Curcumin and Ethanol Effects in Trembler-J Schwann

Subjects: Biochemistry & Molecular Biology

Contributor: Lucia Vázquez Alberdi , Gonzalo Rosso , , Joaquina Farias , Miguel Calero , Alejandra Elizabeth Kun Gonzalez

Charcot-Marie-Tooth (CMT) syndrome is the most common progressive human motor and sensory peripheral neuropathy. CMT type 1E is a demyelinating neuropathy affecting Schwann cells due to peripheral-myelin-protein-22 (PMP22) mutations, modelized by Trembler-J mice. Curcumin, a natural polyphenol compound obtained from turmeric (*Curcuma longa*), exhibits dose- and time-varying antitumor, antioxidant and neuroprotective properties, however, the neurotherapeutic actions of curcumin remain elusive. Here, the researchers propose curcumin as a possible natural treatment capable of enhancing cellular detoxification mechanisms, resulting in an improvement of the neurodegenerative Trembler-J phenotype.

CMT1E	Trembler-J	curcumin	ethanol	Hsps	autophagy
-------	------------	----------	---------	------	-----------

1. Introduction

The group of human hereditary peripheral neuropathies, known as Charcot-Marie-Tooth disease (CMT), has a prevalence of 1/2500 ^[1]. Within the CMT, demyelinating neuropathies (CMT1) have mutations that alter the structural integrity of myelin ^[2].

The mutations affecting the *pmp22* gene, play a central pathognomonic role in the CMT disease, representing between 60% and 70% of total myelinopathies ^[3]. PMP22 is highest expressed in the Schwann cells as a 160-amino-acid myelin glycoprotein, of 22 kDa, with four transmembrane domains, representing approximately 5% of the total compact myelin proteins ^[4]. However, central expression of PMP22 has also been signaled ^{[5][6][7][8][9]}.

However, PMP22 is ubiquitously expressed in various tissues and organs in addition to the nervous system. PMP22 has been reported to play a role in adhesion and proliferation regulation ^[10], and described in epithelial cells, where it localizes with tight junctions and forms complexes with integrins and P2X7 channels ^{[11][12][13]}. It has been suggested that the regulation of PMP22 expression may also be increased by the action of steroid hormones ^{[3][14]}, which has contributed to the exploration of hormonal therapies in the treatment of the CMT1A phenotype ^[14] ^{[15][16]}.

The expression of human PMP22 has been also reported during the proliferative and secretory phase of the menstrual cycle, and PMP22 has been shown to colocalize with alpha-6-integrins both in vitro and in human tissue samples. Thus, PMP22 appears to be associated in the endometrium with both cell adhesion and endometrial

differentiation [17]. Up to 5% of all CMT1s integrate the group of CMT1E, myelinopathies caused by different point mutations in the *pmp22* [18].

The Trembler J (TrJ/+) mouse is an animal model of CMT1E ^{[19][20][21][22]}, carrying the same spontaneous mutation in *pmp22* as that found in a human family ^[23]. Under normal conditions, only 20% of PMP22 is inserted into the membrane, with chaperone assistance, at the cost of high energy expenditure (by the synthesis of unused PMP22 and for the maintenance of the proteasome machinery in charge of eliminating the protein surplus) ^{[24][25]}. In disease, the percentage of myelin that is inserted is even lower, so it is common to find intracellular PMP22 aggregates in the Schwann cells (SCs) of TrJ/+ mice ^[26], interfering with the regular protein transport. Thus, the peripheral nerve fibers from the neurodegenerative phenotype in TrJ/+ show altered autophagic-lysosomal pathways, PMP22 cytoplasmic aggregation, increased ribosome and translational activity ^{[21][27][28][29][30][31]}.

One of the mechanisms underlying cellular stress situations is the response by Heat Shock Proteins (Hsps) ^{[32][33]} ^{[34][35][36]}. Heat Shock Factor 1 (HSF1), the main regulator of the Hsps, is activated by mTOR under stress ^{[34][37]} and its inhibition prevents autophagosome formation ^[38]. Hsp27, recognized for its dual role in normal situations and tumor processes ^{[39][40][41][42]}, has also been pointed out as a possible target of action for neurodegenerative diseases ^[43]. In stressful situations, such as the accumulation of intracellular proteins, Hsp27 activates and modulates serine/threonine protein kinase B (PKB/AKT) action, mTOR main activator ^{[44][45][46]}. It has also been reported that Hsp70, another member of the Hsps family, assists in the processing of PMP22 aggregates in TrJ/+ through the Golgi apparatus and their release into Rab7-positive vesicles to the lysosome ^[47]. It has been observed, both in vivo and in vitro, that there is co-localization of Hsps with PMP22 aggregates ^{[21][47]}. In addition, beneficial effects of autophagy, promoted by chaperones and preventing the accumulation of misfolded PMP22, have been reported in TrJ/+ ^[48]. In CMT2, histone deacetylase 6 (HDAC6) has also been signaled as a potential therapeutic target for the amelioration of the neurodegenerative phenotype, reversing motor and sensory deficits induced by Hsp27 activation ^[49].

A possible cellular and molecular perspective on the therapeutics of these until now incurable hereditary conditions may focus on cellular drainage or detoxification promoted on the autophagic-lysosomal and UPS-chaperone pathways, together with inhibition or reduction of the mTOR pathway. Furthermore, decreased energy availability is a common key player for the modulation of these pathways. For this reason, caloric restriction (CR) at the neuromotor level has been proposed as a valid therapeutic approach for the alleviation of neurodegenerative conditions (including peripheral neuropathies) ^{[50][51][52][53]}. The researchers' group demonstrated that dietary CR activates canonical autophagic pathways by decreasing the levels of aggregated PMP22 and increasing ribophagy in TrJ/+ and in wild-type (+/+) nerves (manuscript in preparation).

CR can be emulated, under certain conditions, by effector molecules of the aforementioned pathways ^{[54][55][56][57]} ^[58]. Among them, curcumin, a polyphenol extracted from *Curcuma longa* (Linnaeus, Species Plantarum 1:2. 1753), has shown promising results ^{[59][60]}. This compound is used as an antitumor, anti-inflammatory, antioxidant, among other beneficial effects. This wide spectrum of curcumin applications depends mainly on the dosage used and the time of application of the treatments. For example, at high concentrations, which in culture range from 25 µM to

160 μ M, curcumin is used as a potent anti-tumoral ^{[61][62]}. At low concentrations, it decreases reactive oxygen species (ROS) (in myoblast cell cultures 4 μ M curcumin and SC from 0.001 to 1 μ M curcumin) ^{[63][64]}, show antiinflammatory effects at decreasing signal transducer and activator of transcription 3 (STAT3) activation (in human multipotent adipose tissue-derived stem, 10 μ M curcumin) ^[65], and promotes autophagy by inhibiting acetyltransferases (glioblastoma multiforme cell line, 10 μ M curcumin) ^{[59][66]} and cell regeneration (primary myoblast culture, 1 μ M curcumin) ^{[67][68]}. Interestingly, in TrJ mice, there is evidence that curcumin treatment can improve the neurodegenerative phenotype ^[69]. On the other hand, CR activates the autophagy process, prevents the formation, and promotes the elimination of PMP22 aggregates in cultured SCs ^[70]. However, little is known about the effect of curcumin and ethanol (EtOH) used as curcumin vehicle in the modulation of PMP22 aggregates, and whether this effect could activate effector molecules in SCs ameliorating the neurodegenerative condition of TrJ mice.

2. Current Insights

PMP22 expression has been studied both in vivo, using different animal models ^{[19][25][48][71][72][73][74]}, and in vitro in SC cultures ^{[24][63][75]}, respectively. These approaches denote different but complementary physiological conditions. While in vivo approaches allow understanding how and where the main expression of this protein is located in SCs arrested in G0, the in vitro studies allow the evaluation of the expression in those cells that are in a proliferative state. One of the contributions of the researchers' work lies in the evaluation of the basal culture conditions for the expression of PMP22 in TrJ/+ SCs, compared with that of +/+, discriminating the nuclear and cytoplasmic compartments. Overall, in both cellular domains, PMP22 expression was higher in TrJ/+ SCs compared to +/+ SCs. This result is in line, not only with works reporting the existence of cytoplasmic aggregates of PMP22 in TrJ/+ nerves ^{[29][69][73][74]} but also in agreement with previous work from the researchers' group, which determines the PMP22 expression in +/+ and TrJ/+ SCs inside the nucleus ^{[8][9]}.

The low dose of curcumin treatment was applied as a possible strategy to stimulate cellular detoxification pathways, and thus alleviate the neurodegenerative phenotype. The researchers' curcumin treatment had an additional effect caused by the EtOH vehicle (used to let cur-cumin solubilization for ulterior cell entry), in both +/+ and TrJ/+ SCs. This collateral effect, observed after six days of treatment, led the researchers to inquire about the use and, more importantly, about the validation of the vehicle so that it could be used without specific effects. In this sense, the recommended vehicles for solubilization of curcumin by the manufacturer are ethanol and dimethyl sulfoxide (DMSO). Although in many papers the effects of the sol-vents on culture viability are not shown ^{[52][64][65]} [76] (or are not discussed ^{[62][77][78]}), if they do not present cytotoxic effects, the reality is that both vehicles produce effects that vary widely depending on the cell type and duration of treatment ^{[77][79][80][81]}. The pre-established idea of the absence of the effects of these vehicles is recurrent in the literature. However, the contribution of the vehicle prevents the researchers from clearly discriminating the real effect and pharmacological potential of curcumin. For this reason, it must be settled as this constitutes a central point, to circumscribe the results only to the applied treatment. The researchers have tested the impact of DMSO as a vehicle of curcumin on fibroblast from +/+ sciatic nerves and the researchers' preliminary results seem to indicate an equivalent effect com-pared to that of EtOH. In

the researchers' primary results, different concentrations of DMSO were test-ed and in all cases, after five days of DMSO exposure, the viability and proliferation were significant differences compared with the untreated control. In future work, the researchers will seek to determine other strategies that allow solubilizing and targeting curcumin, evaluating at each step the cell viability to corroborate a negligible effect of the vehicle.

The impact of curcumin on the studied pathways in +/+ SC could not be analyzed in the TrJ/+ genotype, because all markers showed no significant difference between EtOH control and curcumin treatment. The results highlight the lability of the TrJ/+ genotype, expressed in its poor capacity to recover from the impact of the vehicle. In the literature, ethanol has been reported to increase ROS and mitochondrial dysfunction ^{[82][83][84]}. In zebrafish, at concentrations of 1% ethanol ^[82], a differential effect in mitochondrial function, with acute and chronic treatment, has been described. However, different performances showed that the mechanisms triggered are also dependent on administration protocols. Since the MTT assay is based on the conversion of tetrazolium to formazan by mitochondria in Schwann cells. Therefore, the increase in the ethanol control relative to the negative control could be due to dehydrogenases' functionality changes in response to the vehicle, rather than the normal increase in viability. In this sense, the fact that both genotypes showed equivalent responses in ethanol control to the negative control, supports this hypothesis.

In addition, the results found by the researchers' group show mitochondrial differences in the nerves of +/+ and TrJ/+ mice. From the analysis of electron microscopy images, the researchers obtained the number of mitochondria per fiber in the axonal and SC domains. These results show that there are differences in the number of mitochondria when comparing SCs of +/+ vs. TrJ/+ fibers. The morphological analysis considering the largest and smallest diameters of mitochondria, also showed apparent differences between +/+ and TrJ/+. Although there is a correlation between both parameters for the two genotypes, the equations representing the linear correlation are different. Furthermore, when looking at the genome expression, a qPCR analysis of the cytochrome b gene transcript level shows a higher amount of the transcript in +/+ compared to TrJ/+.

Thus, the study of the vehicle takes on particular relevance for in vitro approaches to the autophagy/mTOR and chaperone pathways in the TrJ/+ neurodegenerative genotype.

In the present work, the effect of ethanol was visualized equally between +/+ and TrJ/+ SCs, and the percentage of viability was calculated taking the ethanol control as 100%. The latter allowed the researchers to obtain the effect of curcumin to analyze the data. Despite the side effect of ethanol, the researchers were able to determine a 0.25 μ M curcumin as the lowest concentration that showed no difference in viability to the ethanol control.

From the study of the heat-stress markers' response pathway, the researchers were able to establish in +/+ SC the HSF1 and Hsp27 expression concordant with that reported in the literature. The HSF1 functions as a transcription factor, which under stress situations is activated and translocated to the nucleus, inducing the expression of the pathway's effectors such as Hsp27 ^{[85][86][87][88]} Furthermore, curcumin treatment allowed the researchers to observe an increase in HSF1 at the nuclear level and a decrease at the cytoplasmic level, together with an

increase in Hsp27, indicating a possible activation of this pathway. In addition, HDAC6 expression increased after treatment with curcumin. This increase of the protein, a member of basal autophagy in-volved in the selective elimination of aberrant protein aggregates ^{[89][90]} suggests that in the wild-type genotype, curcumin treatment may be stimulating this degradation pathway. Conversely, the increase in ribosomal expression under mTOR regulation ^{[91][92]} supports the idea of a favorable nutritional and energetic context in +/+ SC after curcumin treatment.

The impact of curcumin on the studied pathways in +/+ SC could not be analyzed in the TrJ/+ genotype because all of the markers showed no significant difference between the EtOH control and curcumin treatment. These results indicate a TrJ/+ genotype lability that prevents SCs from recovering from the ethanol shock. Thus, the study of the vehicle takes on particular relevance for in vitro approaches to the autophagy/mTOR and chaperone pathways in the TrJ/+ neurodegenerative genotype.

On the horizon of the TrJ/+ in vitro approaches, and its response to the action of different neuroprotective and antiinflammatory agents, such as curcumin, the analysis of the effects of the vehicle itself is an essential, inescapable, and conditional step for the fine-tuning of the experimental strategy to be applied.

3. Conclusions

The researchers' work established a new experimental strategy for obtaining enriched cultures of SC from +/+ and TrJ/+ mice. The researchers were able to determine a curcumin concentration with no effect on viability in both genotypes for an extended period of time, which allowed the researchers to study the expression of key autophagic-pathway markers in the accumulation of PMP22 protein in SCs. The researchers found an intrinsic ethanol effect in +/+ and TrJ/+ SC that was reversed by curcumin treatment in +/+, but not in TrJ/+ SC. These in vitro cultures allow pre-clinical investigations of promising therapeutic strategies or pharmacological compounds such as curcumin, for the alleviation of human-related peripheral neuropathies.

References

- 1. Skre, H. Genetic and clinical aspects of Charcot-Marie-Tooth's disease. Clin. Genet. 1974, 6, 98– 118.
- 2. Vallat, J.M. Dominantly inherited peripheral neuropathies. J. Neuropathol. Exp. Neurol. 2003, 62, 699–714.
- 3. Sereda, M.W.; Nave, K.A. Animal models of Charcot-Marie-Tooth disease type 1A. Neuromolecular Med. 2006, 8, 205–215.
- 4. Li, J.; Parker, B.; Martyn, C.; Natarajan, C.; Guo, J. The PMP22 gene and its related diseases. Mol. Neurobiol. 2013, 47, 673–698.

- 5. Parmantier, E.; Braun, C.; Thomas, J.L.; Peyron, F.; Martinez, S.; Zalc, B. PMP-22 expression in the central nervous system of the embryonic mouse defines potential transverse segments and longitudinal columns. J. Comp. Neurol. 1997, 378, 159–172.
- Parmantier, E.; Cabon, F.; Braun, C.; D'Urso, D.; Müller, H.W.; Zalc, B. Peripheral Myelin Protein-22 is Expressed in Rat and Mouse Brain and Spinal Cord Motoneurons. Eur. J. Neurosci. 1995, 7, 1080–1088.
- Chanson, J.B.; Echaniz-Laguna, A.; Blanc, F.; Lacour, A.; Ballonzoli, L.; Kremer, S.; Namer, I.J.; Lannes, B.; Tranchant, C.; Vermersch, P.; et al. Central nervous system abnormalities in patients with PMP22 gene mutations: A prospective study. J. Neurol. Neurosurg. Psychiatry 2013, 84, 392–397.
- Damián, J.P.; Vázquez Alberdi, L.; Canclini, L.; Rosso, G.; Bravo, S.O.; Martínez, M.; Uriarte, N.; Ruiz, P.; Calero, M.; Di Tomaso, M.V.; et al. Central Alteration in Peripheral Neuropathy of Trembler-J Mice: Hippocampal pmp22 Expression and Behavioral Profile in Anxiety Tests. Biomolecules 2021, 11, 601.
- Di Tomaso, M.V.; Vázquez Alberdi, L.; Olsson, D.; Cancela, S.; Fernández, A.; Rosillo, J.C.; Reyes Ábalos, A.L.; Álvarez Zabaleta, M.; Calero, M.; Kun, A. Colocalization Analysis of Peripheral Myelin Protein-22 and Lamin-B1 in the Schwann Cell Nuclei of Wt and TrJ Mice. Biomolecules 2022, 12, 456.
- Hou, J.; Wang, L.; Zhao, J.; Zhuo, H.; Cheng, J.; Chen, X.; Zheng, W.; Hong, Z.; Cai, J. Inhibition of protein PMP22 enhances etoposide-induced cell apoptosis by p53 signaling pathway in Gastric Cancer. Int. J. Biol. Sci. 2021, 17, 3145–3157.
- Sociali, G.; Visigalli, D.; Prukop, T.; Cervellini, I.; Mannino, E.; Venturi, C.; Bruzzone, S.; Sereda, M.W.; Schenone, A. Tolerability and efficacy study of P2X7 inhibition in experimental Charcot-Marie-Tooth type 1A (CMT1A) neuropathy. Neurobiol. Dis. 2016, 95, 145–157.
- 12. Dong, Y.; Simske, J.S. Vertebrate Claudin/PMP22/EMP22/MP20 family protein TMEM47 regulates epithelial cell junction maturation and morphogenesis. Dev. Dyn. 2016, 245, 653–666.
- Franke, W.W.; Heid, H.; Zimbelmann, R.; Kuhn, C.; Winter-Simanowski, S.; Dörflinger, Y.; Grund, C.; Rickelt, S. Transmembrane protein PERP is a component of tessellate junctions and of other junctional and non-junctional plasma membrane regions in diverse epithelial and epitheliumderived cells. Cell Tissue Res. 2013, 353, 99–115.
- Sereda, M.W.; Meyer Zu Hörste, G.; Suter, U.; Uzma, N.; Nave, K.A. Therapeutic administration of progesterone antagonist in a model of Charcot-Marie-Tooth disease (CMT-1A). Nat. Med. 2003, 9, 1533–1537.
- 15. Maeda, Y.; Kataoka, Y.; Sugaya, A.; Kariya, S.; Kobayashi, K.; Nishizaki, K. Steroid-dependent sensorineural hearing loss in a patient with Charcot-Marie-Tooth disease showing auditory

neuropathy. Auris. Nasus. Larynx 2015, 42, 249–253.

- Zu Horste, G.M.; Prukop, T.; Liebetanz, D.; Mobius, W.; Nave, K.A.; Sereda, M.W. Antiprogesterone therapy uncouples axonal loss from demyelination in a transgenic rat model of CMT1A neuropathy. Ann. Neurol. 2007, 61, 61–72.
- 17. Attardi, L.D.; Reczek, E.E.; Cosmas, C.; Demicco, E.G.; McCurrach, M.E.; Lowe, S.W.; Jacks, T. PERP, an apoptosis-associated target of p53, is a novel member of the PMP-22/gas3 family. Genes Dev. 2000, 14, 1835.
- 18. Bird, T.D. Charcot-Marie-Tooth (CMT) Hereditary Neuropathy Overview; University of Washington: Seattle, WA, USA, 1993.
- 19. Fortun, J.; Dunn, W.A.; Joy, S.; Li, J.; Notterpek, L. Emerging role for autophagy in the removal of aggresomes in Schwann cells. J. Neurosci. 2003, 23, 10672–10680.
- 20. Okamoto, Y.; Pehlivan, D.; Wiszniewski, W.; Beck, C.R.; Snipes, G.J.; Lupski, J.R.; Khajavi, M. Curcumin facilitates a transitory cellular stress response in trembler-J mice. Hum. Mol. Genet. 2013, 22, 4698–4705.
- 21. Suter, U.; Scherer, S.S. Disease mechanisms in inherited neuropathies. Nat. Rev. Neurosci. 2003, 4, 714–726.
- 22. Jaradeh, S.S. Hereditary Neuropathies. J. Clin. Neuromuscul. Dis. 2003, 5, 72-80.
- Valentijn, L.J.; Baas, F.; Wolterman, R.A.; Hoogendijk, J.E.; van den Bosch, N.H.A.; Zorn, I.; Gabreëls-Festen, A.A.W.M.; de Visser, M.; Bolhuis, P.A. Identical point mutations of PMP-22 in Trembler-J mouse and Charcot-Marie-Tooth disease type 1A. Nat. Genet. 1992, 2, 288–291.
- 24. Notterpek, L.; Ryan, M.C.; Tobler, A.R.; Shooter, E.M. PMP22 accumulation in aggresomes: Implications for CMT1A pathology. Neurobiol. Dis. 1999, 6, 450–460.
- Pareek, S.; Notterpek, L.; Snipes, G.J.; Naef, R.; Sossin, W.; Laliberté, J.; Iacampo, S.; Suter, U.; Shooter, E.M.; Murphy, R.A. Neurons promote the translocation of peripheral myelin protein 22 into myelin. J. Neurosci. 1997, 17, 7754–7762.
- Kun, A.; Rosso, G.; Canclini, L.; Bresque, M.; Romeo, C.; Cal, K.; Calliari, A.; Hanuz, A.; Roberto, J.; Roberto, J. The Schwann Cell-Axon Link in Normal Condition or Neuro-Degenerative Diseases: An Immunocytochemical Approach. In Applications of Immunocytochemistry; Dehghani, H., Ed.; InTech: London, UK, 2012; pp. 249–266. ISBN 978-953-51-5235-4.
- 27. Myers, J.K.; Mobley, C.K.; Sanders, C.R. The Peripheral Neuropathy-Linked Trembler and Trembler-J Mutant Forms of Peripheral Myelin Protein 22 are Folding-Destabilized. Biochemistry 2008, 47, 10620–10629.
- 28. Pantera, H.; Shy, M.E.; Svaren, J. Regulating PMP22 expression as a dosage sensitive neuropathy gene. Brain Res. 2020, 1726, 146491.

- 29. Kun, A.; Canclini, L.; Rosso, G.; Bresque, M.; Romeo, C.; Hanusz, A.; Cal, K.; Calliari, A.; Sotelo Silveira, J.; Sotelo, J.R. F-actin distribution at nodes of Ranvier and Schmidt-Lanterman incisures in mammalian sciatic nerves. Cytoskeleton 2012, 69, 486–495.
- Rosso, G.; Negreira, C.; Sotelo, J.R.; Kun, A. Myelinating and demyelinating phenotype of Trembler-J mouse (a model of Charcot-Marie-Tooth human disease) analyzed by atomic force microscopy and confocal microscopy. J. Mol. Recognit. 2012, 25, 247–255.
- 31. Notterpek, L.; Shooter, E.M.; Snipes, G.J. Upregulation of the endosomal-lysosomal pathway in the Trembler-J neuropathy. J. Neurosci. 1997, 17, 4190–4200.
- 32. Khalil, A.A.; Kabapy, N.F.; Deraz, S.F.; Smith, C. Heat shock proteins in oncology: Diagnostic biomarkers or therapeutic targets? Biochim. Biophys. Acta-Rev. Cancer 2011, 1816, 89–104.
- 33. Borges, T.J.; Wieten, L.; Van Herwijnen, M.J.C.; Broere, F.; Van der Zee, R.; Bonorino, C.; Van Eden, W. The anti-inflammatory mechanisms of Hsp70. Front. Immunol. 2012, 3, 95.
- 34. Chou, S.D.; Prince, T.; Gong, J.; Calderwood, S.K. mTOR is essential for the proteotoxic stress response, HSF1 activation and heat shock protein synthesis. PLoS ONE 2012, 7, e39679.
- 35. Morimoto, R.I. Regulation of the heat-shock transcriptional response: Cross talk between a family of heat-shock factors, molecular chaperones, and negative regulators. Genes Dev. 1998, 12, 3788–3796.
- Christie, M.; Chang, C.W.; Róna, G.; Smith, K.M.; Stewart, A.G.; Takeda, A.A.S.; Fontes, M.R.M.; Stewart, M.; Vértessy, B.G.; Forwood, J.K.; et al. Structural Biology and Regulation of Protein Import into the Nucleus. J. Mol. Biol. 2016, 428, 2060–2090.
- 37. Home, T.; Jensen, R.A.; Rao, R. Heat shock factor 1 in protein homeostasis and oncogenic signal integration. Cancer Res. 2015, 75, 907–912.
- 38. Watanabe, Y.; Tsujimura, A.; Taguchi, K.; Tanaka, M. HSF1 stress response pathway regulates autophagy receptor SQSTM1/p62-associated proteostasis. Autophagy 2017, 13, 133–148.
- 39. Schäfer, C.; Seeliger, H.; Bader, D.C.; Assmann, G.; Buchner, D.; Guo, Y.; Ziesch, A.; Palagyi, A.; Ochs, S.; Laubender, R.P.; et al. Heat shock protein 27 as a prognostic and predictive biomarker in pancreatic ductal adenocarcinoma. J. Cell. Mol. Med. 2012, 16, 1776–1791.
- 40. Garrido, C.; Brunet, M.; Didelot, C.; Zermati, Y.; Schmitt, E.; Kroemer, G. Heat shock proteins 27 and 70: Anti-apoptotic proteins with tumorigenic properties. Cell Cycle 2006, 5, 2592–2601.
- 41. Arrigo, A.-P.; Gibert, B. HspB1, HspB5 and HspB4 in Human Cancers: Potent Oncogenic Role of Some of Their Client Proteins. Cancers 2014, 6, 333–365.
- 42. Choi, S.; Lee, H.-J.; Jin, Y.B.; Jang, J.; Kang, G.; Lee, M.; Kim, C.-H.; Kim, J.; Yoon, S.S.; Lee, Y.; et al. MMP9 processing of HSPB1 regulates tumor progression. PLoS ONE 2014, 9, e85509.

- 43. Vidyasagar, A.; Wilson, N.A.; Djamali, A. Heat shock protein 27 (HSP27): Biomarker of disease and therapeutic target. Firogenes Tissue Repairs 2012, 5, 1–5.
- 44. Konishi, H.; Matsuzaki, H.; Tanaka, M.; Takemura, Y.; Kuroda, S.; Ono, Y.; Kikkawa, U. Activation of protein kinase B (Akt/RAC-protein kinase) by cellular stress and its association with heat shock protein Hsp27. FEBS Lett. 1997, 410, 493–498.
- 45. Wu, R.; Kausar, H.; Johnson, P.; Montoya-Durango, D.E.; Merchant, M.; Rane, M.J. Hsp27 regulates Akt activation and polymorphonuclear leukocyte apoptosis by scaffolding MK2 to Akt signal complex. J. Biol. Chem. 2007, 282, 21598–21608.
- 46. Rane, M.J.; Pan, Y.; Singh, S.; Powell, D.W.; Wu, R.; Cummins, T.; Chen, Q.; McLeish, K.R.; Klein, J.B. Heat shock protein 27 controls apoptosis by regulating Akt activation. J. Biol. Chem. 2003, 278, 27828–27835.
- Chittoor-Vinod, V.G.; Lee, S.; Judge, S.M.; Notterpek, L. Inducible HSP70 is critical in preventing the aggregation and enhancing the processing of PMP22. ASN Neuro 2015, 7, 1759091415569909.
- 48. Fortun, J.; Verrier, J.D.; Go, J.C.; Madorsky, I.; Dunn, W.A.; Notterpek, L. The formation of peripheral myelin protein 22 aggregates is hindered by the enhancement of autophagy and expression of cytoplasmic chaperones. Neurobiol. Dis. 2007, 25, 252–265.
- 49. Benoy, V.; Vanden Berghe, P.; Jarpe, M.; Van Damme, P.; Robberecht, W.; Van Den Bosch, L. Development of Improved HDAC6 Inhibitors as Pharmacological Therapy for Axonal Charcot– Marie–Tooth Disease. Neurotherapeutics 2017, 14, 417–428.
- Novelle, M.G.; Davis, A.; Price, N.L.; Ali, A.; Fürer-Galvan, S.; Zhang, Y.; Becker, K.; Bernier, M.; de Cabo, R. Caloric restriction induces heat shock response and inhibits B16F10 cell tumorigenesis both in vitro and in vivo. Aging (Albany. N. Y.) 2015, 7, 233–240.
- 51. Li, J.; Zhang, C.-X.; Liu, Y.-M.; Chen, K.-L.; Chen, G. A comparative study of anti-aging properties and mechanism: Resveratrol and caloric restriction. Oncotarget 2017, 8, 65717–65729.
- 52. Soh, J.W.; Marowsky, N.; Nichols, T.J.; Rahman, A.M.; Miah, T.; Sarao, P.; Khasawneh, R.; Unnikrishnan, A.; Heydari, A.R.; Silver, R.B.; et al. Curcumin is an early-acting stage-specific inducer of extended functional longevity in Drosophila. Exp. Gerontol. 2013, 48, 229–239.
- 53. Sinclair, D.A. Toward a unified theory of caloric restriction and longevity regulation. Mech. Ageing Dev. 2005, 126, 987–1002.
- Kume, S.; Uzu, T.; Kashiwagi, A.; Koya, D. SIRT1, a Calorie Restriction Mimetic, in a New Therapeutic Approach for Type 2 Diabetes Mellitus and Diabetic Vascular Complications. Endocr. Metab. Immune Disord. Drug Targets 2012, 10, 16–24.

- 55. Willcox, B.J.; Willcox, D.C. Caloric restriction, caloric restriction mimetics, and healthy aging in Okinawa: Controversies and clinical implications. Curr. Opin. Clin. Nutr. Metab. Care 2014, 17, 51–58.
- Singh, S.; Garg, G.; Singh, A.K.; Bissoyi, A.; Rizvi, S.I. Fisetin, a potential caloric restriction mimetic, attenuates senescence biomarkers in rat erythrocytes. Biochem. Cell Biol. 2019, 97, 480–487.
- 57. Ingram, D.K.; Zhu, M.; Mamczarz, J.; Zou, S.; Lane, M.A.; Roth, G.S.; deCabo, R. Calorie restriction mimetics: An emerging research field. Aging Cell 2006, 5, 97–108.
- 58. Mariño, G.; Pietrocola, F.; Madeo, F.; Kroemer, G. Caloric restriction mimetics: Natural/physiological pharmacological autophagy inducers. Autophagy 2014, 10, 1879–1882.
- 59. Madeo, F.; Pietrocola, F.; Eisenberg, T.; Kroemer, G. Caloric restriction mimetics: Towards a molecular definition. Nat. Rev. Drug Discov. 2014, 13, 727–740.
- Khandelwal, P.; Alam, A.; Choksi, A.; Chattopadhyay, S.; Poddar, P. Retention of Anticancer Activity of Curcumin after Conjugation with Fluorescent Gold Quantum Clusters: An in Vitro and in Vivo Xenograft Study. ACS Omega 2018, 3, 4776–4785.
- 61. Beevers, C.S.; Li, F.; Liu, L.; Huang, S. Curcumin inhibits the mammalian target of rapamycinmediated signaling pathways in cancer cells. Int. J. Cancer 2006, 119, 757–764.
- Watson, J.L.; Hill, R.; Yaffe, P.B.; Greenshields, A.; Walsh, M.; Lee, P.W.; Giacomantonio, C.A.; Hoskin, D.W. Curcumin causes superoxide anion production and p53-independent apoptosis in human colon cancer cells. Cancer Lett. 2010, 297, 1–8.
- Caillaud, M.; Msheik, Z.; Ndong-Ntoutoume, G.M.A.; Vignaud, L.; Richard, L.; Favreau, F.; Faye, P.A.; Sturtz, F.; Granet, R.; Vallat, J.M.; et al. Curcumin–cyclodextrin/cellulose nanocrystals improve the phenotype of Charcot-Marie-Tooth-1A transgenic rats through the reduction of oxidative stress. Free Radic. Biol. Med. 2020, 161, 246–262.
- 64. Barzegar, A.; Moosavi-Movahedi, A.A. Intracellular ROS protection efficiency and free radicalscavenging activity of curcumin. PLoS ONE 2011, 6, e26012.
- Perugini, J.; Di Mercurio, E.; Tossetta, G.; Severi, I.; Monaco, F.; Reguzzoni, M.; Tomasetti, M.; Dani, C.; Cinti, S.; Giordano, A. Biological Effects of Ciliary Neurotrophic Factor on hMADS Adipocytes. Front. Endocrinol. (Lausanne) 2019, 10.
- Kang, S.K.; Cha, S.H.; Jeon, H.G. Curcumin-induced histone hypoacetylation enhances caspase-3-dependent glioma cell death and neurogenesis of neural progenitor cells. Stem Cells Dev. 2006, 15, 165–174.
- 67. Liu, X.; You, L.; Tarafder, S.; Zou, L.; Fang, Z.; Chen, J.; Lee, C.H.; Zhang, Q. Curcumin-releasing chitosan/aloe membrane for skin regeneration. Chem. Eng. J. 2019, 359, 1111–1119.

- Thaloor, D.; Miller, K.J.; Gephart, J.; Mitchell, P.O.; Pavlath, G.K. Systemic administration of the NF-κB inhibitor curcumin stimulates muscle regeneration after traumatic injury. Am. J. Physiol.-Cell Physiol. 1999, 277, C320–C329.
- 69. Khajavi, M.; Shiga, K.; Wiszniewski, W.; He, F.; Shaw, C.A.; Yan, J.; Wensel, T.G.; Snipes, G.J.; Lupski, J.R. Oral Curcumin Mitigates the Clinical and Neuropathologic Phenotype of the Trembler-J Mouse: A Potential Therapy for Inherited Neuropathy. Am. J. Hum. Genet. 2007, 81, 438–453.
- 70. Fortun, J.; Go, J.C.; Li, J.; Amici, S.A.; Dunn, W.A.; Notterpek, L. Alterations in degradative pathways and protein aggregation in a neuropathy model based on PMP22 overexpression. Neurobiol. Dis. 2006, 22, 153–164.
- 71. Zhou, Y.; Bazick, H.; Miles, J.R.; Fethiere, A.I.; Al Salihi, M.O.; Fazio, S.; Tavori, H.; Notterpek, L. A neutral lipid-enriched diet improves myelination and alleviates peripheral nerve pathology in neuropathic mice. Exp. Neurol. 2019, 321, 113031.
- 72. Hara, T.; Nakamura, K.; Matsui, M.; Yamamoto, A.; Nakahara, Y.; Suzuki-Migishima, R.; Yokoyama, M.; Mishima, K.; Saito, I.; Okano, H.; et al. Suppression of basal autophagy in neural cells causes neurodegenerative disease in mice. Nature 2006, 441, 885–889.
- Fortun, J.; Li, J.; Go, J.C.; Fenstermaker, A.; Fletcher, B.S.; Notterpek, L. Impaired proteasome activity and accumulation of ubiquitinated substrates in a hereditary neuropathy model. J. Neurochem. 2005, 92, 1531–1541.
- 74. Ryan, M.C.; Shooter, E.M.; Notterpek, L. Aggresome formation in neuropathy models based on peripheral myelin protein 22 mutations. Neurobiol. Dis. 2002, 10, 109–118.
- Pareek, S.; Suter, U.; Snipes, G.J.; Welcher, A.A.; Shooter, E.M.; Murphy, R.A. Detection and processing of peripheral myelin protein PMP22 in cultured Schwann cells. J. Biol. Chem. 1993, 268, 10372–10379.
- 76. Tello Velasquez, J.; Watts, M.E.; Todorovic, M.; Nazareth, L.; Pastrana, E.; Diaz-Nido, J.; Lim, F.; Ekberg, J.A.K.; Quinn, R.J.; St John, J.A. Low-dose curcumin stimulates proliferation, migration and phagocytic activity of olfactory ensheathing cells. PLoS ONE 2014, 9, e111787.
- 77. Zhao, Z.; Li, X.; Li, Q. Curcumin accelerates the repair of sciatic nerve injury in rats through reducing Schwann cells apoptosis and promoting myelinization. Biomed. Pharmacother. 2017, 92, 1103–1110.
- Abuelba, H.; Cotrutz, C.E.; Stoica, B.A.; Stoica, L.; Olinici, D.; Petreuş, T. In vitro evaluation of curcumin effects on breast adenocarcinoma 2D and 3D cell cultures. Rom. J. Morphol. Embryol. 2015, 56, 71–76.
- 79. Galvao, J.; Davis, B.; Tilley, M.; Normando, E.; Duchen, M.R.; Cordeiro, M.F. Unexpected lowdose toxicity of the universal solvent DMSO. FASEB J. 2014, 28, 1317–1330.

- Ilieva, Y.; Dimitrova, L.; Zaharieva, M.M.; Kaleva, M.; Alov, P.; Tsakovska, I.; Pencheva, T.; Pencheva-El Tibi, I.; Najdenski, H.; Pajeva, I. Cytotoxicity and Microbicidal Activity of Commonly Used Organic Solvents: A Comparative Study and Application to a Standardized Extract from Vaccinium macrocarpon. Toxics 2021, 9, 92.
- 81. Adler, S.; Pellizzer, C.; Paparella, M.; Hartung, T.; Bremer, S. The effects of solvents on embryonic stem cell differentiation. Toxicol. Vitr. 2006, 20, 265–271.
- Müller, T.E.; Nunes, M.E.M.; Rodrigues, N.R.; Fontana, B.D.; Hartmann, D.D.; Franco, J.L.; Rosemberg, D.B. Neurochemical mechanisms underlying acute and chronic ethanol-mediated responses in zebrafish: The role of mitochondrial bioenergetics. Neurochem. Int. 2019, 131, 104584.
- 83. Pereira, R.B.; Andrade, P.B.; Valentão, P. A Comprehensive View of the Neurotoxicity Mechanisms of Cocaine and Ethanol. Neurotox. Res. 2015, 28, 253–267.
- 84. Yang, F.; Luo, J. Endoplasmic reticulum stress and ethanol neurotoxicity. Biomolecules 2015, 5, 2538–2553.
- 85. Gomez-Pastor, R.; Burchfiel, E.T.; Thiele, D.J. Regulation of heat shock transcription factors and their roles in physiology and disease. Nat. Rev. Mol. Cell Biol. 2018, 19, 4–19.
- 86. Arrigo, A.-P. Structure-functions of HspB1 (Hsp27). Methods Mol. Biol. 2011, 787, 105–119.
- 87. Strauch, A.; Haslbeck, M. The function of small heat-shock proteins and their implication in proteostasis. Essays Biochem. 2016, 60, 163–172.
- 88. Huang, C.; Wu, J.; Xu, L.; Wang, J.; Chen, Z.; Yang, R. Regulation of HSF1 protein stabilization: An updated review. Eur. J. Pharmacol. 2018, 822, 69–77.
- Lee, J.Y.; Koga, H.; Kawaguchi, Y.; Tang, W.; Wong, E.; Gao, Y.S.; Pandey, U.B.; Kaushik, S.; Tresse, E.; Lu, J.; et al. HDAC6 controls autophagosome maturation essential for ubiquitinselective quality-control autophagy. EMBO J. 2010, 29, 969–980.
- 90. Richter-Landsberg, C.; Leyk, J. Inclusion body formation, macroautophagy, and the role of HDAC6 in neurodegeneration. Acta Neuropathol. 2013, 126, 793–807.
- 91. Mayer, C.; Grummt, I. Ribosome biogenesis and cell growth: mTOR coordinates transcription by all three classes of nuclear RNA polymerases. Oncogene 2006, 25, 6384–6391.
- 92. Sarbassov, D.D.; Ali, S.M.; Sabatini, D.M. Growing roles for the mTOR pathway. Curr. Opin. Cell Biol. 2005, 17, 596–603.

Retrieved from https://encyclopedia.pub/entry/history/show/51812