Communication in the cancer microenvironment

Subjects: Oncology Contributor: Agnieszka Dominiak

The tumor microenvironment (TME) is a complex system composed of multiple cells, such as non-cancerous fibroblasts, adipocytes, immune and vascular cells, as well as signal molecules and mediators. Tumor cells recruit and reprogram other cells to produce factors that maintain tumor growth. Communication between cancerous and surrounding cells is a two-way process and engages a diverse range of mechanisms that, in consequence, can lead to rapid proliferation, metastasis, and drug resistance, or can serve as a tumors-suppressor, e.g., through tumor–immune cell interaction. Cross-talk within the cancer microenvironment can be direct by cell-to-cell contact via adhesion molecules, electrical coupling, and passage through gap junctions, or indirect through classical paracrine signaling by cytokines, growth factors, and extracellular vesicles. Therapeutic approaches for modulation of cell-cell communication may be a promising strategy to combat tumors. In particular, integrative approaches targeting tumor communication in combination with conventional chemotherapy seem reasonable. Currently, special attention is paid to suppressing the formation of openended channels as well as blocking exosome production or ablating their cargos. However, many aspects of cell-to-cell communication have yet to be clarified, and, in particular, more work is needed in regard to mechanisms of bidirectional signal transfer. Finally, it seems that some interactions in TEM can be not only cancer-specific, but also patient-specific, and their recognition would help to predict patient response to therapy.

Keywords: tumor microenvironment ; communication in cancer ; therapeutic target ; oncology therapy

1. Introduction

Despite many efforts, cancer is one of the main causes of human deaths. According to the World Health Organization, it was responsible for approximately 9.6 million deaths in 2018. It is generally accepted that the fight against cancer must be multidirectional and involve the development of new strategies for preventive action, early diagnosis, and treatment to enhance effectiveness and precision of cancer therapy, increase patients survivability, and improve their quality of life ^{[1][2]} ^[3]. However, current standards therapy often overlooked the assumption that cancer is an "ensemble production". Apart from malignant cells, there are lots of supporting players, including fibroblasts, pericytes, endothelial cells, adipocytes, bone-marrow-derived mesenchymal stem cells, and immune cells. Each of these stromal cell types plays a role in tumor proliferation, metastasis, and treatment failure ^{[4][5]}. The extracellular matrix (ECM) is a highly dynamic structure that surrounds the above-mentioned cells and affects their proliferation and cell–cell communication via the transmission of mechanical signals and cell adhesion ^[G]. ECM constituents mainly derive from the tumor cells themselves but also, to a large degree, from cancer-associated fibroblasts (CAF). High amounts of metalloproteinases in the cancer niche process ECM components and are involved in ECM remodeling, resulting in the release of various signaling molecules with both pro- and anti-tumor activities ^[Z].

Cell communication is required for proper cellular activities or movements, and both failure and excess of this cross-talk can lead to tissue pathology. Normal and cancerous cells dynamically transmit reciprocal information, and, by contacting the tumor stromal cells, acquire a pro-tumoral phenotype that can promote cancer progression. Cells in this microenvironment are also involved in tumor suppression, and, for example, the accumulation of cytotoxic CD8+T cells and Th1 cells in tumor stroma suggests that the immune system fights against cancer. However, some immune cells, such as tumor-associated macrophages, can promote cancer development, indicating that immune cells have a multifaceted role ^[8]. Thus, increasing attention is being paid to fully understand the mechanism of interaction between cancer and the surrounding cells.

Currently, many studies have documented that the vital role in tumor progression plays on a complex system of intercellular communication via direct cell-to-cell contact or through classical paracrine/endocrine signaling. The most common type of signal transition to neighboring or long-distance cells is the secretion of soluble factors into the extracellular space, like cytokines, chemokines, and growth factors. Another way of cell interaction is through adhesion molecules and gap junctions. Recent research has also highlighted that non-cancer cells can donate healthy mitochondria and other organelles by tunnel nanotubes to keep cancer cells alive, but it was also reported that horizontal mitochondrial

transfer is possible from cancer cells to surrounding cells (e.g., from cancer to stromal cells) ^{[9][10][11]}. An important way of cells to cross-talk is membrane vesicle secretion that does not need specific receptors to reach target cells. Moreover, cancerous cells create a hypoxic and acidic microenvironment. Reduction of the pH (ranging between 6.0 and 6.5) can impact surrounding cells and repress their antitumor activity ^[12]. Hypoxia can support cancer growth through the differentiation of fibroblasts into CAFs ^[13]. However, the main mechanism of fibroblast activation is a cross-talk involving Notch and JAK1/STAT3 signaling pathways, and another way is by a range of inflammatory signaling molecules, for instance, IL-1 acting via NF-kB and IL-6 acting through signal STAT transcription factors. Similarly, transforming growth factor β (TGF β) family ligands and lysophosphatidic acid are also involved in activating signals for fibroblasts, while cytokine, a leukemia inhibitory factor (LIF), is known as a sustainer of their invasive phenotype ^{[14][15][16][17]}.

2. Mechanisms of Cellular Communication

During evolution, many modes of cell-to-cell communication have developed. These appearing forms of intercellular communication turn out to be essential for the maintenance of physiological cell functions and cellular response to external stimuli. Due to the fundamental role of cell cross-talk, different cell-to-cell signaling mechanisms parallelly exist. One way of cell signaling is direct intercellular communication, which includes direct cell-to-cell contact by electrical coupling, direct passage through gap junctions, direct receptor–ligand binding of membrane proteins, or a mechanism that relies on nanotubular cell-to-cell connections termed "membrane nanotubes" (MNTs; Figure 1). Cells also communicate with each other indirectly via signal molecules and mediators, as well as transferring biological information by extracellular vesicles (Figure 2).



DIRECT COMMUNICATION

Figure 1. The schematic visualization of direct communication in the tumor microenvironment. The sharing of information between cancer and stromal cells, such as non-cancerous epithelial cells, cancer-associated fibroblasts, immune cells, and others, can occur in different ways, e.g., via gap junction, open-ended cytoplasmic channels (tunnel nanotubes), or through direct ligand–receptor signaling and cell adhesion ^[18]. These dynamic interplays effectively enable the exchange of cellular cargos over short or long (tunnel nanotubes) distances. For instance, via tunnel nanotubes, mitochondria can be donated from one cell to another in two directions. Gap junctions joint the same and different types of cells and consist of connexions that include six connexins. They are able to transfer rather small molecules and metabolites ^[19]. Similarly, physical cell–cell attachments via adhesion proteins are impermeable for macromolecules.

Figure 2. The schematic visualization of indirect communication in the tumor microenvironment. The sharing of information between cancer and stromal cells, such as non-cancerous epithelial cells, cancer-associated fibroblasts, immune cells, and others, can occur in different ways, e.g., via extracellular vesicles or through signaling by cytokines, chemokines, and growth factors as well as metabolites-mediated communication. Exosomes can transfer many bioactive molecules over long and very long distances. Secreted signaling molecules can also travel to remote cells.

2.1. Direct Intercellular Communication

2.1.1. Gap Junctions

Gap junctions (GJs) are major contributors to the maintenance of cell homeostasis. When the cells are a short distance apart, they can be connected with each other using gap junctions, which allow various molecules, ions, and electrical impulses to pass directly between cells. The gap junction channel is composed of two connexons (hemichannels), which connect cells across the intercellular space. The connexons are formed by hexameric oligomers of transmembrane proteins, the connexins (Cxs) ^[20]. Approximately 21 Cxs are identified in humans. Most cell types express multiple connexin isoforms, which allow heteromeric hemichannels and heterotypic gap junctions to form, which could possibly provide a structural basis for the selectivity of GJs and may have overlapping or distinct functions ^[21]. The Cx43, being more widespread than other Cxs, is expressed mainly in a few tissues only. The regulation of GJs can be modulated by connexin-associated proteins, including regulatory phosphatases, cytoskeletal elements, and enzymes. Key regulatory pathways act through phosphorylation/dephosphorylation of Cxs, mainly at the C-terminal domain. Phosphorylation determines disassembly and internalization of the GJ, due to the disengagement of Cxs from the GJ, whereas unphosphorylated proteins remain in GJ ^[22]. Already in the 1970s, scientists have observed a loss of functional GJs in cancer cells and considered that GJs were involved in carcinogenesis ^[23]. A number of biological and chemical substances (also suspected to participate in cancerogenesis), including toxins, organic solvents, pesticides, pharmaceuticals, peroxides, metals, and phthalates, are able to inhibit GJs ^[24].

A number of studies have highlighted the role of Cxs and GJs in carcinogenesis and their involvement in numerous diseases. The loss of GJ or Cxs is associated with abnormal cell proliferation, and the deregulation of GJs is observed in cancer ^{[25][26][27][28][29]}. Additionally, Cxs themselves can promote tumor cell growth and invasiveness, contributing to the overall tumorigenicity ^[30]. Some Cxs (as Cx26, Cx37, and Cx43) may be crucial in metastasis, and intact GJs may act as tumor suppressors in the initial stages of tumorigenesis, controlling cell proliferation. Depending on the state and function of a cell, inhibition of GJs may lead to uncontrolled proliferation (promotion). Re-expression of Cxs in migrating tumor cells was found to promote tumor metastasis ^[31]. It was also recognized that Cxs and GJs can protect cells from cancer ^[21]. Therefore, Cxs can act as both inhibitors of cell proliferation and inducers of cell migration, invasion, and activation of cancer cell migration out of the tumor core ^{[32][33]}.

2.1.2. Ligand-Receptor Pairs and Cell Adhesion

Adjacent cells can communicate through contact-dependent signaling. Physical contact of one cell with a recipient cell is required. Receptors and ligands can occur as membrane-anchored molecules, and their interaction results in effective membrane adhesion. The ligand binds to membrane receptors expressed by other cells, initiating a specific signaling pathway and response in the target cell, but ligands rather do not diffuse from the producing cell ^{[34][35]}. Adhesion molecules take part in juxtacrine stimulation and are receptors that consist of an intercellular domain that interacts with cytoplasmic proteins and an extracellular domain that binds to ligands ^{[36][37]}. Adhesion molecules are commonly divided into integrins, selectins, cadherins, and the members of the immunoglobulin superfamily. Different adhesion molecules bind to different ligands. Cadherins, selectins, and immunoglobulin superfamily members are involved in cell–cell adhesion. Integrins typically joint the cell with its extracellular matrix, but immune cell integrins also bind to soluble ligands and ligands on other cells. Many proteins of ECM, in particular laminins, fibronectin, collagens, and vitronectin, are ligands for integrins. Due to their function, adhesion molecules are part of an adhesive network. Their expression not only changes in cancer cells but also in non-cancerous cells of TME ^[38].

Moreover, the epithelial barriers for cancer cell migration are made by intercellular adhesion complexes, including tight junctions (TJs), adherens junctions (AJs), and desmosomes. These connections are mainly characteristic of epithelial and endothelial cells. Networks called "tight junctions" are formed in areas where the cell membranes, joined by transmembrane proteins and cell cytoskeletons, are fused. These connections mediate ion or other molecules transport and osmosis as well as dictate cell polarity ^{[39][40]}. Up- or downregulation of TJ proteins in cancers results in the loss of cell–cell association, thus can lead to uncontrolled growth and metastasis ^[28]. A correlation between the loss of TJs and tumor differentiation has been observed ^[29]. AJs are mainly composed of cadherins on adjacent cells, cadherin adhesion receptors, and the catenin protein family ^[41]. Downregulation of E-cadherin was found to be associated with progression, promotion, and poor prognosis, and changes in E-cadherin expression were documented in gastric, prostate, liver, and colon cancers ^{[42][43][44]}. In addition, in prostate, gastric, colorectal, cervical, bladder, breast, skin, and endometrial cancer, modifications of the expression of desmosomal elements have also been observed ^{[45][46]}.

2.1.3. Tunnel Nanotubes called "Intercellular Bridges"

Long-distance cell-to-cell communication is possible by the formation of tunneling nanotubes (TNTs) and tumor microtubes (TMs), which allow cells to exchange different chemical and biological material between cells and also facilitate electrical long-range coupling. TNTs are long and thin (50–200 nm) extensions of the cell cytoplasm formed by F-actin filaments that form open-ended channels ^[47]. They occur in many types of cells, e.g., immune cells (like

macrophages), two types of lymphocytes (natural killers and B-cells), and also in dendritic cells. It has been reported that TNTs are formed between endothelial progenitor cells and endothelial cells, renal proximal tubular epithelial cells (RPTEC), retinal pigment epithelial cells, and also between cardiac myocytes ^{[48][49]}. TNTs are also extensively used for cancerous cell communication. Formation of those structures has been observed in vitro in cell lines of colon carcinoma cells, MCF7 and MDA-MB-231 breast cancers, and also in bladder cancer cells ^{[50][51]}. Described nanotubes either connect cancer cells together or cancer cells with normal stromal cells (e.g., B-cell precursor acute lymphoblastic leukemia cells and acute myeloid leukemia cells) ^[52].

TNTs have been recognized as a novel mode of intercellular communication, enabling the transfer of mitochondria, cellular vesicles, miRNAs, viral particles, and some proteins. In cancer cells, TNT formation may correlate with invasiveness. Moreover, factors that can be transferred by TNTs have been shown to promote chemoresistance ^{[53][54]}.

Little is known about TNT biogenesis, but it has been recognized that TNTs contain an array of cytoskeletal filaments of which F-actin is the most abundant. The presence of calcium-sensitive dynamin-related Rho-GTPases (Miro1 and Miro2), KLF 5 kinesin motor protein, and accessory proteins like TRAK 1 and TRAK2 were reported in the structure of TNTs. Myo 19 and Myo 10 permit the efficient shipping of cargo between cells via an actin–myosin-dependent mechanism. The transported cargo includes large organelles like mitochondria and lysosomes ^{[55][56]}. Different conditions may promote TNT formation. Wang et al. demonstrated a significant contribution of p53, EGFR, Akt, PI3K, and also mTOR activation in neurons and astrocytes. They also recognized that cells under stress conditions always develop TNTs with unstressed cells ^[52]]. In this experimental study, hydrogen peroxide, serum depletion, and some cytokines are known as factors that favor TNT formation. These structures, particularly under stress or injury conditions, transfer mitochondria and other cellular components between numerous cell types, including endothelial, epithelial, cardiac, renal, and immune cells ^[58]. TMs are larger and longer-lived (active from several days to a month) than TNTs (active for minutes). TMs are composed of Cx43 gap junction protein and were recognized to promote aggressiveness, invasion, and therapeutic resistance of brain cancer cells ^{[59][60]}.

Both TNTs and TMs, and GJs, allow a direct exchange of cytoplasmic molecules between connected cells, therefore, they participate in key biological processes, including development, immune response, and signaling, and are also involved in the pathogenesis of several diseases, including HIV and neurodegenerative and cancerous diseases ^[61]. However, there are some differences between TNTs (TMs) and GJs. The distance required to establish cellular membrane contact must be short in forming GJs, while TNTs allow long-distance communication. Secondly, these structures transport molecules of varying sizes. GJs are able to transport only small molecules (up to 1.2 kDa), including second messengers and small peptides, whereas TNTs are able to transfer both small molecules as well as mediate the exchange of large organelles and vesicles. Although both TNTs and GJs are known to mediate cell-to-cell interactions, still little is known whether and how these two direct intercellular communication systems interact with each other ^[62].

2.2. Indirect Intercellular Communication

2.2.1. Signaling by Extracellular Vesicles

Normal and cancerous cells secrete small membrane-bound particles in cup-shaped called extracellular vesicles (EVs), which circulate in body fluids and are finally taken up by proximal or distal recipient cells. However, EVs are characterized by a short half-life; a few minutes after infusion, up to 90% of them are eliminated from the bloodstream ^[63]. Based on particle diameter and mechanism of formation, three types of EVs have been identified: exosomes (with 40–120 nm diameter), microvesicles (MVs, with 50–1000 nm diameter), and apoptotic bodies (Abs, with 500–2000 nm diameter) ^[64] ^[65]. EVs carry packages of information located inside or on their surface. Their cargo composition depends on the cell-type origin and varies between cell conditions. As a result of fusion, receptor-mediated internalization or endocytosis of the EVs with the plasmalemma of the recipient cell, transported proteins (enzymes, membrane proteins, heat shock proteins, transcription factors), mRNA, tRNAs, non-coding RNAs (LncRNAs), microRNAs and DNA, and lipids (such as sphingolipids, ceramides, cholesterol, and saturated fatty acids) are released, and affect the recipient cell's metabolism and can induce cell transformation [^{[66][67]}.

Available data indicate that different factors can modify EV release, and so, for example, in response to intracellular Ca^{2+} accumulation or microenvironment acidification, their secretion is increased [68][69]. In the blood of cancer patients, twice more exosomes were found than in the blood of healthy individuals [70][71]. A high amount of cancer-derived exosomes is associated with poor prognosis [72].

Exosome-mediated interaction between cancer and stromal cells leading to the transfer of tumor-derived bioactive molecules can modify the TME. Tumor-derived EVs, called TEVs, play particular roles in cancer initiation, progression, and metastasis, as shown in ovarian^[73], colorectal ^[74], breast ^[75], melanoma ^[76], and prostate cancer ^[77]. Moreover, TEVs

are suppliers of multidrug resistance-associated proteins involved in treatment failure development. Corcoran and colleagues showed that in prostate cancer, EVs convey docetaxel resistance ^[78]. This phenomenon was also observed in lung, breast, and liver cancers ^{[79][80][81]}. Simultaneously, communication via exosomes is bidirectional; EVs can be donated from cancer cells, but can also reach malignant cells. Exosomes are secreted by different cells as intestinal epithelial cells, platelets, mastocytes, antigen-presenting cells, fibroblasts, hepatocytes, and lymphocytes, and EVs of different origins can interact with cancer cells ^[82]. Exosomes derived from CAF can provide nutrients to malignant cells and support tumor growth ^[83]. Apart from the well-known effects of exosomes on the differentiation of surrounded cells, TEVs can also mediate tumor-stem/progenitor cell communication to create pro-tumorigenic microenvironments ^{[84][85]}. It was observed that through transfer of the oncoprotein MET, melanoma-derived exosomes stimulate vasculogenic and hematopoietic bone marrow progenitor cells ^[86]. Tumor-derived exosomes are also involved in tumor immune escape as extracellular vesicles expressing FasL secreted from melanoma cells induce Fas-mediated apoptosis in T-cells. However, it cannot be ruled out that at early stage of cancer, exosomes have antitumor functions ^[87].

2.2.2. Signaling by Cytokines, Chemokines, and Growth Factors

Various cytokines, chemokines, and growth factors are involved in cell–cell cross-talk within tumor and cancer environments, as well as in communication between cancerous and non-cancerous cells ^[89]. Cytokines and chemokines are key players in the development of cancer-related inflammation, with consequent direct and indirect effects on tumor cells ^[89]. Cytokines are secreted in response to different cellular, developmental, and/or environmental stresses and act via cytokine receptors on target cells, affecting certain intracellular signaling pathways to promote a specific cellular response. They affect not only the above-mentioned closely adjacent cells, as well as acting at a distance, and may also affect the cell of their origin. Cytokines can both inhibit cancer development and progression as well as promote its growth and enhance invasion. Interleukins are produced not only by immune cells but also by cancer cells, and cancer cells express interleukin receptors. There is strong evidence that interleukins such as IL-1, IL-4, and IL-6, as well as IL-8 and IL-10, promote tumor development ^{[90][91][92][93][94]}. IL-6 levels are correlated with cancer progression and inversely correlated with patient response to treatment and survival ^[95]. Other interleukins as IL-2 and other members of this family, such as IL-15 and IL-21, as well as IL-12, have been recognized to activate host immune systems against cancer ^{[96][97]}[98].

Tumor necrosis factor α (TNF α) and TGF β signaling affect the tumor environment, tumor progression, and drug resistance ^[99]. Current data underline a complex impact of these factors on tumorigenesis and cancer therapy ^{[100][101][102]}. Tumor necrosis factor-related apoptosis-inducing ligand (TRAIL) is able to selectively induce apoptosis in cancer cells ^[103]. Moreover, interferon-gamma (IFN-gamma) can protect against tumor development, affecting both innate and adaptive host immune responses ^[104].

Chemokines, a large family of cytokines with chemotactic activity, mediate leukocyte migration and are relevant for cancer-related inflammation. The majority of cancer cells produce chemokines, as well as their receptors. In general, chemokine expression at the tumor site promotes leukocyte infiltration and anti-tumor immune responses ^[105], while the expression of chemokine receptors by cancer cells promotes their growth and metastasis. Therefore, tumor-derived chemokines can both inhibit and stimulate tumor growth. Monocyte chemoattractant protein 1 (MCP-1, also known as CCL2) and other chemokines as CXCL1, CXCL3, CXCL5, and CCL8 can act as mediators of angiogenesis and tumor progression ^[106], while those without the ELR motif (CXCL4, CXCL9, CXCL10, CXCL11, and CXCL14) have been shown to inhibit angiogenesis ^[107]. Additionally, F-box proteins, which are subunit recruiting modules of SCF (SKP1-Cullin 1-F-box protein) E3 ligase complexes, influence cell proliferation, invasion, and metastasis, therefore, are involved in cancer development and progression ^[108].

Vascular endothelial growth factor (VEGF) and their receptors—VEGFR-1, VEGFR-2, VEGFR-3, neuropilin (NRP)-1, and NRP-2—affect tumor angiogenesis by upregulation of expression of a variety of growth factors ^[109]. EGF, a member of a family of peptide growth factors and the EGF/EGF receptor signaling pathway, is involved in cell proliferation, differentiation, and migration ^[110]. Expressions of the EGF and its receptor (EGFR) are correlated with tumor growth and metastasis ^[111]. Additionally, placental growth factor (PIGF) is an important regulator involved in controlling angiogenic and inflammatory responses by the formation of PIGF/VEGF homodimer/heterodimer and VEGF-competitive binding to the VEGF receptors and sFlt-1 ^[112]. Furthermore, the enhanced activity of platelet-derived growth factor (PDGF) receptor signaling may promote tumor development ^[113]. Moreover, dysregulation of the IGF-axis (insulin-like growth factor) is involved in the oncogenesis and metastasis of various solid tumors ^[114].

Nowadays, modern tools, including transcriptome analyses, are used to map ligand and receptor protein expression. They also identify cell types and ligand–receptor interactions, as well as cross-talk between TME cells and TME and cancer cells [115][116][117]. Expression of some ligands and receptors has been recognized to be restricted to the specific cell

types, while some are broadly expressed. Recently, based on bulk RNA sequencing and single-cell RNA sequencing data of more than six thousand glioma cells, the intercellular communication between cancer stem-like cells and macrophages via ligand–receptor interactions have been identified ^[118]. Ligand–receptor pairs related to invasion and angiogenesis have been recognized, as well as ligand–receptor pairs associated with clinical outcomes and patient survival risks.

2.2.3. Metabolites-Mediated Communication

Cancer cells during development alter their biochemical pathways towards increased glucose usage and promote its fermentation to lactate regardless of the amount of oxygen. The accumulation of lactate leads to acidification of the microenvironment. It is hypothesized that lactate is a stimulator of M2 tissue-associated macrophage polarization ^[119] and also activates the secretion of VEGF, TGF- β , as well as hypoxia-inducible factors (HIF-1). Moreover, cancer cells via intensified glycolysis limit glucose availability for tumor-infiltrating lymphocytes. In turn, the lack of glucose reduces the effector functions of immune cells ^[120]. To meet the high energy requirements, cancer cells need other fuel sources. *It was documented that exosomes derived from CAF deliver metabolite cargo to prostate cancer cells. Among them, there are acetate, lactate, and TCA cycle metabolites: citrate, pyruvate and fumarate ^[121]. Tumor cells use these energy-rich metabolites in the TCA cycle to produce ATP. Highly glycolytic CAFs can donate their mitochondria to neighboring cancer cells, enhance their respiration, and facilitate tumor growth ^[122]. Additionally, CAF-derived cytokines like CCL5, IL6, and CXCL10 can promote the TCA cycle and thus favor the proliferation of cancer cells ^[123]. In addition to changes in glucose metabolism, cancer cells are characterized by changing the rates of fatty acid synthesis.*

Bioactive lipids and fatty acids are mediators in the cross-talk between cancer cells and stroma [124]. Fatty acids and other lipids can be transported from stroma cells, especially from adipocytes present in the cancer microenvironment, to cancer cells by lipoproteins or exosomes. Fatty acids taken up by cancer cells and transported by fatty acid-binding proteins (FABPs) are used for energy production for rapid tumor growth and synthesis of prostaglandins, known as potent signaling molecules, and other lipid-derived molecules, which may contribute to cancer development [125] Upregulation of FABP4 in omental metastases compared to the primary ovarian tumor was reported [126]. High FABP3 and FABP4 expressions observed in non-small cell lung cancer were associated with tumor metastasis and negatively affect patient survival [127]. Prostaglandins promote tumor growth in the paracrine mode and coordinate complex interactions between tumor cells and surrounding stromal cells [128]. Cyclooxygenase 2-derived prostaglandin E2 is a potent inducer of the angiogenic switch during mammary cancer progression [129]. PGE2 induces increased proliferation, migration, and invasiveness of colorectal carcinoma cells by phosphorylation of epidermal growth factor receptor EGFR and activation of the PI3K/AKT/mTOR signaling pathway [130]. It was shown that prostaglandin E2 (PGE2) induced the activation of suppressor cells of myeloid origin, which in turn promoted tumor progression of breast cancer [131]. Furthermore, PGE2 promoted the differentiation of monocytes into tumor-associated suppressing macrophages in cervical cancer [132]. Tumorderived PGE2 can also activate CAF to enhance the secretion of kynurenine, a tryptophan metabolite, which, in turn, increases cancer cell invasiveness [133]. Leukocyte-derived leukotrienes were reported to selectively expand the group of breast cancer cells with high tumorigenic potential and support metastasis [134].

Enhanced levels of lysophosphatidic acid, found in the blood and ascites of ovarian cancer patients, can promote the formation of inflammatory cytokines, which favor the survival of malignant cells and predetermine their more aggressive behavior ^[135]. Sphingosine-1-phosphate (S1P), an important bioactive sphingolipid, is involved in angiogenesis and lymphangiogenesis, facilitating tumor growth and metastasis ^[136]. High levels of extracellular S1P induced by enhanced expression of a regulatory sphingosine kinase increase migration and efficiency of vessel formation upon combined cultivation of tumor cells and lymphatic endothelial cells ^[137].

References

- 1. Bode, A.M.; Dong, Z. Cancer prevention research—Then and now. Nat. Rev. Cancer 2009, 9, 508–516, doi:10.1038/nrc2646.
- 2. Helsper, C.C.W.; van Erp, N.N.F.; Peeters, P.; de Wit, N.N.J. Time to diagnosis and treatment for cancer patients in the Netherlands: Room for improvement? Eur. J. Cancer 2017, 87, 113–121, doi:10.1016/j.ejca.2017.10.003.
- Schirrmacher, V. From chemotherapy to biological therapy: A review of novel concepts to reduce the side effects of systemic cancer treatment (Review). Int. J. Oncol. 2019, 54, 407–419, doi:10.3892/ijo.2018.4661.
- 4. Bussard, K.M.; Mutkus, L.; Stumpf, K.; Gomez-Manzano, C.; Marini, F.C. Tumor-associated stromal cells as key contributors to the tumor microenvironment. Breast Cancer Res. 2016, 18, 84, doi:10.1186/s13058-016-0740-2.
- Bergfeld, S.A.; DeClerck, Y.A. Bone marrow-derived mesenchymal stem cells and the tumor microenvironment. Cancer Metastasis Rev. 2010, 29, 249–261, doi:10.1007/s10555-010-9222-7.

- 6. Walker, C.; Mojares, E.; Del Rio Hernandez, A. Role of Extracellular Matrix in Development and Cancer Progression. Int. J. Mol. Sci. 2018, 19, doi:10.3390/ijms19103028.
- 7. Naba, A.; Clauser, K.R.; Hoersch, S.; Liu, H.; Carr, S.A.; Hynes, R.O. The matrisome: In silico definition and in vivo characterization by proteomics of normal and tumor extracellular matrices. Mol. Cell. Proteom. 2012, 11, M111.014647, doi:10.1074/mcp.M111.014647.
- Taube, J.M.; Galon, J.; Sholl, L.M.; Rodig, S.J.; Cottrell, T.R.; Giraldo, N.A.; Baras, A.S.; Patel, S.S.; Anders, R.A.; Rimm, D.L.; et al. Implications of the tumor immune microenvironment for staging and therapeutics. Modem Pathol. 2018, 31, 214–234, doi:10.1038/modpathol.2017.156.
- 9. Torralba, D.; Baixauli, F.; Sanchez-Madrid, F. Mitochondria Know No Boundaries: Mechanisms and Functions of Intercellular Mitochondrial Transfer. Front. Cell Dev. Biol. 2016, 4, 107, doi:10.3389/fcell.2016.00107.
- Pasquier, J.; Guerrouahen, B.S.; Al Thawadi, H.; Ghiabi, P.; Maleki, M.; Abu-Kaoud, N.; Jacob, A.; Mirshahi, M.; Galas, L.; Rafii, S.; et al. Preferential transfer of mitochondria from endothelial to cancer cells through tunneling nanotubes modulates chemoresistance. J. Transl. Med. 2013, 11, 94, doi:10.1186/1479-5876-11-94.
- 11. Polak, R.; de Rooij, B.; Pieters, R.; den Boer, M.L. B-cell precursor acute lymphoblastic leukemia cells use tunneling nanotubes to orchestrate their microenvironment. Blood 2015, 126, 2404–2414, doi:10.1182/blood-2015-03-634238.
- De la Cruz-Lopez, K.G.; Castro-Munoz, L.J.; Reyes-Hernandez, D.O.; Garcia-Carranca, A.; Manzo-Merino, J. Lactate in the Regulation of Tumor Microenvironment and Therapeutic Approaches. Front. Oncol. 2019, 9, 1143, doi:10.3389/fonc.2019.01143.
- 13. Petrova, V.; Annicchiarico-Petruzzelli, M.; Melino, G.; Amelio, I. The hypoxic tumour microenvironment. Oncogenesis 2018, 7, 10, doi:10.1038/s41389-017-0011-9.
- Albrengues, J.; Bertero, T.; Grasset, E.; Bonan, S.; Maiel, M.; Bourget, I.; Philippe, C.; Herraiz Serrano, C.; Benamar, S.; Croce, O.; et al. Epigenetic switch drives the conversion of fibroblasts into proinvasive cancer-associated fibroblasts. Nat. Commun. 2015, 6, 10204, doi:10.1038/ncomms10204.
- Albrengues, J.; Bourget, I.; Pons, C.; Butet, V.; Hofman, P.; Tartare-Deckert, S.; Feral, C.C.; Meneguzzi, G.; Gaggioli, C. LIF mediates proinvasive activation of stromal fibroblasts in cancer. Cell Rep. 2014, 7, 1664–1678, doi:10.1016/j.celrep.2014.04.036.
- Foster, C.T.; Gualdrini, F.; Treisman, R. Mutual dependence of the MRTF-SRF and YAP-TEAD pathways in cancerassociated fibroblasts is indirect and mediated by cytoskeletal dynamics. Genes Dev. 2017, 31, 2361–2375, doi:10.1101/gad.304501.117.
- Scharenberg, M.A.; Pippenger, B.E.; Sack, R.; Zingg, D.; Ferralli, J.; Schenk, S.; Martin, I.; Chiquet-Ehrismann, R. TGF-β-induced differentiation into myofibroblasts involves specific regulation of two MKL1 isoforms. J. Cell Sci. 2014, 127, 1079–1091, doi:10.1242/jcs.142075.
- 18. Aasen, T.; Leithe, E.; Graham, S.V.; Kameritsch, P.; Mayán, M.D.; Mesnil, M.; Pogoda, K.; Tabernero, A. Connexins in cancer: Bridging the gap to the clinic. Oncogene 2019, 38, 4429–4451, doi:10.1038/s41388-019-0741-6.
- 19. Qu, Y.; Dahl, G. Function of the voltage gate of gap junction channels: Selective exclusion of molecules. Proc. Natl. Acad. Sci. USA 2002, 99, 697–702, doi:10.1073/pnas.022324499.
- 20. Beckmann, A.; Hainz, N.; Tschernig, T.; Meier, C. Facets of Communication: Gap Junction Ultrastructure and Function in Cancer Stem Cells and Tumor Cells. Cancers (Basel) 2019, 11, doi:10.3390/cancers11030288.
- 21. Aasen, T.; Mesnil, M.; Naus, C.C.; Lampe, P.D.; Laird, D.W. Gap junctions and cancer: Communicating for 50 years. Nat. Rev. Cancer 2016, 16, 775–788, doi:10.1038/nrc.2016.105.
- 22. Sorgen, P.L.; Trease, A.J.; Spagnol, G.; Delmar, M.; Nielsen, M.S. Protein(-)Protein Interactions with Connexin 43: Regulation and Function. Int. J. Mol. Sci. 2018, 19, doi:10.3390/ijms19051428.
- 23. Loewenstein, W.R.; Kanno, Y. Intercellular communication and the control of tissue growth: Lack of communication between cancer cells. Nature 1966, 209, 1248–1249, doi:10.1038/2091248a0.
- Babica, P.; Ctverackova, L.; Lencesova, Z.; Trosko, J.E.; Upham, B.L. Chemopreventive Agents Attenuate Rapid Inhibition of Gap Junctional Intercellular Communication Induced by Environmental Toxicants. Nutr. Cancer 2016, 68, 827–837, doi:10.1080/01635581.2016.1180409.
- 25. Xu, N.; Chen, H.J.; Chen, S.H.; Xue, X.Y.; Chen, H.; Zheng, Q.S.; Wei, Y.; Li, X.D.; Huang, J.B.; Cai, H.; et al. Reduced Connexin 43 expression is associated with tumor malignant behaviors and biochemical recurrence-free survival of prostate cancer. Oncotarget 2016, 7, 67476–67484, doi:10.18632/oncotarget.11231.
- 26. Yeh, E.S.; Williams, C.J.; Williams, C.B.; Bonilla, I.V.; Klauber-DeMore, N.; Phillips, S.L. Dysregulated connexin 43 in HER2-positive drug resistant breast cancer cells enhances proliferation and migration. Oncotarget 2017, 8, 109358–

109369, doi:10.18632/oncotarget.22678.

- 27. Arshad, M.; Conzelmann, C.; Riaz, M.A.; Noll, T.; Gunduz, D. Inhibition of Cx43 attenuates ERK1/2 activation, enhances the expression of Cav1 and suppresses cell proliferation. Int. J. Mol. Med. 2018, 42, 2811–2818, doi:10.3892/ijmm.2018.3828.
- 28. Martin, T.A. The role of tight junctions in cancer metastasis. Semin. Cell Dev. Biol. 2014, 36, 224–231, doi:10.1016/j.semcdb.2014.09.008.
- Ren, J.; Hamada, J.; Takeichi, N.; Fujikawa, S.; Kobayashi, H. Ultrastructural differences in junctional intercellular communication between highly and weakly metastatic clones derived from rat mammary carcinoma. Cancer Res. 1990, 50, 358–362.
- 30. Sinyuk, M.; Mulkearns-Hubert, E.E.; Reizes, O.; Lathia, J. Cancer Connectors: Connexins, Gap Junctions, and Communication. Front. Oncol. 2018, 8, 646, doi:10.3389/fonc.2018.00646.
- 31. Kar, R.; Batra, N.; Riquelme, M.A.; Jiang, J.X. Biological role of connexin intercellular channels and hemichannels. Arch. Biochem. Biophys. 2012, 524, 2–15, doi:10.1016/j.abb.2012.03.008.
- 32. Sin, W.C.; Crespin, S.; Mesnil, M. Opposing roles of connexin43 in glioma progression. Biochim. Biophys. Acta 2012, 1818, 2058–2067, doi:10.1016/j.bbamem.2011.10.022.
- Arora, S.; Heyza, J.R.; Chalfin, E.C.; Ruch, R.J.; Patrick, S.M. Gap Junction Intercellular Communication Positively Regulates Cisplatin Toxicity by Inducing DNA Damage through Bystander Signaling. Cancers (Basel) 2018, 10, doi:10.3390/cancers10100368.
- 34. Mattes, B.; Scholpp, S. Emerging role of contact-mediated cell communication in tissue development and diseases. Histochem. Cell Biol. 2018, 150, 431–442, doi:10.1007/s00418-018-1732-3.
- Bosenberg, M.W.; Massague, J. Juxtacrine cell signaling molecules. Curr. Opin. Cell Biol. 1993, 5, 832–838, doi:10.1016/0955-0674(93)90032-I.
- 36. Cuatrecasas, P. Membrane receptors. Annu. Rev. Biochem. 1974, 43, 169–214, doi:10.1146/annurev.bi.43.070174.001125.
- 37. Sever, R.; Brugge, J.S. Signal transduction in cancer. Cold Spring Harb. Perspect. Med. 2015, 5, doi:10.1101/cshperspect.a006098.
- 38. De Palma, M.; Biziato, D.; Petrova, T.V. Microenvironmental regulation of tumour angiogenesis. Nat. Rev. Cancer 2017, 17, 457–474, doi:10.1038/nrc.2017.51.
- 39. Gonzalez-Mariscal, L.; Tapia, R.; Chamorro, D. Crosstalk of tight junction components with signaling pathways. Biochim. Biophys. Acta 2008, 1778, 729–756, doi:10.1016/j.bbamem.2007.08.018.
- 40. Ikenouchi, J.; Matsuda, M.; Furuse, M.; Tsukita, S. Regulation of tight junctions during the epithelium-mesenchyme transition: Direct repression of the gene expression of claudins/occludin by Snail. J. Cell Sci. 2003, 116, 1959–1967, doi:10.1242/jcs.00389.
- 41. Takeichi, M. Dynamic contacts: Rearranging adherens junctions to drive epithelial remodelling. Nat. Rev. Mol. Cell Biol. 2014, 15, 397–410, doi:10.1038/nrm3802.
- Ling, Z.Q.; Li, P.; Ge, M.H.; Zhao, X.; Hu, F.J.; Fang, X.H.; Dong, Z.M.; Mao, W.M. Hypermethylation-modulated downregulation of CDH1 expression contributes to the progression of esophageal cancer. Int. J. Mol. Med. 2011, 27, 625– 635, doi:10.3892/ijmm.2011.640.
- 43. Mayer, B.; Johnson, J.P.; Leitl, F.; Jauch, K.W.; Heiss, M.M.; Schildberg, F.W.; Birchmeier, W.; Funke, I. E-cadherin expression in primary and metastatic gastric cancer: Down-regulation correlates with cellular dedifferentiation and glandular disintegration. Cancer Res. 1993, 53, 1690–1695.
- 44. Zhai, B.; Yan, H.X.; Liu, S.Q.; Chen, L.; Wu, M.C.; Wang, H.Y. Reduced expression of E-cadherin/catenin complex in hepatocellular carcinomas. World J. Gastroenterol. 2008, 14, 5665–5673, doi:10.3748/wjg.14.5665.
- 45. Shiina, H.; Breault, J.E.; Basset, W.W.; Enokida, H.; Urakami, S.; Li, L.C.; Okino, S.T.; Deguchi, M.; Kaneuchi, M.; Terashima, M.; et al. Functional Loss of the gamma-catenin gene through epigenetic and genetic pathways in human prostate cancer. Cancer Res. 2005, 65, 2130–2138, doi:10.1158/0008-5472.Can-04-3398.
- Dusek, R.L.; Attardi, L.D. Desmosomes: New perpetrators in tumour suppression. Nat. Rev. Cancer 2011, 11, 317– 323, doi:10.1038/nrc3051.
- 47. Roehlecke, C.; Schmidt, M.H.H. Tunneling Nanotubes and Tumor Microtubes in Cancer. Cancers (Basel) 2020, 12, doi:10.3390/cancers12040857.
- 48. Drab, M.; Stopar, D.; Kralj-Iglic, V.; Iglic, A. Inception Mechanisms of Tunneling Nanotubes. Cells 2019, 8, doi:10.3390/cells8060626.

- Ribeiro-Rodrigues, T.M.; Martins-Marques, T.; Morel, S.; Kwak, B.R.; Girao, H. Role of connexin 43 in different forms of intercellular communication—Gap junctions, extracellular vesicles and tunnelling nanotubes. J. Cell Sci. 2017, 130, 3619–3630, doi:10.1242/jcs.200667.
- 50. Desir, S.; Wong, P.; Turbyville, T.; Chen; Shetty, M.; Clark, C.; Zhai, E.; Romin, Y.; Manova-Todorova, K.; Starr, T.K.; et al. Intercellular Transfer of Oncogenic KRAS via Tunneling Nanotubes Introduces Intracellular Mutational Heterogeneity in Colon Cancer Cells. Cancers (Basel) 2019, 11, doi:10.3390/cancers11070892.
- 51. D'Aloia, A.; Berruti, G.; Costa, B.; Schiller, C.; Ambrosini, R.; Pastori, V.; Martegani, E.; Ceriani, M. RalGPS2 is involved in tunneling nanotubes formation in 5637 bladder cancer cells. Exp. Cell Res. 2018, 362, 349–361, doi:10.1016/j.yexcr.2017.11.036.
- Asencio-Barria, C.; Defamie, N.; Saez, J.C.; Mesnil, M.; Godoy, A.S. Direct Intercellular Communications and Cancer: A Snapshot of the Biological Roles of Connexins in Prostate Cancer. Cancers (Basel) 2019, 11, doi:10.3390/cancers11091370.
- Pasquier, J.; Galas, L.; Boulangé-Lecomte, C.; Rioult, D.; Bultelle, F.; Magal, P.; Webb, G.; Le Foll, F. Different modalities of intercellular membrane exchanges mediate cell-to-cell p-glycoprotein transfers in MCF-7 breast cancer cells. J. Biol. Chem. 2012, 287, 7374–7387, doi:10.1074/jbc.M111.312157.
- 54. Thayanithy, V.; Dickson, E.L.; Steer, C.; Subramanian, S.; Lou, E. Tumor-stromal cross talk: Direct cell-to-cell transfer of oncogenic microRNAs via tunneling nanotubes. Transl. Res. 2014, 164, 359–365, doi:10.1016/j.trsl.2014.05.011.
- 55. Nawaz, M.; Fatima, F. Extracellular Vesicles, Tunneling Nanotubes, and Cellular Interplay: Synergies and Missing Links. Front. Mol. Biosci. 2017, 4, 50, doi:10.3389/fmolb.2017.00050.
- 56. Murray, L.M.A.; Krasnodembskaya, A.D. Concise Review: Intercellular Communication Via Organelle Transfer in the Biology and Therapeutic Applications of Stem Cells. Stem Cells 2019, 37, 14–25, doi:10.1002/stem.2922.
- 57. Wang, Y.; Cui, J.; Sun, X.; Zhang, Y. Tunneling-nanotube development in astrocytes depends on p53 activation. Cell Death Differ. 2011, 18, 732–742, doi:10.1038/cdd.2010.147.
- Babenko, V.A.; Silachev, D.N.; Zorova, L.D.; Pevzner, I.B.; Khutornenko, A.A.; Plotnikov, E.Y.; Sukhikh, G.T.; Zorov, D.B. Improving the Post-Stroke Therapeutic Potency of Mesenchymal Multipotent Stromal Cells by Cocultivation With Cortical Neurons: The Role of Crosstalk Between Cells. Stem Cells Transl. Med. 2015, 4, 1011–1020, doi:10.5966/sctm.2015-0010.
- Osswald, M.; Jung, E.; Sahm, F.; Solecki, G.; Venkataramani, V.; Blaes, J.; Weil, S.; Horstmann, H.; Wiestler, B.; Syed, M.; et al. Brain tumour cells interconnect to a functional and resistant network. Nature 2015, 528, 93–98, doi:10.1038/nature16071.
- 60. Osswald, M.; Solecki, G.; Wick, W.; Winkler, F. A malignant cellular network in gliomas: Potential clinical implications. Neuro Oncol. 2016, 18, 479–485, doi:10.1093/neuonc/now014.
- Abounit, S.; Delage, E.; Zurzolo, C. Identification and Characterization of Tunneling Nanotubes for Intercellular Trafficking. Curr. Protoc. Cell Biol. 2015, 67, 12.10.11–12.10.21, doi:10.1002/0471143030.cb1210s67.
- 62. Okafo, G.; Prevedel, L.; Eugenin, E. Tunneling nanotubes (TNT) mediate long-range gap junctional communication: Implications for HIV cell to cell spread. Sci. Rep. 2017, 7, 16660, doi:10.1038/s41598-017-16600-1.
- 63. Liu, S.-L.; Sun, P.; Li, Y.; Liu, S.-S.; Lu, Y. Exosomes as critical mediators of cell-to-cell communication in cancer pathogenesis and their potential clinical application. Transl. Cancer Res. 2019, 8, 298–311.
- 64. Kanada, M.; Bachmann, M.H.; Contag, C.H. Signaling by Extracellular Vesicles Advances Cancer Hallmarks. Trends Cancer 2016, 2, 84–94, doi:10.1016/j.trecan.2015.12.005.
- 65. Lotvall, J.; Hill, A.F.; Hochberg, F.; Buzas, E.I.; Di Vizio, D.; Gardiner, C.; Gho, Y.S.; Kurochkin, I.V.; Mathivanan, S.; Quesenberry, P.; et al. Minimal experimental requirements for definition of extracellular vesicles and their functions: A position statement from the International Society for Extracellular Vesicles. J. Extracell Vesicles 2014, 3, 26913, doi:10.3402/jev.v3.26913.
- 66. Sadallah, S.; Eken, C.; Martin, P.J.; Schifferli, J.A. Microparticles (ectosomes) shed by stored human platelets downregulate macrophages and modify the development of dendritic cells. J. Immunol. 2011, 186, 6543–6552, doi:10.4049/jimmunol.1002788.
- 67. Wu, M.; Wang, G.; Hu, W.; Yao, Y.; Yu, X.F. Emerging roles and therapeutic value of exosomes in cancer metastasis. Mol. Cancer 2019, 18, 53, doi:10.1186/s12943-019-0964-8.
- 68. Savina, A.; Fader, C.M.; Damiani, M.T.; Colombo, M.I. Rab11 promotes docking and fusion of multivesicular bodies in a calcium-dependent manner. Traffic 2005, 6, 131–143, doi:10.1111/j.1600-0854.2004.00257.x.

- 69. Oves, M.; Qari, H.; Felemban, N.; Aslam, A.; Khan, A.A.; Rehan, M.; Tabrez, S.; Ahmed, F.; Haque, A.; Khan, I.; et al. Exosomes: A Paradigm in Drug Development against Cancer and Infectious Diseases. J. Nanomater. 2018, 2018, doi:10.1155/2018/6895464.
- 70. Herreros-Villanueva, M.; Bujanda, L. Glypican-1 in exosomes as biomarker for early detection of pancreatic cancer. Ann. Transl. Med. 2016, 4, 64, doi:10.3978/j.issn.2305-5839.2015.10.39.
- Melo, S.A.; Sugimoto, H.; O'Connell, J.T.; Kato, N.; Villanueva, A.; Vidal, A.; Qiu, L.; Vitkin, E.; Perelman, L.T.; Melo, C.A.; et al. Cancer exosomes perform cell-independent microRNA biogenesis and promote tumorigenesis. Cancer Cell 2014, 26, 707–721, doi:10.1016/j.ccell.2014.09.005.
- 72. Lv, L.H.; Wan, Y.L.; Lin, Y.; Zhang, W.; Yang, M.; Li, G.L.; Lin, H.M.; Shang, C.Z.; Chen, Y.J.; Min, J. Anticancer drugs cause release of exosomes with heat shock proteins from human hepatocellular carcinoma cells that elicit effective natural killer cell antitumor responses in vitro. J. Biol. Chem. 2012, 287, 15874–15885, doi:10.1074/jbc.M112.340588.
- 73. Cheng, L.; Wu, S.; Zhang, K.; Qing, Y.; Xu, T. A comprehensive overview of exosomes in ovarian cancer: Emerging biomarkers and therapeutic strategies. J. Ovarian Res. 2017, 10, 73, doi:10.1186/s13048-017-0368-6.
- 74. Takahashi, K.; Yan, I.K.; Kogure, T.; Haga, H.; Patel, T. Extracellular vesicle-mediated transfer of long non-coding RNA ROR modulates chemosensitivity in human hepatocellular cancer. FEBS Open Bio 2014, 4, 458–467, doi:10.1016/j.fob.2014.04.007.
- 75. Li, X.; Zhang, Z.; Beiter, T.; Schluesener, H.J. Nanovesicular vaccines: Exosomes. Arch. Immunol. Ther. Exp. (Warsz) 2005, 53, 329–335.
- 76. Zhao, H.; Yang, L.; Baddour, J.; Achreja, A.; Bernard, V.; Moss, T.; Marini, J.C.; Tudawe, T.; Seviour, E.G.; San Lucas, F.A.; et al. Tumor microenvironment derived exosomes pleiotropically modulate cancer cell metabolism. Elife 2016, 5, e10250, doi:10.7554/eLife.10250.
- 77. Peinado, H.; Aleckovic, M.; Lavotshkin, S.; Matei, I.; Costa-Silva, B.; Moreno-Bueno, G.; Hergueta-Redondo, M.; Williams, C.; Garcia-Santos, G.; Ghajar, C.; et al. Melanoma exosomes educate bone marrow progenitor cells toward a pro-metastatic phenotype through MET. Nat. Med. 2012, 18, 883–891, doi:10.1038/nm.2753.
- 78. Whiteside, T.L. Exosome and mesenchymal stem cell cross-talk in the tumor microenvironment. Semin. Immunol. 2018, 35, 69–79, doi:10.1016/j.smim.2017.12.003.
- 79. Peinado, H.; Zhang, H.; Matei, I.R.; Costa-Silva, B.; Hoshino, A.; Rodrigues, G.; Psaila, B.; Kaplan, R.N.; Bromberg, J.F.; Kang, Y.; et al. Pre-metastatic niches: Organ-specific homes for metastases. Nat. Rev. Cancer 2017, 17, 302–317, doi:10.1038/nrc.2017.6.
- 80. Liu, Y.; Gu, Y.; Cao, X. The exosomes in tumor immunity. Oncoimmunology 2015, 4, e1027472, doi:10.1080/2162402x.2015.1027472.
- Sethi, G.; Shanmugam, M.K.; Ramachandran, L.; Kumar, A.P.; Tergaonkar, V. Multifaceted link between cancer and inflammation. Biosci. Rep. 2012, 32, 1–15, doi:10.1042/BSR20100136.
- 82. Amedei, A.; Prisco, D.; MM, D.E. The use of cytokines and chemokines in the cancer immunotherapy. Recent Pat. Anticancer Drug Discov. 2013, 8, 126–142.
- Mantovani, A.; Barajon, I.; Garlanda, C. IL-1 and IL-1 regulatory pathways in cancer progression and therapy. Immunol. Rev. 2018, 281, 57–61, doi:10.1111/imr.12614.
- Li, Z.; Chen, L.; Qin, Z. Paradoxical roles of IL-4 in tumor immunity. Cell. Mol. Immunol. 2009, 6, 415–422, doi:10.1038/cmi.2009.53.
- Middleton, K.; Jones, J.; Lwin, Z.; Coward, J.I. Interleukin-6: An angiogenic target in solid tumours. Crit. Rev. Oncol. Hematol. 2014, 89, 129–139, doi:10.1016/j.critrevonc.2013.08.004.
- Ben-Baruch, A. The multifaceted roles of chemokines in malignancy. Cancer Metastasis Rev. 2006, 25, 357–371, doi:10.1007/s10555-006-9003-5.
- 87. Mannino, M.H.; Zhu, Z.; Xiao, H.; Bai, Q.; Wakefield, M.R.; Fang, Y. The paradoxical role of IL-10 in immunity and cancer. Cancer Lett. 2015, 367, 103–107, doi:10.1016/j.canlet.2015.07.009.
- Salgado, R.; Junius, S.; Benoy, I.; Van Dam, P.; Vermeulen, P.; Van Marck, E.; Huget, P.; Dirix, L.Y. Circulating interleukin-6 predicts survival in patients with metastatic breast cancer. Int. J. Cancer 2003, 103, 642–646, doi:10.1002/ijc.10833.
- Lu, X. Impact of IL-12 in Cancer. Curr. Cancer Drug Targets 2017, 17, 682–697, doi:10.2174/1568009617666170427102729.
- 90. Setrerrahmane, S.; Xu, H. Tumor-related interleukins: Old validated targets for new anti-cancer drug development. Mol. Cancer 2017, 16, 153, doi:10.1186/s12943-017-0721-9.

- Choudhry, H.; Helmi, N.; Abdulaal, W.H.; Zeyadi, M.; Zamzami, M.A.; Wu, W.; Mahmoud, M.M.; Warsi, M.K.; Rasool, M.; Jamal, M.S. Prospects of IL-2 in Cancer Immunotherapy. Biomed. Res. Int. 2018, 2018, 9056173, doi:10.1155/2018/9056173.
- 92. Martin, M.; Wei, H.; Lu, T. Targeting microenvironment in cancer therapeutics. Oncotarget 2016, 7, 52575–52583, doi:10.18632/oncotarget.9824.
- Hao, Y.; Baker, D.; Ten Dijke, P. TGF-beta-Mediated Epithelial-Mesenchymal Transition and Cancer Metastasis. Int. J. Mol. Sci. 2019, 20, doi:10.3390/ijms20112767.
- 94. Liu, S.; Chen, S.; Zeng, J. TGF-β signaling: A complex role in tumorigenesis (Review). Mol. Med. Rep. 2018, 17, 699– 704, doi:10.3892/mmr.2017.7970.
- 95. Montfort, A.; Colacios, C.; Levade, T.; Andrieu-Abadie, N.; Meyer, N.; Ségui, B. The TNF Paradox in Cancer Progression and Immunotherapy. Front. Immunol. 2019, 10, 1818, doi:10.3389/fimmu.2019.01818.
- Ion, G.N.D.; Nitulescu, G.M.; Popescu, C.I. Targeting TRAIL. Bioorg Med. Chem. Lett. 2019, doi:10.1016/j.bmcl.2019.07.053.
- 97. Ikeda, H.; Old, L.J.; Schreiber, R.D. The roles of IFN gamma in protection against tumor development and cancer immunoediting. Cytokine Growth Factor Rev. 2002, 13, 95–109.
- 98. Chow, M.T.; Luster, A.D. Chemokines in cancer. Cancer Immunol. Res. 2014, 2, 1125–1131, doi:10.1158/2326-6066.CIR-14-0160.
- 99. Salcedo, R.; Ponce, M.L.; Young, H.A.; Wasserman, K.; Ward, J.M.; Kleinman, H.K.; Oppenheim, J.J.; Murphy, W.J. Human endothelial cells express CCR2 and respond to MCP-1: Direct role of MCP-1 in angiogenesis and tumor progression. Blood 2000, 96, 34–40.
- 100. Payne, A.S.; Cornelius, L.A. The role of chemokines in melanoma tumor growth and metastasis. J. Investig. Dermatol. 2002, 118, 915–922, doi:10.1046/j.1523-1747.2002.01725.x.
- 101. Zheng, N.; Zhou, Q.; Wang, Z.; Wei, W. Recent advances in SCF ubiquitin ligase complex: Clinical implications. Biochim. Biophys. Acta 2016, 1866, 12–22, doi:10.1016/j.bbcan.2016.05.001.
- 102. Carmeliet, P. VEGF as a key mediator of angiogenesis in cancer. Oncology 2005, 69 (Suppl. 3), 4–10, doi:10.1159/000088478.
- 103. Zeng, F.; Harris, R.C. Epidermal growth factor, from gene organization to bedside. Semin. Cell Dev. Biol. 2014, 28, 2– 11, doi:10.1016/j.semcdb.2014.01.011.
- 104. Jutten, B.; Rouschop, K.M. EGFR signaling and autophagy dependence for growth, survival, and therapy resistance. Cell Cycle 2014, 13, 42–51, doi:10.4161/cc.27518.
- 105. Newell, L.F.; Holtan, S.G. Placental growth factor: What hematologists need to know. Blood Rev. 2017, 31, 57–62, doi:10.1016/j.blre.2016.08.004.
- 106. Heldin, C.H. Targeting the PDGF signaling pathway in tumor treatment. Cell Commun. Signal. 2013, 11, 97, doi:10.1186/1478-811X-11-97.
- 107. Vella, V.; Malaguarnera, R.; Nicolosi, M.L.; Morrione, A.; Belfiore, A. Insulin/IGF signaling and discoidin domain receptors: An emerging functional connection. Biochim. Biophys. Acta Mol. Cell Res. 2019, 118522, doi:10.1016/j.bbamcr.2019.118522.
- 108. Ramilowski, J.A.; Goldberg, T.; Harshbarger, J.; Kloppmann, E.; Lizio, M.; Satagopam, V.P.; Itoh, M.; Kawaji, H.; Carninci, P.; Rost, B.; et al. A draft network of ligand–receptor-mediated multicellular signalling in human. Nat. Commun. 2015, 6, 7866, doi:10.1038/ncomms8866.
- 109. Zhou, J.X.; Taramelli, R.; Pedrini, E.; Knijnenburg, T.; Huang, S. Extracting Intercellular Signaling Network of Cancer Tissues using Ligand-Receptor Expression Patterns from Whole-tumor and Single-cell Transcriptomes. Sci. Rep. 2017, 7, 8815, doi:10.1038/s41598-017-09307-w.
- 110. Sarkar, S.; Sabhachandani, P.; Stroopinsky, D.; Palmer, K.; Cohen, N.; Rosenblatt, J.; Avigan, D.; Konry, T. Dynamic analysis of immune and cancer cell interactions at single cell level in microfluidic droplets. Biomicrofluidics 2016, 10, 054115, doi:10.1063/1.4964716.
- 111. Yuan, D.; Tao, Y.; Chen, G.; Shi, T. Systematic expression analysis of ligand-receptor pairs reveals important cell-to-cell interactions inside glioma. Cell Commun. Signal. 2019, 17, 48, doi:10.1186/s12964-019-0363-1.
- 112. Colegio, O.R.; Chu, N.Q.; Szabo, A.L.; Chu, T.; Rhebergen, A.M.; Jairam, V.; Cyrus, N.; Brokowski, C.E.; Eisenbarth, S.C.; Phillips, G.M.; et al. Functional polarization of tumour-associated macrophages by tumour-derived lactic acid. Nature 2014, 513, 559–563, doi:10.1038/nature13490.

- 113. Chang, C.H.; Qiu, J.; O'Sullivan, D.; Buck, M.D.; Noguchi, T.; Curtis, J.D.; Chen, Q.; Gindin, M.; Gubin, M.M.; van der Windt, G.J.; et al. Metabolic Competition in the Tumor Microenvironment Is a Driver of Cancer Progression. Cell 2015, 162, 1229–1241, doi:10.1016/j.cell.2015.08.016.
- 114. Ruiz-Lopez, L.; Blancas, I.; Garrido, J.M.; Mut-Salud, N.; Moya-Jodar, M.; Osuna, A.; Rodriguez-Serrano, F. The role of exosomes on colorectal cancer: A review. J. Gastroenterol. Hepatol. 2018, 33, 792–799, doi:10.1111/jgh.14049.
- 115. Lowry, M.C.; Gallagher, W.M.; O'Driscoll, L. The Role of Exosomes in Breast Cancer. Clin. Chem. 2015, 61, 1457– 1465, doi:10.1373/clinchem.2015.240028.
- 116. Tucci, M.; Mannavola, F.; Passarelli, A.; Stucci, L.S.; Cives, M.; Silvestris, F. Exosomes in melanoma: A role in tumor progression, metastasis and impaired immune system activity. Oncotarget 2018, 9, 20826–20837, doi:10.18632/oncotarget.24846.
- 117. Pan, J.; Ding, M.; Xu, K.; Yang, C.; Mao, L.J. Exosomes in diagnosis and therapy of prostate cancer. Oncotarget 2017, 8, 97693–97700, doi:10.18632/oncotarget.18532.
- 118. Corcoran, C.; Rani, S.; O'Brien, K.; O'Neill, A.; Prencipe, M.; Sheikh, R.; Webb, G.; McDermott, R.; Watson, W.; Crown, J.; et al. Docetaxel-resistance in prostate cancer: Evaluating associated phenotypic changes and potential for resistance transfer via exosomes. PLoS ONE 2012, 7, e50999, doi:10.1371/journal.pone.0050999.
- 119. Kong, J.N.; He, Q.; Wang, G.; Dasgupta, S.; Dinkins, M.B.; Zhu, G.; Kim, A.; Spassieva, S.; Bieberich, E. Guggulsterone and bexarotene induce secretion of exosome-associated breast cancer resistance protein and reduce doxorubicin resistance in MDA-MB-231 cells. Int. J. Cancer 2015, 137, 1610–1620, doi:10.1002/ijc.29542.
- 120. Xiao, X.; Yu, S.; Li, S.; Wu, J.; Ma, R.; Cao, H.; Zhu, Y.; Feng, J. Exosomes: Decreased sensitivity of lung cancer A549 cells to cisplatin. PLoS ONE 2014, 9, e89534, doi:10.1371/journal.pone.0089534.
- 121. Lucchetti, D.; Ricciardi Tenore, C.; Colella, F.; Sgambato, A. Extracellular Vesicles and Cancer: A Focus on Metabolism, Cytokines, and Immunity. Cancers (Basel) 2020, 12, doi:10.3390/cancers12010171.
- 122. Santi, A.; Caselli, A.; Ranaldi, F.; Paoli, P.; Mugnaioni, C.; Michelucci, E.; Cirri, P. Cancer associated fibroblasts transfer lipids and proteins to cancer cells through cargo vesicles supporting tumor growth. Biochim. Biophys. Acta 2015, 1853, 3211–3223, doi:10.1016/j.bbamcr.2015.09.013.
- 123. Curtis, M.; Kenny, H.A.; Ashcroft, B.; Mukherjee, A.; Johnson, A.; Zhang, Y.; Helou, Y.; Batlle, R.; Liu, X.; Gutierrez, N.; et al. Fibroblasts Mobilize Tumor Cell Glycogen to Promote Proliferation and Metastasis. Cell Metab. 2019, 29, 141–155, doi:10.1016/j.cmet.2018.08.007.
- 124. Beloribi-Djefaflia, S.; Vasseur, S.; Guillaumond, F. Lipid metabolic reprogramming in cancer cells. Oncogenesis 2016, 5, e189, doi:10.1038/oncsis.2015.49.
- 125. Shender, V.O.; Pavlyukov, M.S.; Ziganshin, R.H.; Arapidi, G.P.; Kovalchuk, S.I.; Anikanov, N.A.; Altukhov, I.A.; Alexeev, D.G.; Butenko, I.O.; Shavarda, A.L.; et al. Proteome-metabolome profiling of ovarian cancer ascites reveals novel components involved in intercellular communication. Mol. Cell. Proteom. 2014, 13, 3558–3571, doi:10.1074/mcp.M114.041194.
- 126. Nieman, K.M.; Kenny, H.A.; Penicka, C.V.; Ladanyi, A.; Buell-Gutbrod, R.; Zillhardt, M.R.; Romero, I.L.; Carey, M.S.; Mills, G.B.; Hotamisligil, G.S.; et al. Adipocytes promote ovarian cancer metastasis and provide energy for rapid tumor growth. Nat. Med. 2011, 17, 1498–1503, doi:10.1038/nm.2492.
- 127. Tang, Z.; Shen, Q.; Xie, H.; Zhou, X.; Li, J.; Feng, J.; Liu, H.; Wang, W.; Zhang, S.; Ni, S. Elevated expression of FABP3 and FABP4 cooperatively correlates with poor prognosis in non-small cell lung cancer (NSCLC). Oncotarget 2016, 7, 46253–46262, doi:10.18632/oncotarget.10086.
- 128. Wang, D.; Dubois, R.N. Eicosanoids and cancer. Nat. Rev. Cancer 2010, 10, 181–193, doi:10.1038/nrc2809.
- 129. Chang, S.H.; Liu, C.H.; Conway, R.; Han, D.K.; Nithipatikom, K.; Trifan, O.C.; Lane, T.F.; Hla, T. Role of prostaglandin E2-dependent angiogenic switch in cyclooxygenase 2-induced breast cancer progression. Proc. Natl. Acad. Sci. USA 2004, 101, 591–596, doi:10.1073/pnas.2535911100.
- Buchanan, F.G.; Wang, D.; Bargiacchi, F.; DuBois, R.N. Prostaglandin E2 regulates cell migration via the intracellular activation of the epidermal growth factor receptor. J. Biol. Chem. 2003, 278, 35451–35457, doi:10.1074/jbc.M302474200.
- 131. Xiang, X.; Poliakov, A.; Liu, C.; Liu, Y.; Deng, Z.B.; Wang, J.; Cheng, Z.; Shah, S.V.; Wang, G.J.; Zhang, L.; et al. Induction of myeloid-derived suppressor cells by tumor exosomes. Int. J. Cancer 2009, 124, 2621–2633, doi:10.1002/ijc.24249.
- Heusinkveld, M.; de Vos van Steenwijk, P.J.; Goedemans, R.; Ramwadhdoebe, T.H.; Gorter, A.; Welters, M.J.; van Hall, T.; van der Burg, S.H. M2 macrophages induced by prostaglandin E2 and IL-6 from cervical carcinoma are switched to activated M1 macrophages by CD4+ Th1 cells. J. Immunol. 2011, 187, 1157–1165, doi:10.4049/jimmunol.1100889.

- 133. Chen, J.Y.; Li, C.F.; Kuo, C.C.; Tsai, K.K.; Hou, M.F.; Hung, W.C. Cancer/stroma interplay via cyclooxygenase-2 and indoleamine 2,3-dioxygenase promotes breast cancer progression. Breast Cancer Res. 2014, 16, 410, doi:10.1186/s13058-014-0410-1.
- 134. Wculek, S.K.; Malanchi, I. Neutrophils support lung colonization of metastasis-initiating breast cancer cells. Nature 2015, 528, 413–417, doi:10.1038/nature16140.
- 135. Kuwata, S.; Ohkubo, K.; Kumamoto, S.; Yamaguchi, N.; Izuka, N.; Murota, K.; Tsujiuchi, T.; Iwamori, M.; Fukushima, N. Extracellular lipid metabolism influences the survival of ovarian cancer cells. Biochem. Biophys. Res. Commun. 2013, 439, 280–284, doi:10.1016/j.bbrc.2013.08.041.
- 136. Visentin, B.; Vekich, J.A.; Sibbald, B.J.; Cavalli, A.L.; Moreno, K.M.; Matteo, R.G.; Garland, W.A.; Lu, Y.; Yu, S.; Hall, H.S.; et al. Validation of an anti-sphingosine-1-phosphate antibody as a potential therapeutic in reducing growth, invasion, and angiogenesis in multiple tumor lineages. Cancer Cell 2006, 9, 225–238, doi:10.1016/j.ccr.2006.02.023.
- 137. Anelli, V.; Gault, C.R.; Snider, A.J.; Obeid, L.M. Role of sphingosine kinase-1 in paracrine/transcellular angiogenesis and lymphangiogenesis in vitro. FASEB J. 2010, 24, 2727–2738, doi:10.1096/fj.09-150540.

Retrieved from https://encyclopedia.pub/entry/history/show/9549