Okara and Gut

Subjects: Gastroenterology & Hepatology Contributor: Mohammed Sharif Swallah

Okara is a white-yellow fibrous residue consisting of the insoluble fraction of the soybean seeds remaining after extraction of the aqueous fraction during the production of tofu and soymilk, and is generally considered a waste product. It is packed with a significant number of proteins, isoflavones, soluble and insoluble fibers, soyasaponins, and other mineral elements, which are all attributed with health merits. With the increasing production of soy beverages, huge quantities of this by-product are produced annually, which poses significant disposal problems and financial issues for producers. Extensive studies have been done on the biological activities, nutritional values, and chemical composition of okara as well as its potential utilization. Owing to its peculiar rich fiber composition and low cost of production, okara might be potentially useful in the food industry as a functional ingredient or good raw material and could be used as a dietary supplement to prevent varied ailments such as prevention of diabetes, hyperlipidemia, obesity, as well as to stimulate the growth of intestinal microbes and production of microbe-derived metabolites (xenometabolites), since gut dysbiosis (imbalanced microbiota) has been implicated in the progression of several complex diseases.

Keywords: dietary fiber; gut microbiota

1. Introduction

Soybean is labeled as one of the essential crops across the globe and is native to Asia, and has been under cultivation for over thousands of years there. However, the present leading producers are found on the other side of the Pacific Ocean, thus north and south America [1]. America, Brazil, Argentina, and China are currently the leading world producers and consumers of soybean, respectively [2]. Epidemiological reports unveil a strong connection between regular and optimal intake of soybeans with numerous health-promoting functions, such as lowering the risk of several cancer forms (colorectal cancer, prostate cancer, breast cancer, bone health, etc.), cardiovascular diseases, cognitive function, type II diabetes, renal function, atherosclerosis, menopausal symptom, and coronary heart diseases by lowering the levels of (LDL) low-density lipoproteins [3][4][5]. Soybeans are usually processed to obtain protein isolates and other end-products like soymilk and soybean curd (tofu)[6], which are both known traditional Asian food products, however are now consumed across the globe owing to the nutritional and health-promoting claims. A large sum of fibrous residue called okara is produced after soymilk and soybean curd production processes, i.e., obtained after extraction of the aqueous fraction. Okara is a white-yellow material, which consists of the insoluble parts of the seeds remaining in the filter sack when pureed soybean seeds are filtered during soymilk production. Okara is abundant and nutritionally valuable, and has been used in the vegetarian diets of Western countries since the 20th century $\boxed{\mathbb{Z}[8]}$. Studies on the nutritional and non-nutritional constituent of okara reveal that it is packed with a significant number of proteins, isoflavones, soluble and insoluble fibers, soyasaponins, and other mineral elements, which are all attributed with health merits [9][10][11]. Owing to its high dietary fiber content, okara supplementation yielded a decrease in body weight, beneficial properties on lipid metabolism, protected the gut environment in terms of antioxidant status, as well as prebiotic effects [8][12]. Typically, close to 1.2 kg of wet okara is obtained from 1 kg of dry soybean processed for soymilk or tofu. This makes okara a cheap source of fiberrich food. However, it is normally used as fertilizers or is landfilled, animal feed, or discarded as waste due to its high susceptibility to spoilage, extra cost of production, undesirable flavor, and grittiness in texture attributes, which are all caused by its high moisture content. Its valorization will be essential, to aid in utilizing the untapped precious nutrients as well as serving to eliminate the socio-environmental and economic problems caused by this waste disposal [13] [14] [15] [16]. Moreover, most of the valorization research on okara focused more on physical attributes rather than the possible health attributes[17].

In recent years, there has been a growing interest in using by-products or waste biomasses from the food industry as new sources of functional ingredients such as prebiotics. Its value addition would reduce the environmental effect from their decomposition and as well increase their benefit $\frac{[18][19]}{[19]}$. Due to the peculiar dietary fiber composition of okara, it might be potentially useful in the food industry as a functional ingredient. In this sense, it could be used to increase the dietary fiber content in varied cereal products $\frac{[7][8]}{[19]}$. Dietary fiber is evidenced to positively impact the gut health by fostering the growth

of select gut microbes, as well as the production of microbe-derived metabolites (xenometabolites), and non-gastrointestinal related conditions such as non-alcoholic fatty liver disease, diabetes, and cardiovascular disease^[20]. Gut dysbiosis or poor gut health has been implicated in the progression of chronic kidney disease^[21]. This concept has given rise to terms such as the gut-kidney axis^[22] and the gut-liver axis^[23]. Nowadays, numerous studies have been undertaken aiming to uncover the functional and therapeutic effect of okara, as well as facilitate its effective utilization. Hence, this review seeks to compile scientific research on the bioactive compounds in soybean residue (okara) and discuss the possible prebiotic impact of this fiber-rich residue as a functional diet on eubiosis/dysbiosis condition of the gut, as well as the consequential influence on liver and kidney functions, to facilitate a detailed knowledge base for further exploration, implementation, and development.

Dietary Fiber in Nutrition

The adoption of varied plant-based components including dietary fiber, polysaccharides, and prebiotics seems to be on the rise and is gaining more and more attention owing to its supposed health effects. However, there are variations in the nutritional composition of these varied fiber-containing feed/foodstuffs[24], and consumers have developed much interest in the association between the carbohydrate/fiber content of these foods and the possible glycemic effect mediated post ingestion[25]. For instance, an extruded cereal food product is regarded to reflect a high glycemic index similar to or more than that of bread, and it has been evidenced in multiple reports that an individual rate of glycemic response is dictated by the extent and rate of starch hydrolysis after ingestion and can be manipulated through daily meal dietary fiber supplementation [26][27]. Dietary fiber describes (non-digestible carbohydrates of plant origin) food-derived components that are resilient to digestion/hydrolysis by the core enzyme machinery that is present in animals/humans' gut. Fibers are the residual parts of a plant that are safe for consumption and it includes compounds such as cellulose, lignin, cell wall polysaccharides, oligosaccharides, and other related compounds, e.g., phenolic compounds[28][29][30]. Dietary fiber is tagged as the seventh (7th) important food nutrient for organisms and is subcategorized into two types, thus, soluble dietary fiber (SDF) and insoluble dietary fiber (IDF)[13], which are all made of dense indigestible polysaccharides. The most widely acknowledged classification for dietary fiber has been to distinguish dietary components based on solubility in a buffer at a definite pH, and/or fermentability in an in vitro system with an enzyme solution serving as human alimentary enzymes. Further classification is based on fermentability such as less fermentable/water-insoluble (i.e., lignin, cellulose and hemicellulose) and well fermentable/water-soluble (i.e., gums, pectins and mucilages)[31]. Recent research proposes dietary fiber classification by size/density plots, however, the traditional and most convenient ways for dietary fiber classification still remains via solubility to water. Numerous studies have demonstrated support of dietary fiber to influence the increase in cholesterol- and sodium cholate-binding capacity as well as influence the decrease in blood pressure, protect against several cancers, including colorectal cancer, breast cancer and prostatic cancer, prevent gastrointestinal problems [32][33], ameliorate constipation (fecal softening and bulking, improve regularity and/or frequency), depicting an anti-inflammatory effect on the digestive tract as well as aid partially in lipid substitution, blood glucose regulation, and/or blood cholesterol reduction [29][34][35]. Hence, it is justifiable that dietary fiber influences the performance and functions of the gastrointestinal tract and consequently reflects on human/animals' health [36][37]. High dietary fiber consumption is suggested to inhibit the bioavailability of some essential nutritional components including, but not limited to, vitamins and other minerals, and may impact the rate of food digestion, energy metabolism as well as gut microbial composition, which may, in turn, result in the production of short-chain fatty acid that is responsible for (10-30%) the total energy requirement of the host [37][38][39], and on the other hands, aid in the detoxification of the host digestive tract[40]. Besides, dietary fiber, which comprises polysaccharides that skips enzymatic digestion, acts essentially as a substrate to the intestinal microbiota and is suggested to impact the host-microbial community and immunity [41]. Dietary fiber deprivation in a mice experiment yielded to the alteration in mucus-eroding microbiota, disruption of the intestinal barrier, mucus layer depletion, and lethal colitis[42].

2. Valorization of Soybean Residue (Okara) as a Functional Food

As discussed earlier, okara contains high levels of dietary fiber and proteins, and significant amounts of isoflavones as well as mineral elements, which merits it a high nutritional value and a potential prebiotic function. Hence, it is potentially useful as a functional ingredient with health-promoting effects^[8]. Specifically, the incorporation of soybean-derived ingredients into a variety of products with the motive of providing beneficial properties and functions to the body has been the focus over the years, and have aroused much interest from the food industry. These are termed 'functional foods'^[43]. Okara has been used in food production for human consumption as well as application in animal nutrition for several years mainly in Japan and China, both in its processed and raw forms, to more easily provide a fair intake of the nutritional claim for the fiber and protein. Okara can be a partial substitute for soy flour, wheat flour, and other food-producing components so as to boost the protein and fiber content^[44]. The rich amount of proteins, carbohydrates, and other forms of nutrients embedded in okara make it a potential substrate for microbial fermentation. Bacteria, yeasts, and

fungal fermentation of soybean residue are suggested to decrease the content of raw fiber, increase the content of proteins, soluble fiber, isoflavones, and amino acids, and decomposing phytic acid, resulting to an upgrade in the processing properties as well as nutritional value^[45]. The use of soybean residue in different food formulation such as drinks, bread, sausages, pancakes, candies, biscuits, cake, and nutritional flour, has been studied and evidenced earlier in numerous reports^{[46][47][48][49]}.

2.1. Application of Soybean Residue in Human Nutrition

The rich solvent-binding properties of okara make it an ideal low-cost ingredient with which to encourage yields in bakery and meat products^[50]. It has been evidenced to have a positive influence on the shelf life of chocolate cookies mainly at an optimal concentration of 5%, and as well prevent syneresis in cheese ravioli filling during defrosting and freezing. Besides, its insipid taste enables it to be used at relatively high levels without adversely influencing the texture or taste profiles of the formed products^[1].

The study by Park et al. [51], on the influence of okara and additives including, soy flour, starch, and hydroxypropyl methylcellulose on the quality and nutritional value of cookies were reviewed. The results displayed that okara-enriched cookies had higher levels of carbohydrate (35.3%), fat (25.7%), protein (11.6%), and ash (6.3%) in contrast with wheat cookies containing carbohydrate (59.6%), fat (20.2%), protein (15.2%), and ash (2.2%). A cookie containing okara depicted lower carbohydrate and higher ash content. Hydroxypropyl methylcellulose supplemented cookies portrayed a higher water holding capacity, i.e., three times higher than that of control, which enhanced the performance of the dough and the quality of enriched cookies. Cookies supplemented with additives, mainly, soy flour and hydroxypropyl methylcellulose, portrayed reduced water activities, which enhanced the storage life and hardness in consort with significant improvement in the crispness of okara-enriched cookies. Suda et al. [52] incorporated okara powder (containing 50% dietary fiber, 0.45% calcium, and 21.3% vegetable protein) into bread and pancake ingredients, intending to develop fortified foods for medical use. Three dissimilar types of bread were processed, which include the 10% okara bread, and other additives, as well as preservatives, were needed to encourage yeast fermentation and enable storage at room temperature. After effective freezing without preservatives, the flavors of all three bread were altered. A soft pancake was prepared using a mixture of pancake powders with 20% okara. The pancake free from preservatives was suitable for refrigerator storage before being eaten, while both the fresh pancakes and those comprising the preservatives made with 20% okara were accepted as supplements for the elderly hospitalized patients. The study showed that the soft pancake supplemented with okara and 40% water was much use as a fitting supplement for dietary fiber, calcium, and vegetable protein than the bread supplemented with okara. The quality of okara supplemented bread is suggested to significantly increased with the addition of enzymes (lipase, glucose oxidase, and pentosanase). The incorporation of 4-8% soybean powder has been evidenced to yield a quality increase^[53]. The application of 5% soybean dietary fiber on bread, treated with 1% NaOH for an hour and 1% HCl at 60 °C for 2 h, depicted an appearance and quality similar to that of a normal bread[1][54].

A recent study by Kang et al. [15] on the quality attributes of rice noodles enriched with different levels of okara flour (0-20%) was reviewed. The authors recorded that the adhesiveness, hardness, and cooking loss of the noodles increase with an increasing amount of okara, while swelling index, cohesiveness score, and water absorption significantly decreased. Among all samples, noodles fortified with 10% okara flour demonstrated the lowest score for the predicted glycemic index. The incorporation of alginate with a CaCl₂ coating upgraded the cooking properties without affecting the in vitro starch digestibility of soybean residue-enriched rice noodles. The findings recommended that noodles with good quality attributes and reduced in vitro starch digestibility scores could be developed through incorporating okara flour of up to 10% level. Again, a study on the cooking quality of soybean by-product okara fiber fortified noodle reported that the noodles were of good cooking quality when okara fiber was supplemented at 9% (particle size of 100 mesh), 4% salt, and 0.25% sodium alginate^[55]. Pan et al. [46] studied the impact of okara and vital gluten on the physical and chemical properties of noodles. The results portrayed that a higher level of okara (10-15%) significantly decreased the tensile strength, extensibility, elasticity, and optimum cooking time of the noodles. Noodles enriched with okara displayed an improved flavonoids content, total phenolic, and antioxidant and/or free radical-scavenging activity. The results showed that 6% wheat gluten and 5-10% okara powder incorporation produces noodles with good texture, cooking, and sensory properties. Hence, soybean dietary fiber supplementation in food formulation can be the best alternative in achieving food products with a low glycemic index.

In the study by Katayama and Wilson $[\underline{56}]$, a soybean-based snack was formulated using partially dried okara (44.3% moisture) prepared from lipoxygenase-free soybeans and a commercially dried okara powder (7.7% moisture) made from regular soybeans (lipoxygenase-present). The authors also utilized commercially low linolenic acid soybean oil and commercially low saturated soybean oil in the same recipe and determine the best recipe for baked or a deep-fried soybased food product. At the end of the study, the authors observed that the baked foods made from the partially dried

okara-free lipoxygenase powder and the commercially low-saturated soybean oil, had a taste, appearance, and texture parallel to the reference product (i.e., commercial Japanese okara-based snack). The final product had 11.4% protein and 7.4% dietary fiber, which was 2.0 and 1.5 times higher in contrast with the standard. The amount of calcium was as well higher (4.3 times) than the reference score. Bedani et al. [57] also recorded an increase in the functional and nutritional properties of soy yogurt incorporated with soybean by-product okara. All these findings suggest the commercialization of okara at the industrial level as a potential nutritionally enhanced value-added food product.

2.2. Application of Soybean Residue in Animal Nutrition

Owing to the higher non-fibrous carbohydrates and proteins content of okara, as well as being cheaper than soybean meal, makes it attractive to be used as feed for dairy cattle, goats, sheep, pigs, poultry, and fish, with the aim of substituting part of their normal feed [58][59][60][61].

The by-products of food crops are generally known to possess high moisture content and, the avoidance of the high energy costs of drying, are hence usually stored by silage, a forage storage technique that involves acidifying the plant mass via anaerobic microorganisms, to aid minimize and deny it from being colonized by other microorganisms that will result to the loss of its nutritional value. A famous practice in Japan, with the aim of preventing nutritional loss, curbs this by mixing dry feed with wet by-products to achieve low moisture total mixed ration silage. The mixture of peanut hulls and okara can therefore have a synergistic influence on the silage mix to form the right dry matter fermentable carbohydrates for ideal silage fermentation. Silage with peanut hulls/okara ratio 22:78 has been evidenced to decrease fiber content and lignification, as well as improve the efficiency of both in-vitro ruminal fermentation and silage fermentation models after 8 weeks^[58].

Meeting the required protein profile in young pigs especially for organic pig producers is a challenge. Thus, the price of organic feed is significantly higher (i.e., 4 times higher), and has limited availability. Okara is suggested as an alternative and a potential source of organic protein, and its intake in up to 25% of young pigs' diets depicted no effect on average daily food intake, average daily profit, and gain/feed ratio in contrast with the control [62]. Wang et al. [61][63] also substituted dried okara for soybean meal in dairy cattle and yellow cattle for a duration of 30 days. The authors recorded no significant differences in the production of milk, feed consumption, milk-fat content, and daily profit between both groups. However, the feeding cost of the group substituted with dried okara was significantly reduced.

In addition, okara can also be used in the production of microbial proteins, which are synthesized via solid-state fermentation. During the process of fermentation, the mold degrades the residual fiber into low molecular weight carbohydrates, which are well along used by yeasts to synthesize proteins. Moreover, some anti-nutritional factors (including, but not limited to, trypsin inhibitors, lectin, and saponin) can be further decomposed or reduced through fermentation [64]. Pan et al. [65] studied the changes in microbial production via mixed culture solid-state fermentation on okara. The authors observed a doubled crude protein content in contrast with the original material when using wheat bran and okara (ratio 2:8) as substrate, and *Trichoderma viride*, *Aspergillus niger*, *Saccharomyces cerevisiae*, and *Candida utilis* (1:1:1:3) as a mixed crop, after fermentation for three days at 32 °C. <u>Table 3</u> and <u>Table 4</u> presents summarized studies on the effect of okara supplementation on nutrition, as well as the impact of its fortification on food properties and function respectively (in vivo/in vitro).

3. Physicochemical and Prebiotic Influence of Dietary Fiber (Emphasis on Okara-Derived Fiber) on Gut and Associated Tissues: Gut, Liver, and Kidneys

Numerous shreds of evidence are indicating that the gut microorganisms can directly influence the physiological conditions in human/animals such as improving the host immune system, improving the roles of the intestinal barrier, stimulating the defensive mechanisms against pathogens as well as increasing the defensive mechanisms against inflammatory bowel diseases, producing biological metabolites, regulating autoimmunity, regulating diabetes and preventing obesity, and destroying cancer cells. The host gut–microbiota interactions are dynamic and highly dictated by several environmental conditions particularly diet [66]. In addition, the gut is reported to have dual and opposing roles of enabling nutrients to enter the body while denying the entry of harmful substances. Both gastrointestinal barrier function and nutrient absorption have been demonstrated to be altered by dietary fiber. For instance, dietary fiber-induced changes to the gastrointestinal barrier by increasing mucins and the cell that produce them "goblet cells" [67][68]. Mucins are large glycoproteins that, along with lipids, antibodies, bacteria, ions, proteins, antimicrobial peptides, and water, form what is termed as mucus [69]. Mucus protects the gut epithelium from mechanical stress, to prevent the translocation of harmful substances as well as to lubricate the intestine and to facilitate easy transportation of digested material. A study comparing a standard rodent chow/diet (i.e., fiber from corn, oats, and wheat comprising 4.3% of the diet by weight) with a

chow devoid of any fiber displayed that mice fed with a fiber-deficient diet had a thinner mucus layer, hence enabling microbes to get in closer proximity to the gastrointestinal epithelium^[70]. Therefore, insufficient amounts of dietary fiber in the gut may encourage bacteria to degrade the host mucus layer (i.e., breaking down one of the host's physical barriers) in an attempt to provide themselves with substrates needed to survive.

3.1. Physicochemical Role of Dietary Fiber in The Gastrointestinal Tract

The main role of the gastrointestinal tract is the absorption of nutrients from ingested foods. This absorption is preceded by a series of digestive processes within the different gut compartments. These processes are governed by the secretion of enzymes and associated co-factors, as well as through maintenance of gut lumen at optimal pH conditions for digestion. Classically, the consumption of dietary fiber has been postulated to affect the uptake of nutrients in diverse methods. The physicochemical factors of dietary fiber, including fermentation, bulking ability, viscosity and gel formation, binding ability, solubility, and water-holding capacity, have been proved to influence nutrient absorption. A plethora of in vivo and in vitro studies have been carried out in the past decades to aid illuminate the physicochemical interactions between dietary fiber and these nutrients However, it appears that water-soluble dietary fiber, which is either viscous or gel-forming under intestinal/gastric conditions reduces the rate of absorption than low molecular weight and low viscous fibers. The small intestine is the main absorptive area in the gut, which, per the dietary stand-point, involves in the absorption of the subunits of digestible macronutrients (i.e., monosaccharides from carbohydrates, amino acids and some di/tripeptides from proteins, and fatty acids/glycerol from di/triglycerides), as well as minerals and vitamins, and other micronutrients [71].

Dietary fiber differs in form depending on their biochemical and physiological properties, and hence affect the bioavailability of nutrients, composition of microbiota and gastrointestinal functions. Soluble dietary fiber portrayed an increased viscosity and reduced starch digestibility $\frac{[72]}{}$, as well as suppressed the activity of α -amylase in the gastrointestinal tract, which consequently reduced the rapid rise in postprandial blood glucose levels [73][74]. The consumption of soluble viscous fibers, together with other dietary changes like reduced fat intake, is effective in reducing cholesterol levels [75]. Sasaki and Kohyama [76] studied the influence of various soluble dietary fiber on starch digestibility. They recorded a strong connection between apparent viscosity at low shear rates and inhibition of starch digestibility. However, insoluble dietary fiber decreased the digestibility of starch by non-specific enzyme adsorption. Nagano et al. [77] systematically reviewed the improved functional and physicochemical properties of okara, by nanocellulose technologies used in food development. The authors hypothesized that nanocellulose technologies may improve physicochemical roles, hence influencing the gut microbiota community. Their study demonstrated an increased viscosity, dispersion ability, as well as the specific surface area of cellulose and okara. The increased viscosity triggered the suppression of αamylase activity whereas increased dispersion ability and specific surface area of okara resulted in improved SCFA production of human dominant gut bacteria [77]. Bifidobacteria and lactobacilli are the most important health-promoting bacteria of the gut microbiota. Hence, both are common targets for dietary interventions to promote health. Other bacteria including enterococci, eubacteria streptococci, and Bacteroides can be classified as potentially harmful or beneficial to health depending on the species [78][79]. The healthy bacteria exert beneficial functions to the host through their metabolisms such as SCFA formation (mainly propionate, acetate, and butyrate), absence of toxin production as well as the synthesis of defensins or vitamins [78]. Desai et al. $\frac{|42|}{|42|}$ researched the consequential impact of dietary fiber deprivation on the gut microbiota using a gnotobiotic mouse model. The authors observed that during intermittent or chronic dietary fiber deficiency, the gut microbiota turns to the host's secreted mucus glycoproteins as a nutrient source, consequently yielding to an erosion of the colonic mucus barrier. Thereby, promoting greater epithelial access and lethal colitis by the mucosal pathogen, Citrobacter rodentium. The study disclosed intricate pathways linking diet, the gut microbiota, and intestinal barrier dysfunction, which could be ameliorated via dietary therapeutics.

3.2. The Use of Soybean Residue as a Prebiotic

The consumption of okara as prebiotics/non-digestible food components can selectively stimulate the composition and/or activity of one or a limited number of gastrointestinal microbes, that can confer health merits to the host [80], as well as non-gastrointestinal related conditions such as cardiovascular disease [81], chronic kidney disease (CDK)[82], non-alcoholic fatty liver disease (NAFLD), and diabetes [83]. The prebiotic influence of okara has been researched in in-vitro studies using *Lactobacillus acidophilus* and *Bifidobacterium bifidum* [84][85]. Okara provided a surface for bacteria cell adhesion, thereby enabling substrate uptake as well as cell growth. The degree of fermentation of the residue was about 3.6 times more in *B. bifidum* compared with *L. acidophilus* [84]. Treatment with β -glucanase (Ultraflo L®) improved the soluble dietary fiber content in okara, subsequently enhancing its fermentation by *B. bifidum* [86]. The conversion of okara insoluble dietary fibers to soluble fibers was again observed when *Streptococcus thermophilus* and *Lactobacillus delbrueckii* subspecies *bulgaricus* were both used [87].

There are numerous studies on the dietary effects of yogurt fortified with soybean residue/okara, produced by lactic fermentation of soymilk and soy milk residue, on lipid and cholesterol levels in rats^[88]. Okara fortified yogurt was prepared by mixing dried okara with soymilk, in a ratio of 1:2, and the mixture was then fermented with *L. delbrueckii* subspecies *delbrueckii*. The formed product was freeze-dried and then incorporated into the rat's diets. Regardless of their diet, rats fed with dried okara and soymilk yogurt portrayed a significant and consistent lower level of total plasmatic cholesterol in contrast with the control group and another group fed with only soymilk yogurt. The authors concluded that the use of okara provided additional merits to soymilk, thus the fiber-rich residue facilitated the excretion of bile acids via their absorption to fecal matter, therefore encouraging the hypocholesterolemic effect^[88].

Furthermore, fermentation is suggested to play a crucial role in the cholesterol-lowering effect as it triggered the production of bioactive peptides through the enzymatic hydrolysis of soy proteins [88]. The DNA microarray analyses result also confirmed that the consumption of soybean residue and soymilk yogurt reduced the synthesis of lipids and cholesterol, and up-regulated the β -oxidation of fatty acids and cholesterol disintegration [89]. In summary, these studies demonstrated that both the supplementation of okara and its subsequent fermentation in a soymilk yogurt matrix depicted hypocholesterolemic effects.

The organoleptic qualities and textural profiles of okara fortified yogurt were then assessed. Soymilk with only dried soybean residue or with insulin and dried okara was fermented with a commercial yogurt starter culture that contained L. acidophilus, Bifidobacterium animalis subspecies lactis, and Streptococcus thermophilus [57]. These yogurts exhibited significantly greater physical stability and were ranked lower in hedonic tests, probably owing to the relatively large size of the dried okara. However, the addition of insulin seems to increase palatability, thus the acceptance score was the greatest for the yogurt with both insulin and okara added.

3.3. Prebiotic Status of Soybean Residue on Gut Microbiome

Research on human gastrointestinal microbiota labeled as the "second brain", or "forgotten organ" has increased exponentially in recent years following the latest advances in technology^[90]. The gut is the core immune organ in the body, which harbors approximately 70–80% of the body's immune cells and has been tagged as the largest source of inflammation evidenced to contribute to diseases like NAFLD and CDK ^{[20][91][92]}. It has been proved that the gut microbiota produces not only metabolites that can impact host physiology, but rather with metabolites that can also contribute an essential role in the host's immune system and metabolism via a complex set of chemical interactions as well as signaling pathways^{[93][94][95]}.

Prebiotics are non-digestible parts of food termed carbohydrates that act as fibers. Unaltered, they reach the colon where they are used by the gastrointestinal microorganisms, they serve as food for "good" intestinal bacteria, and encourage their growth, colonization as well as sustainability in the digestive tract. Among prebiotics or fiber types, are the most important galactooligosaccharides, and oligosaccharides, which are referred to as bifidogenic substances; relating to their ability to selectively enhance the growth of *Bifidobacterium* spp. (*B. breve, B. longum, B. infantis, B. pseudolongum, B. lactis*) and *Lactobacillus* spp. (*L. plantarum, L. casei, L. reuteri, L. acidophilus*)^[96]. The most commonly used prebiotics in human studies include, but are not limited to, Fructooligosaccharide, Galactooligosaccharide, Xylo-oligosaccharide, Arabinoxylan-oligosaccharides, and Soybean oligosaccharides^[96].

Pérez-López et al. [98] studied the prebiotic effect of okara on gastrointestinal microbiota in high-fat-fed rats. To improve the solubility of the okara and achieving a soluble dietary fiber-enriched residue, the authors applied high hydrostatic pressure treatment together with the Ultraflo® L enzyme in the preparation. Wistar rats were fed with a high-fat diet supplemented with 20% treated okara for four weeks. The authors observed a reduction in plasma triglycerides (1.4-times), weight loss, and an enhancement in the metabolism of amino-acid (1.2-times lower urea) in contrast with the control group. Regarding its prebiotic status, okara enhanced the release of short-chain fatty acids (SCFAs) and improved magnesium and calcium absorption. Furthermore, quantitative PCR analysis of selected bacterial groups affirmed that okara protected the values of several beneficial groups in gastrointestinal microbiota and restored dysbiosis caused by a high-fat diet. Thus, preventing bacterial drop triggered by a high-fat diet. The authors concluded that the supplementation of okara exerted health-promoting functions in vivo and hence could be used as a prebiotic as well as a functional ingredient in foods.

Villanueva et al. $\frac{[12]}{}$ as well studied the effect of high-fat diets supplemented with 13% or 20% okara on lipid profiles of the liver, plasma, and feces in male golden Syrian hamsters after three (3) weeks of feeding. The diet did not yield any significant differences in body weight gain or feed intake (p > 0.05). However, plasma levels of triglycerides, total cholesterol, and VLDL- plus LDL cholesterol in hamsters fed with 20% okara decreased significantly (p < 0.05) compared with the control group. Though, no significant differences (p > 0.05) were observed in HDL- and LDL-cholesterol plasma levels in all the experimental groups. Triglycerides, total lipids, total and esterified cholesterol concentrations in the liver

were decreased by 20% okara supplemented diet. Regarding hamsters fed with 13% okara fortified diets, the mean values of the triglyceride, total lipid, and cholesterol in the liver and plasma reduced as compared to the control group, but the differences were not statistically significant. Besides, all assayed okara supplemented diets depicted increased fecal excretion of total lipids, free cholesterol, triglycerides, and total nitrogen (p < 0.05) in contrast with their respective controls. The authors suggested that the main components of okara, i.e., dietary fiber and protein, could be attributed with the total lipids and cholesterol decreased in the liver and plasma, and the fecal output increase in high-fat-fed hamsters, and hence might play an essential role in the prevention of hyperlipidemia and could as well be used as value-added ingredient for functional food preparation.

In 2016, Villanueva et al. studied the potential prebiotic and fat-lowering effects of enzymatically treated okara in high-cholesterol-fed Wistar rats^[6]. The authors observed a significant reduction in the serum and liver triglyceride levels (p < 0.01) of rats fed with enzymatically treated okara. Total lipids, bile acids, and triglycerides were significantly (p < 0.001) higher in the feces of rats fed with enzymatically treated okara diet. However, the pH of fecal contents was reduced (p < 0.001), probably owing to the significant increase in the production of short-chain fatty acids in the treated group compared to the control group (i.e., Total SCFA (mmol/g) of 229.2 \pm 37.0 for the control group and 568.2 \pm 56.5 for the treated group). In addition, the enzymatically treated okara yielded reduced triglycerides in liver and serum, and improved the excretion of total lipids, bile acids, and triglycerides, increasing the lipid profile in high cholesterol-fed rats. The fiber content is suggested to aid improve intestinal transit via increasing fecal bulk. The authors concluded that the reduced pH and improved SCFA production affirmed the occurrence of fiber fermentation, hence signifying a potential prebiotic effect.

3.4. Gut, Liver, and Kidney Responses to the Prebiotic Effects of Dietary Fiber in General

In the microbial process of prebiotic fermentation in the large intestine, SCFAs such as butyrate, propionic acid, acetic acid, vitamin K, and vitamin B12 are formed, which are then absorbed by the gastrointestinal mucosa and distributed via the lymphatic and vascular system to cells of an organism^[99]. The resulting butyrate is metabolized directly in the gastrointestinal epithelium, where it acts as a regulator of cell division and growth. Propionate is used in the liver and as well serves as a precursor used for suppressing cholesterol synthesis. Acetate is predominantly metabolized in the muscle cells, heart, kidneys, and brain. The production of SCFA decreases the pH of the environment, hence encouraging cell differentiation and growth of intestinal epithelial cells as well as re-supporting the microflora. In addition, fermentation in the intestine is suggested to produce the final form of the degradation of substances like simple gases: Carbon dioxide, hydrogen, methane, and hydrogen sulfide $^{[96]}$. Figure 1 presents the schematic overview of the major prebiotic effects of dietary fiber on the gut, liver, and kidney $^{[20]}$.

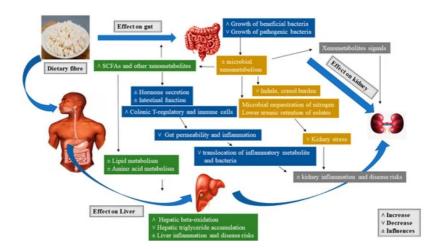


Figure 1. Schematic overview of the major prebiotic effects of dietary fiber on gut, liver, and kidney.

3.4.1. Gut Responses

The increase in the production of short-chain fatty acids (SCFAs) resulting from the fermentation of dietary fiber has been shown to boost gut barrier function via increasing gut cell proliferation and diversity $\frac{[20][100]}{[20][100]}$. SCFAs decrease gastrointestinal pH, which may alter the gut microbiota by obstructing the growth of pathogens and decrease the expression of microbial virulence genes $\frac{[101]}{[102]}$. Again, it has been evidenced that epithelial cell lines metabolize the SCFA butyrate, yielding oxygen reduction that results in the stabilization of transcription factor, hypoxia-inducible factor-1 alpha (Hif-1 α) $\frac{[102]}{[102]}$. This transcription factor has been implicated in gastrointestinal barrier function via regulating apoptosis $\frac{[103]}{[103]}$ and inflammation $\frac{[104]}{[102]}$. A microarray study confirms that high-amylose resistant starch could improve hormones as well as structure and function of the gut via increasing the levels of Hif-1 α expression together with genes related to cell growth, differentiation, apoptosis, and proliferation in the cecal tissue of rats supplemented with 30% resistant starch in contrast

with rats fed an equal amount of energy from a low-fiber diet [105]. A recent study by Zhang et al. [106] evaluated the effects of prebiotics on bioavailability and biotransformation of ginsenosides in rats after a two-week prebiotic (i.e., galactooligosaccharide, fructooligosaccharide, and fibersol-2) intervention. The results displayed that area under concentration-time curve and peak plasma concentration of ginsenoside and its intermediate metabolites in the prebiotic intervention groups were improved at various degrees than those in the control group. In addition, the gut microbiota significantly responded to the prebiotic treatment at both functional and taxonomical levels. The gut metagenomic analysis as well unveiled the functional gene amelioration for polyketide/terpenoid metabolism, gluconeogenesis, glycolysis, and propanoate metabolism, etc. The authors concluded that prebiotics may selectively endorse the proliferation of certain bacterial stains having glycoside hydrolysis capacity, hence, subsequently encouraging bioavailability and biotransformation of primary ginsenosides in vivo.

The tight junction proteins are another key component of the gut barrier affected by dietary fiber. A study by Cani et al. [107] observed that feeding a standard rodent chow supplemented with (10%) fructooligosaccharides improved the expression of genes in the jejunal tight junction proteins and Zonula occludens-1 (ZO-1), decreased intestinal permeability, and reduced plasma lipopolysaccharides (LPS) concentrations. Again, Fukunaga et al. [108] studied the effect of soluble fiber pectin on the production of fecal short-chain fatty acid, intestinal cell proliferation, and microbial population. The results showed that rats fed 2.5% pectin for two weeks displayed increased plasma glucagon-like peptide-2 (GLP-2) and increased cecal SCFAs. Butyrate, in another Caco-2 cell culture model, was again demonstrated to activate AMPactivated protein kinase, enabling tight junction protein assembly as well as improved barrier function shown via improved transepithelial electrical resistance $\frac{[109]}{}$. Ohata et al. $\frac{[110]}{}$, also used the Caco-2 cells culture model and observed that butyrate improved lipoxygenase activity by hindering histone deacetylation, which caused transepithelial electrical resistance to increase. Apart from the alteration of intestinal immune function and physical barriers to minimize injury from microbe-derived proinflammatory factors, prebiotic can as well protect organs like the kidney and liver from metabolic abuses. It has long been evidenced that constant consumption of nondigestible carbohydrates instead of digestible carbohydrates, minimizes the increase in blood glucose and insulin^[20]. Mithieux et al.^[111] researched intestinal gluconeogenesis, which is another carbohydrate regulatory pathway influenced by dietary fiber consumption. The gastrointestinal production of glucose is understood to improve glucose sensing in the portal vein, resulting in reductions of hepatic glucose production as well as influenced signaling to the brain, yielding an increase in satiety. However, microbiota-generated metabolites of prebiotic (e.g., propionate), are suggested to act as a gluconeogenic precursor [112]. In summary, this highlights the merits of microbe-derived SCFAs in supporting the physical components of the gut barrier (mucosal layer, tight junctions, and cellularity) and influencing host immune factors, which will consequentially impact functions of tissues such as the liver and kidneys.

3.4.2. Liver Responses

Indeed, it is known that blood from the gut reaches the liver via the portal vein, and hence is justifiable that this organ is a target of gut-derived factors altered by diet and microbiome changes. However, prebiotic fiber is being considered an ideal treatment and management option for NAFLD and related obesity and insulin resistance[21]. The hepatic effects of prebiotic fiber as discussed earlier include influencing gut ecology, thus gut permeability, systemic inflammation, as well as circulating gastrointestinal-derived hormone and metabolite signals. Endorsing the association between gut health and liver, patients with NAFLD were seen to portray an altered gut microbiota^[91] and improved gut permeability^[113]. Dietary fibers have been demonstrated to hinder the translocation of bacterial products such as LPS[114]. Thus, this will aid to decrease hepatic exposure to LPS and other associated microbe-derived proinflammatory products. This may hinder the tendency of fatty liver progressing to the inflammatory form termed as non-alcoholic steatohepatitis (NASH)[20]. This transition is thought to occur in two stages known as the "two-hit" hypothesis. The "first hit" is the state where the liver is susceptible to metabolic insults or the "second hit", owing to liver fat accumulation. The "second hit" is assumed to come from a variety of sources, such as bacterial overgrowth, and is suggested to induces hepatic inflammation [115]. Cortez-Pinto et al. [116] evaluated the effect of different dietary patterns in NASH patients. The study revealed that these patients consumed low fiber, more fat, and less carbohydrate than the healthy controls. In addition, another study discussed the therapeutic effects of whole grains derived dietary fiber and demonstrated its merits associated with metabolic syndrome (MtS) and NAFLD after consumption. Besides the reduction of liver fat, whole grains derived dietary fiber could as well reduce inflammation $\frac{[117][118][119]}{}$.

The proposed effects of prebiotic fibers on lipid metabolism are attributed to inhibition of de novo fatty acid synthesis (FAS) [120][121][122]. Prebiotic fibers are thought to downregulate hepatic lipogenic enzymes, mainly FAS, by increasing the production of SCFA propionate [123]. Again, due to their unique modes of action regarding cholesterol metabolism specifically linked with their fermentability and modulation of the microflora. As reported earlier, SCFA has been implicated in cholesterol metabolism. Levrat et al. [124] studied the role of dietary propionic acid on hypocholesterolemic rats. Inulin supplementation yielded an increase in caecum SCFA compared to the control diet. However, the concentrations of SCFA

measured in the portal vein indicate that the liver is exposed to high concentrations of SCFA, specifically propionic acid. The authors concluded that the presence of propionic acid in the liver reduces cholesterolemic responses. Another study reported the effect of prebiotic fiber on serum lipids and hepatic gene expression in JCR: LA-cp rats. The authors reported that prebiotic fibers caused a decrease in cholesterol levels by improved cholesterol excretion in the form of bile as well as hinder liver triacylglycerol accumulation. The authors recommended the use of prebiotic fibers as dietary treatments for hypercholesterolemia [122].

3.4.3. Kidney Responses

Apart from the liver, the kidney is another essential organ affected by a significant intake of dietary fiber. Dietary fiber has been proven in numerous reports to decrease nitrogen burden as well as systemic inflammatory abuse in CKD. Similar to NAFLD, in vivo CKD experiment often depict an altered gut microbiota^{[125][126]}, intestinal inflammation, increased intestinal permeability as well as an increment in plasma concentrations of microbe- derived metabolites (e.g., p-cresol sulfate and indoxyl sulfate) ^{[20][127][128]}. It has been shown by epidemiological research that a sufficient intake of dietary fiber decreases all potential causes of mortality in patients diagnosed with CKD^[129]. Although the mechanism is not fully understood, it is however attributed to the role of dietary fiber in maintaining substrate delivery to the lower gut, which in the long run modifies bacterial metabolism. Certainly, if a sufficient amount of nondigestible carbohydrates is unable to reach the colon, then substrates like amino acids will be fermented, leading to the formation of potentially harmful metabolites like p-cresol and indoles, which will consequentially stress the kidney^[130].

A known mechanism highlighting the effects of dietary fiber on the kidney include the reduction in nitrogen load on both liver and kidneys via increasing microbial biomass. The microbial generated biomass aids to sequester nitrogen in the gut and decrease the amount that reaches the portal circulation[20][131]. In 2019, Adams et al. studied the effects of dietary supplementation of prebiotic fiber on nitrogen metabolism, intestinal morphometry, and blood biochemistry in weaning piglets. The inclusion resulted in an increase in nitrogen metabolism by increasing the digestibility and utilization of nitrogen as well as decreasing fecal and urinary content of nitrogen. Again, Mardinoglu et al.[132] compared the metabolism of amino acids in conventional and germ-free mice, and discovered that the concentrations of amino acids entering the liver portal vein in the conventional mice were low; the reduction in the portal amino acids was associated with increased demand in nitrogen for microbial synthesis [133]. The formation of SCFAs is another proposed mechanism by which dietary fiber can impact kidney function. Thus, SCFAs are suggested to influence kidney blood flow by stimulating olfactory receptor 78 (Olfr-78), a G-protein-coupled receptor found in the renal juxtaglomerular apparatus that encourages renin secretion, which is responsible for regulating blood pressure [134]. Blood pressure regulation is an essential component in managing the progression of CKD[135]. In summary, dietary fiber is suggested to improve kidney function as well as decrease the occurrence of CKD via causing shifts in the microbiome, which will consequentially maintain and improve the gut barrier, altering the metabolism of uremic solute and microbial nitrogen, as well as controlling renal blood flow.

References

- 1. Colletti, A.; Attrovio, A.; Boffa, L.; Mantegna, S.; Cravotto, G. Valorisation of By-Products from Soybean (Glycine max (L.) Merr.) Processing. Molecules 2020, 25, 2129.
- 2. Nishinari, K.; Fang, Y.; Guo, S.; Phillips, G. Soy proteins: A review on composition, aggregation and emulsification. Food Hydrocoll. 2014, 39, 301–318.
- 3. Kerwin, S. Soy saponins and the anticancer effects of soybeans and soy-based foods. Curr. Med. Chem. Anti-Cancer Agents 2004, 4, 263–272.
- 4. Hwang, Y.W.; Kim, S.Y.; Jee, S.H.; Kim, Y.N.; Nam, C.M. Soy food consumption and risk of prostate cancer: A meta-analysis of observational studies. Nutr. Cancer 2009, 61, 598–606.
- 5. Messina, M.; Watanabe, S.; Setchell, K.D. Report on the 8th international symposium on the role of soy in health promotion and chronic disease prevention and treatment. J. Nutr. 2009, 139, 796S–802S.
- Villanueva-Suárez, M.-J.; Pérez-Cózar, M.-L.; Mateos-Aparicio, I.; Redondo-Cuenca, A. Potential fat-lowering and prebiotic effects of enzymatically treated okara in high-cholesterol-fed Wistar rats. Int. J. Food Sci. Nutr. 2016, 67, 828–833.
- 7. O'Toole, D.K. Characteristics and use of okara, the soybean residue from soy milk production a review. J. Agric. Food Chem. 1999, 47, 363–371.
- 8. Jiménez-Escrig, A.; Tenorio, M.D.; Espinosa-Martos, I.; Rupérez, P. Health-promoting effects of a dietary fiber concentrate from the soybean byproduct okara in rats. J. Agric. Food Chem. 2008, 56, 7495–7501.

- 9. Singh, B.; Singh, J.P.; Singh, N.; Kaur, A. Saponins in pulses and their health promoting activities: A review. Food Chem. 2017, 233, 540–549.
- 10. Lu, F.; Liu, Y.; Li, B. Okara dietary fiber and hypoglycemic effect of okara foods. Bioact. Carbohydr. Diet. Fibre 2013, 2, 126–132.
- 11. Zhong-Hua, L.; Hong-Lian, G.; Rui-Ling, L.; Jin-Hui, Z. Extraction and Antioxidant Activity of Soybean Saponins from Lowtemperature Soybean Meal by MTEH. Open Biotechnol. J. 2015, 9,178–184. doi: 10.2174/1874070701509010178.
- 12. Villanueva, M.; Yokoyama, W.; Hong, Y.; Barttley, G.; Rupérez, P. Effect of high-fat diets supplemented with okara soybean by-product on lipid profiles of plasma, liver and faeces in Syrian hamsters. Food Chem. 2011, 124, 72–79.
- 13. Hwang, D.; Charchoghlyan, H.; Lee, J.S.; Kim, M. Bioactive compounds and antioxidant activities of the Korean lotus leaf (Nelumbo nucifera) condiment: Volatile and nonvolatile metabolite profiling during fermentation. Int. J. Food Sci. Technol. 2015, 50, 1988–1995.
- 14. Hu, Y.; Piao, C.; Chen, Y.; Zhou, Y.; Wang, D.; Yu, H.; Xu, B. Soybean residue (okara) fermentation with the yeast Kluyveromyces marxianus. Food Biosci. 2019, 31, 100439.
- 15. Kang, M.J.; Bae, I.Y.; Lee, H.G. Rice noodle enriched with okara: Cooking property, texture, and in vitro starch digestibility. Food Biosci. 2018, 22, 178–183.
- 16. Nagai, T.; Te Li, L.; Ma, Y.L.; Sarkar, P.K.; Nout, R.; Park, K.Y.; Jeong, J.K.; Lee, J.E.; Im Lee, G.; Lee, C.H. Diversity of plant-based food products involving alkaline fermentation. In Handbook Indigenous Foods Involving Alkaline Fermentation; CRC Press: Boca Raton, FL, USA,2014; pp. 7–187.
- 17. Chan, L.Y.; Takahashi, M.; Lim, P.J.; Aoyama, S.; Makino, S.; Ferdinandus, F.; Ng, S.Y.C.; Arai, S.; Fujita, H.; Tan, H.C. Eurotium cristatum fermented okara as a potential food ingredient to combat diabetes. Sci. Rep. 2019, 9, 1–9.
- 18. Rastall, R.A.; Gibson, G.R. Recent developments in prebiotics to selectively impact beneficial microbes and promote intestinal health. Curr. Opin. Biotechnol. 2015, 32, 42–46.
- 19. Abdallah, A.; Zhang, P.; Zhong, Q.; Sun, Z., Application of traditional Chinese herbal medicine by-products as dietary feed supplements and antibiotic replacements in animal production. Curr. Drug Metab. 2019, 20 (1), 54-64..
- 20. Kieffer, D.A.; Martin, R.J.; Adams, S.H. Impact of dietary fibers on nutrient management and detoxification organs: Gut, liver, and kidneys. Adv. Nutr. 2016, 7, 1111–1121.
- 21. Parnell, J.A.; Raman, M.; Rioux, K.P.; Reimer, R.A. The potential role of prebiotic fibre for treatment and management of non-alcoholic fatty liver disease and associated obesity and insulin resistance. Liver Int. 2012, 32, 701–711.
- 22. Rossi, M.; Johnson, D.W.; Campbell, K.L. The kidney–Gut axis: Implications for nutrition care. J. Ren. Nutr. 2015, 25, 399–403.
- 23. Compare, D.; Coccoli, P.; Rocco, A.; Nardone, O.; De Maria, S.; Cartenì, M.; Nardone, G. Gut–liver axis: The impact of gut microbiota on non alcoholic fatty liver disease. Nutr. Metab. Cardiovasc. Dis. 2012, 22, 471–476.
- 24. Adams, S.; Che, D.; Hailong, J.; Zhao, B.; Rui, H.; Danquah, K.; Qin, G. Effects of pulverized oyster mushroom (Pleurotus ostreatus) on diarrhea incidence, growth performance, immunity, and microbial composition in piglets. J. Sci. Food Agric. 2019, 99, 3616–3627.
- 25. Brennan, M.A.; Monro, J.A.; Brennan, C.S. Effect of inclusion of soluble and insoluble fibres into extruded breakfast cereal products made with reverse screw configuration. Int. J. Food Sci. Technol. 2008, 43, 2278–2288.
- 26. Symons, L.; Brennan, C. The effect of barley β -glucan fiber fractions on starch gelatinization and pasting characteristics. J. Food Sci. 2004, 69, FCT257–FCT261.
- 27. Brennan, C.S. Dietary fibre, glycaemic response, and diabetes. Mol. Nutr. Food Res. 2005, 49, 560–570.
- 28. Lunn, J.; Buttriss, J. Carbohydrates and dietary fibre. Nutr. Bull. 2007, 32, 21-64.
- 29. Chhabra, S. Dietary Fibre-Nutrition and Health Benefits. In Functional Food and Human Health; Springer, Singapore, 2018; pp. 15–25.
- 30. Li, Q.; Yang, S.; Li, Y.; Huang, Y.; Zhang, J. Antioxidant activity of free and hydrolyzed phenolic compounds in soluble and insoluble dietary fibres derived from hulless barley. LWT 2019, 111, 534–540.
- 31. Dhingra, D.; Michael, M.; Rajput, H.; Patil, R. Dietary fibre in foods: A review. J. Food Sci. Technol. 2012, 49, 255-266.
- 32. Chawla, R.; Patil, G. Soluble dietary fiber. Compr. Rev. Food Sci. Food Saf. 2010, 9, 178-196.
- 33. Nsor-Atindana, J.; Zhong, F.; Mothibe, K.J. In vitro hypoglycemic and cholesterol lowering effects of dietary fiber prepared from cocoa (Theobroma cacao L.) shells. Food Funct. 2012, 3, 1044–1050.

- 34. Stewart, M.L.; Schroeder, N.M. Dietary treatments for childhood constipation: Efficacy of dietary fiber and whole grains. Nutr. Rev. 2013, 71, 98–109.
- 35. De Moraes Crizel, T.; Jablonski, A.; de Oliveira Rios, A.; Rech, R.; Flôres, S.H. Dietary fiber from orange byproducts as a potential fat replacer. LWT-Food Sci. Technol. 2013, 53, 9–14.
- 36. Bikker, P.; Dirkzwager, A.; Fledderus, J.; Trevisi, P.; Le Huërou-Luron, I.; Lallès, J.P.; Awati, A. The effect of dietary protein and fermentable carbohydrates levels on growth performance and intestinal characteristics in newly weaned piglets. J. Anim. Sci. 2006, 84, 3337–3345.
- 37. Adams, S.; Che, D.; Qin, G.; Rui, H.; Sello, C.T.; Hailong, J. Interactions of dietary fibre with nutritional components on gut microbial composition, function and health in monogastrics. Curr. Protein Pept. Sci. 2018, 19, 1011–1023.
- 38. Adams, S.; Kong, X.; Che, D.; Qin, G.; Jiang, H. Effects of dietary supplementation of alfalfa (Medicago sativa) fibre on the blood biochemistry, nitrogen metabolism, and intestinal morphometry in weaning piglets. Appl. Ecol. Environ. Res. 2019, 17, 2275–2295.
- 39. Adams, S.; Sello, C.T.; Qin, G.-X.; Che, D.; Han, R. Does dietary fiber affect the levels of nutritional components after feed formulation? Fibers 2018, 6, 29.
- 40. Napolitano, A.; Costabile, A.; Martin-Pelaez, S.; Vitaglione, P.; Klinder, A.; Gibson, G.R.; Fogliano, V. Potential prebiotic activity of oligosaccharides obtained by enzymatic conversion of durum wheat insoluble dietary fibre into soluble dietary fibre. Nutr. Metab. Cardiovasc. Dis. 2009, 19, 283–290.
- 41. Makki, K.; Deehan, E.C.; Walter, J.; Bäckhed, F. The impact of dietary fiber on gut microbiota in host health and disease. Cell Host Microbe 2018, 23, 705–715.
- 42. Desai, M.S.; Seekatz, A.M.; Koropatkin, N.M.; Kamada, N.; Hickey, C.A.; Wolter, M.; Pudlo, N.A.; Kitamoto, S.; Terrapon, N.; Muller, A. A dietary fiber-deprived gut microbiota degrades the colonic mucus barrier and enhances pathogen susceptibility. Cell 2016, 167, 1339–1353.e21.
- 43. Genovese, M.I.; Lajolo, F.M. Isoflavones in soy-based foods consumed in Brazil: Levels, distribution, and estimated intake. J. Agric. Food Chem. 2002, 50, 5987–5993.
- 44. Li, B.; Qiao, M.; Lu, F. Composition, nutrition, and utilization of okara (soybean residue). Food Rev. Int. 2012, 28, 231–252.
- 45. Vong, W.C.; Lim, X.Y.; Liu, S.-Q. Biotransformation with cellulase, hemicellulase and Yarrowia lipolytica boosts health benefits of okara. Appl. Microbiol. Biotechnol. 2017, 101, 7129–7140.
- 46. Pan, W.-C.; Liu, Y.-M.; Shiau, S.-Y. Effect of okara and vital gluten on physico-chemical properties of noodle. Czech J. Food Sci. 2018, 36, 301–306.
- 47. Kamble, D.B.; Singh, R.; Rani, S.; Pratap, D. Physicochemical properties, in vitro digestibility and structural attributes of okara-enriched functional pasta. J. Food Process. Preserv. 2019, 43, e14232.
- 48. Wu, S. Preparation of bean dregs cake. Food Ind. 2003, 24, 23-24.
- 49. Zhao, G.-L.; Kong, J. Enzymolysis bean dregs biscuit development. Food Res. Dev. 2009, 10, 67–69.
- 50. Mateos-Aparicio, I.; Redondo-Cuenca, A.; Villanueva-Suárez, M. Isolation and characterisation of cell wall polysaccharides from legume by-products: Okara (soymilk residue), pea pod and broad bean pod. Food Chem. 2010, 122, 339–345.
- 51. Park, J.; Choi, I.; Kim, Y. Cookies formulated from fresh okara using starch, soy flour and hydroxypropyl methylcellulose have high quality and nutritional value. LWT-Food Sci. Technol. 2015, 63, 660–666.
- 52. Suda, T.; Kido, Y.; Tsutsui, S.; Tsutsui, D.; Fujita, M.; Nakaya, Y. Nutritional evaluation of the new OKARA powder for food processing material. Foods Food Ingred. J. Jpn. 2007, 212, 320.
- 53. Yao, X.; Song, W.; Zhang, Y.; Xiao, W. On the application of enzyme on preparations in bread containing soybeand. Cereal Feed Ind. 2006, 11, 22–23.
- 54. Wang, Y.; Tang, J. Application of soybean fiber on bread. J. Zhengzhou Inst. Technol. 2000, 21, 75–77.
- 55. Sun, X.; Yang, Y. Study on cooking quality of noodle of okara fiber. Grain Process 2010, 1, 57–59.
- 56. Katayama, M.; Wilson, L.A. Utilization of okara, a byproduct from soymilk production, through the development of soybased snack food. J. Food Sci. 2008, 73, S152–S157.
- 57. Bedani, R.; Campos, M.M.; Castro, I.A.; Rossi, E.A.; Saad, S.M. Incorporation of soybean by-product okara and inulin in a probiotic soy yoghurt: Texture profile and sensory acceptance. J. Sci. Food Agric. 2014, 94, 119–125.
- 58. Yang, C.-M. Soybean milk residue ensiled with peanut hulls: Fermentation acids, cell wall composition, and silage utilization by mixed ruminal microorganisms. Bioresour. Technol. 2005, 96, 1419–1424.

- 59. Wong, M.H.; Tang, L.; Kwok, F. The use of enzyme-digested soybean residue for feeding common carp. Biomed. Environ. Sci. 2003, 9, 418–423.
- 60. Yang, C.; Gu, J. Study on the active okara for feeding egg chicken. Feed Ind. 1997, 18, 26-27.
- 61. Wang, Z.; Jiang, W.; Hu, Z.; Wang, L. Feeding effects on finishing cattle by substituting soybean meal by dry bean curd pulp. J. Yellow Cattle Sci. 2004, 30, 15–17.
- 62. Hermann, J.; Honeyman, M.S. Okara: A possible high protein feedstuff for organic pig diets. Anim. Ind. Rep. 2004, 650, 601–604.
- 63. Wang, Z.-H.; Wang, L.-Z.; Chen, Y.-S.; Wu, Z.-F. Contrast trial of substituting dried tofu pulp for soybean meal in dairy diet. China Dairy Cattle 2003, 2, 24–26.
- 64. Junyi, Q.; Fuyuan, Z. Research on the processing of microbial protein feed by mixed culture solid-state fermentation on soybean waste. Feed Ind. 2008, 22, 21–24.
- 65. Pan, T.; Zhang, D.; Zhao, C.; Li, K. Study on microbial protein production by mixed culture solid state fermentation on soybean waste. Chem. Bioeng. 2004, 6, 35–41.
- 66. Adams, S.; Xiangjie, K.; Hailong, J.; Guixin, Q.; Sossah, F.L.; Dongsheng, C. Prebiotic effects of alfalfa (Medicago sativa) fiber on cecal bacterial composition, short-chain fatty acids, and diarrhea incidence in weaning piglets. RSC Adv. 2019, 9, 13586–13599.
- 67. Satchithanandam, S.; Vargofcak-Apker, M.; Calvert, R.J.; Leeds, A.R.; Cassidy, M.M. Alteration of gastrointestinal mucin by fiber feeding in rats. J. Nutr. 1990, 120, 1179–1184.
- 68. Piel, C.; Montagne, L.; Sève, B.; Lallès, J.-P. Increasing digesta viscosity using carboxymethylcellulose in weaned piglets stimulates ileal goblet cell numbers and maturation. J. Nutr. 2005, 135, 86–91.
- 69. Lamont, J.T. Mucus: The front line of intestinal mucosal defense. Ann. N. Y. Acad. Sci. 1992, 664, 190-201.
- 70. Earle, K.A.; Billings, G.; Sigal, M.; Lichtman, J.S.; Hansson, G.C.; Elias, J.E.; Amieva, M.R.; Huang, K.C.; Sonnenburg, J.L. Quantitative imaging of gut microbiota spatial organization. Cell Host Microbe 2015, 18, 478–488.
- 71. Brownlee, I.A. The physiological roles of dietary fibre. Food Hydrocoll. 2011, 25, 238-250.
- 72. Slaughter, S.L.; Ellis, P.R.; Jackson, E.C.; Butterworth, P.J. The effect of guar galactomannan and water availability during hydrothermal processing on the hydrolysis of starch catalysed by pancreatic α-amylase. Biochim. Iophysica Acta Gen. Subj. 2002, 1571, 55–63.
- 73. Dona, A.C.; Pages, G.; Gilbert, R.G.; Kuchel, P.W. Digestion of starch: In vivo and in vitro kinetic models used to characterise oligosaccharide or glucose release. Carbohydr. Polym. 2010, 80, 599–617.
- 74. Grundy, M.M.-L.; Edwards, C.H.; Mackie, A.R.; Gidley, M.J.; Butterworth, P.J.; Ellis, P.R. Re-evaluation of the mechanisms of dietary fibre and implications for macronutrient bioaccessibility, digestion and postprandial metabolism. Br. J. Nutr. 2016, 116, 816–833.
- 75. Gray, J. Dietary Fibre: Definition, Analysis, Physiology Health; International Life Sciences Institute (ILSI), Brussels, 2006, p. 36.
- 76. Sasaki, T.; Kohyama, K. Influence of non-starch polysaccharides on the in vitro digestibility and viscosity of starch suspensions. Food Chem. 2012, 133, 1420–1426.
- 77. Nagano, T.; Arai, Y.; Yano, H.; Aoki, T.; Kurihara, S.; Hirano, R.; Nishinari, K. Improved physicochemical and functional properties of okara, a soybean residue, by nanocellulose technologies for food development–A review. Food Hydrocoll. 2020, 109, 105964.
- 78. Pérez-López, E.; Cela, D.; Costabile, A.; Mateos-Aparicio, I.; Rupérez, P. In vitro fermentability and prebiotic potential of soyabean Okara by human faecal microbiota. Br. J. Nutr. 2016, 116, 1116–1124.
- 79. Roberfroid, M.; Gibson, G.R.; Hoyles, L.; McCartney, A.L.; Rastall, R.; Rowland, I.; Wolvers, D.; Watzl, B.; Szajewska, H.; Stahl, B. Prebiotic effects: Metabolic and health benefits. Br. J. Nutr. 2010, 104, S1–S63.
- 80. Saad, N.; Delattre, C.; Urdaci, M.; Schmitter, J.-M.; Bressollier, P. An overview of the last advances in probiotic and prebiotic field. LWT-Food Sci. Technol. 2013, 50, 1–16.
- 81. Threapleton, D.E.; Greenwood, D.C.; Evans, C.E.; Cleghorn, C.L.; Nykjaer, C.; Woodhead, C.; Cade, J.E.; Gale, C.P.; Burley, V.J. Dietary fibre intake and risk of cardiovascular disease: Systematic review and meta-analysis. BMJ 2013, 347, f6879.
- 82. Bliss, D.Z.; Stein, T.P.; Schleifer, C.R.; Settle, R.G. Supplementation with gum arabic fiber increases fecal nitrogen excretion and lowers serum urea nitrogen concentration in chronic renal failure patients consuming a low-protein diet. Am. J. Clin. Nutr. 1996, 63, 392–398.

- 83. Johnston, K.; Thomas, E.L.; Bell, J.D.; Frost, G.; Robertson, M.D. Resistant starch improves insulin sensitivity in metabolic syndrome. Diabet. Med. 2009, 27, 391–397.
- 84. Espinosa-Martos, I.; Rupérez, P. Indigestible fraction of okara from soybean: Composition, physicochemical properties and in vitro fermentability by pure cultures of Lactobacillus acidophilus and Bifidobacterium bifidum. Eur. Food Res. Technol. 2009, 228, 685–693.
- 85. Bedani, R.; Rossi, E.A.; Saad, S.M.I. Impact of inulin and okara on Lactobacillus acidophilus La-5 and Bifidobacterium animalis Bb-12 viability in a fermented soy product and probiotic survival under in vitro simulated gastrointestinal conditions. Food Microbiol. 2013, 34, 382–389.
- 86. Villanueva-Suárez, M.J.; Pérez-Cózar, M.L.; Redondo-Cuenca, A. Sequential extraction of polysaccharides from enzymatically hydrolyzed okara byproduct: Physicochemical properties and in vitro fermentability. Food Chem. 2013, 141, 1114–1119.
- 87. Tu, Z.; Chen, L.; Wang, H.; Ruan, C.; Zhang, L.; Kou, Y. Effect of fermentation and dynamic high pressure microfluidization on dietary fibre of soybean residue. J. Food Sci. Technol. 2014, 51, 3285–3292.
- 88. Kitawaki, R.; Takagi, N.; Iwasaki, M.; Asao, H.; Okada, S.; Fukuda, M. Plasma cholesterol-lowering effects of soymilk and okara treated by lactic acid fermentation in rats. J. Jpn. Soc. Food Sci. Technol. 2007, 54, 379–382
- 89. Kitawaki, R.; Nishimura, Y.; Takagi, N.; Iwasaki, M.; Tsuzuki, K.; Fukuda, M. Effects of Lactobacillus fermented soymilk and soy yogurt on hepatic lipid accumulation in rats fed a cholesterol-free diet. Biosci. Biotechnol. Biochem. 2009, 73, 1484–1488.
- 90. O'Hara, A.M.; Shanahan, F. The gut flora as a forgotten organ. Embo Rep. 2006, 7, 688-693.
- 91. Alisi, A.; Ceccarelli, S.; Panera, N.; Nobili, V. Causative role of gut microbiota in non-alcoholic fatty liver disease pathogenesis. Front. Cell. Infect. Microbiol. 2012, 2, 132.
- 92. Brun, P.; Castagliuolo, I.; Leo, V.D.; Buda, A.; Pinzani, M.; Palù, G.; Martines, D. Increased intestinal permeability in obese mice: New evidence in the pathogenesis of nonalcoholic steatohepatitis. Am. J. Physiol. Gastrointest. Liver Physiol. 2007, 292, G518–G525.
- 93. Clemente, J.C.; Ursell, L.K.; Parfrey, L.W.; Knight, R. The impact of the gut microbiota on human health: An integrative view. Cell 2012, 148, 1258–1270.
- 94. Hooper, L.V.; Littman, D.R.; Macpherson, A.J. Interactions between the microbiota and the immune system. Science 2012, 336, 1268–1273.
- 95. Nicholson, J.K.; Holmes, E.; Kinross, J.; Burcelin, R.; Gibson, G.; Jia, W.; Pettersson, S. Host-gut microbiota metabolic interactions. Science 2012, 336, 1262–1267.
- 96. Hijová, E.; Bertková, I.; Štofilová, J. Dietary fibre as prebiotics in nutrition. Cent. Eur. J. Public Health 2019, 27, 251–255
- 97. Livingston, K.A.; Sawicki, C.M.; McKeown, N.M.; Obin, M.; Roberts, S.B.; Chung, M. Dietary Fiber and the Human Gut Microbiota: Application of Evidence Mapping Methodology. Nutrients 2017, 9, 125.
- 98. Pérez-López, E.; Veses, A.; Redondo, N.; Tenorio-Sanz, M.; Villanueva, M.; Redondo-Cuenca, A.; Marcos, A.; Nova, E.; Mateos-Aparicio, I.; Rupérez, P. Soybean Okara modulates gut microbiota in rats fed a high-fat diet. Bioact. Carbohydr. Diet. Fibre 2018, 16, 100–107.
- 99. Hijova, E.; Chmelarova, A. Short chain fatty acids and colonic health. Bratisl. Lek. Listy 2007, 108, 354.
- 100. Goodlad, R.; Ratcliffe, B.; Fordham, J.; Wright, N. Does dietary fibre stimulate intestinal epithelial cell proliferation in germ free rats? Gut 1989, 30, 820–825.
- 101. Sun, Y.; O'Riordan, M.X. Regulation of bacterial pathogenesis by intestinal short-chain fatty acids. Adv. Appl. Microbiol. 2013, 85, 93–118.
- 102. Kelly, C.J.; Zheng, L.; Campbell, E.L.; Saeedi, B.; Scholz, C.C.; Bayless, A.J.; Wilson, K.E.; Glover, L.E.; Kominsky, D.J.; Magnuson, A. Crosstalk between microbiota-derived short-chain fatty acids and intestinal epithelial HIF augments tissue barrier function. Cell Host Microbe 2015, 17, 662–671.
- 103. Tambuwala, M.M.; Cummins, E.P.; Lenihan, C.R.; Kiss, J.; Stauch, M.; Scholz, C.C.; Fraisl, P.; Lasitschka, F.; Mollenhauer, M.; Saunders, S.P. Loss of prolyl hydroxylase-1 protects against colitis through reduced epithelial cell apoptosis and increased barrier function. Gastroenterology 2010, 139, 2093–2101.
- 104. Karhausen, J.; Furuta, G.T.; Tomaszewski, J.E.; Johnson, R.S.; Colgan, S.P.; Haase, V.H. Epithelial hypoxia-inducible factor-1 is protective in murine experimental colitis. J. Clin. Investig. 2004, 114, 1098–1106.
- 105. Keenan, M.J.; Martin, R.J.; Raggio, A.M.; McCutcheon, K.L.; Brown, I.L.; Birkett, A.; Newman, S.S.; Skaf, J.; Hegsted, M.; Tulley, R.T. High-amylose resistant starch increases hormones and improves structure and function of the

- gastrointestinal tract: A microarray study. Lifestyle Genom. 2012, 5, 26-44.
- 106. Zhang, X.; Chen, S.; Duan, F.; Liu, A.; Li, S.; Zhong, W.; Sheng, W.; Chen, J.; Xu, J.; Xiao, S. Prebiotics enhance the biotransformation and bioavailability of ginsenosides in rats by modulating gut microbiota. J. Ginseng. Res. 2020, doi:10.1016/j.jgr.2020.08.001.
- 107. Cani, P.D.; Possemiers, S.; Van de Wiele, T.; Guiot, Y.; Everard, A.; Rottier, O.; Geurts, L.; Naslain, D.; Neyrinck, A.; Lambert, D.M. Changes in gut microbiota control inflammation in obese mice through a mechanism involving GLP-2-driven improvement of gut permeability. Gut 2009, 58, 1091–1103.
- 108. Fukunaga, T.; Sasaki, M.; Araki, Y.; Okamoto, T.; Yasuoka, T.; Tsujikawa, T.; Fujiyama, Y.; Bamba, T. Effects of the soluble fibre pectin on intestinal cell proliferation, fecal short chain fatty acid production and microbial population. Digestion 2003, 67, 42–49.
- 109. Peng, L.; Li, Z.-R.; Green, R.S.; Holzman, I.R.; Lin, J. Butyrate enhances the intestinal barrier by facilitating tight junction assembly via activation of AMP-activated protein kinase in Caco-2 cell monolayers. J. Nutr. 2009, 139, 1619–1625.
- 110. Ohata, A.; Usami, M.; Miyoshi, M. Short-chain fatty acids alter tight junction permeability in intestinal monolayer cells via lipoxygenase activation. Nutrition 2005, 21, 838–847.
- 111. Mithieux, G.; Andreelli, F.; Magnan, C. Intestinal gluconeogenesis: Key signal of central control of energy and glucose homeostasis. Curr. Opin. Clin. Nutr. Metab. Care 2009, 12, 419–423.
- 112. De Vadder, F.; Kovatcheva-Datchary, P.; Goncalves, D.; Vinera, J.; Zitoun, C.; Duchampt, A.; Bäckhed, F.; Mithieux, G. Microbiota-generated metabolites promote metabolic benefits via gut-brain neural circuits. Cell 2014, 156, 84–96.
- 113. Miele, L.; Valenza, V.; La Torre, G.; Montalto, M.; Cammarota, G.; Ricci, R.; Masciana, R.; Forgione, A.; Gabrieli, M.L.; Perotti, G. Increased intestinal permeability and tight junction alterations in nonalcoholic fatty liver disease. Hepatology 2009, 49, 1877–1887.
- 114. Lecerf, J.-M.; Dépeint, F.; Clerc, E.; Dugenet, Y.; Niamba, C.N.; Rhazi, L.; Cayzeele, A.; Abdelnour, G.; Jaruga, A.; Younes, H. Xylo-oligosaccharide (XOS) in combination with inulin modulates both the intestinal environment and immune status in healthy subjects, while XOS alone only shows prebiotic properties. Br. J. Nutr. 2012, 108, 1847–1858.
- 115. Paschos, P.; Paletas, K. Non alcoholic fatty liver disease two-hit process: Multifactorial character of the second hit. Hippokratia 2009, 13, 128.
- 116. Cortez-Pinto, H.; Jesus, L.; Barros, H.; Lopes, C.; Moura, M.; Camilo, M. How different is the dietary pattern in non-alcoholic steatohepatitis patients? Clin. Nutr. 2006, 25, 816–823.
- 117. Galisteo, M.; Duarte, J.; Zarzuelo, A. Effects of dietary fibers on disturbances clustered in the metabolic syndrome. J. Nutr. Biochem. 2008, 19, 71–84.
- 118. Federico, A.; Zulli, C.; de Sio, I.; Del Prete, A.; Dallio, M.; Masarone, M.; Loguercio, C. Focus on emerging drugs for the treatment of patients with non-alcoholic fatty liver disease. World J. Gastroenterol. 2014, 20, 16841.
- 119. Duarte, S.M.B.; Stefano, J.T.; Vanni, D.S.; Carrilho, F.J.; Oliveira, C.P.M.S.D. Impact of current diet at the risk of non-alcoholic fatty liver disease (NAFLD). Arq. Gastroenterol. 2019, 56, 431–439.
- 120. Williams, C.M. Effects of inulin on lipid parameters in humans. J. Nutr. 1999, 129, 1471S-1473S.
- 121. Kok, N.; Roberfroid, M.; Delzenne, N. Dietary oligofructose modifies the impact of fructose on hepatic triacylglycerol metabolism. Metab. Clin. Exp. 1996, 45, 1547–1550.
- 122. Parnell, J.A.; Reimer, R.A. Effect of prebiotic fibre supplementation on hepatic gene expression and serum lipids: A dose–response study in JCR: LA-cp rats. Br. J. Nutr. 2010, 103, 1577–1584.
- 123. Delzenne, N.M.; Williams, C.M. Prebiotics and lipid metabolism. Curr. Opin. Lipidol. 2002, 13, 61-67.
- 124. Levrat, M.-A.; Favier, M.-L.; Moundras, C.; Rémésy, C.; Demigné, C.; Morand, C. Role of dietary propionic acid and bile acid excretion in the hypocholesterolemic effects of oligosaccharides in rats. J. Nutr. 1994, 124, 531–538.
- 125. Vaziri, N.D.; Wong, J.; Pahl, M.; Piceno, Y.M.; Yuan, J.; DeSantis, T.Z.; Ni, Z.; Nguyen, T.-H.; Andersen, G.L. Chronic kidney disease alters intestinal microbial flora. Kidney Int. 2013, 83, 308–315.
- 126. Mafra, D.; Fouque, D. Gut Microbiota and Inflammation in Chronic Kidney Disease Patients. Clin. Kidney J. 2015, 8, 332–334.
- 127. Wang, F.; Jiang, H.; Shi, K.; Ren, Y.; Zhang, P.; Cheng, S. Gut bacterial translocation is associated with microinflammation in end-stage renal disease patients. Nephrology 2012, 17, 733–738.

- 128. Wu, I.-W.; Hsu, K.-H.; Lee, C.-C.; Sun, C.-Y.; Hsu, H.-J.; Tsai, C.-J.; Tzen, C.-Y.; Wang, Y.-C.; Lin, C.-Y.; Wu, M.-S. p-Cresyl sulphate and indoxyl sulphate predict progression of chronic kidney disease. Nephrol. Dial. Transplant. 2011, 26, 938–947.
- 129. Krishnamurthy, V.M.R.; Wei, G.; Baird, B.C.; Murtaugh, M.; Chonchol, M.B.; Raphael, K.L.; Greene, T.; Beddhu, S. High dietary fiber intake is associated with decreased inflammation and all-cause mortality in patients with chronic kidney disease. Kidney Int. 2012, 81, 300–306.
- 130. Rossi, M.; Johnson, D.; Xu, H.; Carrero, J.; Pascoe, E.; French, C.; Campbell, K. Dietary protein-fiber ratio associates with circulating levels of indoxyl sulfate and p-cresyl sulfate in chronic kidney disease patients. Nutr. Metab. Cardiovasc. Dis. 2015, 25, 860–865.
- 131. Younes, H.; Alphonse, J.-C.; Behr, S.R.; Demigné, C.; Rémésy, C. Role of fermentable carbohydrate supplements with a low-protein diet in the course of chronic renal failure: Experimental bases. Am. J. Kidney Dis. 1999, 33, 633–646.
- 132. Mardinoglu, A.; Shoaie, S.; Bergentall, M.; Ghaffari, P.; Zhang, C.; Larsson, E.; Bäckhed, F.; Nielsen, J. The gut microbiota modulates host amino acid and glutathione metabolism in mice. Mol. Syst. Biol. 2015, 11, 834.
- 133. Bergen, W.G. Small-intestinal or colonic microbiota as a potential amino acid source in animals. Amino Acids 2015, 47, 251–258.
- 134. Pluznick, J.L.; Protzko, R.J.; Gevorgyan, H.; Peterlin, Z.; Sipos, A.; Han, J.; Brunet, I.; Wan, L.-X.; Rey, F.; Wang, T. Olfactory receptor responding to gut microbiota-derived signals plays a role in renin secretion and blood pressure regulation. Proc. Natl. Acad. Sci. USA 2013, 110, 4410–4415.
- 135. Remuzzi, G.; Perico, N.; Macia, M.; Ruggenenti, P. The role of renin-angiotensin-aldosterone system in the progression of chronic kidney disease. Kidney Int. 2005, 68, S57–S65.

Retrieved from https://encyclopedia.pub/entry/history/show/16313