

Phytoremediation of Cadmium

Subjects: Biology

Contributor: Mirza Hasanuzzaman

Cadmium (Cd) is one of the most toxic metals in the environment, and has noxious effects on plant growth and production. Cd-accumulating plants showed reduced growth and productivity. Therefore, remediation of this non-essential and toxic pollutant is a prerequisite. Plant-based phytoremediation methodology is considered as one a secure, environmentally friendly, and cost-effective approach for toxic metal remediation. Phytoremediating plants transport and accumulate Cd inside their roots, shoots, leaves, and vacuoles. Phytoremediation of Cd-contaminated sites through hyperaccumulator plants proves a ground-breaking and profitable choice to combat the contaminants.

Keywords: heavy metals ; phytoremediation ; antioxidant defense system ; genetic engineering ; microbes ; metallothionein ; omics ; phytochelatins

1. Introduction

Cadmium (Cd) is a non-essential element for plants and humans but is present in many soils in excessive amounts ^{[1][2]}. When it enters into the food chain, it poses a major threat to the living biota. The control of Cd accumulation in plants is complicated by the fact that most of the essential nutrient transporters, such as copper (Cu), manganese (Mn), iron (Fe), and zinc (Zn), also facilitate Cd uptake ^[2]. Cd stress alters plant growth, as evident from a reduced dry matter yield and leaf area, and stunted growth ^{[3][4][5][6]}. Cd affects plant growth at both the morphological and physiological level ^[7]. At the whole plant level, Cd toxicity includes leaf chlorosis, a delay in the growth rate, and inhibition of respiration and photosynthesis ^[8], increased oxidative damage, and decreased nutrient uptake ability ^[9].

Generally, Cd occurs in sedimentary rocks (0.3 mg kg⁻¹), lithosphere (0.2 mg kg⁻¹), and soil (0.53 mg kg⁻¹) ^[10]. Cd enrichment in soil occurs from both anthropogenic and natural sources ^[11]. Geologically weathering of rocks is the major natural source of Cd contaminants ^{[12][13]}, while primary anthropogenic sources of Cd include agrochemicals, manufacturing, vehicular emission, irrigation wastewater, smelting, and mining ^{[14][15]}. Moreover, improper and uncontrolled waste disposal practices, sea spray, windblown dust, forest fires, and volcanic eruption also increase the Cd level in soil ^{[12][13][14]}.

Apart from this, Cd toxicity has been reported to damage human physiology by various means, such as Cd-contaminated water and food. For example, Cd exposure influences the human male reproductive organs/system and deteriorates spermatogenesis and semen quality, especially sperm motility and hormonal synthesis/release. Based on experimental and human studies, it also impairs female reproduction and the reproductive hormonal balance and affects menstrual cycles ^{[16][17]}. In animals, experimental studies revealed that Cd and Cd compounds (referred to as Cd) by multiple routes of exposure prompt benign and malignant tumor formation at various sites in many species of experimental animals ^[18]. Besides, environmental Cd contact can cause pancreatic cancer in animals ^[19].

Efficient and economical remediation of contaminated urban and agriculture land is a pressing need for sustainable agriculture development prospects. Different methods, like biological, chemical, and physical, have been used for the remediation of heavy metal contaminants from soil. Some of them face limitations due to mechanical limitations, logistical problems, time, and cost. Soil remediation techniques include physical, chemical, and biological remediation; electrokinetics; and phytoremediation. Physical remediation includes both the soil high replacement method and thermal desorption method. In the soil replacement method, clean soil is used for partial and full replacement of contaminated soil ^{[20][21]}. Importing new soil dilutes the contaminated soil; however, this practice is only useful for small-scale severely contaminated soil.

Chemical remediation is a mechanical process used for leaching the contaminated soil by using liquids enriched with solvents, freshwater, and chelating agents ^[20]. Researchers found that ethylenediaminetetraacetic acid (EDTA) is an effective chelating agent for soil washing. Recent studies have shown that biosurfactants, such as sophorolipids, saponin, and rhamnolipids, can efficiently remove Cd from contaminated soils ^[22]. According to Juwarkar et al. ^[23], more than 92%

of Cd was removed with 0.1% di-rhamnolipid. In chemical oxidation, oxidants, such as Fe²⁺-activated peroxy monosulfate, are used to degrade and oxidize the contaminant particles [24]. Bioremediation is the use of microorganisms, plants, and microbial or plant enzymes to treat the contaminated soil through natural biodegradation. Some Cd-removing microorganisms include *Aspergillus niger* (fungus) [25], *Pleurotus ostreatus* [26], *Spergillus versicolor* [27], *Fomitopsis pinicola*, *Pseudomonas aeruginosa*, *Streptomyces*, and *Bacillus* [28]. Electrokinetics is another technique to remediate heavy metals from soil by using an electrical current [29][30]. In this context, Shen et al. [31] reported an accumulation of 99% of total Cd at the cathode after remediating soil with approaching anode electro-kinetics. Similarly, Li et al. [32] reported a removal of 97.32% of the total Cd through electroosmotic and electromigration, which was positively correlated with the electric voltage. Recently, it has become the best technique and seems to be a promising alternative to conventional approaches.

Phytoremediation is a cost-effective and eco-friendly technique for remediating soils. Notably, phytoremediating plants uptake and accumulate Cd inside their roots, shoots, leaves, and vacuoles. Still, it takes a long time to provide fruitful results because phytoremediation is still under the investigation and progress phase, and several technical barriers have to be overcome. In the present review, we illustrate the recent advancements in the physiological, biochemical, and molecular mechanisms associated with Cd phytoremediation. Additionally, the potential of omics and genetic engineering approaches are outlined for the efficient remediation of a Cd-contaminated environment.

2. Plant Responses to Cadmium Toxicity

The toxic effects of Cd on plant growth and metabolism differ among plant species [3]. The Cd concentration in plants is a direct function of its presence in the soil. An increase in Cd concentration in the growth medium led to a subsequent rise in its accumulation in different parts of the plants [33][34]. It might alter the plant growth and metabolism even if present in a minute amount [33][35]. Cd application in basil seeds delayed the germination period from 4.66 to around 7–10 days [5]. Several other studies have reported high Cd accumulation in wheat seedling roots compared to shoots [36][37].

In some studies, Cd toxicity is linked with the low dry matter accumulation in roots [4], turning them black [38], and leads to a reduction in lateral root growth [39]. It has further been associated with root development with the mature apoplastic pathway, enhanced porosity, and few root tips per surface area in rice plants [40]. In the genus *Citrus*, seedlings wilted and turned yellow, followed by eventual death under cadmium chloride (CdCl₂) treatment [41]. In contrast, tomato plants were reported to endure short-term exposure to high (250 μM) CdCl₂ concentrations [42]. Increased exposure to Cd in carrots and radish significantly inhibited the development of radicals due to increased Cd accumulation in roots [43]. Moreover, a considerably high metal concentration was found in parsley seedlings under Cd stress, but the plants did not show any visual stress symptoms [44].

The morphological, biochemical, and physiological effects on plant growth are more pronounced with a high concentration of Cd [39]. An evaluation of the morpho-physiological growth parameters of tomatoes showed that at high Cd concentrations, the root-shoot growth decreased at a relative rate, which could be attributed to the lesser water content in the seedlings due to reduced imbibition. The literature suggests that Cd stress decreased the root-shoot length in wheat [45], peas [46], *Corchorus capsularis* [47], and *Suaeda glauca* [48]. Further, it has been documented to reduce the dry weight in wheat [49], maize [50], and tomatoes [51].

Wu et al. [52] described an increase in the net photosynthesis rate by Cd application, which translated to an increase in the net biomass. On the contrary, Cd toxicity has been reported to inhibit plant growth by decreasing the water-use efficiency (WUE) and the net rate of photosynthesis [53]. A significant reduction in the leaf area and dry mass in female *Populus cathayana* under Cd stress has been observed [54]. In contrast, growth inhibition and a disturbance in photosynthetic performance has been reported in Cd-stressed tomato [55] and cucumber [56]. Cd exposure has further been reported to decrease the stomatal conductance and net rate of photosynthesis in rapeseed [57].

Cd toxicity also affects the plasma membrane of plants, which could be attributed to electrolyte leakage [58], and membrane proteins like H-ATPase inhibition [42]. It has been known to affect the DNA repair system [59]; thus, the stability of the genomic template was distinctly reduced in *Phaseolus vulgaris* [60] and peas [61] in response to the direct application of Cd.

Reactive oxygen species (ROS) generation in response to Cd-induced oxidative stress affects the electron transport and leaks electrons to molecular oxygen [62]. Cd was found to be toxic for peanuts at a higher dosage, marked by the production and accumulation of ROS in the cytosol. Moreover, it damaged the integrity and selective transport system of plasma membranes, leading to metal transport in cells [63]. Furthermore, the overproduction of ROS in wheat seedlings

upon Cd exposure, marked by an increase in the hydrogen peroxide (H₂O₂) content and malondialdehyde (MDA) level, was linked to genotoxicity [64]. Being sessile, plants try to elude its harmful effects by adopting various defense mechanisms, which include antioxidant activation and other mechanisms of metal homeostasis [65]. In response, plants have developed enzymatic and non-enzymatic antioxidant mechanisms. Increased activities of catalase (CAT), superoxide dismutase (SOD), ascorbate peroxidase (APX), and peroxidase (POD) were found against increased Cd stress in *Brassica juncea* [66]. In another study, the glutamate-mediated alleviation of Cd toxicity reduced ROS-induced membrane lipid peroxidation, metal uptake, and translocation to rice shoots, and improved the chlorophyll biosynthesis [67].

3. Phytoremediation Processes and Their Salient Features

Phytoremediation refers to the biological cleaning of the environment (soil, water, and air) by plants. Plants make a symbiotic association with microorganisms, which helps in the remediation of the soil, particularly from heavy metals and organic pollutants. Phytoremediation is generally considered as a green technology because of its excellent decontamination ability of heavy metals with a minimum influx of secondary waste to the environment. Alternatively, phytoremediation is highly acceptable among the general public due to its ease of application, low cost, and environmentally friendly nature [1][2]. However, hampered growth activities, such as reduced biomass and increased sensitivity to Cd, were observed in the plants involved in phytoremediation processes [6].

Phytoremediation involves various processes, such as phytoextraction, phytoaccumulation, phytovolatilization, phytostabilization, and phytotransformation. The phytoextraction and phytoaccumulation processes work in association. For instance, during phytoextraction, plants uptake heavy metals, such as Cd, Zn, nickel (Ni), chromium (Cr), and other minerals and nutrients from the soil. After this, these elements accumulate in the shoots and leaves with the help of the phytoaccumulation mechanism [6]. Many plants species have been reported previously for their high accumulation capacity; these are potential candidates for phytoremediation.

In Cd phytoremediation, plants are often used to absorb or translocate Cd into harvestable plant parts. Plants have evolved many diverse adaptations to maintain normal growth even under high Cd-contaminated soils, which also includes detoxification mechanisms [68]. The Cd concentration in plant parts shows the following trend: root > stem > leaves [69]. Many techniques are being used to increase the efficiency of Cd phytoremediation (Table 1).

Table 1. Types of phytoremediation approaches and their specific methods. Abbreviations are explained in the text.

Types	Process	Mechanism	Plants	References
	Bioaugmentation-assisted phytoextraction	Combined with mycorrhiza	<i>Suaeda salsa</i> and <i>Trichoderma asperellum</i>	[70]
Phytoextraction/ Phytoaccumulation	Chelated-assisted phytoextraction	Chelates like EDTA, SDS, and EGTA	<i>Fagopyrum esculentum</i>	[71]
	Phytomining	Phytoextraction for commercial use, like silver (Ag), Ni	<i>Alyssum murale</i> , <i>Odontarrhena chalcidica</i>	[72]
Phyostabilization	Organic fertilizers, biochar	Immobilization of Cd by using biomolecules	<i>Viola surinamensis</i> , <i>Boehmeria nivea</i>	[73][74]
	Biosorption	Metals are absorbed bound in cells, used for phytoremediation	<i>Lythrum salicaria</i>	[75]
Phytofiltration	Rhizofiltration	Metals are absorbed and bound on only roots	<i>Micranthemum umbrosum</i>	[76]
	Blastofiltration	Metals are absorbed and bound on only seedlings	<i>Moringa Oliefera</i> , <i>Cucumis melo</i> , <i>Abelmoschus esculentus</i> , <i>Ricinus communis</i>	[77][78]
	Caulofiltration	Metals are bond and absorbed on excised plant	<i>Berkheya coddii</i>	[79]

Types	Process	Mechanism	Plants	References
Phytostimulation	Fungi, bacteria	Phytoremediation with the intervention of microorganisms in different terms to remediate soil with organic pollutants	<i>Rumex K-1 (Rumex patientia × R. timschmicus)</i> <i>Viola baoshanensis</i> . <i>Vertiveria zizanioides</i>	[80][81]

3.1. Phytoextraction

This technique is used to absorb inorganic and organic contaminants through the stem and roots. Plants that are already growing in the ecosystem should be chosen for this technique. After harvest, they are exposed to another method known as composition, or burned in an incinerator [82]. Hyperaccumulator families, such as *Scrophulariaceae*, *Lamiaceae*, *Asteraceae*, *Euphorbiaceae*, and *Brassicaceae*, are essential for this technique. Moreover, some particular plant species, like *Celosia argentea* [83], *Salix mucronata* [84], *Cassia alata* [85], *Vigna unguiculata*, *Solanum melonaena*, *Momordica charantia* [86], *Nicotiana tabacum*, *Kummerowia striata* [87], and *Swietenia macrophylla* [88], may be used as potential plant choices to increase the process of Cd phytoextraction. Moreover, a sub-division of phytoextraction, known as chelate-assisted phytoextraction, is also used as a possible solution for metals that have no hyperaccumulator species. Several amino polycarboxylic acid and chelating agents have been applied to soil to increase the solubility of trace elements. For instance, EDTA-assisted phytoextraction of Cd was preferred by Farid et al. [89]. Similarly, citric acid was used as a chelating agent to increase the Cd uptake ability of jute mallow (*Corchorus olitorius*) [90].

Phytoextraction helps to reduce metalloid toxicity by improving substrate geochemistry for future colonization of native plants [91]. It is an effective, affordable, environmentally friendly, and potentially cost-effective technique for remediating soils [92]. Despite the generally agreed advantages of phytoextraction, there are some disadvantages, such as the time required for the remediation of highly contaminated soils may be decades [93], and a limitation for mine waste applications [94]. Mostly hyperaccumulator plants have developed the capacity to accumulate only one metal and may be sensitive to the presence of other elements [93].

3.2. Phytostabilization

There has been a progressing shift from phytoextraction to phytostabilization. Phytostabilization is the ability of plants to store and immobilized heavy metals by binding with biomolecules; this process prevents metal transport, and converts them into less toxic substances [95]. Most of the plants growing on contaminated soils are not hyperaccumulators but work as excluders. An excluder transforms the metals and metalloids into a less toxic mobile form without extracting them from the soil and accumulates these compounds in roots by absorption or precipitation within the rhizosphere [96]. Recently, promising results of *Viola surinamensis* for Cd phytostabilization have been documented [97]. Likewise, *Miscanthus x giganteus* [73], and oats and white mustard [98] also have phytostabilization potential for Cd. In another example, the putative role of Fe-Si-Ca, organic fertilizers, and coconut shell biochar has been reported to enhance the phytostabilization ability of *Boehmeria nivea* L. for Cd [99].

Phytostabilization is one emerging ecofriendly phytotechnology, which immobilizes the environmental toxins [100]. Roots take part in phytostabilization, so the metal availability is reduced to the plants, thus reducing the exposure to the other tropic level of the environment [101]. At the same time, the major disadvantage is the fact that pollutant remains in the soil or in the root system, generally in the rhizosphere [74].

3.3. Phytofiltration

Phytofiltration is categorized as rhizofiltration that includes blastofiltration (use seedlings) and caulofiltration (use of excised plant) (Table 1) [102]. Rhizofiltration is the remediation of water in which roots effectively absorb contaminants [103]. In rhizofiltration, contaminant clings or assimilates to the roots, and can be transported to the plants. This method is mostly used to sterilize underground wastes or polluted water. Mostly radioactive substances or metals are removed by this method. Abhilash et al. [104] used the phytofiltration technique to increase the Cd uptake from water by using *Limncharis flava* as an experimental plant. Islam et al. [76] reported the phytofiltration capability of *Micranthemum umbrosum* to remove Cd and arsenic (As) from a hydroponic system. In another experiment, the rhizofiltration potential of *Arunda donax* for Zn and Cd removal, it and recommended the use of the rhizofiltration technique for Cd elimination [105].

It is a cost-effective technique, and plants act as solar-driven pumps to extract the contaminants from the environment [103][76]. However, any contaminant below the rooting depth is not extracted. It is a time-consuming technique and will not

suffice for the extraction of both organic and metal contaminants [76][106].

3.4. Phytostimulation

Phytostimulation is a technique used to boost the process of phytoremediation by stimulating the root-released compounds to enhance microbial activities. These exudates enhance microbial growth by fulfilling their nutrient requirements. This process is being used in rhizoremediation technologies. It is a low-cost technique for Cd removal and other organic compounds [107]. Another method is the addition of resistant microbial inoculants into the soil, which can cause the accumulation of heavy metals, including Cd [108].

It is a more effective technique for converting toxic contaminants into non-toxic chemicals. Both in situ and ex situ practices can be done with low-cost treatments [109]. Microbes are able to help limit the growth of plant pathogens and increase nitrogen (N) fixation [110]. However, it is a more time-consuming technique, and the use of volatile and biodegradable compounds ex situ is not an easy practice. The process is sensitive to the level of toxicity in soil, and in some cases, incomplete breakdown of the organic compounds is observed. Moreover, well-controlled monitoring is required for this technique [109].

References

1. Dhaliwal, S.S.; Singh, J.; Taneja, P.K.; Mandal, A. Remediation techniques for removal of heavy metals from the soil contaminated through different sources: A review. *Environ. Sci. Pollut. Res.* 2020, 27, 1319–1333.
2. Huang, R.; Dong, M.; Mao, P.; Zhuang, P.; Paz-Ferreiro, J.; Li, Y.; Li, Y.; Hu, X.; Netherway, P.; Li, Z. Evaluation of phytoremediation potential of five Cd (hyper) accumulators in two Cd contaminated soils. *Sci. Total Environ.* 2020, 721, 137581.
3. Baliardini, C.; Meyer, C.-L.; Salis, P.; Saumitou-Laprade, P.; Verbruggen, N. Cation EXCHANGER1 cosegregates with cadmium tolerance in the metal hyperaccumulator *Arabidopsis halleri* and plays a role in limiting oxidative stress in *Arabidopsis* spp. *Plant Physiol.* 2015, 169, 549–559.
4. Borges, K.L.R.; Salvato, F.; Alcântara, B.K.; Nalin, R.S.; Piotto, F.Â.; Azevedo, R.A. Temporal dynamic responses of roots in contrasting tomato genotypes to cadmium tolerance. *Ecotoxicology* 2018, 27, 245–258.
5. Fattahi, B.; Arzani, K.; Souri, M.K.; Barzegar, M. Effects of cadmium and lead on seed germination, morphological traits, and essential oil composition of sweet basil (*Ocimum basilicum* L.). *Ind. Crops Prod.* 2019, 138, 111584.
6. Shah, V.; Daverey, A. Phytoremediation: A multidisciplinary approach to clean up heavy metal contaminated soil. *Environ. Technol. Innov.* 2020, 18, 100774.
7. Shanying, H.; Xiaoe, Y.; Zhenli, H.; Baligar, V.C. Morphological and physiological responses of plants to cadmium toxicity: A review. *Pedosphere* 2017, 27, 421–438.
8. Navarro-León, E.; Oviedo-Silva, J.; Ruiz, J.M.; Blasco, B. Possible role of HMA4a TILLING mutants of *Brassica rapa* in cadmium phytoremediation programs. *Ecotoxicol. Environ. Saf.* 2019, 180, 88–94.
9. Mohamed, A.A.; Castagna, A.; Ranieri, A.; di Toppi, L.S. Cadmium tolerance in *Brassica juncea* roots and shoots is affected by antioxidant status and phytochelatin biosynthesis. *Plant Physiol. Biochem.* 2012, 57, 15–22.
10. Kabata-Pendias, A. *Trace Elements in Soils and Plants*; CRC Press: Boca Raton, FL, USA, 2010.
11. Pan, L.-B.; Ma, J.; Wang, X.-L.; Hou, H. Heavy metals in soils from a typical county in Shanxi Province, China: Levels, sources and spatial distribution. *Chemosphere* 2016, 148, 248–254.
12. Khan, S.; Rehman, S.; Khan, A.Z.; Khan, M.A.; Shah, M.T. Soil and vegetables enrichment with heavy metals from geological sources in Gilgit, northern Pakistan. *Ecotoxicol. Environ. Saf.* 2010, 73, 1820–1827.
13. Liu, Y.; Xiao, T.; Ning, Z.; Li, H.; Tang, J.; Zhou, G. High cadmium concentration in soil in the Three Gorges region: Geogenic source and potential bioavailability. *Appl. Geochem.* 2013, 37, 149–156.
14. Khan, S.; Munir, S.; Sajjad, M.; Li, G. Urban park soil contamination by potentially harmful elements and human health risk in Peshawar City, Khyber Pakhtunkhwa, Pakistan. *J. Geochem. Explor.* 2016, 165, 102–110.
15. Nawab, J.; Khan, S.; Aamir, M.; Shamshad, I.; Qamar, Z.; Din, I.; Huang, Q. Organic amendments impact the availability of heavy metal (loid) s in mine-impacted soil and their phytoremediation by *Penisetum americanum* and *Sorghum bicolor*. *Environ. Sci. Pollut. Res.* 2016, 23, 2381–2390.
16. Kumar, S.; Sharma, A. Cadmium toxicity: Effects on human reproduction and fertility. *Rev. Environ. Health* 2019, 34, 327–338.

17. Genchi, G.; Sinicropi, M.S.; Lauria, G.; Carocci, A.; Catalano, A. The effects of cadmium toxicity. *Int. J. Environ. Res. Public Health* 2020, 17, 3782.
18. Huff, J.; Lunn, R.M.; Waalkes, M.P.; Tomatis, L.; Infante, P.F. Cadmium-induced cancers in animals and in humans. *Int. J. Occup. Environ. Health* 2007, 13, 202–212.
19. Djordjevic, V.R.; Wallace, D.R.; Schweitzer, A.; Boricic, N.; Knezevic, D.; Matic, S.; Grubor, N.; Kerkez, M.; Radenkovic, D.; Bulat, Z.; et al. Environmental cadmium exposure and pancreatic cancer: Evidence from case control, animal and in vitro studies. *Environ. Int.* 2019, 128, 353–361.
20. Yao, Z.; Li, J.; Xie, H.; Yu, C. Review on remediation technologies of soil contaminated by heavy metals. *Procedia Environ. Sci.* 2012, 16, 722–729.
21. Paz-Ferreiro, J.; Gascó, G.; Méndez, A.; Reichman, S.M. Soil Pollution and Remediation. *Int. J. Environ. Res. Public Health* 2018, 15, 1657.
22. Maity, J.P.; Huang, Y.M.; Fan, C.-W.; Chen, C.-C.; Li, C.-Y.; Hsu, C.-M.; Chang, Y.-F.; Wu, C.-I.; Chen, C.-Y.; Jean, J.-S. Evaluation of remediation process with soapberry derived saponin for removal of heavy metals from contaminated soils in Hai-Pu, Taiwan. *J. Environ. Sci.* 2013, 25, 1180–1185.
23. Juwarkar, A.A.; Nair, A.; Dubey, K.V.; Singh, S.; Devotta, S. Biosurfactant technology for remediation of cadmium and lead contaminated soils. *Chemosphere* 2007, 68, 1996–2002.
24. Guo, J.; Zhou, Y. Transformation of heavy metals and dewaterability of waste activated sludge during the conditioning by Fe²⁺-activated peroxydisulfate oxidation combined with rice straw biochar as skeleton builder. *Chemosphere* 2020, 238, 124628.
25. Ren, W.-X.; Li, P.-J.; Geng, Y.; Li, X.-J. Biological leaching of heavy metals from a contaminated soil by *Aspergillus niger*. *J. Hazard. Mater.* 2009, 167, 164–169.
26. Kapahi, M.; Sachdeva, S. Mycoremediation potential of *Pleurotus* species for heavy metals: A review. *Bioresour. Bioprocessing* 2017, 4, 32.
27. Fazli, M.M.; Soleimani, N.; Mehrasbi, M.; Darabian, S.; Mohammadi, J.; Ramazani, A. Highly cadmium tolerant fungi: Their tolerance and removal potential. *J. Environ. Health Sci. Eng.* 2015, 13, 19.
28. Bagot, D.; Lebeau, T.; Jezequel, K. Microorganisms for remediation of cadmium-contaminated soils. *Environ. Chem. Lett.* 2006, 4, 207–211.
29. Tang, J.; He, J.; Liu, T.; Xin, X.; Hu, H. Removal of heavy metal from sludge by the combined application of a biodegradable biosurfactant and complexing agent in enhanced electrokinetic treatment. *Chemosphere* 2017, 189, 599–608.
30. Virkutyte, J.; Sillanpää, M.; Latostenmaa, P. Electrokinetic soil remediation—Critical overview. *Sci. Total Environ.* 2002, 289, 97–121.
31. Shen, Z.; Chen, X.; Jia, J.; Qu, L.; Wang, W. Comparison of electrokinetic soil remediation methods using one fixed anode and approaching anodes. *Environ. Pollut.* 2007, 150, 193–199.
32. Li, X.; Yang, Z.; He, X.; Liu, Y. Optimization analysis and mechanism exploration on the removal of cadmium from contaminated soil by electrokinetic remediation. *Sep. Purif. Technol.* 2020.
33. Azizollahi, Z.; Ghaderian, S.M.; Ghotbi-Ravandi, A.A. Cadmium accumulation and its effects on physiological and biochemical characters of summer savory (*Satureja hortensis* L.). *Int. J. Phytoremediat.* 2019, 21, 1241–1253.
34. Yu, S.; Sheng, L.; Zhang, C.; Deng, H. Physiological response of *Arundo donax* to cadmium stress by Fourier transform infrared spectroscopy. *Spectrochim. Acta A* 2018, 198, 88–91.
35. Luo, J.S.; Yang, Y.; Gu, T.; Wu, Z.; Zhang, Z. The *Arabidopsis* defensin gene AtPDF2.5 mediates cadmium tolerance and accumulation. *Plant Cell Environ.* 2019, 42, 2681–2695.
36. Gajewska, E.; Skłodowska, M. Differential effect of equal copper, cadmium and nickel concentration on biochemical reactions in wheat seedlings. *Ecotoxicol. Environ. Saf.* 2010, 73, 996–1003.
37. Popova, L.P.; Maslenkova, L.T.; Yordanova, R.Y.; Ivanova, A.P.; Krantev, A.P.; Szalai, G.; Janda, T. Exogenous treatment with salicylic acid attenuates cadmium toxicity in pea seedlings. *Plant Physiol. Biochem.* 2009, 47, 224–231.
38. Yamada, M.; Malambane, G.; Yamada, S.; Suharsono, S.; Tsujimoto, H.; Moseki, B.; Akashi, K. Differential physiological responses and tolerance to potentially toxic elements in biodiesel tree *Jatropha curcas*. *Sci. Rep.* 2018, 8, 1–10.
39. Meena, M.; Aamir, M.; Kumar, V.; Swapnil, P.; Upadhyay, R. Evaluation of morpho-physiological growth parameters of tomato in response to Cd induced toxicity and characterization of metal sensitive NRAMP3 transporter protein. *Environ. Exp. Bot.* 2018, 148, 144–167.

40. Huang, L.; Li, W.C.; Tam, N.F.Y.; Ye, Z. Effects of root morphology and anatomy on cadmium uptake and translocation in rice (*Oryza sativa* L.). *J. Environ. Sci.* 2019, 75, 296–306.
41. Chun, C.-P.; Zhou, W.; Ling, L.-L.; Cao, L.; Fu, X.-Z.; Peng, L.-Z.; Li, Z.-G. Uptake of cadmium (Cd) by selected citrus rootstock cultivars. *Sci. Hortic.* 2020, 263, 109061.
42. Gallego, S.M.; Pena, L.B.; Barcia, R.A.; Azpilicueta, C.E.; Iannone, M.F.; Rosales, E.P.; Zawoznik, M.S.; Groppa, M.D.; Benavides, M.P. Unravelling cadmium toxicity and tolerance in plants: Insight into regulatory mechanisms. *Environ. Exp. Bot.* 2012, 83, 33–46.
43. Zheng, R.-L.; Li, H.-F.; Jiang, R.-F.; Zhang, F.-S. Cadmium accumulation in the edible parts of different cultivars of radish, *Raphanus sativus* L.; and carrot, *Daucus carota* var. *sativa*, grown in a Cd-contaminated soil. *Bull. Environ. Cont. Toxicol.* 2008, 81, 75–79.
44. Ulusu, Y.; Öztürk, L.; Elmastaş, M. Antioxidant capacity and cadmium accumulation in parsley seedlings exposed to cadmium stress. *Russ. J. Plant Physiol.* 2017, 64, 883–888.
45. Li, L.-Z.; Tu, C.; Peijnenburg, W.J.; Luo, Y.-M. Characteristics of cadmium uptake and membrane transport in roots of intact wheat (*Triticum aestivum* L.) seedlings. *Environ. Pollut.* 2017, 221, 351–358.
46. Rahman, M.F.; Ghosal, A.; Alam, M.F.; Kabir, A.H. Remediation of cadmium toxicity in field peas (*Pisum sativum* L.) through exogenous silicon. *Ecotoxicol. Environ. Saf.* 2017, 135, 165–172.
47. Saleem, M.H.; Fahad, S.; Khan, S.U.; Ahmar, S.; Khan, M.H.U.; Rehman, M.; Maqbool, Z.; Liu, L. Morpho-physiological traits, gaseous exchange attributes, and phytoremediation potential of jute (*Corchorus capsularis* L.) grown in different concentrations of copper-contaminated soil. *Ecotoxicol. Environ. Saf.* 2020, 189, 109915.
48. Zhang, X.; Li, M.; Yang, H.; Li, X.; Cui, Z. Physiological responses of *Suaeda glauca* and *Arabidopsis thaliana* in phytoremediation of heavy metals. *J. Environ. Manag.* 2018, 223, 132–139.
49. Kaya, C.; Okant, M.; Ugurlar, F.; Alyemeni, M.N.; Ashraf, M.; Ahmad, P. Melatonin-mediated nitric oxide improves tolerance to cadmium toxicity by reducing oxidative stress in wheat plants. *Chemosphere* 2019, 225, 627–638.
50. Lian, J.; Zhao, L.; Wu, J.; Xiong, H.; Bao, Y.; Zeb, A.; Tang, J.; Liu, W. Foliar spray of TiO₂ nanoparticles prevails over root application in reducing Cd accumulation and mitigating Cd-induced phytotoxicity in maize (*Zea mays* L.). *Chemosphere* 2020, 239, 124794.
51. Carvalho, M.E.; Piotto, F.A.; Gaziola, S.A.; Jacomino, A.P.; Jozefczak, M.; Cuypers, A.; Azevedo, R.A. New insights about cadmium impacts on tomato: Plant acclimation, nutritional changes, fruit quality and yield. *Food Energy Secur.* 2018, 7, e00131.
52. Wu, M.; Luo, Q.; Zhao, Y.; Long, Y.; Liu, S.; Pan, Y. Physiological and biochemical mechanisms preventing Cd toxicity in the new hyperaccumulator *Abelmoschus manihot*. *J. Plant Growth Regul.* 2018, 37, 709–718.
53. Ahmed, H.; Häder, D.-P. Rapid ecotoxicological bioassay of nickel and cadmium using motility and photosynthetic parameters of *Euglena gracilis*. *Environ. Exp. Bot.* 2010, 69, 68–75.
54. Liu, M.; Bi, J.; Liu, X.; Kang, J.; Korpelainen, H.; Niinemets, Ü.; Li, C. Microstructural and physiological responses to cadmium stress under different nitrogen levels in *Populus cathayana* females and males. *Tree Physiol.* 2020, 40, 30–45.
55. Alyemeni, M.N.; Ahanger, M.A.; Wijaya, L.; Alam, P.; Bhardwaj, R.; Ahmad, P. Selenium mitigates cadmium-induced oxidative stress in tomato (*Solanum lycopersicum* L.) plants by modulating chlorophyll fluorescence, osmolyte accumulation, and antioxidant system. *Protoplasma* 2018, 255, 459–469.
56. Sun, H.; Wang, X.; Shang, L.; Zhou, Z.; Wang, R. Cadmium accumulation and its effects on nutrient uptake and photosynthetic performance in cucumber (*Cucumis sativus* L.). *Philipp. Agric. Sci.* 2017, 100, 263–270.
57. Rossi, L.; Bagheri, M.; Zhang, W.; Chen, Z.; Burken, J.G.; Ma, X. Using artificial neural network to investigate physiological changes and cerium oxide nanoparticles and cadmium uptake by *Brassica napus* plants. *Environ. Pollut.* 2019, 246, 381–389.
58. Iannone, M.F.; Rosales, E.P.; Groppa, M.D.; Benavides, M.P. Reactive oxygen species formation and cell death in catalase-deficient tobacco leaf disks exposed to cadmium. *Protoplasma* 2010, 245, 15–27.
59. Bertin, G.; Averbeck, D. Cadmium: Cellular effects, modifications of biomolecules, modulation of DNA repair and genotoxic consequences (a review). *Biochimie* 2006, 88, 1549–1559.
60. Gjorgjeva, D.; Kadifkova-Panovska, T.; Mitrev, S.; Kovacevik, B.; Kostadinovska, E.; Bačeva, K.; Stafilov, T. Assessment of the genotoxicity of heavy metals in *Phaseolus vulgaris* L. as a model plant system by Random Amplified Polymorphic DNA (RAPD) analysis. *J. Environ. Sci. Health* 2012, 47, 366–373.

61. Surgun-Acar, Y. Determination of heavy metal-induced DNA damage in *Pisum Sativum*, L. at the molecular and population level. *J. Anim. Plant Sci.* 2018, 28, 1825–1834.
62. Ahmad, P.; Ahanger, M.A.; Alyemeni, M.N.; Wijaya, L.; Alam, P. Exogenous application of nitric oxide modulates osmolyte metabolism, antioxidants, enzymes of ascorbate-glutathione cycle and promotes growth under cadmium stress in tomato. *Protoplasma* 2018, 255, 79–93.
63. Dong, Y.; Chen, W.; Liu, F.; Wan, Y. Physiological responses of peanut seedlings to exposure to low or high cadmium concentration and the alleviating effect of exogenous nitric oxide to high cadmium concentration stress. *Plant Biosyst.* 2020, 154, 405–412.
64. Çatav, Ş.S.; Genç, T.O.; Oktay, M.K.; Küçükakyüz, K. Cadmium toxicity in wheat: Impacts on element contents, antioxidant enzyme activities, oxidative stress, and genotoxicity. *Bull. Environ. Cont. Toxicol.* 2020, 104, 71–77.
65. Barman, F.; Majumdar, S.; Arzoo, S.H.; Kundu, R. Genotypic variation among 20 rice cultivars/landraces in response to cadmium stress grown locally in West Bengal, India. *Plant Physiol. Biochem.* 2020, 148, 193–206.
66. Irfan, M.; Ahmad, A.; Hayat, S. Effect of cadmium on the growth and antioxidant enzymes in two varieties of *Brassica juncea*. *Saudi J. Biol. Sci.* 2014, 21, 125–131.
67. Lee, H.J.; Abdula, S.E.; Jang, D.W.; Park, S.-H.; Yoon, U.-H.; Jung, Y.J.; Kang, K.K.; Nou, I.S.; Cho, Y.-G. Overexpression of the glutamine synthetase gene modulates oxidative stress response in rice after exposure to cadmium stress. *Plant Cell Rep.* 2013, 32, 1521–1529.
68. Viehweger, K. How plants cope with heavy metals. *Bot. Stud.* 2014, 55, 35.
69. Ahmadpour, P.; Soleimani, M. Cadmium accumulation and translocation in *Jatropha curcas* grown in contaminated soils. *JWSS-Isfahan Uni. Technol.* 2015, 19, 179–190.
70. Li, X.; Zhang, X.; Wang, X.; Yang, X.; Cui, Z. Bioaugmentation-assisted phytoremediation of lead and salinity co-contaminated soil by *Suaeda salsa* and *Trichoderma asperellum*. *Chemosphere* 2019, 224, 716–725.
71. Braud, A.M.; Gaudin, P.; Hazotte, A.; Le Guern, C.; Lebeau, T. Chelate-assisted phytoextraction of lead using *Fagopyrum esculentum*: Laboratory vs. field experiments. *Int. J. Phytoremediat.* 2019, 21, 1072–1079.
72. Bani, A.; Echevarria, G. Can organic amendments replace chemical fertilizers in nickel agromining cropping systems in Albania? *Int. J. Phytoremediat.* 2019, 21, 43–51.
73. Zgorelec, Z.; Bilandzija, N.; Knez, K.; Galic, M.; Zuzul, S. cadmium and Mercury phytostabilization from soil using *Miscanthus× giganteus*. *Sci. Rep.* 2020, 10, 1–10.
74. Ghosh, M.; Singh, S. A review on phytoremediation of heavy metals and utilization of it's by products. *Asian. J. Energy Environ.* 2005, 6, 18.
75. Bingöl, N.A.; Özmal, F.; Akın, B. Phytoremediation and Biosorption Potential of *Lythrum salicaria* L. for Nickel Removal from Aqueous Solutions. *Pol. J. Environ. Stud.* 2017, 26.
76. Islam, M.S.; Saito, T.; Kurasaki, M. Phytofiltration of arsenic and cadmium by using an aquatic plant, *Micranthemum umbrosum*: Phytotoxicity, uptake kinetics, and mechanism. *Ecotoxicol. Environ. Saf.* 2015, 112, 193–200.
77. da Conceição Gomes, M.A.; Hauser-Davis, R.A.; de Souza, A.N.; Vitória, A.P. Metal phytoremediation: General strategies, genetically modified plants and applications in metal nanoparticle contamination. *Ecotoxicol. Environ. Saf.* 2016, 134, 133–147.
78. Nasr, M. Phytomanagement in Egypt: A Sustainable Approach for Clean Environment Coupled with Meeting Future Energy Demand. In *Waste Management in MENA Regions*; Springer: Berlin/Heidelberg, Germany, 2020; pp. 93–109.
79. Mesjasz-Przybyłowicz, J.; Nakonieczny, M.; Migula, P.; Augustyniak, M.; Tarnawska, M.; Reimold, U.; Koeberl, C.; Przybyłowicz, W.; Głowacka, E. Uptake of cadmium, lead nickel and zinc from soil and water solutions by the nickel hyperaccumulator *Berkheya coddii*. *Acta Biol. Cracoviensia Ser. Bot.* 2004, 46, 75–85.
80. Ojuederie, O.B.; Babalola, O.O. Microbial and plant-assisted bioremediation of heavy metal polluted environments: A review. *Int. J. Environ. Res. Public Health* 2017, 14, 1504.
81. Zhuang, P.; Ye, Z.; Lan, C.; Xie, Z.; Shu, W. Chemically assisted phytoextraction of heavy metal contaminated soils using three plant species. *Plant Soil* 2005, 276, 153–162.
82. Suman, J.; Uhlik, O.; Viktorova, J.; Macek, T. Phytoextraction of heavy metals: A promising tool for clean-up of polluted environment? *Front. Plant Sci.* 2018, 9, 1476.
83. Yu, G.; Liu, J.; Long, Y.; Chen, Z.; Sunahara, G.I.; Jiang, P.; You, S.; Lin, H.; Xiao, H. Phytoextraction of cadmium-contaminated soils: Comparison of plant species and low molecular weight organic acids. *Int. J. Phytoremediat.* 2020, 22, 383–391.

84. El-Mahrouk, E.-S.M.; Eisa, E.A.-H.; Hegazi, M.A.; Abdel-Gayed, M.E.-S.; Dewir, Y.H.; El-Mahrouk, M.E.; Naidoo, Y. Phytoremediation of cadmium-, copper-, and lead-contaminated soil by *Salix mucronata* (Synonym *Salix safsaf*). *HortScience* 2019, 54, 1249–1257.
85. Silva, J.; Fernandes, A.; Junior, M.S.; Santos, C.; Lobato, A. Tolerance mechanisms in *Cassia alata* exposed to cadmium toxicity—potential use for phytoremediation. *Photosynthetica* 2018, 56, 495–504.
86. Ali, S.Y.; Banerjee, S.N.; Chaudhury, S. Phytoextraction of cadmium and lead by three vegetable-crop plants. *Plant Sci. Today* 2016, 3, 298–303.
87. Liu, L.; Li, Y.; Tang, J.; Hu, L.; Chen, X. Plant coexistence can enhance phytoextraction of cadmium by tobacco (*Nicotiana tabacum* L.) in contaminated soil. *J. Environ. Sci.* 2011, 23, 453–460.
88. Fan, K.-C.; Hsi, H.-C.; Chen, C.-W.; Lee, H.-L.; Hseu, Z.-Y. Cadmium accumulation and tolerance of mahogany (*Swietenia macrophylla*) seedlings for phytoextraction applications. *J. Environ. Manag.* 2011, 92, 2818–2822.
89. Farid, M.; Ali, S.; Shakoor, M.B.; Bharwana, S.A.; Rizvi, H.; Ehsan, S.; Tauqeer, H.M.; Iftikhar, U.; Hannan, F. EDTA assisted phytoremediation of cadmium, lead and zinc. *Int. J. Agron. Plant Prod.* 2013, 4, 2833–2846.
90. Hassan, M.; Dagari, M.; Babayo, A. Effect of citric acid on cadmium ion uptake and stress response of hydroponically grown jute mallow (*Corchorus olitorius*). *J. Environ. Anal. Toxicol.* 2016, 6.
91. Anderson, C.; Brooks, R.; Stewart, R.; Simcock, R.; Robinson, B. The phytoremediation and phytomining of heavy metals. In *Proceedings of the Pacrim International Congress on Earth Science, Exploration and Mining Around Pacific Rim, Bali, Indonesia, 10–13 October 1999*; pp. 127–135.
92. Ranieri, E.; Moustakas, K.; Barbaferi, M.; Ranieri, A.C.; Herrera-Melián, J.A.; Petrella, A.; Tommasi, F. Phytoextraction technologies for mercury-and chromium-contaminated soil: A review. *J. Chem. Technol. Biotechnol.* 2020, 95, 317–327.
93. Ernst, W.H. Phytoextraction of mine wastes—Options and impossibilities. *Geochemistry* 2005, 65, 29–42.
94. Robinson, B.; Anderson, C.; Dickinson, N. Phytoextraction: Where's the action? *J. Geochem. Explor.* 2015, 151, 34–40.
95. Cheraghi, M.; Lorestani, B.; Khorasani, N.; Yousefi, N.; Karami, M. Findings on the phytoextraction and phytostabilization of soils contaminated with heavy metals. *Biol. Trace Elem. Res.* 2011, 144, 1133–1141.
96. Dalvi, A.A.; Bhalerao, S.A. Response of plants towards heavy metal toxicity: An overview of avoidance, tolerance and uptake mechanism. *Ann. Plant Sci.* 2013, 2, 362–368.
97. Andrade Júnior, W.V.; de Oliveira Neto, C.F.; Santos Filho, B.G.d.; do Amarante, C.B.; Cruz, E.D.; Okumura, R.S.; Barbosa, A.V.C.; de Sousa, D.J.P.; Teixeira, J.S.S.; Botelho, A.D.S. Effect of cadmium on young plants of *Virola surinamensis*. *AoB Plants* 2019, 11.
98. Boros-Lajszner, E.; Wyszowska, J.; Kucharski, J. Application of white mustard and oats in the phytostabilisation of soil contaminated with cadmium with the addition of cellulose and urea. *J. Soil Sediment.* 2020, 20, 931–942.
99. Lan, M.-M.; Liu, C.; Liu, S.-J.; Qiu, R.-L.; Tang, Y.-T. Phytostabilization of Cd and Pb in Highly polluted Farmland soils using ramie and amendments. *Int. J. Environ. Res. Public Health* 2020, 17, 1661.
100. Sheoran, V.; Sheoran, A.; Poonia, P. Phytostabilization of metalliferous mine waste. *J. Indus. Pollut. Control* 2013, 29, 183–192.
101. Wong, M.H. Ecological restoration of mine degraded soils, with emphasis on metal contaminated soils. *Chemosphere* 2003, 50, 775–780.
102. Sarma, H. Metal hyperaccumulation in plants: A review focusing on phytoremediation technology. *J. Environ. Sci. Technol.* 2011, 4, 118–138.
103. Galal, T.M.; Eid, E.M.; Dakhil, M.A.; Hassan, L.M. Bioaccumulation and rhizofiltration potential of *Pistia stratiotes* L. for mitigating water pollution in the Egyptian wetlands. *Int. J. Phytoremediat.* 2018, 20, 440–447.
104. Abhilash, P.; Pandey, V.C.; Srivastava, P.; Rakesh, P.; Chandran, S.; Singh, N.; Thomas, A. Phytofiltration of cadmium from water by *Limnocharis flava* (L.) Buchenau grown in free-floating culture system. *J. Hazard. Mater.* 2009, 170, 791–797.
105. Důřešová, Z.; Šušnovská, A.; Horník, M.; Pipíška, M.; Gubišová, M.; Gubiš, J.; Hostin, S. Rhizofiltration potential of *Arundo donax* for cadmium and zinc removal from contaminated wastewater. *Chem. Pap.* 2014, 68, 1452–1462.
106. Olguín, E.J.; Sánchez-Galván, G. Heavy metal removal in phytofiltration and phycoremediation: The need to differentiate between bioadsorption and bioaccumulation. *New Biotech.* 2012, 30, 3–8.
107. Jia, H.; Wang, H.; Lu, H.; Jiang, S.; Dai, M.; Liu, J.; Yan, C. Rhizodegradation potential and tolerance of *Avicennia marina* (Forsk.) Vierh in phenanthrene and pyrene contaminated sediments. *Mar. Pollut. Bull.* 2016, 110, 112–118.

108. Yanai, J.; Zhao, F.-J.; McGrath, S.P.; Kosaki, T. Effect of soil characteristics on Cd uptake by the hyperaccumulator *Thlaspi caerulescens*. *Environ. Pollut.* 2006, 139, 167–175.
 109. Sharma, H.D.; Reddy, K.R. *Geoenvironmental Engineering: Site Remediation, Waste Containment, and Emerging Waste Management Technologies*; John Wiley and Sons: Hoboken, NJ, USA, 2004.
 110. Chen, M.; Xu, P.; Zeng, G.; Yang, C.; Huang, D.; Zhang, J. Bioremediation of soils contaminated with polycyclic aromatic hydrocarbons, petroleum, pesticides, chlorophenols and heavy metals by composting: Applications, microbes and future research needs. *Biotechnol. Adv.* 2015, 33, 745–755.
-

Retrieved from <https://encyclopedia.pub/entry/history/show/30078>