Med. Sci. Monit.* 2010, 16, CS33–CS39.
46. Ghabach, B.; Anderson, W.F.; Curtis, R.E.; Huycke, M.M.; Lavigne, J.A.; Dores, G.M. Adenoid cystic carcinoma of the breast in the United States (1977 to 2006): A population-based cohort study. *Breast Cancer Res.* 2010, 12, R54.
47. Brogi, E.; Hoda, S.A.; Koerner, F.C.; Rosen, P.P. (Eds.) *Rosen's Diagnosis of Breast Pathology by Needle Core Biopsy*, 4th ed.; Wolters Kluwer Health: Philadelphia, PA, USA, 2017.
48. Elimimian, E.B.; Samuel, T.A.; Liang, H.; Elson, L.; Bilani, N.; Nahleh, Z.A. Clinical and demographic factors, treatment patterns, and overall survival associated with rare triple-negative

breast carcinomas in the US. *JAMA Netw. Open* 2021, 4, e214123.

49. Kulkarni, N.; Pezzi, C.M.; Greif, J.M.; Klimberg, V.S.; Bailey, L.; Korourian, S.; Zuraek, M. Rare breast cancer: 933 adenoid cystic carcinomas from the National Cancer Data Base. *Ann. Surg. Oncol.* 2013, 20, 2236–2241.
50. Li, N.; Xu, L.; Zhao, H.; El-Naggar, A.K.; Sturgis, E.M. A comparison of the demographics, clinical features, and survival of patients with adenoid cystic carcinoma of major and minor salivary glands versus less common sites within the Surveillance, Epidemiology, and End Results registry. *Cancer* 2012, 118, 3945–3953.
51. Khanfir, K.; Kallel, A.; Villette, S.; Belkacémi, Y.; Vautravers, C.; Nguyen, T.; Miller, R.; Li, Y.X.; Taghian, A.G.; Boersma, L.; et al. Management of adenoid cystic carcinoma of the breast: A rare cancer network study. *Int. J. Rad. Oncol. Biol. Phys.* 2012, 82, 2118–2124.
52. Thompson, K.; Grabowski, J.; Saltzstein, S.L.; Sadler, G.R.; Blair, S.L. Adenoid cystic carcinoma: Is axillary staging necessary in all cases? Results from the California Cancer Registry. *Breast J.* 2011, 17, 485–489.
53. Coates, J.M.; Martinez, S.R.; Bold, R.J.; Chen, S.L. Adjuvant radiation therapy is associated with improved survival for adenoid cystic carcinoma of the breast. *J. Surg. Oncol.* 2010, 102, 342–347.
54. Arpino, G.; Clark, G.M.; Mohsin, S.; Bardou, V.J.; Elledge, R.M. Adenoid cystic carcinoma of the breast: Molecular markers, treatment and clinical outcome. *Cancer* 2002, 94, 2119–2127.
55. Treitl, D.; Radkani, P.; Rizer, M.; El Hussein, S.; Paramo, J.C.; Mesko, T.W. Adenoid cystic carcinoma of the breast, 20 years of experience in a single center with review of literature. *Breast Cancer* 2018, 25, 28–33.
56. Zhang, W.; Fang, Y.; Zhang, Z.; Wang, J. Management of adenoid cystic carcinoma of the breast: A single-institution study. *Front. Oncol.* 2021, 11, 621012.
57. Kim, M.; Lee, D.W.; Im, J.; Suh, K.J.; Keam, B.; Moon, H.G.; Im, S.A.; Han, W.; Park, I.A.; Noh, D.Y. Adenoid cystic carcinoma of the breast: A case series of six patients and literature review. *Cancer Res. Treat.* 2014, 46, 93–97.
58. Millar, B.A.; Kerba, M.; Youngson, B.; Lockwood, G.A.; Liu, F.F. The potential role of breast conservation surgery and adjuvant breast radiation for adenoid cystic carcinoma of the breast. *Breast Cancer Res. Treat.* 2004, 87, 225–232.
59. Defaud-Hénon, F.; Tunon-de-Lara, C.; Fournier, M.; Marty, M.; Velasco, V.; de Mascarel, I.; MacGrogan, G. Le carcinome adénoïde kystique du sein: Étude clinique, histologique, immunohistochimique et revue de la littérature. *Ann. Pathol.* 2010, 30, 7–16.
60. Franzese, C.; Zei, G.; Masoni, T.; Cecchini, S.; Monteleone, E.; Livi, L.; Biti, G. Adenoid cystic carcinoma of the breast. The double face of an exocrine gland carcinoma. *Strahlenther. Onkol.*

2013, 189, 1049–1050.

61. Yiğit, S.; Etit, D.; Hayrullah, L.; Atahan, M.K. Androgen receptor expression in adenoid cystic carcinoma of breast: A subset of seven cases. *Eur. J. Breast Health* 2019, 16, 44–47.
62. Marco, V.; Garcia, F.; Rubio, I.T.; Soler, T.; Ferrazza, L.; Roig, I.; Mendez, I.; Andreu, X.; Mínguez, C.G.; Tresserra, F. Adenoid cystic carcinoma and basaloid carcinoma of the breast: A clinicopathological study. *Rev. Esp. Patol.* 2021, 54, 242–249.
63. Herzberg, A.J.; Bossen, E.H.; Walther, P.J. Adenoid cystic carcinoma of the breast metastatic to the kidney. A clinically symptomatic lesion requiring surgical management. *Cancer* 1991, 68, 1015–1020.
64. Koller, M.; Ram, Z.; Findler, G.; Lipshitz, M. Brain metastasis: A rare manifestation of adenoid cystic carcinoma of the breast. *Surg. Neurol.* 1986, 26, 470–472.
65. Vranić, S.; Bilalović, N.; Lee, L.M.; Kruslin, B.; Lilleberg, S.L.; Gatalica, Z. PIK3CA and PTEN mutations in adenoid cystic carcinoma of the breast metastatic to kidney. *Hum. Pathol.* 2007, 38, 1425–1431.
66. Silva, I.; Tome, V.; Oliveira, J. Adenoid cystic carcinoma of the breast with cerebral metastatisation: A clinical novelty. *Case Rep.* 2011, 2011, bcr0820114692.
67. Mhamdi, H.A.; Kourie, H.R.; Jungels, C.; Aftimos, P.; Belbaraka, R.; Piccart-Gebhart, M. Adenoid cystic carcinoma of the breast—An aggressive presentation with pulmonary, kidney, and brain metastases: A case report. *J. Med. Case Rep.* 2017, 11, 303.
68. Slodkowska, E.; Xu, B.; Kos, Z.; Bane, A.; Barnard, M.; Zubovits, J.; Iyengar, P.; Faragalla, H.; Turbin, D.; Williams, P.; et al. Predictors of outcome in mammary adenoid cystic carcinoma: A multi-institutional study. *Am. J. Surg. Pathol.* 2020, 44, 214–223.
69. Vranic, S.; Frkovic-Grazio, S.; Lamovec, J.; Serdarevic, F.; Gurjeva, O.; Palazzo, J.; Bilalovic, N.; Lee, L.M.; Gatalica, Z. Adenoid cystic carcinomas of the breast have low Topo II α expression but frequently overexpress EGFR protein without EGFR gene amplification. *Hum. Pathol.* 2010, 41, 1617–1623.
70. Vranic, S.; Gatalica, Z.; Deng, H.; Frkovic-Grazio, S.; Lee, L.M.; Gurjeva, O.; Wang, Z.Y. ER- α 36, a novel isoform of ER- α 66, is commonly over-expressed in apocrine and adenoid cystic carcinomas of the breast. *J. Clin. Pathol.* 2011, 64, 54–57.
71. Kim, J.; Geyer, F.C.; Martelotto, L.G.; Ng, C.K.; Lim, R.S.; Selenica, P.; Li, A.; Pareja, F.; Fusco, N.; Edelweiss, M.; et al. MYBL1 rearrangements and MYB amplification in breast adenoid cystic carcinomas lacking the MYB-NFIB fusion gene. *J. Pathol.* 2018, 244, 143–150.
72. Martelotto, L.G.; De Filippo, M.R.; Ng, C.K.; Natrajan, R.; Fuhrmann, L.; Cyrta, J.; Piscuoglio, S.; Wen, H.C.; Lim, R.S.; Shen, R.; et al. Genomic landscape of adenoid cystic carcinoma of the

- breast. *J. Pathol.* 2015, 237, 179–189.
73. Stolnicu, S.; Alvarado-Cabrero, I. (Eds.) *Practical Atlas of Breast Pathology*; Springer International Publishing, Springer Nature: Cham, Switzerland, 2018; pp. 263–292.
 74. Lakhani, S.R.; Ellis, I.O.; Schnitt, S.J.; Tan, P.H.; van de Vijver, M.J. (Eds.) *WHO Classification of Tumours of the Breast*, 4th ed.; International Agency for Research of Cancer: Lyon, France, 2012.
 75. Gobbi, H.; Simpson, J.F.; Borowsky, A.; Jensen, R.A.; Page, D.L. Metaplastic breast tumors with a dominant fibromatosis-like phenotype have a high risk of local recurrence. *Cancer* 1995, 85, 2170–2182.
 76. Sneige, N.; Yaziji, H.; Mandavilli, S.R.; Perez, E.R.; Ordonez, N.G.; Gown, A.M.; Ayala, A. Low-grade (fibromatosis-like) spindle cell carcinoma of the breast. *Am. J. Surg. Pathol.* 2001, 25, 1009–1016.
 77. Schafernak, K.T.; Policarpio-Nicolas, M.L.; Wiley, E.L.; Laskin, W.B.; Diaz, L.K. A 59-year-old woman with a spindle cell lesion of the breast. Low-grade (fibromatosis-like) spindle cell carcinoma of the breast. *Arch. Pathol. Lab. Med.* 2006, 130, e81–e83.
 78. Rekhi, B.; Shet, T.M.; Badwe, R.A.; Chinoy, R.F. Fibromatosis-like carcinoma-an unusual phenotype of a metaplastic breast tumor associated with a micropapilloma. *World J. Surg. Oncol.* 2007, 5, 24.
 79. Podetta, M.; D'Ambrosio, G.; Ferrari, A.; Sgarella, A.; Dal Bello, B.; Fossati, G.S.; Zonta, S.; Silini, E.; Dionigi, P. Low-grade fibromatosis-like spindle cell metaplastic carcinoma: A basal-like tumor with a favorable clinical outcome. Report of two cases. *Tumori* 2009, 95, 264–267.
 80. Lamovec, J.; Gasljevic, G. Keloid type of fibromatosis-like metaplastic carcinoma of the breast with transformation into biphasic tumour in recurrences and lymph node metastases. *Histopathology* 2010, 57, 318–320.
 81. Nonnis, R.; Paliogiannis, P.; Giangrande, D.; Marras, V.; Trignano, M. Low-grade fibromatosis-like spindle cell metaplastic carcinoma of the breast: A case report and literature review. *Clin. Breast Cancer* 2012, 12, 147–150.
 82. Rito, M.; Schmitt, F.; Pinto, A.E.; Andre, S. Fibromatosis-like metaplastic carcinoma of the breast has a claudin-low immunohistochemical phenotype. *Virchows Arch.* 2014, 465, 185–191.
 83. Takano, E.A.; Hunter, S.M.; Campbell, I.G.; Fox, S.B. Low-grade fibromatosis-like spindle cell carcinomas of the breast are molecularly exiguous. *J. Clin. Pathol.* 2015, 68, 362–367.
 84. Pinilla Pagnon, I.; Pérez Mies, B.; Tulio Martinez, M.; Peña Jaimes, L.; Roldan Cabanillas, A.M.; Romio de las Heras, E.; Blázquez Ortiz, J.M.; Sánchez Monforte, J.; Delgado Moya, M.Á.; Rubio Marín, D. Metaplastic breast carcinoma “fibromatosis like”, associated with intraductal papilloma. Case report and literature review. *Hum. Pathol. Case Rep.* 2017, 9, 15–18.

85. Zhu, H.; Li, K.; Dong, D.D.; Fu, J.; Liu, D.D.; Wang, L.; Xu, G.; Song, L.H. Spindle cell metaplastic carcinoma of breast: A clinicopathological and immunohistochemical analysis. *Asia Pac. J. Clin. Oncol.* 2017, 13, e72–e78.
86. Zhao, Y.; Gong, X.; Li, N.; Zhu, B.; Yu, D.; Jin, X. Fibromatosis-like metaplastic carcinoma of breast: A challenge for clinicopathologic diagnosis. *Int. J. Clin. Exp. Pathol.* 2018, 11, 3691–3696.
87. Victoor, J.; Bourgain, C.; Vander Borght, S.; Vanden Bempt, I.; De Rop, C.; Floris, G. Fibromatosis-like metaplastic carcinoma: A case report and review of the literature. *Diagn. Pathol.* 2020, 15, 20.
88. Takatsuka, D.; Ogura, H.; Asano, Y.; Nakamura, A.; Koizumi, K.; Shiiya, N.; Baba, S. A difficult-to-diagnose fibromatosis-like metaplastic carcinoma of the breast: A case report. *Surg. Case Rep.* 2021, 7, 16.
89. Dwyer, J.B.; Clark, B.Z. Low-grade fibromatosis-like spindle cell carcinoma of the breast. *Arch. Pathol. Lab. Med.* 2015, 139, 552–557.
90. Gobbi, H.; Simpson, J.F.; Jensen, R.A.; Olson, S.J.; Page, D.L. Metaplastic spindle cell breast tumors arising within papillomas, complex sclerosing lesions, and nipple adenomas. *Mod. Pathol.* 2003, 16, 893–901.
91. Rakha, E.A.; Coimbra, N.D.M.; Hodi, Z.; Juneinah, E.; Ellis, I.O.; Lee, A.H.S. Immunoprofile of metaplastic carcinomas of the breast. *Histopathology* 2017, 70, 975–985.
92. Norkowski, E.; Masliah-Plancho, J.; Le Guellec, S.; Trassard, M.; Courrèges, J.B.; Charron-Barra, C.; Terrier, P.; Bonvalot, S.; Coindre, J.M.; Laé, M. Lower rate of CTNNB1 mutations and higher rate of APC mutations in desmoid fibromatosis of the breast: A series of 134 tumors. *Am. J. Surg. Pathol.* 2020, 44, 1266–1273.
93. Bataillon, G.; Fuhrmann, L.; Girard, E.; Menet, E.; Laé, M.; Capovilla, M.; Treilleux, I.; Arnould, L.; Penault-Llorca, F.; Rouzier, R.; et al. High rate of PIK3CA mutations but no TP53 mutations in low-grade adenosquamous carcinoma of the breast. *Histopathology* 2018, 73, 273–283.
94. Rosen, P.P.; Ernsberger, D. Low-grade adenosquamous carcinoma. A variant of metaplastic mammary carcinoma. *Am. J. Surg. Pathol.* 1987, 11, 351–358.
95. Yang, G.Z.; Liang, S.H.; Shi, X.H. A novel collision tumour of myofibroblastoma and low-grade adenosquamous carcinoma in breast. *Diagn. Pathol.* 2020, 15, 76.
96. Cheng, E.; D'Alfonso, T.M.; Arafah, M.; Marrero Rolon, R.; Ginter, P.S.; Hoda, S.A. Subareolar sclerosing ductal hyperplasia. *Int. J. Surg. Pathol.* 2017, 25, 4–11.
97. Ohashi, R.; Sangen, M.; Namimatsu, S.; Takei, H.; Naito, Z. IMP3 contributes to poor prognosis of patients with metaplastic breast carcinoma: A clinicopathological study. *Ann. Diagn. Pathol.* 2017, 31, 30–35.

98. Senger, J.L.; Meiers, P.; Kanthan, R. Bilateral synchronous low-grade adenosquamous carcinoma of the breast: A Case report with review of the current literature. *Int. J. Surg. Case Rep.* 2015, 14, 53–57.
99. Tan, Q.T.; Chuwa, E.W.; Chew, S.H.; Lim-Tan, S.K.; Lim, S.H. Low-grade adenosquamous carcinoma of the breast: A diagnostic and clinical challenge. *Int. J. Surg.* 2015, 19, 22–26.
100. Bataillon, G.; Collet, J.F.; Voillemot, N.; Menet, E.; Vincent-Salomon, A.; Klijanienko, J. Fine-needle aspiration of low-grade adenosquamous carcinomas of the breast: A report of three new cases. *Acta Cytol.* 2014, 58, 427–431.
101. Scali, E.P.; Ali, R.H.; Hayes, M.; Tyldesley, S.; Hassell, P. Low-grade adenosquamous rare disease. *Can. Assoc. Radiol. J.* 2013, 64, 339–344.
102. Agrawal, A.; Saha, S.; Ellis, I.O.; Bello, A.M. Adenosquamous carcinoma of breast in a 19 years old woman: A case report. *World J. Surg. Oncol.* 2010, 8, 44.
103. Ho, B.C.-S.; Tan, H.W.; Lee, V.K.-M.; Tan, P.H. Preoperative and intraoperative diagnosis of low-grade adenosquamous carcinoma of the breast: Potential diagnostic pitfalls. *Histopathology* 2006, 49, 603–611.
104. Ferrara, G.; Nappi, O.; Wick, M.R. Fine-needle aspiration cytology and immunohistology of low-grade adenosquamous carcinoma of the breast. *Diagn. Cytopathol.* 1999, 20, 13–18.
105. Shizawa, S.; Sasano, H.; Suzuki, T.; Ishii, H.; Takeda, T.; Nagura, H. Low-grade adenosquamous carcinoma of the breast: A case report with cytologic findings and review of the literature. *Pathol. Int.* 1997, 47, 264–267.
106. Foschini, M.P.; Pizzicannella, G.; Peterse, J.L.; Eusebi, V. Adenomyoepithelioma of the breast associated with low-grade adenosquamous and sarcomatoid carcinomas. *Virchows Arch.* 1995, 427, 243–250.
107. Van Hoesen, K.H.; Drudis, T.; Cranor, M.L.; Erlandson, R.A.; Rosen, P.P. Low-grade adenosquamous carcinoma of the breast—a clinicopathologic study of 32 cases with ultrastructural analysis. *Am. J. Surg. Pathol.* 1993, 17, 248–258.
108. Ye, R.P.; Liao, Y.H.; Xia, T.; Kuang, R.; Long, H.A.; Xiao, X.L. Breast mucoepidermoid carcinoma: A case report and review of literature. *Int. J. Clin. Exp. Pathol.* 2020, 13, 3192–3199.
109. Di Tommaso, L.; Foschini, M.P.; Ragazzini, T.; Magrini, E.; Fornelli, A.; Ellis, I.O.; Eusebi, V. Mucoepidermoid carcinoma of the breast. *Virchows Arch.* 2004, 444, 13–19.
110. Bean, G.R.; Krings, G.; Otis, C.N.; Solomon, D.A.; García, J.J.; van Zante, A.; Camelo-Piragua, S.; van Ziffle, J.; Chen, Y.Y. CRTC1-MAML2 fusion in mucoepidermoid carcinoma of the breast. *Histopathology* 2019, 74, 463–473.

111. Camelo-Piragua, S.I.; Habib, C.; Kanumuri, P.; Lago, C.E.; Mason, H.S.; Otis, C.N. Mucoepidermoid carcinoma of the breast shares cytogenetic abnormality with mucoepidermoid carcinoma of the salivary gland: A case report with molecular analysis and review of the literature. *Hum. Pathol.* 2009, 40, 887–892.
112. Pareja, F.; Weigelt, B.; Reis-Filho, J.S. Problematic breast tumors reassessed in light of novel molecular data. *Mod. Pathol.* 2021, 34 (Suppl. 1), 38–47.
113. Yan, M.; Gilmore, H.; Harbhajanka, A. Mucoepidermoid carcinoma of the breast with MAML2 rearrangement: A case report and literature review. *Int. J. Surg. Pathol.* 2020, 28, 787–792.
114. Basbug, M.; Akbulut, S.; Arikanoglu, Z.; Sogutcu, N.; Firat, U.; Kucukoner, M. Mucoepidermoid carcinoma in a breast affected by burn scars: Comprehensive literature review and case report. *Breast Care* 2011, 6, 293–297.
115. Patchefsky, A.S.; Frauenhoffer, C.M.; Krall, R.A.; Cooper, H.S. Low-grade mucoepidermoid carcinoma of the breast. *Arch. Pathol. Lab. Med.* 1979, 103, 196–198.
116. Burghel, G.J.; Abu-Dayyeh, I.; Babouq, N.; Wallace, A.; Abdelnour, A. Mutational screen of a panel of tumor genes in a case report of mucoepidermoid carcinoma of the breast from Jordan. *Breast J.* 2018, 24, 1102–1104.
117. Sherwell-Cabello, S.; Maffuz-Aziz, A.; Rios-Luna, N.P.; Pozo-Romero, M.; Lopez-Jimenez, P.V.; Rodriguez-Cuevas, S. Primary mucoepidermoid carcinoma of the breast. *Breast J.* 2017, 23, 753–755.
118. Cheng, M.; Geng, C.; Tang, T.; Song, Z. Mucoepidermoid carcinoma of the breast: Four case reports and review of the literature. *Medicine* 2017, 96, e9385.
119. Fujino, M.; Mori, D.; Akashi, M.; Yamamoto, H.; Aibe, H.; Matake, K.; Shirahane, K. Mucoepidermoid carcinoma of the breast found during treatment of lymphoma. *Case Rep. Oncol.* 2016, 9, 806–814.
120. Hornychova, H.; Ryska, A.; Betlach, J.; Bohac, R.; Cizek, T.; Tomsova, M.; Obermannova, R. Mucoepidermoid carcinoma of the breast. *Neoplasma* 2007, 54, 168–172.
121. Horii, R.; Akiyama, F.; Ikenaga, M.; Iwase, T.; Sakamoto, G. Muco-epidermoid carcinoma of the breast. *Pathol. Int.* 2006, 56, 549–553.
122. Tjalma, W.A.; Verslegers, I.O.; De Loecker, P.A.; Van Marck, E.A. Low and high grade mucoepidermoid carcinomas of the breast. *Eur. J. Gynaecol. Oncol.* 2002, 23, 423–425.
123. Fisher, E.R.; Palekar, A.S.; Gregorio, R.M.; Paulson, J.D. Mucoepidermoid and squamous cell carcinomas of breast with reference to squamous metaplasia and giant cell tumors. *Am. J. Surg. Pathol.* 1983, 7, 15–27.

124. Li, L.J.; Wu, N.; Li, F.X.; Li, L.M.; Wei, L.J.; Liu, J.T. Clinicopathologic and molecular characteristics of 44 patients with pure secretory breast carcinoma. *Cancer Biol. Med.* 2019, 16, 139–146.
125. McDivitt, R.W.; Stewart, F.W. Breast carcinoma in children. *JAMA* 1966, 195, 388–390.
126. Tavassoli, F.A.; Norris, H.J. Secretory carcinoma of the breast. *Cancer* 1980, 45, 2404–2413.
127. Ghilli, M.; Mariniello, M.D.; Scatena, C.; Dosa, L.; Traficante, G.; Tamburini, A.; Caporalini, C.; Buccoliero, A.M.; Facchini, F.; Colizzi, L.; et al. Male secretory breast cancer: Case in a 6-year-old boy with a peculiar gene duplication and review of the literature. *Breast Cancer Res. Treat.* 2018, 170, 445–454.
128. Li, D.; Xiao, X.; Yang, W.; Shui, R.; Tu, X.; Lu, H.; Shi, D. Secretory breast carcinoma: A clinicopathological and immunophenotypic study of 15 cases with a review of the literature. *Mod. Pathol.* 2012, 25, 567–575.
129. Xu, J.; Weisman, P. Dedifferentiated secretory breast carcinoma with fibrosarcomatous features harboring an ETV6-NTRK3 fusion in both components. *Genes Chromosomes Cancer* 2021, 60, 447–451.
130. Hoda, R.S.; Brogi, E.; Pareja, F.; Nanjangud, G.; Murray, M.P.; Weigelt, B.; Reis-Filho, J.S.; Wen, H.Y. Secretory carcinoma of the breast: Clinicopathologic profile of 14 cases emphasising distant metastatic potential. *Histopathology* 2019, 75, 213–224.
131. Yang, Y.; Wang, Z.Y.; Pan, G.Q.; Li, S.M.; Wu, Y.Y.; Liu, L. Pure secretory carcinoma in situ: A case report and literature review. *Diagn. Pathol.* 2019, 14, 95.
132. Shui, R.; Cheng, Y.; Bai, Q.; Yang, W. Secretory breast carcinoma with a papillary-predominant pattern: An unusual morphological variant. *Histopathology* 2017, 71, 488–493.
133. Cimino-Mathews, A. Novel uses of immunohistochemistry in breast pathology: Interpretation and pitfalls. *Mod. Pathol.* 2021, 34 (Suppl. 1), 62–77.
134. Stenzinger, A.; van Tilburg, C.M.; Tabatabai, G.; Länger, F.; Graf, N.; Griesinger, F.; Heukamp, L.C.; Hummel, M.; Klingebiel, T.; Hettmer, S.; et al. Diagnosis and therapy of tumors with NTRK gene fusion. *Pathologe* 2021, 42, 103–115.
135. Toll, A.; Joneja, U.; Palazzo, J. Pathologic spectrum of secretory and mucinous breast lesions. *Arch. Pathol. Lab. Med.* 2016, 140, 644–650.
136. Harrison, B.T.; Fowler, E.; Krings, G.; Chen, Y.Y.; Bean, G.R.; Vincent-Salomon, A.; Fuhrmann, L.; Barnick, S.E.; Chen, B.; Hosfield, E.M.; et al. Pan-TRK immunohistochemistry: A useful diagnostic adjunct for secretory carcinoma of the breast. *Am. J. Surg. Pathol.* 2019, 43, 1693–1700.
137. Zaborowski, M.; Gill, A.J. Is secretory breast carcinoma underdiagnosed? In the era of targeted therapy should there be a low threshold to screen for NTRK immunohistochemistry in triple

negative breast cancers? *Pathology* 2019, 51, 653–655.

138. Horowitz, D.P.; Sharma, C.S.; Connolly, E.; Gidea-Addeo, D.; Deutsch, I. Secretory carcinoma of the breast: Results from the survival, epidemiology and end results database. *Breast* 2012, 21, 350–353.
139. Jacob, J.D.; Hodge, C.; Franko, J.; Pezzi, C.M.; Goldman, C.D.; Klimberg, V.S. Rare breast cancer: 246 invasive secretory carcinomas from the National Cancer Data Base. *J. Surg. Oncol.* 2016, 113, 721–723.
140. Lee, S.G.; Jung, S.P.; Lee, H.Y.; Kim, S.; Kim, H.Y.; Kim, I.; Bae, J.W. Secretory breast carcinoma: A report of three cases and a review of the literature. *Oncol. Lett.* 2014, 8, 683–686.
141. Shukla, N.; Roberts, S.S.; Baki, M.O.; Mushtaq, Q.; Goss, P.E.; Park, B.H.; Gundem, G.; Tian, K.; Geiger, H.; Redfield, K.; et al. Successful targeted therapy of refractory pediatric ETV6-NTRK3 fusion-positive secretory breast carcinoma. *JCO Precis. Oncol.* 2017, 2017, PO.17.00034.
142. Sheshe, A.A.; Imam, M.I. Secretory carcinoma of the breast in a 20-year-old male: Case report and review of literature. *Niger. J. Surg.* 2018, 24, 135–137.
143. Benabu, J.-C.; Stoll, F.; Koch, A.; Moliere, S.; Bellocq, J.-P.; Mathelin, C. De-escalating systemic therapy in triple negative breast cancer: The example of secretory carcinoma. *J. Obstet. Hum. Reprod.* 2018, 47, 163–165.
144. Garlick, J.W.; Olson, K.A.; Downs-Kelly, E.; Bucher, B.T.; Matsen, C.B. Secretory breast carcinoma in an 8-year-old girl: A case report and literature review. *Breast J.* 2018, 24, 1055–1061.
145. Pohlodek, K.; Meciarova, I.; Grossmann, P.; Martinek, P.; Kinkor, Z. Secretory carcinoma of the breast: A case report. *Int. J. Surg. Case Rep.* 2019, 56, 74–77.
146. Novochadlo Klüppel, E.; Rodrigues da Costa, L.; Marquette Tognolo, C.; do Nascimento, A.; Grignet Ribeiro, M.; Girardi Fachin, C. Secretory breast carcinoma in a male child: Case report and literature review. *Int. J. Surg. Case Rep.* 2020, 73, 310–314.
147. Gohara, T.; Komura, M.; Asano, A.; Emura, T.; Obana, K.; Kikuchi, T.; Yonekawa, H.; Komuro, H.; Kodaka, T.; Terawaki, K.; et al. A case of secretory breast cancer in a 6 year-old girl: Is it possible to make a correct preoperative diagnosis? *Breast Cancer* 2020, 27, 785–790.
148. Altundag, K. Secretory carcinoma of the breast in postmenopausal women. *J. BUON* 2020, 25, 1266.
149. Solomon, J.P.; Benayed, R.; Hechtman, J.F.; Ladanyi, M. Identifying patients with NTRK fusion cancer. *Ann. Oncol.* 2019, 30 (Suppl. 8), viii16–viii22.
150. Eusebi, V.; Damiani, S.; Ellis, I.O.; Azzopardi, J.G.; Rosai, J. Breast tumor resembling the tall cell variant of papillary thyroid carcinoma: Report of 5 cases. *Am. J. Surg. Pathol.* 2003, 27, 1114–

1118.

151. Cameselle-Teijeiro, J.; Abdulkader, I.; Barreiro-Morandeira, F.; Ruiz-Ponte, C.; Reyes-Santías, R.; Chavez, E.; Sobrinho-Simões, M. Breast tumor resembling the tall cell variant of papillary thyroid carcinoma: A case report. *Int. J. Surg. Pathol.* 2006, 14, 79–84.
152. Tosi, A.L.; Ragazzi, M.; Asioli, S.; Del Vecchio, M.; Cavalieri, M.; Eusebi, L.H.; Foschini, M.P. Breast tumor resembling the tall cell variant of papillary thyroid carcinoma: Report of 4 cases with evidence of malignant potential. *Int. J. Surg. Pathol.* 2007, 15, 14–19.
153. Villegas, S.L.; Nekljudova, V.; Pfarr, N.; Engel, J.; Untch, M.; Schrodi, S.; Holms, F.; Ulmer, H.U.; Fasching, P.A.; Weber, K.E.; et al. Therapy response and prognosis of patients with early breast cancer with low positivity for hormone receptors—An analysis of 2765 patients from neoadjuvant clinical trials. *Eur. J. Cancer* 2021, 148, 159–170.
154. Bhargava, R.; Florea, A.V.; Pelmus, M.; Jones, M.W.; Bonaventura, M.; Wald, A.; Nikiforova, M. Breast tumor resembling tall cell variant of papillary thyroid carcinoma: A solid papillary neoplasm with characteristic immunohistochemical profile and few recurrent mutations. *Am. J. Clin. Pathol.* 2017, 147, 399–410.
155. Foschini, M.P.; Asioli, S.; Foreid, S.; Cserni, G.; Ellis, I.O.; Eusebi, V.; Rosai, J. Solid papillary breast carcinomas resembling the tall cell variant of papillary thyroid neoplasms: A unique invasive tumor with indolent behavior. *Am. J. Surg. Pathol.* 2017, 41, 887–895.
156. Chiang, S.; Weigelt, B.; Wen, H.C.; Pareja, F.; Raghavendra, A.; Luciano, G.; Martelotto, L.G.; Burke, K.A.; Basili, T.; Li, A.Q.; et al. IDH2 mutations define a unique subtype of breast cancer with altered nuclear polarity. *Cancer Res.* 2016, 76, 7118–7129.
157. Lozada, J.R.; Basili, T.; Pareja, F.; Alemar, B.; Paula, A.D.C.; Gularte-Merida, R.; Giri, D.D.; Querzoli, P.; Cserni, G.; Rakha, E.A.; et al. Solid papillary breast carcinomas resembling the tall cell variant of papillary thyroid neoplasms (solid papillary carcinomas with reverse polarity) harbour recurrent mutations affecting IDH2 and PIK3CA: A validation cohort. *Histopathology* 2018, 73, 339–344.
158. Alsadoun, N.; MacGrogan, G.; Truntzer, C.; Lacroix-Triki, M.; Bedgedjian, I.; Koeb, M.H.; El Alam, E.; Medioni, D.; Parent, M.; Wuither, P.; et al. Solid papillary carcinoma with reverse polarity of the breast harbors specific morphologic, immunohistochemical and molecular profile in comparison with other benign or malignant papillary lesions of the breast: A comparative study of 9 additional cases. *Mod. Pathol.* 2018, 31, 1367–1380.
159. Zhong, E.; Scognamiglio, T.; D'Alfonso, T.; Song, W.; Tran, H.; Baek, I.; Hoda, S.A. Breast tumor resembling the tall cell variant of papillary thyroid carcinoma: Molecular characterization by next-generation sequencing and histopathological comparison with tall cell papillary carcinoma of thyroid. *Int. J. Surg. Pathol.* 2019, 27, 134–141.

160. Pareja, F.; da Silva, E.M.; Frosina, D.; Geyer, F.C.; Lozada, J.R.; Basili, T.; Da Cruz, P.A.; Zhong, E.; Derakhshan, F.; D'Alfonso, T.; et al. Immunohistochemical analysis of IDH2 R172 hotspot mutations in breast papillary neoplasms: Applications in the diagnosis of tall cell carcinoma with reverse polarity. *Mod. Pathol.* 2020, 33, 1056–1064.
161. Zhang, X.; Wu, H.; Wang, Z.; Zhou, Y.; Mao, F.; Lin, Y.; Shen, S.; Liang, Z.; Sun, Q. Tall cell carcinoma of the breast with reverse polarity: Case report with gene sequencing and literature review. *Gland Surg.* 2021, 10, 837–843.
162. Fiche, M.; Cassagnau, E.; Aillet, G.; Bailly, J.; Chupin, M.; Classe, J.M.; Bodic, M.F. Métastase mammaire d'un carcinome papillaire à cellules hautes de la thyroïde. . *Ann. Pathol.* 1998, 18, 130–132.

Retrieved from <https://encyclopedia.pub/entry/history/show/38856>