

Gamma-Delta T Cells

Subjects: [Virology](#)

Contributor: SHIVKUMAR BIRADAR

Recent advances in $\gamma\delta$ T cell biology have focused on the unique attributes of these cells and their role in regulating innate and adaptive immunity, promoting tissue homeostasis, and providing resistance to various disorders. Numerous bacterial and viral pathogens, including human immunodeficiency virus-1 (HIV), greatly alter the composition of $\gamma\delta$ T cells *in vivo*. Despite the effectiveness of antiretroviral therapy (ART) in controlling HIV and restoring health in those affected, $\gamma\delta$ T cells are dramatically impacted during HIV infection and fail to reconstitute to normal levels in HIV-infected individuals during ART for reasons that are not clearly understood. Importantly, their role in controlling HIV infection, and the implications of their failure to rebound during ART are also largely unknown and understudied.

Gamma-Delta T Cells, $\gamma\delta$ T cells, Human Immunodeficiency Virus

1. Introduction

Although conventional alpha-beta ($\alpha\beta$) T cells have been studied intensively as a current “known” in the field of immunology, with the basis of their recognition of processed peptides in the context of MHC molecules being well-understood, far less attention has been paid to gamma delta ($\gamma\delta$) T cells. This is in part due to their relatively limited numbers in the blood and spleen, and because they reside primarily in tissues less readily accessible for study. They have important early innate immune functions, recognizing relatively conserved pathogen-associated molecules, simple metabolites, and stress ligands expressed on infected and transformed cells. They can mount rapid and direct cytolytic responses while producing cytokines and chemokines to promote the function and mobilization of other immune effector cells [1]. They are widely distributed throughout the body [2], particularly within epithelial sites, and have the capability to recognize a variety of self-, as well as non-self-antigens without regard to MHC constraint. Their prospective utilization or strategic targeting in novel immunotherapeutic approaches to treat a variety of chronic diseases is quite attractive. This is especially true in instances where classical $\alpha\beta$ T cell immune escape may be of particular concern. However, because their frequency and function can be severely altered in the setting of diseases such as HIV, and because there are still so many “unknowns” regarding the basic biology of $\gamma\delta$ T cells in health and disease, there are hurdles to be addressed before their therapeutic potential can be fully appreciated.

2. $\gamma\delta$ T Cells in HIV-1 Infection

While $\gamma\delta$ T cells have been described to provide protective immunity against tumors of epithelial [3][4] and hematological origin [5][6], they have also been explored in the setting of various chronic viral [7][8][9][10][11] and

bacterial diseases [12][13], as well as malaria [14][15]. Furthermore, $\gamma\delta$ T cells contribute to the pathogenesis and regulation of autoimmune diseases, including rheumatoid arthritis and psoriasis [16][17]. For the remainder of this paper, we have chosen to focus our attention on the function of these cells in the setting of chronic HIV-1 infection, and to shed some light on the immunotherapeutic potential of $\gamma\delta$ T cells for the treatment of HIV/AIDS.

2.1. Impact of HIV on $\gamma\delta$ T Cells

In healthy individuals, Vy9V δ 2 T cells contribute to 90–95% of the total $\gamma\delta$ population in the peripheral blood, and the remaining 5–10% are V δ 2^{neg}, such as V δ 1 and V δ 3s [18]. In HIV-infected individuals, Vy9V δ 2 T cells are drastically depleted, and the V δ 2:V δ 1 ratio is inverted in peripheral blood [18]. Interestingly, Vy9V δ 2 cell depletion occurs very early during the infection and correlates with viral load and CD4+ T cell depletion [19]. HIV preferentially depletes phosphoantigen responsive Vy9V δ 2 cells with Vy9-Jy1.2 TCR rearrangement [20]. Although the precise mechanism of depletion of V δ 2 cells in HIV-infected individuals is not understood, some studies indicate that α 4 β 7 and CCR5 receptors form a complex on Vy9V δ 2 cells, which facilitates the binding of the bV3 loop of HIV gp120 to CCR5 in the absence of expression of the CD4 co-receptor. This interaction leads to p38 kinase activation and induces apoptosis in these cells [21]. Additionally, HIV infection leads to loss of Th17 CD4+ T cells, important for maintaining epithelial barrier integrity in the gut [22]. Depletion results in dysregulation of mucosal immunity and allows microbial translocation into the circulation, resulting in systemic immune activation [23] and expansion of V δ 1 cell numbers in viremic patients [24]. Moreover, the Vy9V δ 2 subset from HIV-infected individuals fails to proliferate or produce cytokines in response to mycobacterial infection in vitro, suggesting that Vy9V δ 2 cells are functionally inactive [25]. This functional anergy in residual Vy9V δ 2 cells is also characterized by their decrease in responsiveness to phosphoantigens and lytic activity toward the Daudi lymphoma cell line [26].

V δ 1 cells are enriched in the gut mucosa, where they help in maintaining tissue homeostasis. HIV infection further increases the frequency of V δ 1 cells in the mucosa, similar to the increased levels observed in peripheral blood of HIV-infected individuals [27]. Despite immunologic control of viral replication, elite controllers (EC) also display elevated levels of V δ 1 cells in the gut mucosa [28]. Few studies demonstrated that HIV-mediated disruption of gut epithelial barrier leads to translocation of gut microbiota, which activates V δ 1 T cells to produce pro-inflammatory cytokines and exacerbate the chronic inflammation [24][29]. The increase in V δ 1 T cell numbers seen in EC strongly correlated with the gut viral load, implying that viral replication in the mucosa and disruption in the gut epithelial barrier integrity and microbial translocation may contribute to enhanced V δ 1 T cell proliferation, also along with other immune activation events and disease progression. However, further studies are needed to better understand the causation and mechanism of V δ 1 T cell expansion in the gut during HIV infection and the net impact of this on disease progression.

2.2. Impact of $\gamma\delta$ T Cell Perturbations on Immune Control of HIV

As previously alluded to, much of the impact of $\gamma\delta$ T cells on immune function in health and disease is mediated through their bi-directional cross-talk with other immune cells [30]. In healthy individuals, activated $\gamma\delta$ T cells induce the maturation and differentiation of DCs and B cells into functional APCs, but in HIV-infected individuals, this

immunomodulatory capacity of $\gamma\delta$ T cells is compromised [31]. Typically, upon activation, Vy9V δ 2 T cells can drive the upregulation of CD80, CD86, HLA-DR, and CD40 surface expression on APCs, enhancing their capacity to induce primary $\alpha\beta$ T cell responses [32]. HIV infection alters the ability of APCs to process and present antigens, inhibiting the ability of Vy9V δ 2 T cells to effectively interact with and positively impact the phenotype and function of APCs [31].

HIV infection may also impair the APC function of $\gamma\delta$ T cells and their ability to induce $\alpha\beta$ T cell responses [33]. Activated $\gamma\delta$ T cells produce large quantities of chemokines, including the macrophage inflammatory protein (MIP)-1 α /CCL3, MIP-1 β /CCL4, and CCL5/RANTES, which binds to, and can compete with the HIV coreceptor CCR5 to block HIV infection of the target cells. Again, alteration of $\gamma\delta$ T cells in HIV infection may lead to reduced protection against new infection of target cells. V δ 2 T cells cultured in the presence of HMBPP and IL-21 express B cell-attracting chemokine CXCL13, and the CXCL13 receptor CXCR5. This promotes B cell somatic mutation, productive rearrangement, and maturation in the germinal center, thereby promoting antibody production by B cells [34][35][36]. Due to the loss of $\gamma\delta$ T cells in HIV infected individuals, it is likely that these crucial helper functions will also be compromised.

Numerous studies have demonstrated the protective role of $\gamma\delta$ T cells in inducing cytotoxicity in HIV-infected cells and controlling HIV replication [37][38][39]. However, $\gamma\delta$ T cells can also contribute to negative outcomes in HIV infection. $\gamma\delta$ T cells from HIV-infected individuals express high levels of the inhibitory receptor TIGIT, and they are the primary inflammatory driver in ART-suppressed HIV infection, and they contribute to age-associated morbidity and mortality [40]. This negative outcome is particularly highlighted in respiratory conditions, where alveolar immune cell homeostasis is disrupted in HIV-infected adults, characterized by the increased infiltration of $\gamma\delta$ T cells and other immune cells in broncho-alveolar lavage fluid, resulting in an enhanced susceptibility of these individuals to lower respiratory tract infections [41].

2.3. $\gamma\delta$ T Cell Contribution to HIV Reservoirs

Even though current ART effectively suppresses HIV replication, the integrated viral genome remains transcriptionally silent in host chromatin, representing a major challenge towards efforts to eradicate infection [42]. Resting memory CD4+ T lymphocytes have long been considered the major reservoir site for HIV [43]. However, recent studies demonstrate that other cell types can also contribute to the latent HIV reservoir, including $\gamma\delta$ T cells [28].

In one study, replication-competent HIV could be obtained from highly purified V δ 2 T cells from HIV-infected people during ART, thus identifying a previously unrecognized latent HIV reservoir within V δ 2 T cells at an unexpectedly high frequency [44]. Although the precise mechanism is not known, $\gamma\delta$ T cells can be infected by the CXCR4-tropic laboratory clone, HIV_{LAI} [26]. Possible mechanisms of infection include the binding of HIV envelope glycoproteins to the highly expressed CCR5/ α 4 β 7 receptor on Vy9V δ 2 T cells, leading to infection through a CD4-independent pathway [21]. Alternatively, HIV infection has been shown to induce immune activation and subsequent upregulation of CD4 receptors on some $\gamma\delta$ T cells, potentially making them more susceptible to HIV infection [44]. Preliminary

findings from our group, utilizing a humanized mouse model, suggest that activated Vy9V δ 2 cells can indeed serve as early targets for HIV infection and play a critical role in the early stages of viral dissemination [45]. Depletion of these cells during early infection would likely negatively impact immune defenses, particularly due to the important roles they serve by producing chemokines that can act as competitive inhibitors to block HIV entry and to recruit other immune effector cells to promote HIV clearance [8]. However, the increased presence and survival of activated $\gamma\delta$ T cells, which can also serve as targets for HIV infection, could contribute to enhancement in viral production and rebound. This raises more questions about the role of $\gamma\delta$ T cells in the initial sequelae of HIV infection and their potential contribution to the HIV reservoir.

2.4. Impact of Anti-Retroviral Therapy on $\gamma\delta$ T Cells in HIV-Infection

Although ART is very effective in restoring CD4+ T cell counts and suppressing the virus below detectable levels, it fails to fully restore standard frequencies of Vy9V δ 2 T cells in HIV-infected individuals [18]. Long-term ART partially restored the Jy1.2 repertoire of the V δ 2 subset [20]. These Vy9V δ 2 cells are highly activated [46] but are functionally compromised, with diminished cytokine production, cytotoxicity, and proliferation [47]. At the same time, expanded V δ 1 levels remain stable during ART in peripheral blood and mucosal sites, and they produce pro-inflammatory cytokines and express the inhibitory molecule CD279 (PD-1) [48]. Whether long-term ART impacts the number of V δ 1 T cells and/or their activation status is not clearly understood. Studies suggest that the loss of Vy9V δ 2 cells in viral controllers is significantly lower than untreated or ART-treated individuals and that $\gamma\delta$ 17 cells are highly preserved. This preserved $\gamma\delta$ population may be responsible for preventing microbial translocation and controlling chronic systemic immune activation [49][50].

2.5. $\gamma\delta$ T Cells in HIV Immunotherapy

$\gamma\delta$ T cells are the first line of defense against many pathogens, but their number and functions are severely altered in many infectious diseases, including HIV. Despite the long-term ART, $\gamma\delta$ T cells are not reconstituted to the original frequency in HIV-infected individuals. However, in elite controllers, the Vy9V δ 2 T cell numbers are maintained at normal levels, implying that immunotherapy using Vy9V δ 2 T cells might recapitulate the immune status seen with those capable of naturally controlling HIV infection [49]. Several efforts were made to develop suitable methods for activating and expanding $\gamma\delta$ T cells *in vitro* and *in vivo*. *In vitro* methods involve stimulating PBMCs with bisphosphonates, such as IPP, HMBPP, and zoledronate [51]. Zoledronate blocks the metabolic conversion of IPP, allowing this phosphoantigen to accumulate until stimulatory levels are reached, resulting in the selective activation and expansion of Vy9V δ 2 T cells [51]. Bisphosphonate-mediated expansion of $\gamma\delta$ T cells is a rapid means to generate large quantities of these cells for adoptive cell therapy. On the other hand, the delta one T (DOT) cell subtype of $\gamma\delta$ T cells can be expanded using OKT-3 monoclonal antibody and a cytokine cocktail consisting of rIL-4, rIFN- γ , rIL-21, and rIL-1 β [52]. A high yield of DOT cells could be obtained by treating PBMCs with Con-A and rIL-2 and rIL-4 [53]. Alternative strategies for expanding $\gamma\delta$ T cells that do not respond to pAg or N-BP involve the use of agonistic monoclonal antibodies (mAb). Using $\gamma\delta$ TCR-specific antibodies, low levels of expansion of V δ 1, and V δ 2 T cells has been achieved, but it has not been very successful to date. However, 20.1, an agonistic Ab specific for CD277 (a member of BTN3 subfamily), mimics pAg-induced Vy9V δ 2 cell activation.

This antibody may simulate a conformational change in the CD277 molecule to activate and expand Vy9Vδ2 T cells [54].

References

1. Gu, S.; Nawrocka, W.; Adams, E.J. Sensing of Pyrophosphate Metabolites by V γ 9V δ 2 T-cells. *Front. Immunol.* 2014, 5, 688.
2. Caccamo, N.; Dieli, F.; Wesch, D.; Jomaa, H.; Eberl, M. Sex-specific phenotypical and functional differences in peripheral human Vy9/V δ 2 T-cells. *J. Leukoc. Biol.* 2006, 79, 663–666.
3. Liu, C.; Emami, S.N.; Pettersson, J.; Ranford-Cartwright, L.; Faye, I.; Parmryd, I. V γ 9V δ 2 T-cells proliferate in response to phosphoantigens released from erythrocytes infected with asexual and gametocyte stage *Plasmodium falciparum*. *Cell Immunol.* 2018, 334, 11–19.
4. Eloso, M.M.; van der Heyde, H.C.; vande Waa, J.A.; Manning, D.D.; Weidanz, W.P. Inhibition of *Plasmodium falciparum* In Vitro by human gamma delta T-cells. *J. Immunol.* 1994, 153, 1187–1194.
5. Correia, D.V.; Fogli, M.; Hudspeth, K.; da Silva, M.G.; Mavilio, D.; Silva-Santos, B. Differentiation of human peripheral blood V δ 1+ T-cells expressing the natural cytotoxicity receptor NKp30 for recognition of lymphoid leukemia cells. *Blood* 2011, 118, 992–1001.
6. Lanca, T.; Correia, D.V.; Moita, C.F.; Raquel, H.; Neves-Costa, A.; Ferreira, C.; Ramalho, J.S.; Barata, J.T.; Moita, L.F.; Gomes, A.Q.; et al. The MHC class Ib protein ULBP1 is a nonredundant determinant of leukemia/lymphoma susceptibility to gammadelta T-cell cytotoxicity. *Blood* 2010, 115, 2407–2411.
7. Hu, C.; Qian, L.; Miao, Y.; Huang, Q.; Miao, P.; Wang, P.; Yu, Q.; Nie, H.; Zhang, J.; He, D.; et al. Antigen-presenting effects of effector memory V γ 9V δ 2 T-cells in rheumatoid arthritis. *Cell Mol. Immunol.* 2012, 9, 245–254.
8. Poles, M.A.; Barsoum, S.; Yu, W.; Yu, J.; Sun, P.; Daly, J.; He, T.; Mehandru, S.; Talal, A.; Markowitz, M.; et al. Human immunodeficiency virus type 1 induces persistent changes in mucosal and blood gammadelta T-cells despite suppressive therapy. *J. Virol.* 2003, 77, 10456–10467.
9. Halary, F.; Pitard, V.; Dlubek, D.; Krzysiek, R.; de la Salle, H.; Merville, P.; Dromer, C.; Emilie, D.; Moreau, J.F.; Dechanet-Merville, J. Shared reactivity of V δ 2(neg) { γ { δ } T-cells against cytomegalovirus-infected cells and tumor intestinal epithelial cells. *J. Exp. Med.* 2005, 201, 1567–1578.

10. Ehl, S.; Schwarz, K.; Enders, A.; Duffner, U.; Pannicke, U.; Kuhr, J.; Mascart, F.; Schmitt-Graeff, A.; Niemeyer, C.; Fisch, P. A variant of SCID with specific immune responses and predominance of gamma delta T-cells. *J. Clin. Investig.* 2005, 115, 3140–3148.
11. De Paoli, P.; Gennari, D.; Martelli, P.; Basaglia, G.; Crovatto, M.; Battistin, S.; Santini, G. A subset of gamma delta lymphocytes is increased during HIV-1 infection. *Clin. Exp. Immunol.* 1991, 83, 187–191.
12. Spencer, C.T.; Abate, G.; Sakala, I.G.; Xia, M.; Truscott, S.M.; Eickhoff, C.S.; Linn, R.; Blazevic, A.; Metkar, S.S.; Peng, G.; et al. Granzyme A produced by gamma(9)delta(2) T-cells induces human macrophages to inhibit growth of an intracellular pathogen. *PLoS Pathog.* 2013, 9, e1003119.
13. Xi, X.; Han, X.; Li, L.; Zhao, Z. Identification of a new tuberculosis antigen recognized by gammadelta T-cell receptor. *Clin. Vaccine Immunol.* 2013, 20, 530–539.
14. Liu, C.; Emami, S.N.; Pettersson, J.; Ranford-Cartwright, L.; Faye, I.; Parmryd, I. Vgamma9Vdelta2 T-cells proliferate in response to phosphoantigens released from erythrocytes infected with asexual and gametocyte stage Plasmodium falciparum. *Cell Immunol.* 2018, 334, 11–19.
15. Elloso, M.M.; van der Heyde, H.C.; vande Waa, J.A.; Manning, D.D.; Weidanz, W.P. Inhibition of Plasmodium falciparum In Vitro by human gamma delta T-cells. *J. Immunol.* 1994, 153, 1187–1194.
16. Hu, C.; Qian, L.; Miao, Y.; Huang, Q.; Miao, P.; Wang, P.; Yu, Q.; Nie, H.; Zhang, J.; He, D.; et al. Antigen-presenting effects of effector memory Vgamma9Vdelta2 T-cells in rheumatoid arthritis. *Cell Mol. Immunol.* 2012, 9, 245–254.
17. Cai, Y.; Shen, X.; Ding, C.; Qi, C.; Li, K.; Li, X.; Jala, V.R.; Zhang, H.G.; Wang, T.; Zheng, J.; et al. Pivotal role of dermal IL-17-producing gammadelta T-cells in skin inflammation. *Immunity* 2011, 35, 596–610.
18. Poles, M.A.; Barsoum, S.; Yu, W.; Yu, J.; Sun, P.; Daly, J.; He, T.; Mehandru, S.; Talal, A.; Markowitz, M.; et al. Human immunodeficiency virus type 1 induces persistent changes in mucosal and blood gammadelta T-cells despite suppressive therapy. *J. Virol.* 2003, 77, 10456–10467.
19. Li, H.; Peng, H.; Ma, P.; Ruan, Y.; Su, B.; Ding, X.; Xu, C.; Pauza, C.D.; Shao, Y. Association between Vgamma2Vdelta2 T-cells and disease progression after infection with closely related strains of HIV in China. *Clin. Infect. Dis.* 2008, 46, 1466–1472.
20. Chaudhry, S.; Cairo, C.; Venturi, V.; Pauza, C.D. The gammadelta T-cell receptor repertoire is reconstituted in HIV patients after prolonged antiretroviral therapy. *Aids* 2013, 27, 1557–1562.

21. Li, H.; Pauza, C.D. HIV envelope-mediated, CCR5/alpha4beta7-dependent killing of CD4-negative gammadelta T-cells which are lost during progression to AIDS. *Blood* 2011, 118, 5824–5831.
22. Brenchley, J.M.; Paiardini, M.; Knox, K.S.; Asher, A.I.; Cervasi, B.; Asher, T.E.; Scheinberg, P.; Price, D.A.; Hage, C.A.; Kholi, L.M.; et al. Differential Th17 CD4 T-cell depletion in pathogenic and nonpathogenic lentiviral infections. *Blood* 2008, 112, 2826–2835.
23. Brenchley, J.M.; Price, D.A.; Schacker, T.W.; Asher, T.E.; Silvestri, G.; Rao, S.; Kazzaz, Z.; Bornstein, E.; Lambotte, O.; Altmann, D.; et al. Microbial translocation is a cause of systemic immune activation in chronic HIV infection. *Nat. Med.* 2006, 12, 1365–1371.
24. Li, Z.; Li, W.; Li, N.; Jiao, Y.; Chen, D.; Cui, L.; Hu, Y.; Wu, H.; He, W. gammadelta T-cells are involved in acute HIV infection and associated with AIDS progression. *PLoS ONE* 2014, 9, e106064.
25. Poccia, F.; Boullier, S.; Lecoeur, H.; Cochet, M.; Poquet, Y.; Colizzi, V.; Fournie, J.J.; Gougeon, M.L. Peripheral V gamma 9/V delta 2 T-cell deletion and anergy to nonpeptidic mycobacterial antigens in asymptomatic HIV-1-infected persons. *J. Immunol.* 1996, 157, 449–461.
26. Wallace, M.; Scharko, A.M.; Pauza, C.D.; Fisch, P.; Imaoka, K.; Kawabata, S.; Fujihashi, K.; Kiyono, H.; Tanaka, Y.; Bloom, B.R.; et al. Functional gamma delta T-lymphocyte defect associated with human immunodeficiency virus infections. *Mol. Med.* 1997, 3, 60–71.
27. Nilssen, D.E.; Müller, F.; Oktedalen, O.; Frøland, S.S.; Fausa, O.; Halstensen, T.S.; Brandtzaeg, P. Intraepithelial gamma/delta T-cells in duodenal mucosa are related to the immune state and survival time in AIDS. *J. Virol.* 1996, 70, 3545–3550.
28. Imlach, S.; Leen, C.; Bell, J.E.; Simmonds, P. Phenotypic analysis of peripheral blood gammadelta T lymphocytes and their targeting by human immunodeficiency virus type 1 In Vivo. *Virology* 2003, 305, 415–427.
29. Klatt, N.R.; Funderburg, N.T.; Brenchley, J.M. Microbial translocation, immune activation, and HIV disease. *Trends Microbiol.* 2013, 21, 6–13.
30. Biradar, S.; Mailliard, R.B.; et al. Adoptive transfer of allogeneic expanded gamma delta T cells promotes HIV replication in a humanized mouse model. (Manuscript in preparation).
31. Sacchi, A.; Rinaldi, A.; Tumino, N.; Casetti, R.; Agrati, C.; Turchi, F.; Bordoni, V.; Cimini, E.; Martini, F. HIV infection of monocytes-derived dendritic cells inhibits Vgamma9Vdelta2 T-cells functions. *PLoS ONE* 2014, 9, e111095.
32. Brandes, M.; Willimann, K.; Moser, B. Professional antigen-presentation function by human gammadelta T-cells. *Science* 2005, 309, 264–268.

33. Poccia, F.; Battistini, L.; Cipriani, B.; Mancino, G.; Martini, F.; Gougeon, M.L.; Colizzi, V. Phosphoantigen-reactive V γ 9V δ 2 T lymphocytes suppress in vitro human immunodeficiency virus type 1 replication by cell-released antiviral factors including CC chemokines. *J. Infect. Dis.* 1999, 180, 858–861.
34. Brandes, M.; Willimann, K.; Lang, A.B.; Nam, K.H.; Jin, C.; Brenner, M.B.; Morita, C.T.; Moser, B. Flexible migration program regulates gamma delta T-cell involvement in humoral immunity. *Blood* 2003, 102, 3693–3701.
35. Bansal, R.R.; Mackay, C.R.; Moser, B.; Eberl, M. IL-21 enhances the potential of human gammadelta T-cells to provide B-cell help. *Eur. J. Immunol.* 2012, 42, 110–119.
36. Caccamo, N.; Battistini, L.; Bonneville, M.; Poccia, F.; Fournié, J.J.; Meraviglia, S.; Borsellino, G.; Krocze, R.A.; La Mendola, C.; Scotet, E.; et al. CXCR5 identifies a subset of V γ 9V δ 2 T-cells which secrete IL-4 and IL-10 and help B cells for antibody production. *J. Immunol.* 2006, 177, 5290–5295.
37. Cai, Y.; Shen, X.; Ding, C.; Qi, C.; Li, K.; Li, X.; Jala, V.R.; Zhang, H.G.; Wang, T.; Zheng, J.; et al. Pivotal role of dermal IL-17-producing gammadelta T-cells in skin inflammation. *Immunity* 2011, 35, 596–610.
38. Siegers, G.M.; Dhamko, H.; Wang, X.H.; Mathieson, A.M.; Kosaka, Y.; Felizardo, T.C.; Medin, J.A.; Tohda, S.; Schueler, J.; Fisch, P.; et al. Human V δ 1 y δ T-cells expanded from peripheral blood exhibit specific cytotoxicity against B-cell chronic lymphocytic leukemia-derived cells. *Cytotherapy* 2011, 13, 753–764.
39. Mingari, M.C.; Varese, P.; Bottino, C.; Melioli, G.; Moretta, A.; Moretta, L. Clonal analysis of CD4-CD8- human thymocytes expressing a T-cell receptor gamma/delta chain. Direct evidence for the de novo expression of CD8 surface antigen and of cytolytic activity against tumor targets. *Eur. J. Immunol.* 1988, 18, 1831–1834.
40. Belkina, A.C.; Starchenko, A.; Drake, K.A.; Proctor, E.A.; Pihl, R.M.F.; Olson, A.; Lauffenburger, D.A.; Lin, N.; Snyder-Cappione, J.E. Multivariate Computational Analysis of Gamma Delta T-cell Inhibitory Receptor Signatures Reveals the Divergence of Healthy and ART-Suppressed HIV+ Aging. *Front. Immunol.* 2018, 9, 2783.
41. Mwale, A.; Hummel, A.; Mvaya, L.; Kamng'ona, R.; Chimbayo, E.; Phiri, J.; Malamba, R.; Kankwatira, A.; Mwandumba, H.C.; Jambo, K.C. B cell, CD8 (+) T-cell and gamma delta T-cell infiltration alters alveolar immune cell homeostasis in HIV-infected Malawian adults. *Wellcome Open Res.* 2017, 2, 105.
42. Stevenson, M.; Stanwick, T.L.; Dempsey, M.P.; Lamonica, C.A. HIV-1 replication is controlled at the level of T-cell activation and proviral integration. *Embo J.* 1990, 9, 1551–1560.

43. Siliciano, J.D.; Kajdas, J.; Finzi, D.; Quinn, T.C.; Chadwick, K.; Margolick, J.B.; Kovacs, C.; Gange, S.J.; Siliciano, R.F. Long-term follow-up studies confirm the stability of the latent reservoir for HIV-1 in resting CD4+ T-cells. *Nat. Med.* 2003, 9, 727–728.
44. Soriano-Sarabia, N.; Archin, N.M.; Bateson, R.; Dahl, N.P.; Crooks, A.M.; Kuruc, J.D.; Garrido, C.; Margolis, D.M. Peripheral Vgamma9Vdelta2 T-cells Are a Novel Reservoir of Latent HIV Infection. *PLoS Pathog.* 2015, 11, e1005201.
45. Biradar, S.; Mailliard, R.B.; et al. Adoptive transfer of allogeneic expanded gamma delta T cells promotes HIV replication in a humanized mouse model. (Manuscript in preparation).
46. Negash, M.; Tsegaye, A.; Wassie, L.; Howe, R. Phenotypic and functional heterogeneity of peripheral gammadelta T-cells in pulmonary TB and HIV patients in Addis Ababa, Ethiopia. *BMC Infect. Dis.* 2018, 18, 464.
47. Cummings, J.S.; Cairo, C.; Armstrong, C.; Davis, C.E.; Pauza, C.D. Impacts of HIV infection on Vgamma2Vdelta2 T-cell phenotype and function: A mechanism for reduced tumor immunity in AIDS. *J. Leukoc. Biol.* 2008, 84, 371–379.
48. Olson, G.S.; Moore, S.W.; Richter, J.M.; Garber, J.J.; Bowman, B.A.; Rawlings, C.A.; Flagg, M.; Corleis, B.; Kwon, D.S. Increased frequency of systemic pro-inflammatory Vdelta1(+) gammadelta T-cells in HIV elite controllers correlates with gut viral load. *Sci. Rep.* 2018, 8, 16471.
49. Riedel, D.J.; Sajadi, M.M.; Armstrong, C.L.; Cummings, J.S.; Cairo, C.; Redfield, R.R.; Pauza, C.D. Natural viral suppressors of HIV-1 have a unique capacity to maintain gammadelta T-cells. *Aids* 2009, 23, 1955–1964.
50. Chevalier, M.F.; Bhatnagar, N.; Didier, C.; Lopez-Gonzalez, M.; Pavie, J.; Bollens, D.; Duvivier, C.; Collias, L.; Jung, C.; Scott-Algara, D.; et al. gammadelta T-cell subsets in HIV controllers: Potential role of Tgammadelta17 cells in the regulation of chronic immune activation. *Aids* 2019, 33, 1283–1292.
51. Roelofs, A.J.; Jauhainen, M.; Monkkonen, H.; Rogers, M.J.; Monkkonen, J.; Thompson, K. Peripheral blood monocytes are responsible for gammadelta T-cell activation induced by zoledronic acid through accumulation of IPP/DMAPP. *Br. J. Haematol.* 2009, 144, 245–250.
52. Almeida, A.R.; Correia, D.V.; Fernandes-Platzgummer, A.; da Silva, C.L.; da Silva, M.G.; Anjos, D.R.; Silva-Santos, B. Delta One T-cells for Immunotherapy of Chronic Lymphocytic Leukemia: Clinical-Grade Expansion/Differentiation and Preclinical Proof of Concept. *Clin. Cancer Res.* 2016, 22, 5795–5804.
53. Siegers, G.M.; Dhamko, H.; Wang, X.H.; Mathieson, A.M.; Kosaka, Y.; Felizardo, T.C.; Medin, J.A.; Tohda, S.; Schueler, J.; Fisch, P.; et al. Human Vδ1 γδ T-cells expanded from peripheral blood exhibit specific cytotoxicity against B-cell chronic lymphocytic leukemia-derived cells. *Cytotherapy* 2011, 13, 753–764.

-
54. Gu, S.; Borowska, M.T.; Boughter, C.T.; Adams, E.J. Butyrophilin3A proteins and $\text{V}\gamma 9\text{V}\delta 2$ T-cell activation. *Semin. Cell Dev. Biol.* 2018, 84, 65–74.
-

Retrieved from <https://encyclopedia.pub/entry/history/show/15048>